

SUPPLEMENTARY DATA

Table 1: Primers used for Real Time PCR

CDX2	sense	GCCTGTCACCAGAGCTTCTC
	antisense	AGACCAACAACCCAAACAGC
$\beta$ actin	sense	ACTGGAACGGTGAAGGTGAC
	antisense	GTGGACTTGGGAGAGGACTG
Rps 11	sense	GCCGAGACTATCTGCACTAC
	antisense	ATGTCCAGCCTCAGAACTTC
$\alpha$ tubulin	sense	GCCTGGACCACAAGTTTGAC
	antisense	TGAAATTCTGGGAGCATGAC

Table 2: Correlation between *in vitro* and *ex vivo* findings

	IEC lines + LPL <i>in vitro</i>						<i>ex vivo</i> studies (crypt staining)					
	IAP	Akt	ERK 1/2	p38	JNK	CDX2	IAP	Akt	ERK 1/2	p38	JNK	CDX2
Nor	+	+	+	+	+	+	-	-	-	-	-	-
Inact CD	+++	+	++	++	++	++	++	++	-	+	+	+
Act CD	ND	ND	ND	ND	ND	ND	+++	+++	+	+++	+	+
Inact UC	ND	+	++	+	++	ND	-	++	++	-	+	+
Act UC	ND	ND	ND	ND	ND	ND	-	+	++	-	+	+

ND = Not Determined.

#### Supplementary figure legends

Figure 1: MAPK pathway activation in T84 cells co-cultured with LPL. Immunoblotting for P-ERK1/2, P-p38 MAPK, P-JNK, and ERK2 in lysates obtained from T84 cells co-cultured with freshly isolated Nor, CD or UC LPL for 30 min to 3 hours. The CTL lane contains T84 alone. EGF (10 nM; 15min) was used as a positive control. These data are representative of 3 experiments. The percentage of MAPK activity related to the activity induced by EGF was quantified after densitometric analysis.

Figure 2: Nor, inactive and active CD, and inactive and active UC colonic tissue sections were immunostained using anti-phospho ERK1/2, anti-phospho p38 MAPK, and anti-phospho JNK antibodies. The slides were counter-stained with Mayer's Hematoxylin solution, and examined with a Zeiss Axioskop Light Microscope at 20X magnification. The insets represent 100X magnification of the crypt epithelial cells to document nuclear localization. These data are representative of 4 experiments.



