## Supplementary material

for

## A modified fluorescent intercalator displacement assay for RNA ligand discovery

Papa Nii Asare-Okai and Christine S. Chow\*

Department of Chemistry, Wayne State University, Detroit, MI 48202

## **Figures**



Figure S1: Fluorescence titration data of the A-site RNA (3  $\mu$ M) with TO-PRO is shown (buffer conditions were 100 mM KCl, 20 mM Tris, pH 7).



Figure S2: A Job plot of TO-PRO binding to the A-site RNA (3  $\mu$ M) is given (buffer conditions were 100 mM KCl, 20 mM Tris, pH 7).



Figure S3: ESI-MS spectra showing a titration of Tat peptide into equimolar concentrations of TAR RNA and TO-PRO complex (1  $\mu$ M). Buffer conditions are 150 mM ammonium acetate, pH 7.

## Tables

**Table S1:** Quantification of the ESI-MS peak areas of the TO-PRO:A-site RNA complex after titration with  $0-15 \mu$ M of paromomycin.

[paromomycin] µM	fraction bound (TO-PRO) <sup>a</sup>
0	0.29
1	0.25
3	0.20
6	0.19
10	0.13
15	0.04

<sup>a</sup>The fraction bound of TO-PRO was calculated by dividing the peak area corresponding to A-site RNA/TO-PRO complex by the peak area corresponding to total RNA (peak area of total RNA is equal to the summation of the peak area for free RNA, RNA/TO-PRO complex, and RNA/paromomycin complex).

**Table S2:** Quantification of the ESI-MS peak areas of TO-PRO:A-site RNA complex after titration with  $0-100 \mu$ M of chloramphenicol.

[chloramphenicol]	fraction bound	
μΜ	(TO-PRO) <sup>a</sup>	
0	0.23	
5	0.26	
40	0.29	
100	0.26	

<sup>a</sup>The fraction bound of TO-PRO was calculated by dividing the peak area corresponding to A-site RNA/TO-PRO complex by the peak area corresponding to total RNA (peak area of total RNA is equal to the summation of the peak area for free RNA, RNA/TO-PRO complex, and RNA/chloramphenicol complex).

**Table S3:** FID assay of Tat peptide utilizing pre-bound equimolar concentrations of TAR RNA and TO-PRO (1  $\mu$ M each). Buffer conditions were 150 mM ammonium acetate, pH 7. The data shown is an average of three separate experiments.

	TAR		relative
dye	RNA	Tat	fluorescence
(µM)	(µM)	(µM)	(%)
1	0	0	0.3
1	1	0	100
1	1	2	86
1	1	10	49
1	1	19	35
1	1	31	23

**Table S4:** Quantification of the ESI-MS peak areas of TO-PRO:TAR RNA complex after titration with  $0-25 \mu$ M Tat peptide.

[Tat], μM	fraction bound (TO-PRO) <sup>a</sup>
0	0.42
3	0.35
15	0.13
25	0.05

<sup>a</sup>The fraction bound of TO-PRO was calculated by dividing the peak area corresponding to TAR RNA/TO-PRO complex by the peak area corresponding to total RNA (peak area of total RNA is equal to the summation of the peak area for free RNA, RNA/TO-PRO complex, and RNA/Tat complex).

**Table S5:** Comparison of the FID results and dissociation constants for DHR23 and paromomycin binding to H69 and A-site RNA.

	DHR23		paromomycin	
	fluorescence (%)	$\frac{K_d}{(\mu M)^a}$	fluorescence (%)	$\frac{K_d}{(\mu M)^a}$
A-site				
RNA	54	19	14	1.3
H69	83	43	36	21

<sup>a</sup> Dissociation constants were obtained from ESI-MS experiments as described previously [25] and in the Materials and Methods section.