Supporting Information

Yazdi et al. 10.1073/pnas.1008155107

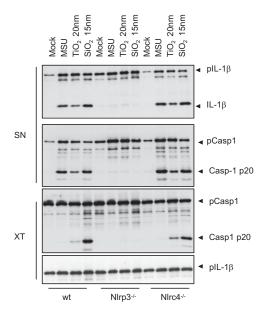


Fig. S1. Nano-TiO₂ and nano-SiO₂ activate the Nlrp3 inflammasome, but not the Nlrc4 inflammasome. Caspase-1 cleavage and IL-1 β secretion was impaired in Nlrp3-deficient, but not in Nlrc4-deficient murine bone marrow-derived macrophages upon treatment with nano-TiO₂ (200 μ g/mL) or nano-SiO₂ (200 μ g/mL).

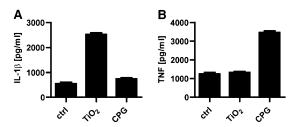


Fig. S2. Nano-TiO₂ activates IL-1 β secretion in murine dendritic cells (DCs). (A) TiO₂ (200 μ g/mL) exposure evoked robust secretion of IL-1 β after LPS-priming (10 ng/mL) and (B) low levels of TNF in unprimed cells compared with the TLR9 ligand CpG (2.5 μ g/mL).

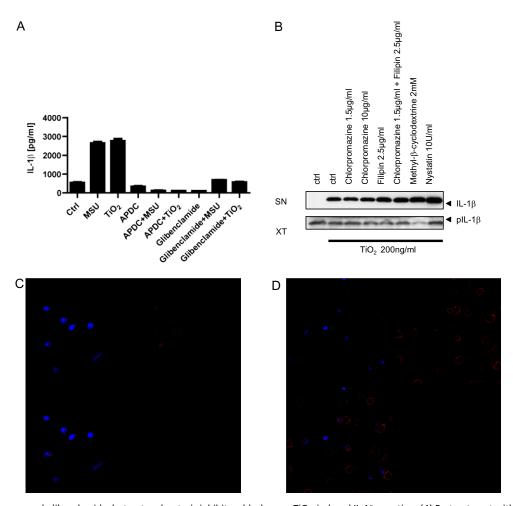


Fig. S3. ROS scavengers and glibenclamide, but not endocytosis-inhibitors block nano-TiO₂—induced IL-1β secretion. (A) Pretreatment with the ROS scavenger APDC (100 μM) and the ATP-sensitive potassium inhibitor glibenclamide (50 μM) impairs nano-TiO₂ (200 μg/mL)-induced IL-1β release from bone marrow-derived DCs, whereas (B) lipid-raft extraction by methyl-β-cyclodextrin (MBCD; 2× 10 mM wash, 2 mM during treatment), blocking clathrin-mediated endocytosis by chlorpromazine-hydrochloride (1.5 μg/mL or 10 μg/mL) and blocking caveolin-dependent endocytosis by either filipin (2.5 μg/mL or 2.5 μg/mL in combination with chlorpromazine-hydrochloride, 1.5 μg/mL) or nystatin (10 U/mL) did not alter TiO₂-induced IL-1β secretion in THP1 cells. The effective lipid-raft depletion was demonstrated by the lack of membranous cholera toxin B staining (C) after treatment with MBCD, (D) compared with the untreated control.

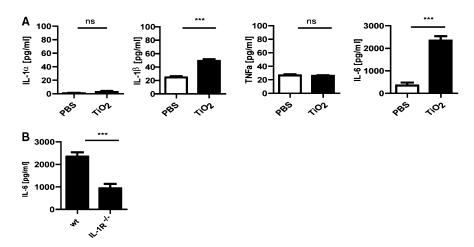


Fig. S4. TiO₂ i.p. injection induces IL-1β and IL-1 receptor-dependent IL-6 secretion. (A) I.p. injection of 1.5 mg TiO₂ into the peritoneal cavity led to significant IL-1β and IL-6 secretion, whereas IL-1α and TNF remained unchanged. (B) IL-6 release into the peritoneal cavity was drastically dimished in IL-1R^{-/-} mice compared with wild-type controls, indicating the dependence of IL-6 secretion on IL-1 signaling (*P < 0.05; **P < 0.01; ***P < 0.001; NS, nonsignificant, P = 0.001; mice per group).