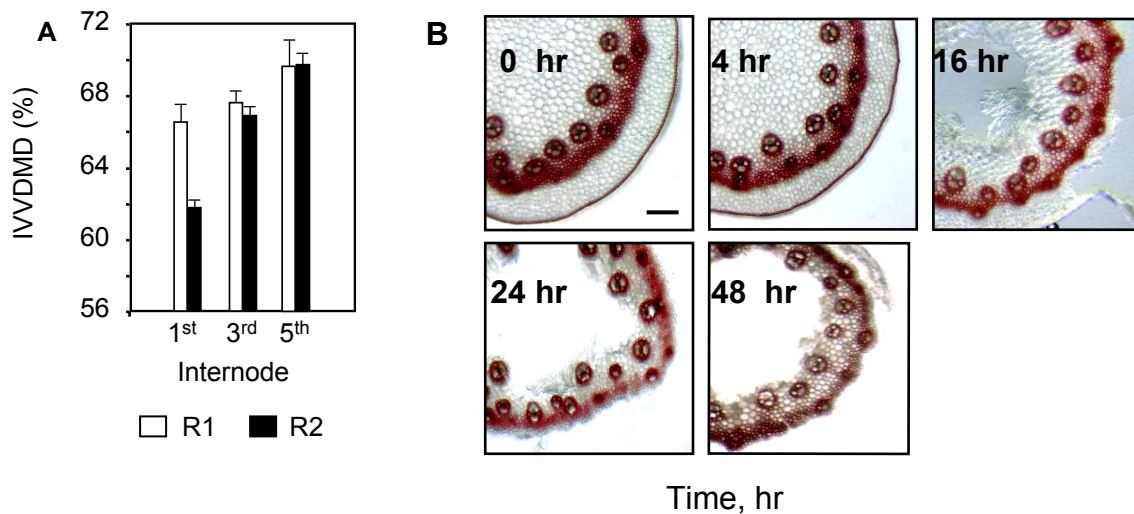


Supplementary Figure 1. Quantification of lignin content and composition in perennial ryegrass stems

(A) Acetyl bromide soluble lignin content in developing stems. The error bars are based on two biological and three technical replicates. Developmental stages as for Figure 1. Internodes were analysed separately and at each stage, results from the basal internode are shown on the left-hand column and those from the apical internode in the right-hand column. Results from the remaining internodes are shown in the other columns. CWR: cell wall residues.

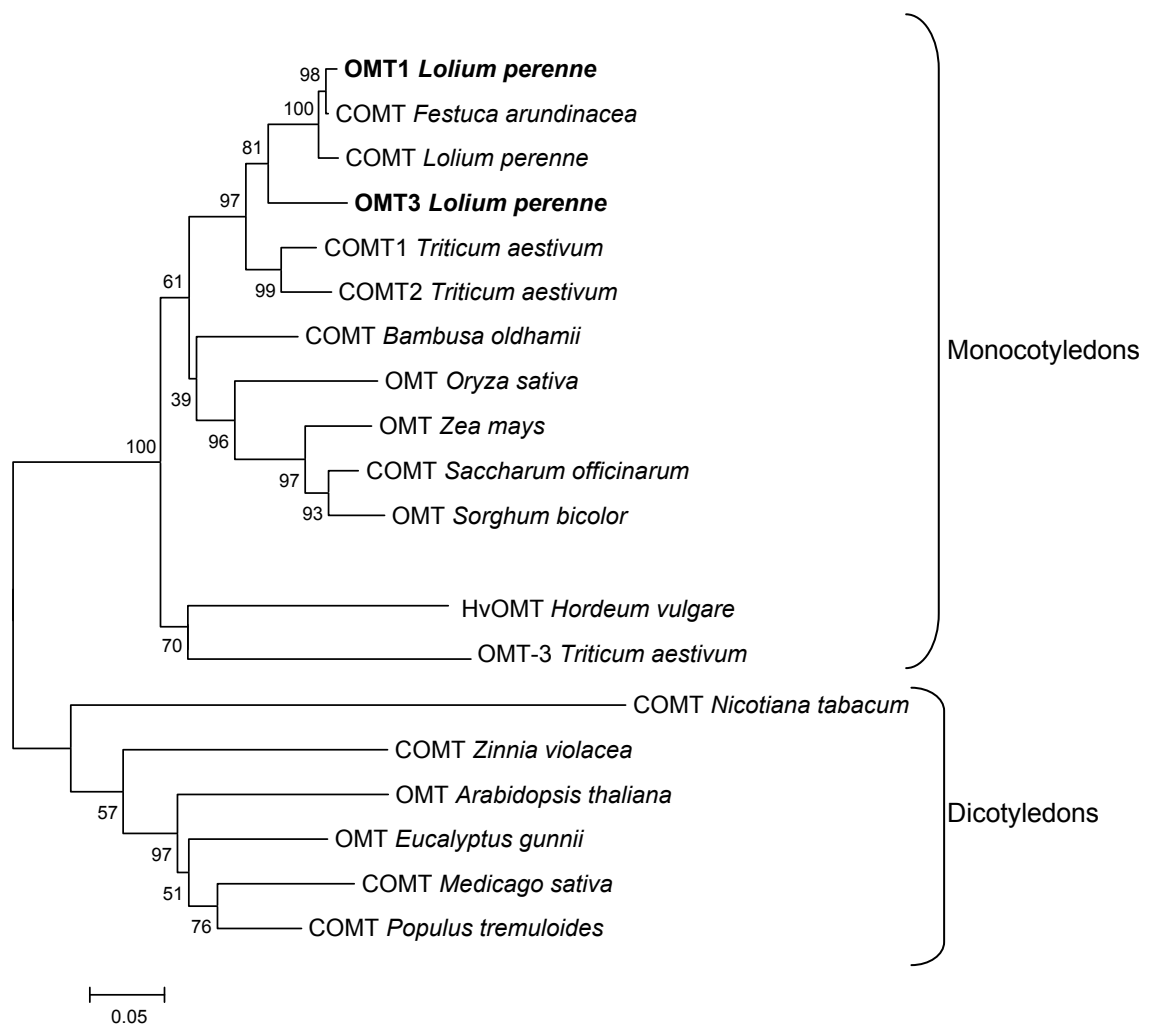
(B) Syringyl: Guaiacyl (S/G) monolignol ratio measured in different internodes of stems at the R3 stage. Internodes were numbered from the base to the apex of the stem.



Supplementary Figure 2. Assessment of the digestibility of perennial ryegrass stems

(A) Estimation of *in vivo* dry matter digestibility (IVVDMD) of 1st, 3rd and 5th internodes of stems at the R1 and R2 stages, measured using near-infrared spectroscopy (NIRS). IVVDMD is a regression of the pepsin-cellulose digestibility compared to *in vivo* digestibility of known standards of similar class. white bars: R1 stage, black bars: R2 stage.

(B) Mäule staining of sections taken from the 1st internode of stems at the E3 stage incubated *in vitro* with bovine rumen fluid over a 48 h time course. Each micrograph is representative of at least 4 stems from different plants. Bars = 160 μm for all sections

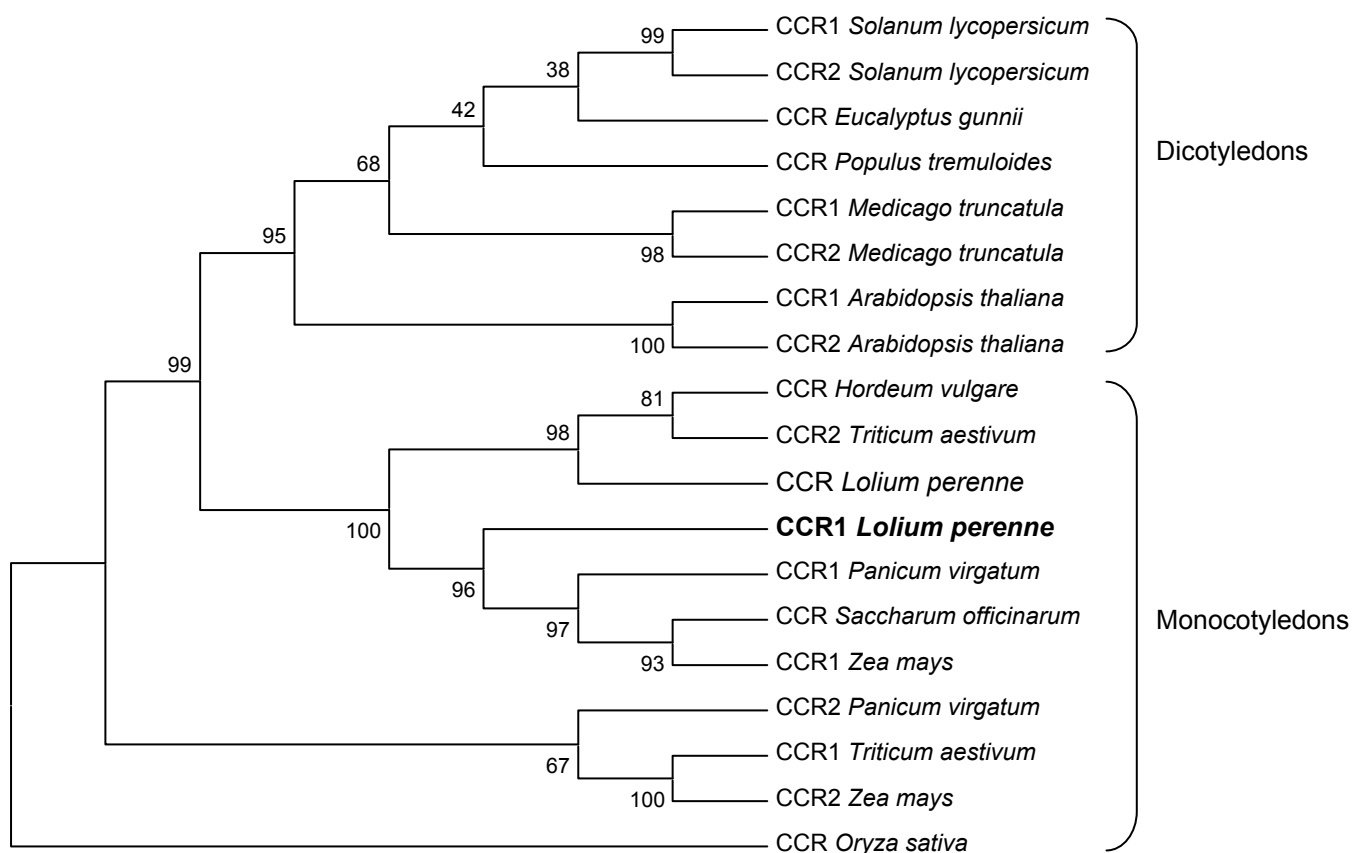


Supplementary Figure 3. Phylogenetic tree showing the relationships between the deduced amino acid sequences of plant caffeic acid O-methyltransferases. The Swissprot accession numbers are as follows: AAD10253, AAK68907.1, AAC18623, AAD10255, ABP63535.1, AAP23942.1, ABS18316.1, ABB90678.1, AAB03364.1, CAA13175.1, AAL57301.1, BAC54275.1, ABP93669.1, AAL91506.1, AAA86718.1, AAB96879.1, CAA52814.1, AAB46623.1 and AAB61731.1.

		*	20	*	40	*	60	*	
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At OMT	:	MGSTA-ETQLTPVQVT	DDEAALFAMQLASASVLPMLKSALELDLLEIMAK--NG--SPMSPTIEIASKLP	:	65				
Pt COMT	:	MGSTG-ETQMTPTQV	SDEEAHLFAMQLASASVLPMLKTAIELDLLEIMAK--AGPGAFLSTSEIASHLP	:	67				
Ta COMT	:	MGSTAADM----	AASADEEACMYALQLVSSSILPMTLKNKNAIELGLLETLV--AAGGKL-LTPAEVAAKLP	:	63				
Zm OMT	:	MGSTAGDV----	AAVVDEEACMYAMQLASSSILPMTLKNKNAIELGLLEVLQKEAGGKAALAPEEVVARM	:	66				
Fa COMT	:	MGSTAADM----	AASADEDACMFALQLASSSILPMTLKNKNAIELGLLEILV--AAGGKS-LTPTEVAAKLP	:	63				
Lp OMT1	:	MGSTAADM----	AASADEDACMFALQLASSSILPMTLKNKNAIELGLLEILV--AAGGKS-LTPTEVAAKLP	:	63				
Lp OMT3	:	MGSTAAEI----	AASADEEACLYALQLGSSSILPMTLKNKNTIELGLLETLM--AAGGKS-LTPTEVAAKLP	:	63				
		80	*	100	*	120	*	140	
Ms COMT	:	TT--NPDAPV-MLDRMLRLL	LACYIILTCSV--RTQQDGKVVRLYGLATVAKYLVKNEDGVSISALNLMNQ	:	132				
At OMT	:	TK--NPEAPV-MLDRILRLL	TSYSVLTCSN--RKLSDGVERIYGLGPVCKYLTKNEDGVSIAALCLMNQ	:	130				
Pt COMT	:	TK--NPDAPV-MLDRILRLL	SYSILTCSL--KDLDPDGKVERLYGLAPVCKFLTKNEDGVSISPLCLMNQ	:	132				
Ta COMT	:	STA-NPAAAD-MVDRMLRLL	SYNVVSCTM--EEGKDGRLSRRYRAAPVCKFLTPNEDGVSMAALALMNQ	:	129				
Zm OMT	:	AAPSDFAAAAAMVDRMLRLL	SADYDVVRCQM--ED-RDGRYERRYSAAPVCKWLTLPNEDGVSMAALALMNQ	:	133				
Fa COMT	:	SAA-NPEAPD-MVDRILRLL	SYNVVTCLV--EEGKDGRLSRSYGAPVCKFLTLPNEDGVSMAALALMNQ	:	129				
Lp OMT1	:	SAA-NPEAPD-MVDRILRLL	SYNVVTCLV--EEGKDGRLSRSYGAPVCKFLTLPNEDGVSMAALALMNQ	:	129				
Lp OMT3	:	CAAKNPEAPD-MVDRMLRLL	SYNVVSCLV--EEGTDGRLSRRYGAAPVCKFLTLPNEDGVSMAALALMNQ	:	130				
		*	160	*	180	*	200	*	
Ms COMT	:	DKVLMESWYHLKDAVLDGGI	IPFNKAYGMTAFEYHGTDPRENFVNKMGMSDHSITITMKKIL-ETYTGFEGL	:	201				
At OMT	:	DKVLMESWYHLKDAILDGGI	IPFNKAYGMSAFEYHGTDPRENFVNNGMSNHSITITMKKIL-ETYKGFEG	:	199				
Pt COMT	:	DKVLMESWYHLKDAILDGGI	IPFNKAYGMTAFEYHGTDPRENFVNKMGMSDHSITITMKKIL-ETYKGFEG	:	201				
Ta COMT	:	DKVLMESWYHLKDAVLDGGI	IPFNKAYGMSAFEYHGTDPRENFVNEGMKNHSIIITKKLL-EVYKGFEG	:	198				
Zm OMT	:	DKVLMESWYHLKDAVLDGGI	IPFNKAYGMTAFEYHGTDPRENFVNEGMKNHSVIIITKKLL-DFYTGFEGL	:	202				
Fa COMT	:	DKVLMESWYHLKDAVLDGGI	IPFNKAYGMSAFEYHGTDPRENFVNEGMKNHSIIITKKLL-ELYHGFEG	:	198				
Lp OMT1	:	DKVLMESWYHLKDAVLDGGI	IPFNKAYGMSAFEYHGTDPRENFVNEGMKNHSIIITKKLL-ELYHGFEG	:	198				
Lp OMT3	:	DKVLMESWYHLKDAVLDGGI	IPFNKAYGMSAFEYHGTDPRENFVNEAMKNNSIIITKKLL-QLYDGGQGL	:	199				
		220	*	240	*	260	*	280	
Ms COMT	:	KSLVDVGGGTGAVINTIV	SKYPTIKGINFDLPHVIEDAPSYPGVEHVGGDMFVSVIPKA-DAVFMKWI	CHD	:	270			
At OMT	:	TSLVDVGGGTICATLKMIV	SKYPNLKGINFDLPHVIEDAPSHPGIEHVGGDMFVSVIPKG-DAIFM	KWICH	:	268			
Pt COMT	:	TSLVDVGGGTGAVNTIV	SKYPSIKGINFDLPHVIEDAPSYPGVEHVGGDMFVSVIPKA-DAVFM	KWICH	:	270			
Ta COMT	:	GTIVDVGGGIVGATVGA	ITAAYPKIKGINFDLPHVISEAQPFPGVTHVGGDMFQKVP	PSG-DAILMKWIL	HD	:	267		
Zm OMT	:	STLVDVGGGIVGATLHA	ITSRPHISGVNFDLPHVISEAPFPFPGVRHVGGDMFASVPAG-DAILM	KWIL	HD	:	271		
Fa COMT	:	GSLVDVGGGIVGATVA	AAIAAHYPTIKGVNFDLPHVISEAPQFPVTHVGGDMFKEVPSG-DAILM	KWIL	HD	:	267		
Lp OMT1	:	GTLVDVGGGIVGATVA	AAIAAHYPTIKGVNFDLPHVISEAPQFPVTHVGGDMFKEVPSG-DTILM	KWIL	HD	:	267		
Lp OMT3	:	GTLVDVGGGIVGATVA	AATAHYPTIKGINFDLPHVISEAPFPVTHVGGDMFKKVP	LG-DAILMKWIL	HD	:	268		
		*	300	*	320	*	340	*	
Ms COMT	:	MSDEHCLKFLKNCYEALPDN	---GKVIVAEICILPVAP-DSSLATKGVVHIDV-IMLAHNP	GGKERTQ	KEF	:	335		
At OMT	:	MSDEHCVKFLKNCYESLPED	---GKVILAEICILPETP-DSSLSTKQVVHDC-IMLAHNP	GGKERT	KEF	:	333		
Pt COMT	:	MSDAHCLKFLKNCYDALPEN	---GKVILVEICILPVAP-DTSLATKGVVHIDV-IMLAHNP	GGKERT	KEF	:	335		
Ta COMT	:	MSDEHCATLLKNCYDALPAH	---GKVVLVEICILPVNP-EATPKAQGVFHVDM-IMLAHNP	GGRERY	REF	:	332		
Zm OMT	:	MSDAHCATLLKNCYDALPEN	---GKVIVVEICVLPVNT-EATPKAQGVFHVDM-IMLAHNP	GGKERY	REF	:	336		
Fa COMT	:	MSDQHCATLLKNCYDALPAH	---GKVVLVEICILPVNP-EANPSSQGVFHVDM-IMLAHNP	GGRERY	REF	:	332		
Lp OMT1	:	MSDQHCATLLKNCYDALPAH	---GKVVLVEICILPVNP-EANPSSQGVFHVDM-IMLAHNP	GGRERY	REF	:	332		
Lp OMT3	:	MSDQHCATLLKNCYDALPMH	---GKVVLVEICILPVNP-EAKPSSQGVFHVDM-IMLAHNP	GGRERY	REF	:	333		
		360	*	380					
Ms COMT	:	EDLAKGAGFGFKVHCNAFN	TYIMEFLKKV	:	365				
At OMT	:	EALAKASGFKGKVVCD	AFVGNLIELLKKL	:	363				
Pt COMT	:	EGLAKGAGFGFEVMCCA	FNTHVIEFRKKA	:	365				
Ta COMT	:	EALAKGAGFKAIKTTYI	YANAWAIEFTK--	:	360				
Zm OMT	:	RELAKGAGFSGFKATYI	YANAWAIEFIK--	:	364				
Fa COMT	:	QALARGAGFTGVKSTYI	YANAWAIEFTK--	:	360				
Lp OMT1	:	QALARGAGFTGVKSTYI	YANAWAIEFTK--	:	360				
Lp OMT3	:	EALARGAGFAGFKSTYI	YANAWAIEFTK--	:	361				

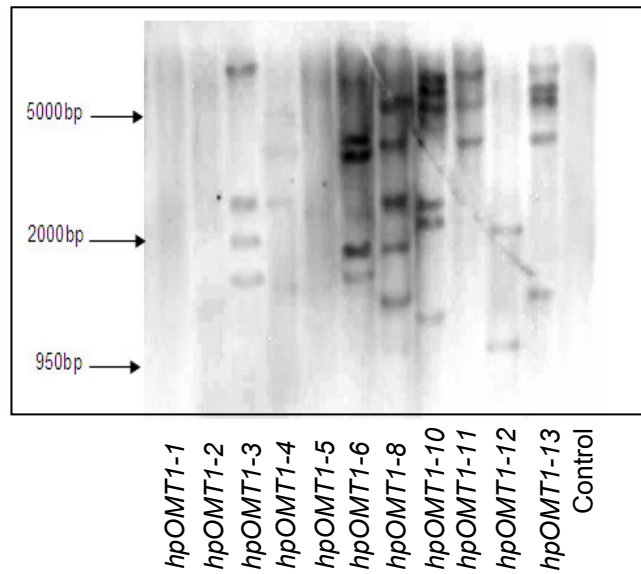
Supplementary Figure 4. Multiple sequence alignment of caffeic acid O-methyltransferases from perennial ryegrass and other plant species.

Residues involved in SAM binding, catalytic activity and substrate specificity are highlighted in blue, red and yellow, according to structural analysis of Zubieta et al. (2002). Lp: *Lolium perenne*, Ms: *Medicago sativa*, At: *Arabidopsis thaliana*, Pt: *Populus tremuloides*, Ta: *Triticum aestivum*, Zm: *Zea mays*, Fa: *Festuca arundinacea*. The accession numbers are as follows. Swissprot AAB46623.1, AAB96879.1, AAB61731.1, ABP63535.1, AAB03364.1, AAK68907.1, AF033538 AF033540



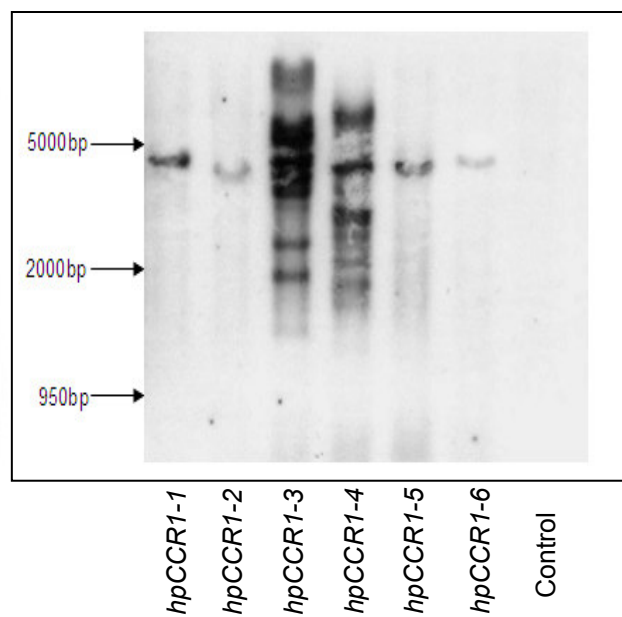
Supplementary Figure 5. Phylogenetic tree showing the relationships between the deduced amino acid sequences of plant cinnamoyl-CoA-reductases.

Genbank accession numbers are as follows: CAA13176.1, NP_001105488.1, ACZ74580.1, AAL47182.1, AAG09817.1, AAX08107.1, AAN71760.1, AAG46037.1, NP_178197.1, AAF43141.1, CAA56103.1, AAY41879.1, AAY41880.1, CAD21520.1, ACZ74585.1, NP_001105715.1, ABE01883.1, Mt CCR1 (TC 128336), Mt CCR2 (TC 132256),

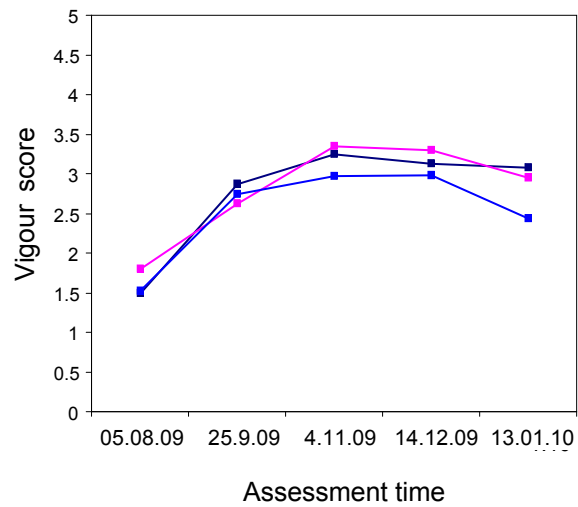


Supplementary Figure 6. Southern hybridization analysis of transgene copy number in perennial ryegrass plants containing an *hpOMT1* construct.

Genomic DNA was extracted from 11 transgenic lines and 1 wild type control plant, digested with *EcoRI* and hybridized to a probe that was complementary to the maize *Ubiquitin* promoter present in the transgene.

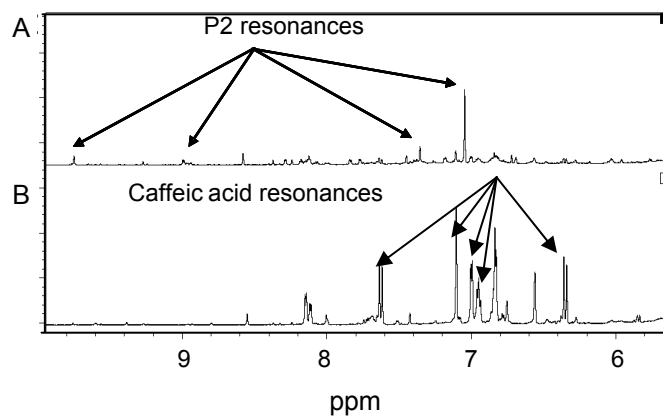


Supplementary Figure 7. Southern hybridization analysis of transgene copy number in perennial ryegrass plants containing an *hpCCR1* construct. Genomic DNA was extracted from 6 transgenic lines and 1 wild type control plant, digested with *EcoRI* and hybridized with a probe that was complementary to the maize *Ubiquitin* promoter present in the transgene cassette



Supplementary Figure 8. Phenotypic analysis of the plants grown under field conditions.

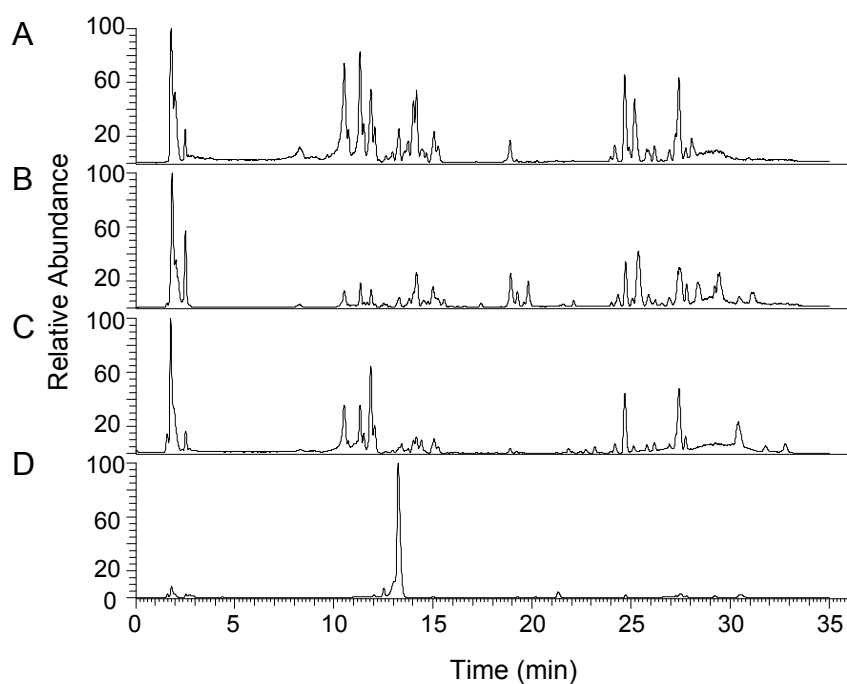
Transgenic *hpCCR1-1* (blue line), *hpOMT1-3* (red line) and control plants (black line) were assessed for vigour growth (scores 1-5) under field condition from August 2009 till January 2010.



Supplementary Figure 9. NMR analysis of aqueous soluble metabolites from *hpCCR1-1* and control plants. Expansion of the aromatic region of the ¹H NMR spectrum (acquired in MeOD for maximum phenolic solubility).

A. Control plants

B. *hpCCR1-1* plants



Supplementary Figure 10. LCMS chromatograms of a *hpCCR1-1* mutant compared to control in both positive and negative acquisition modes.

(A) *hpCCR1-1* ESI negative base peak chromatogram

(B) Control ESI negative base peak chromatogram

(C) *hpCCR1-1* ESI positive base peak chromatogram

(D) Control ESI positive base peak chromatogram

Supplementary Table 1 Homology between members of COMT proteins from perennial ryegrass and tall fescue

	Lp OMT1	Lp OMT3	La COMT
Lp OMT1		90%	98%
Lp OMT3	90%		90.3%

The Swissprot accession numbers are as follows:

Lp OMT1: AF033538

Lp OMT3: AF033540

La COMT: AAK68907

Supplementary Table 2 Homology between members of CCR proteins from monocots

	Lp CCR1	Lp CCR	Ta CCR1	Hv CCR	So CCR	Zm CCR1	Pv CCR1
Lp CCR1		75%	80%	80%	82%	85%	89%
Lp CCR	75%		77%	78%	77%	77%	79%

The Swissprot accession numbers are as follows:

Lp CCR1: AAL47182

Lp CCR: AAG09817.1

Ta CCR1: ABE01883.1

Hv CCR: AAN71760.1

So CCR: CAA13176.1

Zm CCR1: NP_001105488.1

Pv CCR1: ACZ74580

Supplementary Table 3 List of primers

Gene	Primer sequence (5'-3')	
<i>OMT1</i> (qRT-PCR)	Forward	GGCCATCGAGTTCACCAAGTAG
	Reverse	GGCAGCAGCCAGCAAGAT
<i>OMT3</i> (qRT-PCR)	Forward	TCTCTCGCTCATACTACCAAAAACA
	Reverse	ATCTCTGCCTGTGTCTGCTTCTC
<i>CCR1</i> (qRT-PCR)	Forward	GATGTCTGAACCAGAAGCTCCA
	Reverse	GCTAGGGTTTCCTTGTCCGC
<i>His</i> (qRT-PCR)	Forward	TGCTTGCCCTTCAGGAGGCT
	Reverse	ATCCTCCTGGCAAGCTGAATGA
<i>GAPDH</i> (qRT-PCR)	Forward	TGGTGCCAAGAAGGTCATCAT
	Reverse	GACCATCAACAGTCTTCTGG
<i>Tub</i> (qRT-PCR)	Forward	AGTTGCCCCCAGTGATGGTA
	Reverse	GCCACGATACACAAACAAGCA
<i>CCR1</i> (<i>hpCCR1</i>)	Forward	CGGGATCCTTCAGGCCGGTGAGCC
	Reverse	CTAGCTTATTTGTAGAGATGGTGCTTAAGG
<i>OMT1</i> (<i>hpOMT1</i>)	Forward	GGGGACAAGTTTGTACAAAAAAGCAGGCTTCATGGGCTCC
	Reverse	GGGGACCACTTTGTACAAGAAAGCTGGGTCCTTGGTGC
<i>hph</i> (probe)	Forward	CGCATAACAGCGGTCATTGACTGGAGC
	Reverse	GCTGGGGCGTCGGTTTCCACTATCGG
<i>Ubi</i> (probe)	Forward	CCAAGCGAAGCAGACGGCAC
	Reverse	GGACCGGAACGCCGATCTAG

Supplemental Methods. Phylogenetic analysis.

Plant sequences shown in Supplemental Data Set 1 and Supplemental Data Set 2 were aligned using ClustalW program. The phylogenetic tree was constructed using the neighbor-joining method in the MEGA4.0.2 package (Tamura et al. 2007)

Supplemental Reference.

Tamura, K., Dudley, J., Nei, M., and Kumar, S. (2007). MEGA4: Molecular Evolutionary Genetics Analysis (MEGA) software version 4.0. *Mol Biol Evol* **24**, 1596-1599.