Supporting Information

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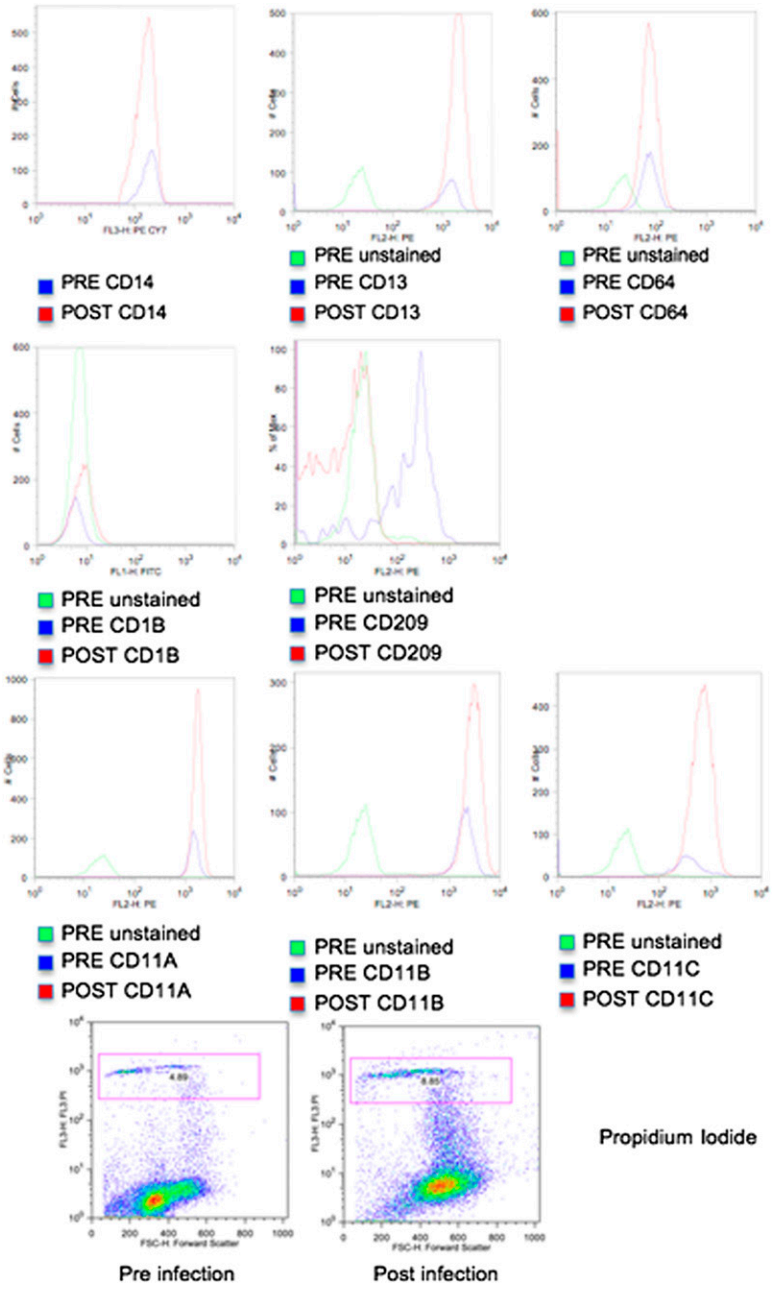

Fig. S1. Isolation of monocytes does not affect their differentiation status. Whole peripheral blood mononuclear cell populations (PRE; blue) and $CD14^+$ monocytes isolated from these populations (POST; red) were analyzed by flow cytometry using the indicated antibodies to analyze cell surface markers. Similar concentrations of dead cells were observed before and after isolation (5% and 9%, respectively, calculated by propidium idodine staining). Unstained control sorts are depicted in green.

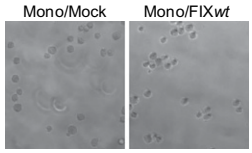

Fig. S2. Infection with the FIX clinical isolate does not alter the morphology of cultured monocytes. Cells were visualized by phase microscopy.

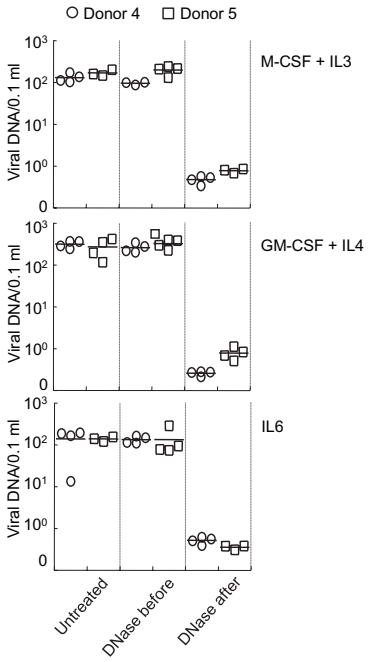

Fig. S3. Viral DNA in the medium of differentiated monocytes is protected from digestion with DNase I. Supernatants from monocytes (Fig. 5*B*, donors 4 and 5) were collected. Viral DNA was isolated with no prior treatment (untreated), after treatment with DNase I but before virions were disrupted with SDS and proteinase K (DNase before), or after treatment with DNase I but after virions were disrupted (DNase after). DNA in the samples was then quantified by quantitative PCR (qPCR) using a primer amplifying a portion of the UL123 ORF. DNA samples were analyzed in quadruplicate; each determination is presented (circles for donor 4 and squares for donor 5), and the mean is indicated with a horizontal bar.

Table S1. Primers used for quantitative PCR assays

Gene	Primer sequence
UL138	5′-TGCGCATGTTTCTGAGCTAC-3′
	5′-ACGGGTTTCAACAGATCGAC-3′
UL82 (LUNA PCR round 1)	5′-ATGACCTCTCCTCCACACC-3′
	5′-GACGCTATATTTAGGGCTTCC-3′
UL81 (LUNA PCR round 2)	5′-GAGCCTTGACGACTTGGTAC-3′
	5′-GGAAAAACACGCGGGGGA-3′
UL122	5′-ATGGTTTTGCAGGCTTTGATG-3′
	5′-ACCTGCCCTTCACGATTCC-3′
UL123	5′-GCCTTCCCTAAGACCACCAAT-3′
	5′-ATTTTCTGGGCATAAGCCATAATC-3′
UL54	5′-CCCTCGGCTTCTCACAACAAT-3′
	5′-CGAGTTAGTCTTGGCCATGCAT-3′
UL94	5-′AGAGCCGGAAACACCGC-3′
	5′-TCCGGACTAAAGATCGTCGAA-3′
GAPDH	5′-ACCCACTCCTCCACCTTTGAC-3′
	5′-CTGTTGCTGTAGCCAAATTCGT -3′
β-actin	5′-CATTGCCGACGGATGCA-3′
	5′-GCCGATCCACACGGAGTACT-3′
vIL-10	5′-TGTTGAGGCGGTATCTGGAGA-3′
	5′-CCGTCTTGAGTCCGGGATAG-3′
US28	5′-TTTGGTGGATCTTTGCCGTG-3′
	5′-ACGAAAGCACCAAGCATGAGTTC-3′