

Asymmetric Construction of Rings A–D of Daphnicyclidin Type Alkaloids

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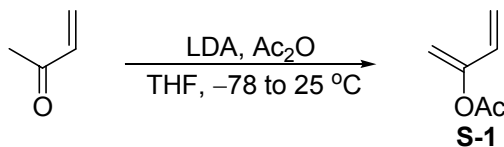
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Materials and Methods. Unless stated otherwise, reactions were conducted in flame-dried glassware under an atmosphere of nitrogen using anhydrous solvents (either freshly distilled or passed through activated alumina columns). All commercially obtained reagents were used as received unless additional purification is stated in the procedure. Reaction temperatures were controlled using an IKAmag temperature modulator, and unless stated otherwise, reactions were performed at room temperature (rt, approximately 25 °C). Thin-layer chromatography (TLC) was conducted with E. Merck silica gel 60 F254 pre-coated plates, (0.25 mm) and visualized by exposure to UV light (254 nm) or stained with ceric ammonium molybdate or potassium permanganate. Flash column chromatography was performed using normal phase silica gel (60 Å, 230-240 mesh, Merck KGA). ¹H NMR spectra were recorded on Bruker spectrometers (at 500 or 600 MHz) and are reported relative to deuterated solvent signals. Data for ¹H NMR spectra are reported as follows: chemical shift (δ ppm), multiplicity, coupling constant (Hz) and integration. ¹³C NMR spectra were recorded on Bruker Spectrometers (at 125 or 150 MHz). Data for ¹³C NMR spectra are reported in terms of chemical shift. IR spectra were recorded on an Applied System REACT-IR 1000 spectrometer or a Varian 640-IR FT-IR spectrometer and are reported in terms of frequency of absorption (cm⁻¹). Optical

rotations were measured with a Jasco P-1010 polarimeter. High resolution mass spectra were obtained from the UC Irvine Mass Spectrometry Facility.

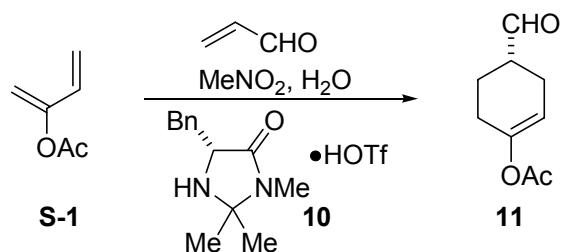
Experimental Procedures



1-Methyleneprop-2-enyl acetate (S-1).¹ A solution of *n*-butyllithium (145 mL, 276 mmol, 1.9 M in hexanes) was added dropwise to a solution of diisopropylamine (42.0 mL, 300 mmol) in THF (190 mL) over 40 min at $-78\text{ }^{\circ}\text{C}$, ensuring that the temperature did not rise above $-60\text{ }^{\circ}\text{C}$ (internal measure). The cooling bath was removed, and the solution was allowed to warm to $-25\text{ }^{\circ}\text{C}$ over 15 min. The solution was then recooled to $-78\text{ }^{\circ}\text{C}$. A solution of freshly distilled methyl vinyl ketone (20.8 mL, 250 mmol) in THF (20 mL) was added over 30 min. After maintaining the reaction for 15 min at $-78\text{ }^{\circ}\text{C}$, acetic anhydride (30.7 mL, 325 mmol) was added over 20 min, resulting in a viscous yellow suspension. The cooling bath and the reaction mixture were allowed to slowly warm to rt and then stirred at rt for 15 h. Pentane (225 mL) was then added, along with saturated aqueous sodium bisulfate (300 mL). The layers were separated, and the aqueous layer was extracted with pentane (225 mL). The combined organic extracts were washed with brine ($2 \times 100\text{ mL}$), dried over MgSO_4 , filtered, and then concentrated to a volume of $\sim 120\text{ mL}$ in vacuo. The mixture was then further concentrated by short path distillation (1 atm) to $\sim 60\text{ mL}$ total volume. Distillation of this residue under partial vacuum (~ 40 torr, $55\text{ }^{\circ}\text{C}$ at thermometer head) afforded **S-1** (13.2 g, 47%) as a colorless oil, contaminated with THF and *N,N*-diisopropylacetamide (approximately 85% pure by ^1H NMR analysis). This material was used in the subsequent Diels-Alder reaction without further purification.² ^1H NMR (400 MHz, CDCl_3) δ 6.27 (ddd, $J = 17.2, 10.9, 0.3\text{ Hz}$, 1H), 5.29 (dq, $J = 17.2, 1.5, 0.6\text{ Hz}$, 1H), 5.18 (dddd, $J = 10.9, 1.5, 0.8, 0.8\text{ Hz}$, 1H), 5.03 (m, 1H), 4.94 (ddd, $J = 1.6, 1.6, 0.6\text{ Hz}$, 1H), 2.23 (s, 3H).

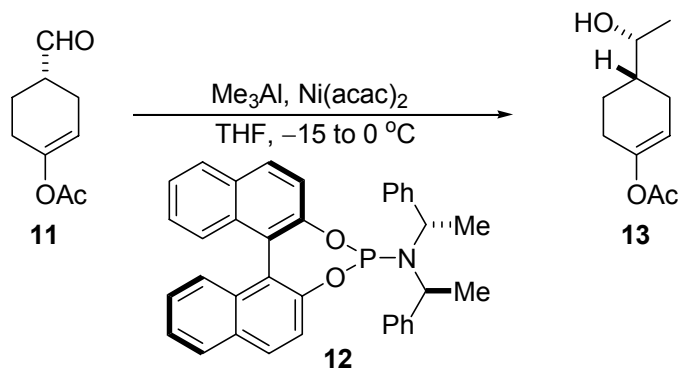
¹ Kowalski, M. D. Studies Toward the Enantioselective Total Synthesis of Manzamine A. Ph.D. Dissertation, University of California, Irvine, CA 2008.

² For an alternative preparation of **S-1**, see: Hagemeyer, H. J.; Hull, D. C. *Ind. Eng. Chem.* **1949**, *41*, 2920–2924.



(S)-4-Formylcyclohex-1-enyl acetate (11). Prepared by the general method of MacMillan.^{3b} Freshly distilled acrolein (1.2 mL, 18 mmol) was added to a solution of acetoxydiene **S-1** (1.2 g, ~85% pure, 9 mmol) and catalyst **10**³ (0.082 g, 0.22 mmol) in water-saturated nitromethane (9 mL). Aliquots of the reaction mixture were diluted with CDCl₃ and analyzed directly by ¹H NMR spectroscopy, with no starting material detected after 21 h. The reaction mixture was then filtered through a plug of silica gel (3 x 9 cm) and eluted with 100 mL each of 20%, 30%, and 50% ethyl acetate in hexanes. The product-containing fractions were concentrated in vacuo to give a solution of the Diels-Alder product and nitromethane. The nitromethane was removed in vacuo at approximately 10 torr pressure. The residue was purified by flash column chromatography (silica gel, gradient: 1.5:1 to 1:1 hexanes:Et₂O), which separated the desired product from a slightly faster-eluting impurity. Aldehyde **11** (1.17 g, 77%) was isolated as a colorless oil: [α]²⁷₅₈₉ -60, [α]²⁷₅₇₇ -63, [α]²⁷₅₄₆ -71, [α]²⁷₄₃₅ -130, [α]²⁷₄₀₅ -162 (*c* 1.85, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 9.70 (s, 1H), 5.41 (s, 1H), 2.57–2.52 (m, 1H), 2.37–2.35 (m, 2H), 2.34–2.28 (m, 1H), 2.22–2.15 (m, 1H), 2.13–2.10 (m, 1H), 2.11 (s, 3H), 1.85–1.79 (m, 1H); ¹³C NMR (125 MHz, CDCl₃) δ 203.1 (CH), 169.2 (C), 148.1 (C), 111.9 (CH), 45.2 (CH), 25.4 (CH₂), 23.0 (CH₂), 22.2 (CH₂), 20.9 (CH₃); IR (film) 2931, 2848, 2719, 1748, 1721, 1692, 1553, 1439, 1366, 1218, 1119, 1040, 909 cm⁻¹; HRMS (ESI) *m/z* calcd for C₉H₁₃O₃ [(M+H)⁺]: 169.0864; found: 169.0866. The ee of the enantiomeric product **ent-11**, prepared using catalyst **ent-10**, was previously determined to be 91% by ¹H NMR analysis of a Mosher ester derivative.¹

³ Catalyst **10** was prepared from its hydrochloride salt by a literature procedure: a) Jen, W. S.; Wiener, J. J. M.; MacMillan, D. W. C. *J. Am. Chem. Soc.* **2000**, *122*, 9874–9875. The hydrochloride salt of **10** was in turn prepared from (*R*)-phenylalanine methyl ester hydrochloride by a literature procedure: b) Ahrendt, K. A.; Borths, C. J.; MacMillan, D. W. C. *J. Am. Chem. Soc.* **2000**, *122*, 4243–4244.



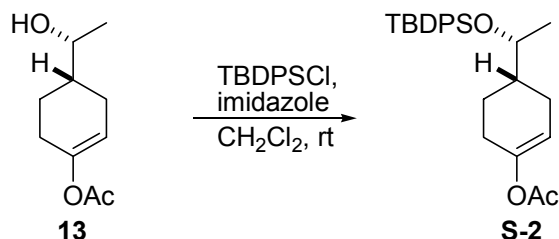
4-((1*R*)-1-Hydroxyethyl)(4*S*)cyclohex-1-enyl acetate (13). Prepared by the general procedure of Woodward.⁴ Phosphoramidite ligand **12**⁵ (0.361 g, 0.669 mmol), Ni(acac)₂ (0.086 g, 0.34 mmol) and THF (105 mL) were combined in a round-bottomed flask. The solution was cooled to 0 °C and stirred for 10 min at 0 °C. Aldehyde **11** (5.63 g, 33.5 mmol) was added, along with a THF rinse (5 mL). The solution was cooled to –20 °C, and trimethylaluminum (34 mL, 67 mmol, 2.0 M in toluene) was added dropwise by addition funnel, maintaining a temperature below –8 °C (internal measure). The solution was warmed to 0 °C and stirred for 1.5 h at 0 °C. The reaction mixture was then transferred through a wide-bored cannula into a stirring, biphasic mixture of aqueous 3 N HCl (110 mL) and Et₂O (160 mL), keeping the temperature below 12 °C. The mixture was then allowed to warm to rt and stirred at rt for 30 min. The layers were separated and the aqueous portion was extracted with Et₂O (160 mL). The combined organic portions were diluted with hexanes (120 mL) and washed with 1 N HCl (120 mL) and brine (120 mL). The combined aqueous washes were then extracted with Et₂O (2 × 160 mL). The combined organic portions were dried over MgSO₄, filtered and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, gradient: 3:7 EtOAc:hexanes to 1:1 EtOAc:hexanes) to give **13** (5.80 g, 94%, 5.7:1 dr as determined by ¹H NMR analysis) as a colorless oil. Because the diastereomeric alcohols were not separable at this stage, the mixture was characterized, with only the signals for the major isomer reported:⁶ ¹H NMR (500 MHz, CDCl₃) δ 5.39–5.38 (m, 1H), 3.74 (appt quint, *J* = 6.0 Hz, 1H), 2.32–2.22 (m, 2H), 2.14–2.10 (m, 1H), 2.12 (s, 3H), 2.05–1.99 (m, 1H), 1.83–1.79 (m, 1H), 1.65–1.59 (m, 1H), 1.59–1.50 (m, 1H), 1.32 (br s, 1H), 1.22 (d, *J* = 6.5

⁴ Biswas, K.; Prieto, O.; Goldsmith, P. J.; Woodward, S. *Angew. Chem., Int. Ed.* **2005**, *44*, 2232–2234.

⁵ Prepared by a literature procedure: Rimkus, A.; Sewald, N. *Synthesis* **2004**, 135–146.

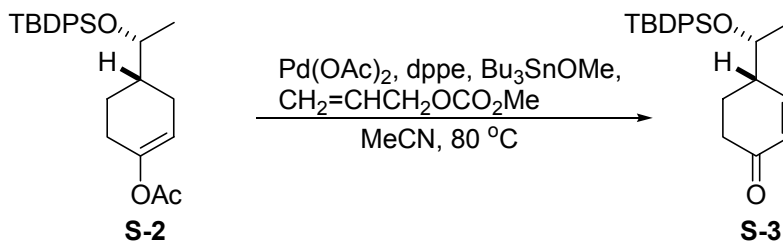
⁶ Optical rotation not determined because this product is a mixture of epimers.

Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 169.5 (C), 148.1 (C), 113.2 (CH), 70.9 (CH), 40.3 (CH), 26.6 (CH_2), 25.1 (CH_2), 25.0 (CH_2), 21.1 (CH_3), 20.9 (CH_3); IR (film) 3400, 2971, 2929, 2896, 1752, 1694, 1439, 1370, 1223, 1113, 913 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{10}\text{H}_{16}\text{O}_3\text{Na}$ $[(\text{M}+\text{Na})^+]$: 207.0997; found: 207.1003.



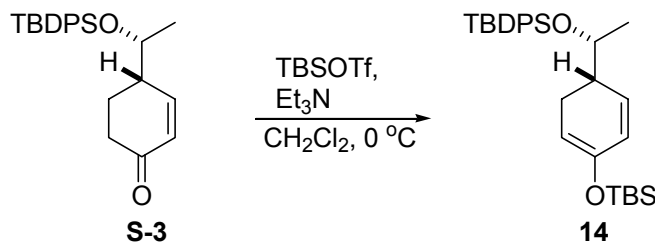
4-[(1*R*)-1-(2,2-Dimethyl-1,1-diphenyl-1-silapropoxy)ethyl](4*S*)cyclohex-1-enyl

acetate (S-2). *tert*-Butyldiphenylsilyl chloride (14.0 mL, 54.0 mmol) was added to a solution of secondary alcohol **13** (9.28 g, 51.5 mmol) and imidazole (7.20 g, 106 mmol) in CH_2Cl_2 (100 mL), resulting in the formation of a precipitate. The suspension was stirred for 16 h at rt, then diluted with CH_2Cl_2 (100 mL) and quenched with saturated aqueous ammonium chloride (100 mL). The layers were separated, and the aqueous portion was extracted with CH_2Cl_2 (2 \times 200 mL). The combined organic extracts were washed with brine (100 mL), dried over MgSO_4 , filtered and concentrated in vacuo. Purification of the residue by silica gel flash column chromatography (gradient: 1:49 EtOAc:hexanes to 1:9 EtOAc:hexanes) gave **S-2** (21.8 g, 100%) as a colorless oil and a 5.7:1 mixture of diastereomers. Because these stereoisomers were not separable at this stage, the mixture was characterized, with only the signals for the major isomer being reported:⁶ ^1H NMR (500 MHz, CDCl_3) δ 7.68 (m, 4H), 7.46–7.37 (m, 6H), 5.37–5.35 (m, 1H), 3.84–3.80 (m, 1H), 2.26–2.24 (m, 1H), 2.17–1.99 (m, 3H), 2.13 (s, 3H), 1.82–1.78 (m, 1H), 1.71–1.64 (m, 1H), 1.57 (qd, $J = 12.0, 5.5$ Hz, 1H), 1.06 (s, 9H), 1.01 (d, $J = 6.5$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 169.5 (C), 148.2 (C), 135.92 (CH), 135.89 (CH), 134.9 (C), 134.1 (C), 129.5 (CH), 129.4 (CH), 127.5 (CH), 127.3 (CH), 113.5 (CH), 72.3 (CH), 40.9 (CH), 27.03 (CH_3), 26.99 (CH_2), 25.1 (CH_2), 24.8 (CH_2), 21.1 (CH_3), 20.4 (CH_3), 19.4 (C); IR (film) 3072, 2960, 2933, 2894, 2858, 1756, 1694, 1428, 1362, 1223, 1208, 1034, 1007 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{26}\text{H}_{34}\text{O}_3\text{SiNa}$ $[(\text{M}+\text{Na})^+]$: 445.2175; found: 445.2164.

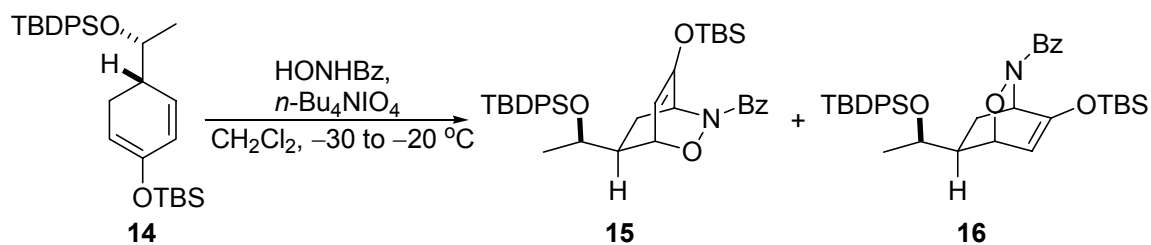


4-[(1*R*)-1-(2,2-Dimethyl-1,1-diphenyl-1-silapropoxy)ethyl](4*S*)cyclohex-2-en-1-one (S-3). Prepared by the general procedure of Tsuji.⁷ Allyl methyl carbonate (8.39 mL, 73.8 mmol), 1,2-bis(diphenylphosphino)ethane (1.47 g, 3.69 mmol), and palladium (II) acetate (0.829 g, 3.69 mmol) were added to a solution of enol acetate **S-2** (15.6 g, 36.9 mmol) in MeCN (185 mL). The resultant suspension was stirred for 10 min at rt. Tributyltin methoxide (3.2 mL, 11 mmol) was added and the suspension was heated at 80 °C for 3 h. The suspension was allowed to cool to rt and was filtered through a silica gel plug, eluting with EtOAc. The organic solution was concentrated in vacuo and the residue was purified by silica gel flash column chromatography (gradient: 1:9 EtOAc:hexanes to 1:3 EtOAc:hexanes) to give enone **S-3** (12.7 g, 91%) as a colorless oil and a 5.7:1 mixture of diastereomers. Because these stereoisomers were not separable at this stage, the mixture was characterized, with only the signals for the major isomer being reported:⁶ ¹H NMR (500 MHz, CDCl₃) δ 7.70–7.67 (m, 4H), 7.46–7.38 (m, 6H), 6.95 (dt, *J* = 10.0, 2.0 Hz, 1H), 6.01 (dd, *J* = 10.0, 2.5 Hz, 1H), 3.94 (appt quint, *J* = 6.0 Hz, 1H), 2.53–2.48 (m, 2H), 2.32 (ddd, *J* = 17.0, 14.0, 5.0 Hz, 1H), 2.07–2.04 (m, 1H), 1.89–1.81 (m, 1H), 1.09 (d, *J* = 6.0 Hz, 3H), 1.07 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 199.9 (C), 151.7 (CH), 135.85 (CH), 135.78 (CH), 134.2 (C), 133.7 (C), 129.8 (CH), 129.71 (CH), 129.68 (CH), 127.7 (CH), 127.5 (CH), 71.5 (CH), 44.1 (CH), 37.3 (CH₂), 27.0 (CH₃), 24.5 (CH₂), 20.5 (CH₃), 19.3 (C); IR (film) 3072, 3049, 2960, 2933, 2889, 2858, 1683, 1472, 1428, 1389, 1362, 1187, 1136, 1106, 1084, 1009 cm⁻¹; HRMS (ESI) *m/z* calcd for C₂₄H₃₀O₂SiNa [(M+Na)⁺]: 401.1913; found: 401.1911.

⁷ Minami, I.; Takahashi, K.; Shimizu, I.; Kimura, T.; Tsuji, J. *Tetrahedron* **1986**, *42*, 2971–2977.



1-{(1*R*)-1-[(1*S*)-4-(1,1,2,2-Tetramethyl-1-silapropoxy)cyclohexa-2,4-dienyl]ethoxy}-2,2-dimethyl-1,1-diphenyl-1-silapropane (14). Triethylamine (6.6 mL, 47 mmol) was added to a solution of enone **S-3** (8.98 g, 23.7 mmol) in CH₂Cl₂ (120 mL) at 0 °C, followed by dropwise addition of *tert*-butyldimethylsilyl trifluoromethanesulfonate (5.99 mL, 26.1 mmol). The solution was maintained at 0 °C for 1 h, then methanol (1.5 mL) was added and the solution was concentrated in vacuo. The oily residue was extracted with pentane (4 × 25 mL), and decanted, and subsequent concentration of the combined organic extracts in vacuo afforded **14** (11.9 g, 102%) as a yellow oil and a 5.7:1 mixture of diastereomers. The material was used in the subsequent reaction without further purification. The mixture was characterized, with only the signals for the major isomer being reported:⁶ ¹H NMR (500 MHz, CDCl₃) δ 7.68 (m, 4H), 7.44–7.36 (m, 6H), 5.72 (dd, *J* = 10.0, 3.0 Hz, 1H), 5.67 (dt, *J* = 10.0, 2.0 Hz, 1H), 4.85 (br s, 1H), 3.87 (appt quint, *J* = 6.0 Hz, 1H), 2.47–2.42 (m, 1H), 2.26 (ddd, *J* = 16.5, 14.0, 3.5 Hz, 1H), 2.17 (ddd, *J* = 16.5, 9.0, 5.5 Hz, 1H), 1.06 (s, 9H), 1.04 (d, *J* = 6.5 Hz, 3H), 0.93 (s, 9H), 0.13 (s, 6H); ¹³C NMR (125 MHz, CDCl₃) δ 148.0, 135.88, 135.85, 134.7, 134.2, 130.1, 129.5, 129.4, 127.5, 127.4, 126.3, 102.0, 71.7, 41.4, 27.0, 25.7, 22.9, 20.3, 19.4, 18.1, –4.47, –4.52; IR (film) 3072, 3051, 2958, 2931, 2889, 2858, 1652, 1590, 1472, 1463, 1428, 1362, 1254, 1111, 1086, 1007 cm⁻¹; HRMS (ESI) *m/z* calcd for C₃₀H₄₅O₂Si₂ [(M+H)⁺]: 493.2958; found: 493.2961.



8-[(1*R*)-1-(2,2-Dimethyl-1,1-diphenyl-1-silapropoxy)ethyl](1*S*,4*S*,8*S*)-2-aza-3-oxa-6-(1,1,2,2-tetramethyl-1-silapropoxy)bicyclo[2.2.2]oct-5-en-2-yl phenyl ketone (15) and

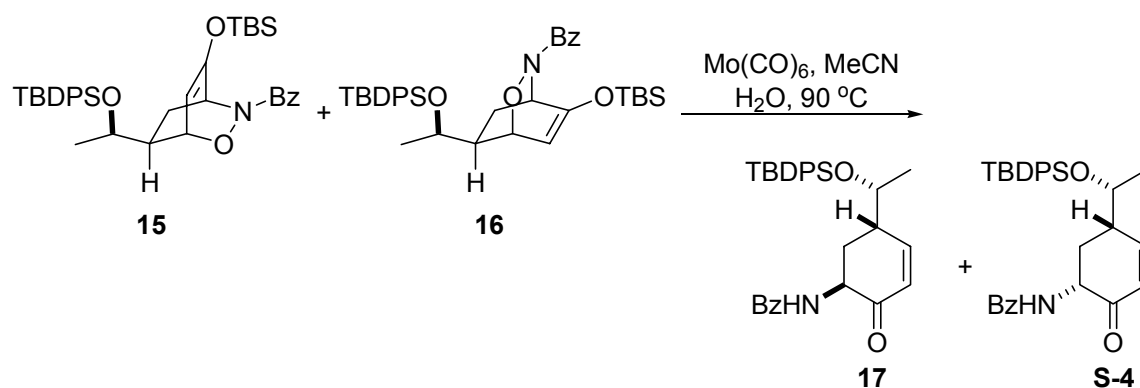
8-[(1*R*)-1-(2,2-dimethyl-1,1-diphenyl-1-silapropoxy)ethyl](8*S*,1*R*,4*R*)-2-aza-3-oxa-6-(1,1,2,2-tetramethyl-1-silapropoxy)bicyclo[2.2.2]oct-5-en-2-yl phenyl ketone (16).

Benzohydroxamic acid (3.58 g, 26.1 mmol) was added to a solution of dienoxysilane **14** (11.7 g, 23.7 mmol) in CH₂Cl₂ (130 mL) and the resultant yellow suspension was cooled to –35 °C. A solution of tetra-*n*-butylammonium periodate (11.3 g, 26.1 mmol) in CH₂Cl₂ (30 mL) was added dropwise by syringe, maintaining a temperature below –30 °C (internal measure), resulting in an orange solution. The reaction was maintained at –30 to –20 °C for 2.5 h, then the mixture was poured into a biphasic mixture of Et₂O (300 mL) and 2 M aqueous sodium thiosulfate (150 mL). The layers were separated and the aqueous portion was extracted with Et₂O (300 mL). The combined organic extracts were washed with 2 M aqueous sodium thiosulfate (150 mL) and water (2 × 225 mL). The organic layer was then diluted with hexanes (500 mL), washed with brine (2 × 225 mL), dried over Na₂SO₄, filtered and concentrated in vacuo. The residue was purified by silica gel flash column chromatography (gradient: 1:9 EtOAc:hexanes to 1:4 EtOAc:hexanes) to give **15** and **16** (9.55 g, 64%, 6:1 dr) as a colorless foam. The isomers were chromatographically separable for characterization purposes. The undesired diastereomer from the Ni-catalyzed methylation step was also separated chromatographically at this stage. **16** elutes first, followed by **15**, which is then followed by the undesired diastereomer.

15: [α]²⁷₅₈₉ +37, [α]²⁷₅₇₇ +39, [α]²⁷₅₄₆ +44, [α]²⁷₄₃₅ +73, [α]²⁷₄₀₅ +89 (*c* 1.67, CHCl₃); ¹H NMR (500 MHz, C₆D₆) δ 8.10 (br d, *J* = 7.0 Hz, 2H), 7.71–7.69 (m, 4H), 7.24–7.18 (m, 6H), 7.11–7.06 (m, 3H), 5.28 (br s, 1H), 5.22 (dd, *J* = 6.0, 3.5 Hz, 1H), 4.93 (dd, *J* = 6.0, 2.5 Hz, 1H), 3.27 (dq, *J* = 10.0, 6.0 Hz, 1H), 2.44–2.39 (m, 1H), 1.90 (ddd, *J* = 13.0, 9.0, 4.0 Hz, 1H), 1.05 (s, 9H), 0.99 (s, 9H), 0.75 (d, *J* = 6.0 Hz, 3H), 0.65 (ddd, *J* = 13.0, 5.0, 2.0 Hz, 1H), 0.21 (s, 3H), 0.11 (s, 3H); ¹³C NMR (125 MHz, C₆D₆, 323 K) δ 171.1 (C), 157.1 (C, broad at rt), 136.3 (CH), 136.2 (CH), 135.1 (C), 134.9 (C), 134.1 (C), 130.9 (CH), 130.1 (CH), 129.94 (CH), 129.92 (CH), 100.3 (CH), 75.9 (CH), 73.1 (CH) 53.5 (CH, broad at rt), 46.2 (CH), 27.2 (CH₃), 26.2 (CH₂), 25.8 (CH₃), 20.6 (CH₃), 19.5 (C), 18.2 (C), –4.3 (CH₃), –4.5 (CH₃), 3 missing carbons attributed to overlap with solvent; IR (film) 3072, 2958, 2931, 2896, 2860, 1638, 1472, 1428, 1376, 1341, 1254, 1233, 1219,

1104, 1077 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{37}\text{H}_{49}\text{O}_4\text{Si}_2\text{Na}$ $[(\text{M}+\text{Na})^+]$: 650.3098; found: 650.3110.

16: $[\alpha]^{29}_{589} +28$, $[\alpha]^{29}_{577} +28$, $[\alpha]^{29}_{546} +32$, $[\alpha]^{29}_{435} +60$, $[\alpha]^{29}_{405} +73$ (c 1.10, CHCl_3); ^1H NMR (500 MHz, C_6D_6) δ 8.12 (br d, $J = 7.0$ Hz, 2H), 7.87–7.81 (m, 4H), 7.28–7.14 (m, 9H), 5.31 (d, $J = 7.0$ Hz, 1H), 5.19 (br s, 1H), 5.13 (dd, $J = 7.0, 2.5$ Hz, 1H), 4.08 (dq, $J = 10.0, 6.0$ Hz, 1H), 1.55 (ddd, $J = 10.0, 10.0, 4.0$ Hz, 1H), 1.38 (ddd, $J = 13.0, 11.0, 3.0$ Hz, 1H), 1.15 (s, 9H), 1.03–1.09 (m, 1H), 0.99 (s, 9H), 0.84 (d, $J = 6.0$ Hz, 3H), 0.20 (s, 3H), 0.12 (s, 3H); ^{13}C NMR (125 MHz, C_6D_6 , 323 K) δ 170.7 (C), 157.4 (C, broad at rt), 136.3 (CH), 136.2 (CH), 135.5 (C), 135.0 (C), 134.1 (C), 131.0 (CH), 130.1 (CH), 129.8 (CH), 128.5 (CH), 102.7 (CH), 74.4 (CH), 71.8 (CH), 53.5 (CH, broad at rt), 46.9 (CH), 27.3 (CH_3), 25.8 (CH_3), 25.1 (CH_2), 22.1 (CH_3), 19.8 (C), 18.2 (C), -4.4 (CH_3), -4.6 (CH_3), 3 missing carbons attributed to overlap with solvent; IR (film) 3072, 2958, 2931, 2896, 2860, 1642, 1472, 1428, 1389, 1378, 1362, 1254, 1237, 1136, 1111 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{37}\text{H}_{49}\text{O}_4\text{Si}_2\text{Na}$ $[(\text{M}+\text{Na})^+]$: 650.3098; found: 650.3093.



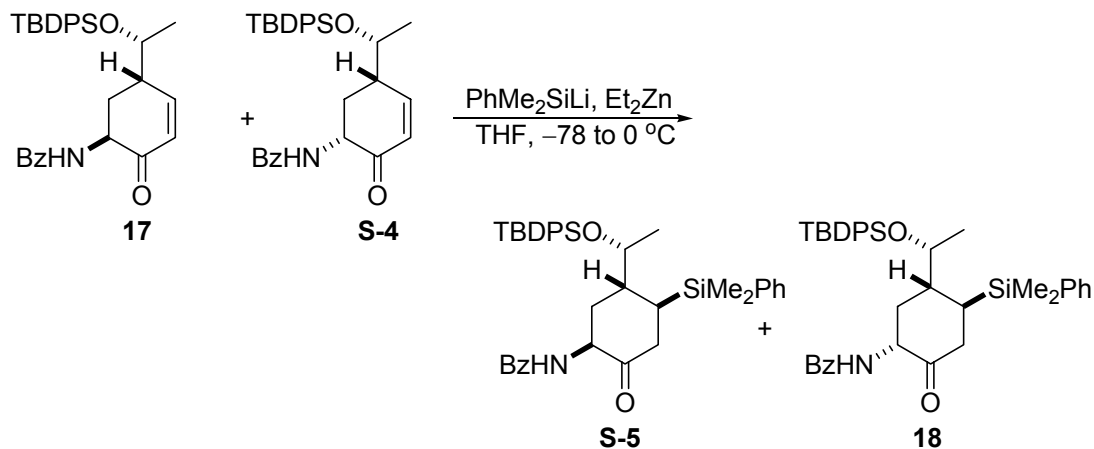
N-{5-[(1*R*)-1-(2,2-Dimethyl-1,1-diphenyl-1-silapropoxy)ethyl](1*S*,5*S*)-2-oxocyclohex-3-enyl}benzamide (**17**) and *N*-{5-[(1*R*)-1-(2,2-dimethyl-1,1-diphenyl-1-silapropoxy)ethyl](5*S*,1*R*)-2-oxocyclohex-3-enyl}benzamide (**S-4**). Prepared by the general procedure of Ghosez.⁸ Molybdenum hexacarbonyl (1.59 g, 6.04 mmol) was added to a mixture of **15** and **16** (3.79 g, 6.04 mmol) in acetonitrile (60 mL) and water (6 mL), resulting in a cloudy suspension. The flask containing the mixture was lowered into an oil bath that had been pre-heated to $90\text{ }^\circ\text{C}$ and the solution was vigorously stirred. The

⁸ Cabanal-Duvillard, I.; Berrien, J.-F.; Ghosez, L.; Husson, H.-P.; Royer, J. *Tetrahedron* **2000**, *56*, 3763–3769.

cloudy suspension became a yellow solution within 9 min. After 38 min at 90 °C, the yellow solution became dark brown and analysis of the reaction mixture by TLC indicated that no starting material remained. The reaction was removed from the oil bath and allowed to cool to rt. The mixture was then filtered through a silica gel plug, eluting with 1:1 EtOAc:hexanes. The filtrate was then concentrated in vacuo. The residue was purified by silica gel flash column chromatography (gradient: 3:17 EtOAc:hexanes to 1:3 EtOAc:hexanes) to give the diastereomeric enones **17** (elutes first) and **S-4** (elutes second) (2.44 g, 81%, 5:1 dr) as a brown foam.

17: $[\alpha]^{24}_{589} -46$, $[\alpha]^{24}_{577} -47$, $[\alpha]^{24}_{546} -54$, $[\alpha]^{24}_{435} -79$, $[\alpha]^{24}_{405} -74$ (*c* 0.91, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 7.82 (d, *J* = 7.5 Hz, 2H), 7.73–7.71 (m, 4H), 7.52 (t, *J* = 7.5 Hz, 1H), 7.46–7.38 (m, 8H), 7.07 (d, *J* = 4.0 Hz, 1H), 7.04 (dd, *J* = 10.0, 5.0 Hz, 1H), 5.99 (d, *J* = 10.0 Hz, 1H), 4.64 (dt, *J* = 13.5, 5.0 Hz, 1H), 4.32 (appt quint, *J* = 6.0 Hz, 1H), 2.85 (dd, *J* = 13.5, 5.0 Hz, 1H), 2.70–2.67 (m, 1H), 1.92 (td, *J* = 13.5, 6.5 Hz, 1H), 1.40 (d, *J* = 6.0 Hz, 3H), 1.08 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 196.6 (C), 167.3 (C), 153.1 (CH), 135.9 (CH), 135.8 (CH), 134.0 (C), 133.9 (C), 133.4 (C), 131.7 (CH), 129.9 (CH), 129.7 (CH), 128.5 (CH), 127.7 (CH), 127.5 (CH), 127.1 (CH), 127.0 (CH), 71.6 (CH), 52.6 (CH), 44.1 (CH), 30.6 (CH₂), 27.0 (CH₃), 22.5 (CH₃), 19.3 (C); IR (film) 3344, 3072, 3054, 2962, 2933, 2894, 2858, 1690, 1648, 1532, 1486, 1474, 1428, 1329, 1111, 1028 cm⁻¹; HRMS (ESI) *m/z* calcd for C₃₁H₃₆NO₃Si [(M+H)⁺]: 498.2465; found: 498.2466.

S-4: ¹H NMR (500 MHz, CDCl₃) δ 7.85 (d, *J* = 7.5 Hz, 2H), 7.69–7.67 (m, 4H), 7.53 (t, *J* = 7.5 Hz, 1H), 7.48–7.38 (m, 8H), 7.15–7.11 (m, 2H), 6.17 (d, *J* = 10.5, 3.0 Hz, 1H), 4.65 (dt, *J* = 13.5, 5.0 Hz, 1H), 3.88 (appt quint, *J* = 6.0 Hz, 1H), 2.83–2.80 (m, 1H), 2.77–2.73 (m, 1H), 1.68 (appt quart, *J* = 12.5 Hz, 1H), 1.12 (d, *J* = 6.0 Hz, 3H), 1.06 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 196.9 (C), 167.3 (C), 152.5 (CH), 135.85 (CH), 135.81 (CH), 134.1 (C), 134.0 (C), 133.4 (C), 131.7 (CH), 129.9 (CH), 129.7 (CH), 128.6 (CH), 127.8 (CH), 127.7 (CH), 127.6 (CH), 127.1 (CH), 71.5 (CH), 55.6 (CH), 44.7 (CH), 32.8 (CH₂), 27.0 (CH₃), 20.9 (CH₃), 19.4 (C); IR (film) 3359, 3072, 2962, 2931, 2892, 2858, 1690, 1656, 1543, 1509, 1484, 1428, 1324, 1218, 1136, 1111, 1067 cm⁻¹; HRMS (ESI) *m/z* calcd for C₃₁H₃₆NO₃Si [(M+H)⁺]: 498.2465; found: 498.2471.



Preparation of phenyldimethylsilyllithium.⁹ Lithium wire (295 mg, 42.5 mmol) was cut into BB-sized pieces, washed with pentane, and placed into a Schlenk flask, along with a magnetic stir bar. THF (25 mL) was added, and the mixture was cooled to 0 °C. Phenyldimethylsilyl chloride (2.10 mL, 12.5 mmol) was added and the mixture was sonicated for 2 h at 0 °C. After 1 h, the mixture became deep red. The mixture was then stirred overnight at 0 °C, then allowed to warm to rt. The prepared reagent was titrated by a total base method by adding the PhMe₂SiLi solution (1.0 mL) to a solution of phenolphthalein (3 mg) in water (10 mL), followed by aqueous HCl (8.0 mL, 0.80 mmol, 0.10 M). Aqueous NaOH (3.4 mL, 0.34 mmol, 0.10 M) was then added dropwise to the solution until a pink color persisted, indicating a concentration of 0.46 M for the PhMe₂SiLi solution.

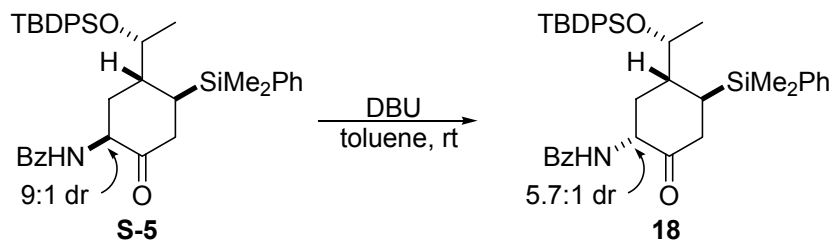
***N*-{5-[(1*R*)-1-(2,2-Dimethyl-1,1-diphenyl-1-silapropoxy)ethyl](4*S*,1*S*,5*R*)-4-(1-methyl-1-phenyl-1-silaethyl)-2-oxocyclohexyl}benzamide (**S-5**) and *N*-{5-[(1*R*)-1-(2,2-dimethyl-1,1-diphenyl-1-silapropoxy)ethyl](4*S*,1*R*,5*R*)-4-(1-methyl-1-phenyl-1-silaethyl)-2-oxocyclohexyl}benzamide (**18**).** Prepared by the general procedure of Fleming.¹⁰ A solution of diethylzinc (6.4 mL, 9.6 mmol, 1.5 M in toluene) was added to THF (40 mL) and the resultant solution was cooled to 0 °C. The previously prepared solution of phenyldimethylsilyllithium (21 mL, 9.6 mmol, 0.46 M in THF) was then added, and the solution was maintained for 15 min at 0 °C, then cooled to -78 °C. A solution of enones **17** and **S-4** (3.00 g, 6.03 mmol) in THF (15 mL with a 5 mL rinse) was added dropwise at -78 °C. The solution was maintained for 15 min at -78 °C, then

⁹ Fleming, I.; Newton, T. W.; Roessler, F. *J. Chem. Soc., Perkin Trans. 1* **1981**, 2527–2532.

¹⁰ Crump, R. A. N. C.; Fleming, I.; Urch, C. J. *J. Chem. Soc., Perkin Trans. 1* **1994**, 701–706.

allowed to slowly warm to 0 °C over 30 min. The reaction was then maintained at 0 °C for 30 min. Saturated aqueous NH₄Cl (60 mL) was then added to the solution, and the mixture was extracted with Et₂O (3 × 100 mL). The combined organic extracts were dried over MgSO₄, filtered and concentrated in vacuo. The residue was purified by silica gel flash column chromatography (gradient: 1:9 EtOAc:hexanes to 1:4 EtOAc:hexanes) to give the ketones **S-5** (elutes first) and **18** (elutes second) (3.09 g, 81%, 9:1 dr) as a colorless foam, along with approximately 0.260 g (approximated by ¹H NMR integration) of dimethylphenylsilanol that co-eluted with **S-5**.

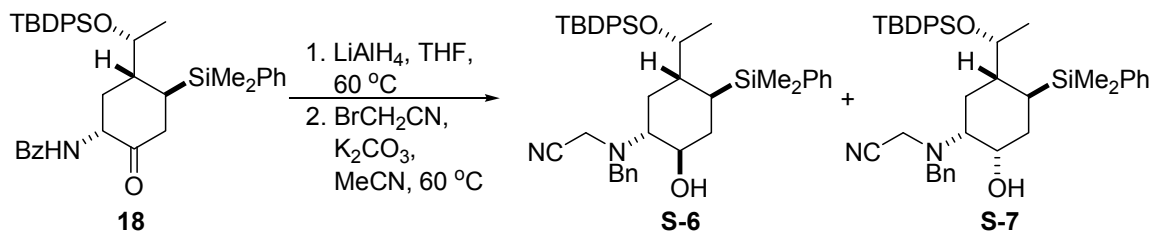
S-5:⁶ ¹H NMR (500 MHz, CDCl₃) δ 7.75 (d, *J* = 7.5 Hz, 2H), 7.23 (d, *J* = 7.0 Hz, 2H), 7.67 (d, *J* = 7.0 Hz, 2H), 7.52–7.31 (m, 14H), 7.01 (br d, *J* = 5.0 Hz, 1H), 4.36 (ddd, *J* = 12.5, 6.0, 5.5 Hz, 1H), 4.25 (dq, *J* = 8.0, 6.0 Hz, 1H), 2.70–2.67 (m, 1H), 2.13–2.07 (m, 2H), 2.02 (br m, 1H), 1.96 (dd, *J* = 14.5, 8.0 Hz, 1H), 1.50 (td, *J* = 13.5, 5.0 Hz, 1H), 1.42 (d, *J* = 6.0 Hz, 3H), 1.08 (s, 9H), 0.29 (s, 3H), 0.27 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 208.3 (C), 166.8 (C), 137.0 (C), 136.1 (CH), 135.9 (CH), 134.0 (C), 133.9 (CH), 133.8 (C), 133.7 (C), 131.7 (CH), 129.8 (CH), 129.7 (CH), 129.2 (CH), 128.5 (CH), 127.83 (CH), 127.78 (CH), 127.5 (CH), 127.0 (CH), 69.1 (CH), 54.7 (CH), 42.6 (CH), 37.1 (CH₂), 33.6 (CH₂), 27.1 (CH₃), 25.1 (CH), 21.4 (CH₃), 19.5 (C), –3.4 (CH₃), –3.5 (CH₃); IR (film) 3346, 3072, 3051, 2958, 2933, 2894, 2858, 1717, 1648, 1511, 1484, 1428, 1378, 1364, 1316, 1252, 1111, 1079 cm⁻¹; HRMS (ESI) *m/z* calcd for C₃₉H₄₈NO₃Si₂ [(M+H)⁺]: 634.3173; found: 634.3178.



***N*-{5-[(1*R*)-1-(2,2-Dimethyl-1,1-diphenyl-1-silapropoxy)ethyl](4*S*,1*R*,5*R*)-4-(1-methyl-1-phenyl-1-silaethyl)-2-oxocyclohexyl}benzamide (**18**)**. A solution of enones **S-5** and **18** (5.29 g, 8.34 mmol) in toluene (55 mL) was cooled to 0 °C and DBU (0.19 mL, 1.3 mmol) was added dropwise. The solution was allowed to warm to rt and maintained at rt for 15 h. The reaction mixture was then filtered through a silica gel plug, eluting

with 1:3 EtOAc:hexanes. The filtrate was concentrated in vacuo and the residue was purified by silica gel flash column chromatography (gradient: 1:19 acetone:hexanes to 1:9 acetone:hexanes) to give the ketones **18** and **S-5** (5.22 g, 99%, 5.7:1 dr) as a colorless foam.

18: $[\alpha]_D^{28}$ $^{28}_{589} +53$, $[\alpha]_D^{28}$ $^{28}_{577} +57$, $[\alpha]_D^{28}$ $^{28}_{546} +65$, $[\alpha]_D^{28}$ $^{28}_{435} +131$, $[\alpha]_D^{29}$ $^{29}_{405} +173$ (*c* 0.58, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.85 (d, *J* = 7.0 Hz, 2H), 7.79–7.77 (m, 2H), 7.64 (d, *J* = 7.0 Hz, 2H), 7.53–7.44 (m, 7H), 7.40 (t, *J* = 7.5 Hz, 2H), 7.30 (t, *J* = 7.5 Hz, 1H), 7.21 (t, *J* = 7.5 Hz, 2H), 7.13 (d, *J* = 7.0 Hz, 2H), 7.03 (br d, *J* = 6.0 Hz, 1H), 4.72 (ddd, *J* = 12.0, 6.0, 6.0 Hz, 1H), 4.04–4.01 (m, 1H), 3.21 (ddd, *J* = 12.5, 6.0, 2.5 Hz, 1H), 2.35 (dd, *J* = 13.5, 3.5 Hz, 1H), 2.22–2.16 (m, 2H), 1.44 (q, *J* = 12.5 Hz, 1H), 1.17 (ddd, *J* = 15.0, 12.0, 3.5 Hz, 1H), 1.06 (s, 9H), 0.68 (d, *J* = 6.0 Hz, 3H), 0.04 (s, 3H), 0.03 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 209.1 (C), 166.7 (C), 137.0 (C), 136.0 (CH), 135.8 (CH), 134.4 (C), 134.2 (C), 134.0 (C), 133.5 (CH), 131.5 (CH), 129.8 (CH), 129.6 (CH), 129.2 (CH), 128.5 (CH), 127.9 (CH), 127.6 (CH), 127.1 (CH), 69.5 (CH), 57.7 (CH), 44.4 (CH), 41.7 (CH₂), 36.7 (CH₂), 30.8 (CH), 27.0 (CH₃), 19.1 (C), 16.6 (CH₃), –3.8 (CH₃), –4.7 (CH₃); IR (film) 3415, 3328, 3072, 3051, 2960, 2933, 2894, 2860, 1717, 1661, 1534, 1513, 1486, 1428, 1362, 1252, 1111, 1069 cm⁻¹; HRMS (ESI) *m/z* calcd for C₃₉H₄₇NO₃Si₂Na [(M+Na)⁺]: 656.2992; found: 656.2994.



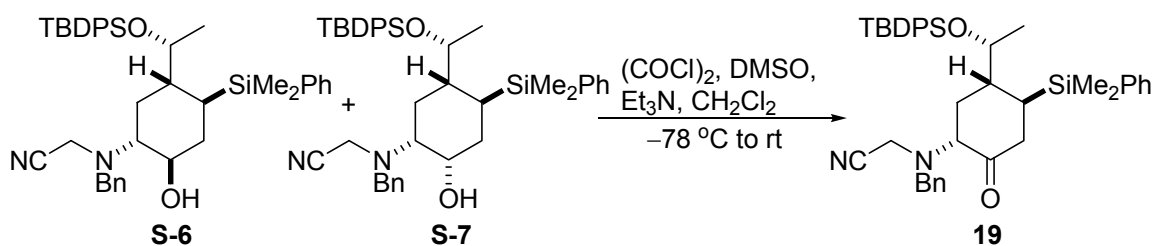
2-({5-[(1*R*)-1-(2,2-Dimethyl-1,1-diphenyl-1-silapropoxy)ethyl](4*S*,1*R*,2*R*,5*R*)-2-hydroxy-4-(1-methyl-1-phenyl-1-silaethyl)cyclohexyl}benzylamino)ethanenitrile (**S-6**) and 2-({5-[(1*R*)-1-(2,2-dimethyl-1,1-diphenyl-1-silapropoxy)ethyl](2*S*,4*S*,1*R*,5*R*)-2-hydroxy-4-(1-methyl-1-phenyl-1-silaethyl)cyclohexyl}benzylamino)ethanenitrile (**S-7**).

Lithium aluminum hydride (9.2 mL, 9.2 mmol, 1.0 M in Et₂O) was added dropwise to a solution of ketone **18** (1.94 g, 3.06 mmol) in THF (20 mL) at 0 °C. The solution was allowed to warm to rt and then heated at 60 °C for 3.5 h. The reaction was then cooled to

0 °C and benzene (10 mL) and sodium fluoride (1.93 g, 45.9 mmol) were added. Water (777 μ L, 43.1 mmol) was then added dropwise 0 °C. The mixture was allowed to warm to rt, then stirred vigorously at rt for 3 h. The suspension was filtered through celite, rinsing with THF (100 mL), and the filtrate was concentrated in vacuo. The resultant amino alcohols were used in the subsequent reaction without further purification. The crude mixture of amino alcohols was dissolved in MeCN (12 mL). Potassium carbonate (1.27 g, 9.18 mmol) was added, followed by bromoacetonitrile (0.62 μ L, 8.9 mmol), and the resultant mixture was heated at 60 °C for 4.5 h. The mixture was then allowed to cool to rt, and water (30 mL) and Et₂O (60 mL) were added. The layers were separated and the aqueous portion was extracted with Et₂O (2 \times 60 mL). The combined organic extracts were diluted with hexanes (50 mL) and washed with saturated aqueous sodium carbonate (20 mL), brine (20 mL), dried over Na₂SO₄, filtered and concentrated in vacuo. The residue was purified by silica gel flash column chromatography (gradient: 1:9 acetone:hexanes to 1:4 acetone:hexanes) to give the nitriles **S-6** and **S-7** (1.94 g, 96%, 4:1 dr) as a colorless solid.

S-6: $[\alpha]^{27}_{589} -2.0$, $[\alpha]^{27}_{577} -2.2$, $[\alpha]^{27}_{546} -3.2$, $[\alpha]^{28}_{435} -8.4$, $[\alpha]^{28}_{405} -12.6$ (*c* 1.36, CHCl₃); ¹H NMR (600 MHz, C₆D₆) δ 7.80 (d, *J* = 7.0 Hz, 2H), 7.76 (d, *J* = 8.0 Hz, 2H), 7.28–7.24 (m, 6H), 7.18–7.06 (m, 10H), 4.11–4.08 (m, 1H), 3.75 (d, *J* = 13.0 Hz, 1H), 3.50 (d, *J* = 13.0 Hz, 1H), 3.15 (ddd, *J* = 10.0, 10.0, 4.5 Hz, 1H), 2.95 (br s, 1H), 2.86 (d, *J* = 17.5 Hz, 1H), 2.76 (br d, *J* = 12.0 Hz, 1H), 2.63 (d, *J* = 17.5 Hz, 1H), 2.33 (ddd, *J* = 12.5, 9.5, 3.0 Hz, 1H), 2.14–2.11 (m, 1H), 1.70–1.64 (m, 1H), 1.25–1.19 (m, 1H), 1.23 (s, 9H), 1.07 (q, *J* = 12.0 Hz, 1H), 0.78 (d, *J* = 6.0 Hz, 3H), 0.49 (ddd, *J* = 14.0, 11.0, 3.0 Hz, 1H), 0.00 (s, 3H), –0.01 (s, 3H); ¹³C NMR (125 MHz, C₆D₆) δ 138.7 (C), 137.7 (C), 136.35 (CH), 136.29 (CH), 134.9 (C), 134.8 (C), 133.9 (CH), 130.0 (CH), 129.9 (CH), 129.7 (CH), 129.1 (CH), 129.0 (CH), 128.5 (CH), 116.9 (C), 70.75 (CH), 70.64 (CH), 69.2 (CH), 53.3 (CH₂), 46.7 (CH), 38.2 (CH₂), 35.5 (CH₂), 27.3 (CH₃), 25.8 (CH), 24.2 (CH₂), 19.4 (C), 17.1 (CH₃), –3.1 (CH₃), –4.4 (CH₃), 3 missing carbons attributed to overlap with solvent and/or accidental equivalence; IR (film) 3500, 3070, 2952, 2931, 2898, 2858, 1590, 1462, 1428, 1382, 1362, 1250, 1111, 1082, 1023 cm⁻¹; HRMS (ESI) *m/z* calcd for C₄₁H₅₂N₂O₂Si₂Na [(M+Na)⁺]: 683.3465; found: 683.3471.

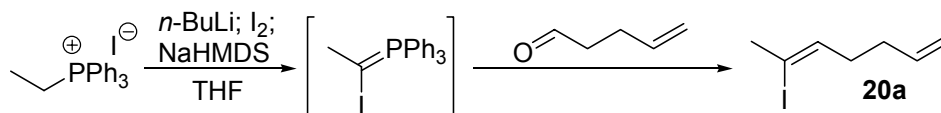
S-7:⁶ ¹H NMR (500 MHz, C₆D₆) δ 7.87 (d, *J* = 7.5 Hz, 2H), 7.77–7.75 (m, 2H), 7.37 (t, *J* = 7.5 Hz, 2H), 7.31–7.07 (m, 14H), 4.18–4.15 (m, 1H), 3.77 (s, 1H), 3.76 (d, *J* = 13.5 Hz, 1H), 3.50 (d, *J* = 13.5 Hz, 1H), 2.98 (d, *J* = 18.0 Hz, 1H), 2.90 (d, *J* = 18.0 Hz, 1H), 2.43 (br d, *J* = 12.0 Hz, 1H), 2.37 (br d, *J* = 12.0 Hz, 1H), 2.23 (s, 1H), 1.95 (br d, *J* = 14.0 Hz, 1H), 1.68 (br t, *J* = 12.0 Hz, 1H), 1.41 (q, *J* = 12.0 Hz, 1H), 1.23 (s, 9H), 1.23–1.20 (m, 1H), 1.00 (t, *J* = 14.0 Hz, 1H), 0.87 (d, *J* = 6.0 Hz, 3H), 0.02 (s, 3H), 0.00 (s, 3H); ¹³C NMR (125 MHz, C₆D₆) δ 139.2, 137.9, 136.4, 136.2, 134.85, 134.76, 134.0, 130.1, 129.9, 129.2, 129.0, 128.9, 115.2, 70.9, 64.7, 62.9, 54.8, 45.8, 37.3, 33.4, 27.3, 23.3, 19.6, 19.4, 17.2, –3.1, –4.5, 4 missing carbons attributed to overlap with solvent; IR (film) 3494, 3070, 2958, 2933, 2896, 2858, 1428, 1382, 1250, 1111, 1082, 1052 cm⁻¹; HRMS (ESI) *m/z* calcd for C₄₁H₅₂N₂O₂Si₂Na [(M+Na)⁺]: 683.3465; found: 683.3461.



2-(Benzyl((1*R*,4*S*,5*R*)-5-((*R*)-1-(*tert*-butyldiphenylsilyloxy)ethyl)-4-

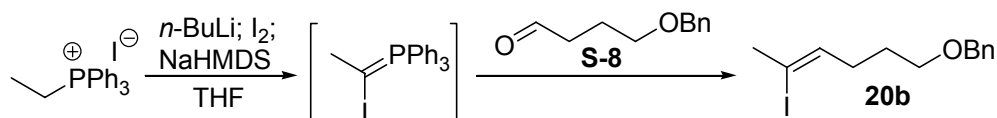
(dimethyl(phenyl)silyl)-2-oxocyclohexyl)amino)acetonitrile (19). A solution of oxalyl chloride (410 μL, 4.70 mmol) in CH₂Cl₂ (2 mL) was added dropwise to a solution of dimethyl sulfoxide (833 μL, 11.7 mmol) in CH₂Cl₂ (14 mL) at –78 °C, maintaining an internal temperature below –70 °C. The solution was stirred for 10 min at –78 °C, and then a solution of amino alcohols **S-6** and **S-7** (1.94 g, 2.93 mmol) in CH₂Cl₂ (14 mL with a 2 mL rinse) was added dropwise, maintaining an internal temperature below –68 °C. The reaction was stirred 15 min at –78 °C and then triethylamine (2.9 mL, 21 mmol) was added dropwise. The reaction was then allowed to warm to –10 °C and saturated aqueous sodium bicarbonate (20 mL) was added. The layers were separated and the organic portion was washed with water (20 mL). The combined aqueous layers were extracted with CH₂Cl₂ (2 × 40 mL). The combined organic extracts were dried over Na₂SO₄, filtered and concentrated in vacuo. The residue was purified by silica gel flash column chromatography (gradient: 1:19 acetone:hexanes to 1:9 acetone:hexanes) to give ketone **19** (1.80 g, 93%) as a colorless foam: [α]₅₈₉²⁴ +51.4, [α]₅₇₇²⁴ +53.6, [α]₅₄₆²⁴ +61.7,

$[\alpha]_{435}^{24} +120$ (*c* 2.82, CH₂Cl₂); ¹H NMR (500 MHz, CDCl₃) δ 7.68 (d, *J* = 7.0 Hz, 2H), 7.62 (d, *J* = 7.0 Hz, 2H), 7.50–7.28 (m, 12H), 7.20 (t, *J* = 7.5 Hz, 2H), 7.12 (d, *J* = 7.0 Hz, 2H), 4.04–4.00 (m, 1H), 3.90 (d, *J* = 13.5 Hz, 1H), 3.86 (d, *J* = 13.5 Hz, 1H), 3.71 (d, *J* = 18.0 Hz, 1H), 3.67 (d, *J* = 18.0 Hz, 1H), 3.42 (dd, *J* = 13.0, 5.5 Hz, 1H), 2.93–2.90 (m, 1H), 2.25 (dd, *J* = 13.5, 3.5 Hz, 1H), 2.04 (t, *J* = 13.5 Hz, 1H), 2.01–1.96 (m, 1H), 1.65 (q, *J* = 12.5 Hz, 1H), 1.12 (ddd, *J* = 15.0, 11.5, 3.5 Hz, 1H), 1.07 (s, 9H), 0.69 (d, *J* = 6.0 Hz, 3H), 0.02 (s, 3H), –0.01 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 210.3 (C), 137.4 (C), 137.1 (C), 135.9 (CH), 135.8 (CH), 134.09 (C), 134.07 (C), 133.5 (CH), 129.8 (CH), 129.7 (CH), 129.1 (CH), 128.8 (CH), 128.6 (CH), 127.9 (CH), 127.8 (CH), 127.7 (CH), 127.7 (CH), 116.8 (C), 69.7 (CH), 67.9 (CH), 55.6 (CH₂), 45.4 (CH), 42.7 (CH₂), 38.7 (CH₂), 31.9 (CH₂), 30.0 (CH), 26.9 (CH₃), 19.2 (C), 16.7 (CH₃), –3.8 (CH₃), –5.0 (CH₃); IR (film) 3070, 2960, 2931, 2894, 2858, 1717, 1590, 1428, 1383, 1252, 1111, 1079 cm⁻¹; HRMS (ESI) *m/z* calcd for C₄₁H₅₀N₂O₂Si₂Na [(M+Na)⁺]: 681.3309; found: 681.3324.



(Z)-6-Iodohepta-1,5-diene (20a). A solution of *n*-butyllithium (9.70 mL, 23.8 mmol, 2.45 M in hexanes) was added dropwise to a suspension of ethyltriphenylphosphonium iodide (9.94 g, 23.8 mmol) in THF (50 mL), ensuring that the temperature remained below 35 °C (internal measure). The resultant red solution was stirred 30 min at rt. In a separate flask, a solution of iodine (6.03 g, 23.8 mmol) in THF (140 mL) was cooled to –78 °C. The red solution of ylide was then transferred into the iodine solution by cannula over 25 min, maintaining a temperature below –65 °C (internal measure). The reaction was then allowed to warm to –10 °C and then re-cooled to –45 °C. A solution of sodium hexamethyldisilazide (23.8 mL, 23.8 mmol, 1.0 M in THF) was added dropwise over 15 min, resulting in a red solution. After 10 min at –40 °C, 4-pentenal (1.18 mL, 11.9 mmol) was quickly added, and the solution was allowed to warm to –10 °C. The reaction was quenched with methanol (1.5 mL), resulting in a brown suspension, and silica gel (90 mL) was added. The suspension was concentrated in vacuo, and the residue was dry loaded onto a silica gel column and eluted with pentane:Et₂O (49:1) to provide **20a**, contaminated with triphenylphosphine. The mixture was re-purified by silica flash column

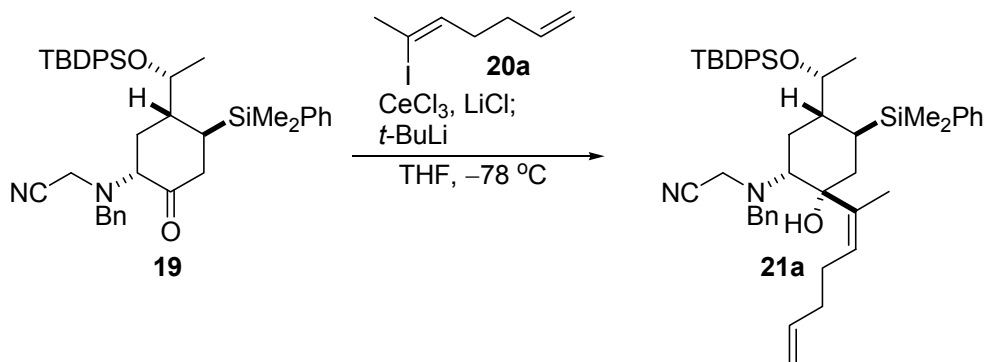
chromatography (49:1 pentane:Et₂O) to provide **20a** (0.980 g, 37%) as a colorless oil: ¹H NMR (500 MHz, CDCl₃) δ 5.83 (dddd, *J* = 16.3, 12.1, 5.9, 5.9 Hz, 1H), 5.44 (t, *J* = 6.3 Hz, 1H), 5.05 (d, *J* = 17.2 Hz, 1H), 5.00 (d, *J* = 10.3 Hz, 1H), 2.51 (s, 3H), 2.24–2.11 (m, 4H); ¹³C NMR (125 MHz, CDCl₃) δ 137.7 (CH), 134.6 (CH), 115.1 (CH₂), 101.3 (C), 35.8 (CH₂), 33.5 (CH₃), 32.4 (CH₂); IR (film): 2921 cm⁻¹; attempts to obtain HRMS data were unsuccessful, presumably due to poor ionization of the molecule.



(Z)-1-((5-Iodohex-4-enyloxy)methyl)benzene (20b). A solution of *n*-butyllithium (38.2 mL, 61.0 mmol, 1.60 M in hexanes) was added dropwise over 45 min to a suspension of ethyltriphenylphosphonium iodide (25.5 g, 61.0 mmol) in THF (140 mL), maintaining a temperature below 35 °C (internal measure). The resultant red solution was stirred 30 min at rt. In a separate flask, a solution of iodine (15.5 g, 61.0 mmol) in THF (340 mL) was cooled to –78 °C. The red solution of ylide was transferred by cannula into the flask containing the iodine solution over 50 min, maintaining a temperature below –65 °C (internal measure), resulting in the formation of a yellow precipitate. The suspension was allowed to warm to –10 °C, then re-cooled to –45 °C. A solution of sodium hexamethyldisilazide (61 mL, 61 mmol, 1.0 M in THF) was added over 20 min, resulting in a red solution. After 10 min at –40 °C, a solution of aldehyde **S-8**¹¹ (5.44 g, 30.5 mmol) in THF (15 mL) was quickly added, and the solution was allowed to warm to –15 °C. Methanol (3 mL) was then added, resulting in a brown suspension, and silica gel was added. The suspension was concentrated in vacuo, and the crude material was dry loaded onto a silica gel column and eluted with hexanes:Et₂O (49:1) to provide slightly impure **20b** as a 7:1 mixture of *Z* and *E* isomers. The 7:1 mixture of olefin isomers was re-purified by silver nitrate impregnated silica gel flash column chromatography (49:1 hexanes:Et₂O) to give **20b** (2.04 g, 21%) as a colorless oil: ¹H NMR (600 MHz, CDCl₃) δ 7.35–7.28 (m, 5H), 5.44 (t, *J* = 7.0 Hz, 1H), 4.52 (s, 2H), 3.50 (t, *J* = 6.5 Hz, 2H), 2.50 (s, 3H), 2.20 (q, *J* = 7.0 Hz, 2H), 1.73 (quint, *J* = 7.0 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 138.5,

¹¹ Moreau, B.; Ginisty, M.; Alberico, D.; Charette, A. B. *J. Org. Chem.* **2007**, *72*, 1235–1240.

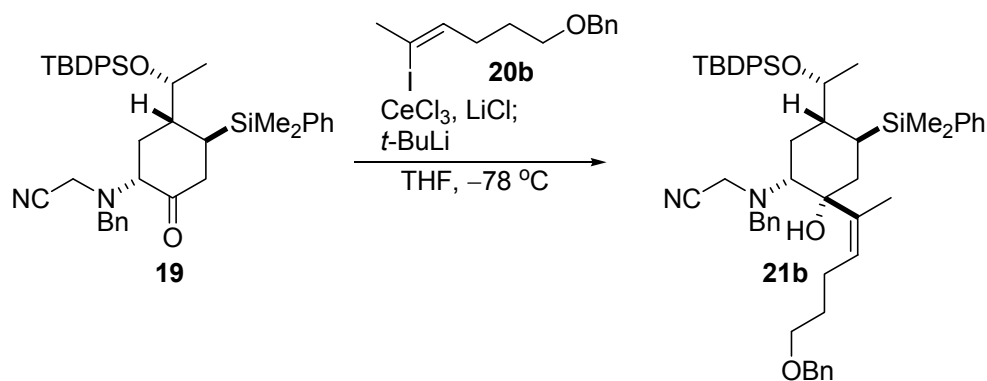
134.8, 128.3, 127.7, 127.5, 101.3, 72.9, 69.6, 33.5, 33.4, 28.4; IR (film) 3064, 3029, 2943, 2916, 2858, 1495, 1455, 1426, 1364, 1272, 1024 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{13}\text{H}_{17}\text{OINa}$ $[(\text{M}+\text{Na})^+]$: 339.0222; found: 339.0220.



2-(Benzyl((1*R*,2*S*,4*S*,5*R*)-5-((*R*)-1-(*tert*-butyldiphenylsilyloxy)ethyl)-4-(dimethyl(phenyl)silyl)-2-((*Z*)-hepta-2,6-dien-2-yl)-2-

hydroxycyclohexyl)amino)acetonitrile (21a). Note: THF used in this reaction was freshly distilled from sodium/benzophenone. Lithium chloride (95 mg, 2.2 mmol) and anhydrous cerium (III) trichloride (275 mg, 1.12 mmol) were added to a round-bottomed flask in a glove box. The flask was removed from the glove box, THF (10 mL) was added, and the suspension was stirred vigorously for 2 h at rt, producing a near colorless solution. A solution of ketone **19** (480 mg, 0.744 mmol) in THF (8 mL with a 2 mL rinse) was added by cannula. The solution was stirred 2 h at rt, then a solution of iodide **20a** (413 mg, 1.86 mmol) in THF (4 mL with a 1 mL rinse) was added. The solution was cooled to $-78\text{ }^{\circ}\text{C}$, and a solution of *tert*-butyllithium (1.4 mL, 2.1 mmol, 1.5 M in pentane) was added dropwise in two portions. The progress of the reaction was monitored by thin layer chromatography after the addition of each portion. Methanol (1.5 mL) was then added at $-78\text{ }^{\circ}\text{C}$, the cold bath was removed, and 10% aqueous AcOH (5 mL), saturated aqueous sodium thiosulfate (5 mL), and Et_2O (30 mL) were added sequentially. After the mixture warmed to rt, Et_2O (30 mL) and water (30 mL) were added, and the layers were separated. The aqueous portion was extracted with Et_2O (60 mL). The combined organic extracts were diluted with hexanes (30 mL), and washed sequentially with saturated aqueous sodium thiosulfate (40 mL), saturated aqueous sodium bicarbonate (40 mL), and brine (40 mL). The organic portion was dried over Na_2SO_4 ,

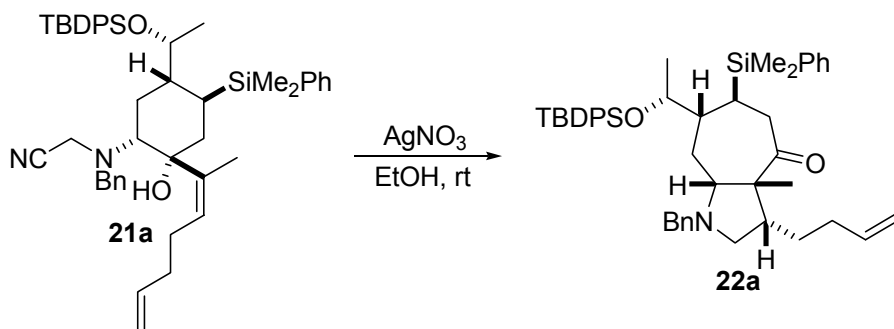
filtered and concentrated in vacuo. The residue was purified by silica gel flash column chromatography (gradient: 1:19 acetone:hexanes to 1:9 acetone:hexanes) to give **21a** (410 mg, 73%) as a pale yellow foam, along with the product derived from *tert*-butyl addition to ketone **19** (36 mg, 7%): $[\alpha]^{24}_{589} +39$, $[\alpha]^{24}_{577} +42$, $[\alpha]^{24}_{546} +47$, $[\alpha]^{24}_{435} +76$ ($c = 0.63$, CH_2Cl_2); ^1H NMR (500 MHz, CDCl_3): δ 7.72 (m, 2H), 7.64 (m, 2H), 7.46 (m, 2H), 7.42 (m, 4H), 7.35–7.26 (m, 6H), 7.19 (m, 2H), 7.14 (m, 2H), 5.77 (dddd, $J = 16.9$, 10.2, 6.7, 6.7 Hz, 1H), 5.19 (ddd, $J = 6.4$, 6.4, 1.0 Hz, 1H), 5.00 (dd, $J = 17.1$, 1.8 Hz, 1H), 4.95 (dd, $J = 10.2$, 0.8 Hz, 1H), 4.06 (d, $J = 13.3$ Hz, 1H), 4.01 (m, 1H), 3.91 (d, $J = 17.5$ Hz, 1H), 3.82 (d, $J = 13.3$ Hz, 1H), 3.50 (d, $J = 17.4$ Hz, 1H), 2.84 (dd, $J = 12.2$, 3.1 Hz, 1H), 2.46 (m, 1H), 2.29 (m, 2H), 2.06 (ddd, $J = 7.1$, 7.1, 7.1 Hz, 2H), 1.96 (s, 1H), 1.69 (s, 3H), 1.66 (ddd, $J = 12.1$, 12.1, 12.1 Hz, 1H), 1.49 (d, $J = 8.3$ Hz, 3H), 1.08 (s, 9H), 0.97 (m, 1H), 0.76 (d, $J = 6.2$ Hz, 3H), 0.00 (s, 3H), -0.05 (s, 3H); ^{13}C NMR (125 MHz, CDCl_3): δ 138.6 (C), 138.4 (CH), 138.1 (C), 136.03 (CH), 135.98 (CH), 134.7 (C), 134.5 (C), 133.6 (CH), 129.7 (CH), 129.6 (CH), 129.0 (CH), 128.8 (CH), 128.6 (CH), 127.8 (CH), 127.6 (CH), 117.7 (C), 115.1 (CH_2), 70.5 (CH), 64.8 (CH), 56.7 (CH_2), 46.2 (CH), 40.1 (CH_2), 39.0 (CH_2), 34.5 (CH_2), 28.7 (CH_2), 27.1 (CH_3), 23.4 (CH_3), 21.8 (CH), 21.6 (C), 19.3 (C), 16.9 (CH_3), -3.3 (CH_3), -4.5 (CH_3), 5 missing carbons attributed to accidental equivalence and/or broadening; IR (film) 3512, 2930 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{48}\text{H}_{63}\text{N}_2\text{O}_2\text{Si}_2$ $[(\text{M}+\text{H})^+]$: 755.4428; found: 755.4431.



2-(Benzyl((1*R*,2*S*,4*S*,5*R*)-2-((*Z*)-6-(benzyloxy)hex-2-en-2-yl)-5-((*R*)-1-(*tert*-butyldiphenylsilyloxy)ethyl)-4-(dimethyl(phenyl)silyl)-2-hydroxycyclohexyl)amino)acetonitrile (21b**). Note: THF used in this reaction was freshly distilled from sodium/benzophenone. Anhydrous cerium (III) trichloride (93 mg,**

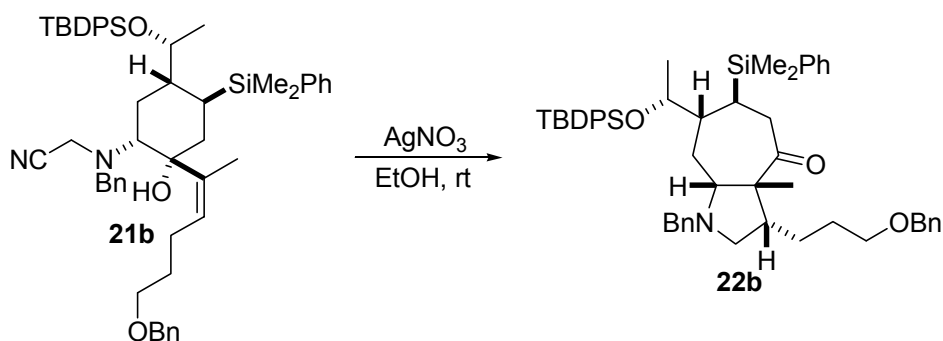
0.38 mmol) and lithium chloride (32 mg, 0.76 mmol) were added to a round-bottomed flask in a glove box. The flask was removed from the glove box, THF (1.6 mL) was added, and the suspension was stirred vigorously and heated at 45 °C for 3 h. The suspension was allowed to cool to rt, and a solution of ketone **19** (165 mg, 0.250 mmol) in THF (1.5 mL with a 0.5 mL rinse) was added by cannula. The suspension was stirred 2.5 h at rt, then a solution of iodide **20b** (151 mg, 0.501 mmol) in THF (3.4 mL) was added. The suspension was cooled to -78 °C, and a solution of *tert*-butyllithium (0.60 mL, 0.90 mmol, 1.5 M in pentane) was added dropwise in three portions. The progress of the reaction was monitored by thin layer chromatography after the addition of each portion. Methanol (0.5 mL) was then added at -78 °C, the cold bath was removed, and 10% aqueous AcOH (1 mL), saturated aqueous sodium thiosulfate (1 mL), and Et₂O (5 mL) were added sequentially. After allowing the mixture to warm to rt, Et₂O (10 mL) and water (10 mL) were added, and the layers were separated. The aqueous portion was extracted with Et₂O (10 mL). The combined organic extracts were diluted with hexanes (10 mL), and the resultant solution was washed sequentially with saturated aqueous sodium thiosulfate (7 mL), saturated aqueous sodium bicarbonate (7 mL), and brine (7 mL). The organic portion was dried over Na₂SO₄, filtered and concentrated in vacuo. The residue was purified by silica gel flash column chromatography (gradient: 1:19 EtOAc:hexanes to 1:9 EtOAc:hexanes) to give **21b** (151 mg, 71%) as a pale yellow foam, along with the product derived from *tert*-butyl addition to ketone **19** (14 mg, 8%): $[\alpha]_{589}^{25} +35.5$, $[\alpha]_{577}^{26} +36.8$, $[\alpha]_{546}^{25} +41.9$, $[\alpha]_{435}^{26} +68.5$ (*c* 2.13, CH₂Cl₂); ¹H NMR (500 MHz, CDCl₃) δ 7.70 (d, *J* = 7.0 Hz, 2H), 7.65 (d, *J* = 7.0 Hz, 2H), 7.47–7.23 (m, 17H), 7.15 (t, *J* = 7.5 Hz, 2H), 7.08 (d, *J* = 7.0 Hz, 2H), 5.20 (t, *J* = 8.0 Hz, 1H), 4.42 (d, *J* = 12.0 Hz, 1H), 4.38 (d, *J* = 12.0 Hz, 1H), 4.09 (d, *J* = 13.5 Hz, 1H), 4.02 (d, *J* = 17.0 Hz, 1H), 4.00–3.94 (m, 1H), 3.77 (d, *J* = 13.5 Hz, 1H), 3.55 (d, *J* = 17.0 Hz, 1H), 3.50–3.45 (m, 1H), 3.44–3.40 (m, 1H), 3.01 (br s, 1H), 2.85–2.70 (br m, 1H), 2.74 (br d, *J* = 12.0 Hz, 1H), 2.41 (br d, *J* = 12.0 Hz, 1H), 2.19–2.14 (m, 1H), 1.81–1.74 (m, 1H), 1.77 (q, *J* = 12.0 Hz, 1H), 1.63 (s, 3H), 1.52–1.46 (m, 3H), 1.39 (dd, *J* = 14.0, 2.5 Hz, 1H), 1.09 (s, 9H), 1.00 (ddd, *J* = 14.0, 11.0, 3 Hz, 1H), 0.75 (d, *J* = 6.0 Hz, 3H), -0.05 (s, 3H), -0.07 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 140.2 (C), 138.7 (C), 138.3 (C), 137.9 (C), 135.93 (CH), 135.89 (CH), 134.60 (C), 134.52 (C), 133.6 (CH), 129.56 (CH), 129.44 (CH), 128.97

(CH), 128.56 (CH), 128.40 (CH), 128.36 (CH), 127.64 (CH), 127.62 (CH), 127.56 (CH), 127.49 (CH), 127.36 (CH), 126.8 (CH), 118.1 (C), 79.7 (C), 72.3 (CH₂), 70.5 (CH), 68.6 (CH₂), 64.2 (CH), 56.9 (CH₂), 46.4 (CH), 40.7 (CH₂), 38.6 (CH₂), 28.9 (CH₂), 27.0 (CH₃), 24.8 (CH₂), 23.1 (CH₃), 21.3 (CH), 20.1 (CH₂), 19.2 (C), 16.8 (CH₃), -3.5 (CH₃), -4.5 (CH₃), missing carbon attributed to accidental equivalence; IR (film) 3400, 3070, 3031, 2956, 2931, 2858, 1455, 1428, 1382, 1250, 1111, 1088 cm⁻¹; HRMS (ESI) *m/z* calcd for C₅₄H₆₈N₂O₃Si₂Na [(M+Na)⁺]: 871.4666; found: 871.4675.



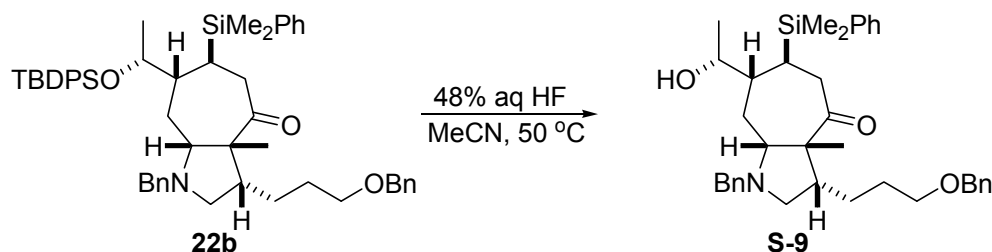
(3*S*,3*aR*,6*S*,7*R*,8*aR*)-1-Benzyl-3-(but-3-enyl)-7-((*R*)-1-(*tert*-butyldiphenylsilyloxy)ethyl)-6-(dimethyl(phenyl)silyl)-3*a*-methyl-octahydrocyclohepta[*b*]pyrrol-4(5*H*)-one (22a**). Silver nitrate (0.191 g, 1.12 mmol) was added to a solution of tertiary allylic alcohol **21a** (0.707 g, 0.936 mmol) in ethanol (10 mL), resulting in a brown suspension. The flask was covered with aluminum foil and the reaction was stirred for 3.5 h at rt. Saturated aqueous ammonium hydroxide (20 mL) was then added, followed by water (20 mL). The mixture was extracted with a 1:3 mixture of hexanes:EtOAc (2 × 100 mL). The combined organic extracts were washed with brine (50 mL), dried over Na₂SO₄, filtered and concentrated in vacuo. The residue was purified by silica gel flash column chromatography (1:19 acetone:hexanes) to give **22a** (0.605 g, 89%) as a colorless foam: $[\alpha]_{589}^{23} +1.18$, $[\alpha]_{577}^{23} +1.23$, $[\alpha]_{546}^{23} +1.75$, $[\alpha]_{435}^{23} +9.32$, $[\alpha]_{405}^{23} +16.8$ ($c = 1.77$, CH₂Cl₂); ¹H NMR (500 MHz, CDCl₃) δ 7.66 (d, $J = 7.4$ Hz, 2H), 7.63 (d, $J = 7.4$ Hz, 2H), 7.44 (m, 2H), 7.40–7.29 (m, 8H), 7.25 (m, 2H), 7.18 (m, 4H), 5.78 (m, 1H), 4.97 (d, $J = 17.2$ Hz, 1H), 4.92 (d, $J = 10.2$ Hz, 1H), 4.08 (d, $J = 13.7$ Hz, 1H), 3.95 (br s, 1H), 3.65 (d, $J = 13.7$ Hz, 1H), 2.77–2.68 (m, 4H), 2.61 (d, $J = 10.3$ Hz, 1H), 2.42 (dd, $J = 10.2, 10.2$ Hz, 1H), 2.08–1.94 (m, 2H), 1.91 (m, 1H), 1.80 (br s, 2H), 1.43 (m, 2H), 1.19 (s, 3H), 1.05 (s, 10H), 0.66 (d, $J = 6.1$ Hz, 3H), 0.13 (s,**

3H), 0.02 (s, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 215.3 (C), 139.6 (CH), 138.9 (C), 138.7 (C), 136.0 (CH), 134.6 (C), 134.4 (C), 133.8 (CH), 129.8 (CH), 129.7 (CH), 128.9 (CH), 128.7 (CH), 128.3 (CH), 127.9 (CH), 127.8 (CH), 127.7 (CH), 126.9 (CH), 114.6 (CH₂), 70.72 (CH), 70.67 (CH), 61.5 (C), 57.9 (CH₂), 55.4 (CH₂), 47.4 (CH), 47.3 (CH), 42.6 (CH₂), 34.4 (CH₂), 33.9 (CH₂), 29.6 (CH₂), 29.0 (CH), 27.9 (CH₃), 27.1 (CH₃), 19.3 (C), 16.9 (CH₃), -2.4 (CH₃), -3.9 (CH₃), missing carbon attributed to accidental equivalence; IR (film) 2930, 1693 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{47}\text{H}_{62}\text{NO}_2\text{Si}_2$ $[(\text{M}+\text{H})^+]$: 728.4319; found: 728.4316.



(3*S*,3*aR*,6*S*,7*R*,8*aR*)-1-Benzyl-3-(3-(benzyloxy)propyl)-7-((*R*)-1-(*tert*-butyldiphenylsilyloxy)ethyl)-6-(dimethyl(phenyl)silyl)-3*a*-methyl-octahydrocyclohepta[*b*]pyrrol-4(5*H*)-one (22b). Silver nitrate (0.108 g, 0.637 mmol) was added to a solution of tertiary allylic alcohol **21b** (0.451 g, 0.531 mmol) in ethanol (5 mL), resulting in a brown suspension. The flask was covered with aluminum foil and the reaction was stirred for 4 h at rt. Saturated aqueous ammonium hydroxide (10 mL) was added, followed by water (10 mL). The mixture was extracted with 1:3 hexanes:EtOAc (2 × 50 mL). The combined organic extracts were washed with water (10 mL), brine (30 mL), dried over Na_2SO_4 and concentrated in vacuo. The residue was purified by silica gel flash column chromatography (1:19 acetone:hexanes) to give **22b** (0.395 g, 90%) as a colorless foam: $[\alpha]^{24}_{589} -2.3$, $[\alpha]^{24}_{577} -2.3$, $[\alpha]^{24}_{546} -2.2$, $[\alpha]^{24}_{435} +2.4$ (c 2.28, EtOH); ^1H NMR (600 MHz, C_6D_6) δ 7.74–7.69 (m, 4H), 7.44 (d, $J = 7.0$ Hz, 2H), 7.31 (d, $J = 7.0$ Hz, 2H), 7.25–7.08 (m, 17H), 4.29 (s, 2H), 4.17 (d, $J = 13.5$ Hz, 1H), 4.10–4.02 (br m, 1H), 3.57 (d, $J = 13.5$ Hz, 1H), 3.30–3.25 (m, 2H), 2.97–2.92 (m, 1H), 2.81 (dd, $J = 11.0$, 4.0 Hz, 1H), 2.77–2.73 (m, 2H), 2.57 (dd, $J = 9.0$, 6.5 Hz, 1H), 2.42 (dd, $J = 11.0$, 8.5 Hz, 1H), 2.25–2.17 (m, 1H), 1.96–1.91 (m, 1H), 1.81–1.78 (m, 1H), 1.70–1.53 (m, 3H), 1.47–

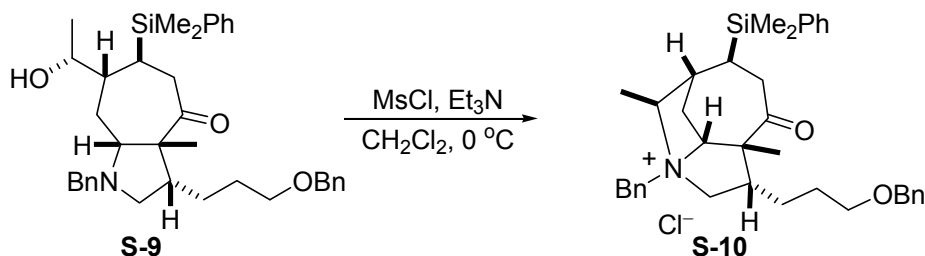
1.43 (m, 1H), 1.20 (s, 3H), 1.14 (s, 9H), 1.05–1.00 (m, 1H), 0.69 (d, $J = 6.0$ Hz, 3H), 0.18 (s, 3H), 0.08 (s, 3H); ^{13}C NMR (125 MHz, C_6D_6) δ 213.7 (C), 140.2 (C), 139.6 (C), 139.1 (C), 136.3 (CH), 134.9 (C), 134.7 (C), 134.0 (CH), 130.0 (CH), 129.9 (CH), 128.9 (CH), 128.7 (CH), 128.6 (CH), 128.4 (CH), 127.7 (CH), 127.4 (CH), 127.2 (CH), 72.7 (CH_2), 71.5 (CH), 71.0 (CH), 70.5 (CH_2), 61.4 (C), 58.4 (CH_2), 56.0 (CH_2), 48.6 (CH), 47.1 (CH), 42.5 (CH_2), 34.3 (br, CH_2), 30.2 (CH_2), 28.8 (br, CH), 28.0 (CH_3), 27.7 (CH_2), 27.2 (CH_3), 19.4 (C), 16.9 (br, CH_3), -2.2 (CH_3), -4.0 (CH_3), 4 missing carbons attributed to overlap with solvent and/or accidental equivalence; IR (film) 3068, 2958, 2931, 2858, 2800, 1698, 1455, 1428, 1382, 1362, 1258, 1111, 1079 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{53}\text{H}_{68}\text{NO}_3\text{Si}_2$ [(M+H) $^+$]: 822.4738; found: 822.4721.



(3*S*,3*aR*,6*S*,7*R*,8*aR*)-1-Benzyl-3-(3-(benzyloxy)propyl)-6-(dimethyl(phenyl)silyl)-7-((*R*)-1-hydroxyethyl)-3*a*-methyl-octahydrocyclohepta[*b*]pyrrol-4(5*H*)-one (S-9).

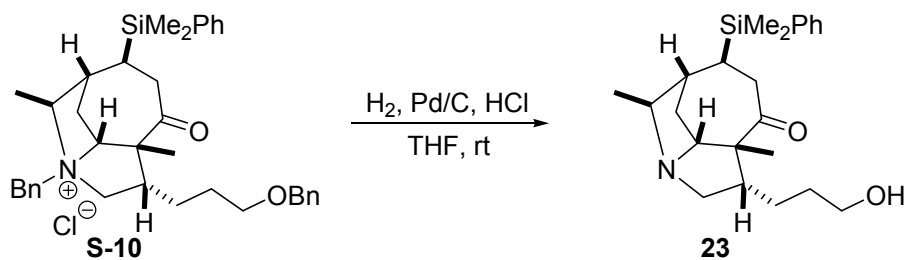
Hydrogen fluoride (0.5 mL, 48% aq) was added to a solution of silyl ether **22b** (0.162 g, 0.197 mmol) in MeCN (2.0 mL) in a Nalgene vial. The solution was heated at 50 °C for 19 h. Additional hydrogen fluoride (0.5 mL, 48% aq) was added and the reaction was maintained at 50 °C for another 7 h. The solution was then cooled to rt and poured into a stirring, biphasic mixture of Et₂O (50 mL) and aq 1 N NaOH (50 mL). The mixture was stirred for 20 min at rt, and then the layers were separated. The aqueous layer was extracted with Et₂O (2 × 50 mL). The combined organic extracts were washed with water (10 mL), brine (30 mL), dried over Na₂SO₄, filtered, and concentrated in vacuo. The residue was purified by silica gel flash column chromatography (gradient: 1:19 acetone:hexanes to 1:9 acetone:hexanes) to give **S-9** (92 mg, 80%) as a colorless foam, along with recovered starting material **22b** (25 mg, 15%): $[\alpha]_{589}^{25} -43$, $[\alpha]_{577}^{25} -44$, $[\alpha]_{546}^{25} -50$, $[\alpha]_{435}^{25} -83$ (c 0.74, EtOH); ^1H NMR (500 MHz, CDCl_3) δ 7.55–7.53 (m, 2H), 7.37–7.27 (m, 13H), 7.00 (br s, 1H), 4.44 (s, 2H), 4.11 (d, $J = 13.0$ Hz, 1H), 3.93 (q,

$J = 6.0$ Hz, 1H), 3.54 (d, $J = 13.0$ Hz, 1H), 3.35 (t, $J = 6.0$ Hz, 2H), 3.15 (dd, $J = 11.0, 5.0$ Hz, 1H), 3.01 (dd, $J = 11.5, 9.0$ Hz, 1H), 2.86–2.81 (m, 2H), 2.54 (dd, $J = 11.0, 6.0$ Hz, 1H), 2.00–1.94 (m, 1H), 1.85 (br dd, $J = 15.0, 5.0$ Hz, 1H), 1.80–1.72 (m, 2H), 1.70–1.66 (m, 1H), 1.48–1.40 (m, 3H), 1.02 (s, 3H), 1.02–0.94 (m, 1H), 0.90 (d, $J = 6.0$ Hz, 3H), 0.40 (s, 3H), 0.36 (s, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 213.5 (C), 138.3 (C), 138.0 (C), 136.8 (C), 134.3 (CH), 129.6 (CH), 129.0 (CH), 128.5 (CH), 128.3 (CH), 127.62 (CH), 127.55 (CH), 127.54 (CH), 127.48 (CH), 73.2 (CH), 72.9 (CH_2), 71.6 (br, CH), 70.0 (CH_2), 61.8 (C), 60.8 (CH_2), 56.5 (CH_2), 47.3 (CH), 43.3 (CH_2), 42.1 (CH), 32.9 (br, CH_2), 29.0 (CH_2), 25.9 (CH_2), 24.0 (CH_3), 21.5 (CH), 20.7 (br, CH_3), –3.3 (CH_3), –4.2 (CH_3); IR (film) 3382, 3068, 3029, 2966, 2931, 2860, 1690, 1455, 1428, 1372, 1250, 1111, 1077 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{37}\text{H}_{50}\text{NO}_3\text{Si}$ [(M+H) $^+$]: 584.3560; found: 584.3555.



(3*S*,3*aR*,6*S*,7*R*,8*aR*,9*S*)-Octahydro-1-benzyl-3-(3-(benzyloxy)propyl)-3*a*,9-dimethyl-6-(dimethyl(phenyl)silyl)-1,7-methanocyclohepta[*b*]pyrrolium-4(5*H*)-one chloride (S-10). Triethylamine (45 μL , 0.32 mmol) and methanesulfonyl chloride (16 μL , 0.21 mmol) were added sequentially to a solution of alcohol S-9 (0.110 g, 0.188 mmol) in CH_2Cl_2 (1.9 mL) at 0 $^\circ\text{C}$. The solution was stirred for 30 min at 0 $^\circ\text{C}$, then concentrated in vacuo to give S-10, which was used in the subsequent reaction without further purification: ^1H NMR (600 MHz, CDCl_3) δ 7.71 (d, $J = 7.0$ Hz, 2H), 7.52–7.50 (m, 2H), 7.41–7.23 (m, 11H), 5.10 (d, $J = 12.5$ Hz, 1H), 4.91 (d, $J = 12.5$ Hz, 1H), 4.47 (dd, $J = 13.0, 9.0$ Hz, 1H), 4.38 (s, 2H), 3.55 (d, $J = 7.0$ Hz, 1H), 3.33–3.28 (m, 4H), 3.08 (ddd, $J = 16.0, 9.0, 7.0$ Hz, 1H), 2.83 (dd, $J = 9.0, 5.0$ Hz, 1H), 2.58 (t, $J = 12.0$ Hz, 1H), 2.25 (dd, $J = 12.0, 5.0$ Hz, 1H), 1.96–1.90 (m, 1H), 1.72 (br d, $J = 16.0$ Hz, 1H), 1.70 (d, $J = 7.0$ Hz, 3H), 1.66–1.60 (m, 1H), 1.49–1.43 (m, 1H), 1.25–1.18 (m, 1H), 1.09 (dd, $J = 12.0, 5.0$ Hz, 1H), 0.98–0.92 (m, 1H), 0.96 (s, 3H), 0.39 (s, 3H), 0.37 (s, 3H); ^{13}C NMR

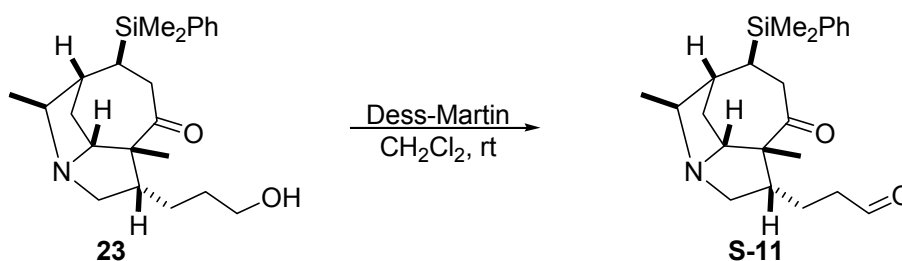
(125 MHz, CDCl₃) δ 212.9 (C), 138.2 (C), 134.9 (C), 133.9 (CH), 133.0 (CH), 130.4 (C), 129.9 (CH), 129.2 (CH), 128.36 (CH), 128.30 (CH), 128.27 (CH), 127.7 (CH), 127.6 (CH), 87.7 (CH), 79.6 (CH), 72.9 (CH₂), 69.5 (CH₂), 66.2 (CH₂), 58.4 (CH₂), 56.4 (C), 49.6 (CH), 42.6 (CH), 37.7 (CH₂), 30.4 (CH), 29.3 (CH₂), 24.2 (CH₂), 23.3 (CH₂), 19.8 (CH₃), 14.9 (CH₃), -4.8 (CH₃), -4.9 (CH₃); HRMS (ESI) m/z calcd for C₃₇H₄₈NO₂Si [M⁺]: 566.3455; found: 566.3438.



(3*S*,3*aR*,6*S*,7*R*,8*aR*,9*S*)-Octahydro-3-(3-hydroxypropyl)-3*a*,9-dimethyl-6-(dimethyl(phenyl)silyl)-1,7-methanocyclohepta[*b*]pyrrol-4(5*H*)-one (**23**).

Concentrated HCl (47 μ L, 0.57 mmol, 12.1 N) was added to a solution of benzylammonium salt **S-10** (0.113 g, 0.188 mmol) in THF (4 mL). Palladium on carbon (0.040 g, 0.038 mmol) was then added and the flask was purged thoroughly with hydrogen. The suspension was stirred at rt under an atmosphere of hydrogen (balloon pressure) for 15 h. The reaction mixture was then filtered through celite, rinsing with methanol (50 mL). The filtrate was concentrated in vacuo and the resultant oil was dissolved in CH₂Cl₂ (30 mL) and stirred vigorously with saturated aqueous sodium carbonate (15 mL) for 20 min. The layers were separated and the aqueous layer was extracted with a 1:9 mixture of MeOH:CH₂Cl₂ (3 \times 30 mL). The combined organic extracts were dried over Na₂SO₄, filtered, and concentrated in vacuo. The residue was purified by silica gel flash column chromatography (gradient: 1:5:95 NH₄OH:MeOH:CH₂Cl₂ to 1:10:90 NH₄OH:MeOH:CH₂Cl₂) to give **23** (0.064 g, 88% over two steps) as a colorless oil: $[\alpha]_D^{23}$ ₅₈₉ +30, $[\alpha]_D^{23}$ ₅₇₇ +31, $[\alpha]_D^{23}$ ₅₄₆ +40, $[\alpha]_D^{23}$ ₄₃₅ +130 (c 0.83, CH₂Cl₂); ¹H NMR (600 MHz, CDCl₃) δ 7.56–7.54 (m, 2H), 7.38–7.37 (m, 3H), 3.71 (dd, J = 10.5, 9.0 Hz, 1H), 3.62–3.55 (m, 2H), 3.36 (br d, J = 8.0 Hz, 1H), 2.80 (appt t, J = 10.5 Hz, 1H), 2.68 (appt q, J = 6.5 Hz, 1H), 2.53 (dd, J = 12.0, 4.0 Hz, 1H), 2.40 (dd, J = 12.0, 7.0 Hz, 1H), 2.22–2.14 (m, 2H), 2.09 (br d, J = 6.0 Hz, 1H), 1.7–1.5 (br s,

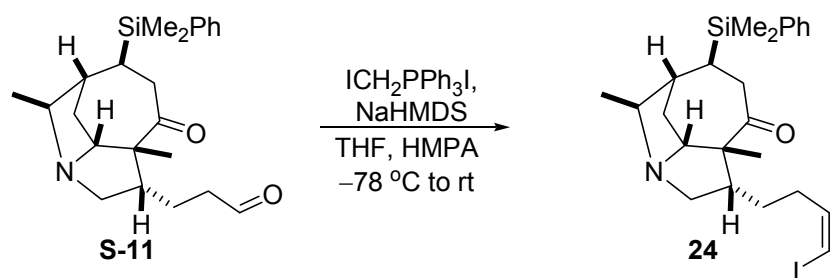
1H), 1.57–1.43 (m, 4H), 1.32–1.26 (m, 1H), 1.24–1.22 (m, 1H), 1.10 (d, $J = 6.5$ Hz, 3H), 1.03 (s, 3H), 0.36 (s, 3H), 0.35 (s, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 214.0 (C), 137.9 (C), 134.0 (CH), 129.1 (CH), 127.8 (CH), 75.0 (CH), 73.9 (CH), 62.8 (CH_2), 60.9 (CH_2), 60.3 (C), 52.7 (CH), 44.7 (CH), 41.3 (CH_2), 32.1 (CH_2), 31.9 (CH), 29.8 (CH_2), 24.8 (CH_2 and CH_3), 21.1 (CH_3), -3.0 (CH_3), -3.2 (CH_3), missing carbon attributed to accidental equivalence; IR (film) 3400, 3070, 3049, 2956, 2929, 2865, 1684, 1457, 1387, 1374, 1320, 1250, 1111, 1081, 1063 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{23}\text{H}_{36}\text{NO}_2\text{Si}$ $[(\text{M}+\text{H})^+]$: 386.2515; found: 386.2502.



3-((3*S*,3*aR*,6*S*,7*R*,8*aR*,9*S*)-Octahydro-3*a*,9-dimethyl-6-(dimethyl(phenyl)silyl)-4(5*H*)-oxo-1,7-methanocyclohepta[*b*]pyrrol-3-yl)propanal (S-11). Dess–Martin periodinane¹² (0.048 g, 0.11 mmol) was added to a solution of alcohol **23** (0.022 g, 0.057 mmol) in water-saturated CH_2Cl_2 (1 mL). The resultant suspension was stirred at rt for 30 min. A solution of 1:9 MeOH: CH_2Cl_2 (2 mL), followed by a solution of sodium thiosulfate (0.28 g) in 80% saturated aqueous sodium bicarbonate (2 mL) were added. The layers were separated and the aqueous portion was extracted with a 1:9 mixture of MeOH: CH_2Cl_2 (3 x 3 mL). The combined organic extracts were dried over Na_2SO_4 , filtered, and concentrated in vacuo. The residue was purified by silica gel flash column chromatography (eluted with acetone) to give aldehyde **S-11** (0.018 g, 82%) as a colorless oil: $[\alpha]^{24}_{589} +38$, $[\alpha]^{24}_{577} +40$, $[\alpha]^{24}_{546} +50$, $[\alpha]^{24}_{435} +154$ (c 0.60, CH_2Cl_2); ^1H NMR (600 MHz, CDCl_3) δ 9.72 (s, 1H), 7.55–7.53 (m, 2H), 7.38–7.37 (m, 3H), 3.63 (dd, $J = 11.0, 9.0$ Hz, 1H), 3.32 (d, $J = 8.0$ Hz, 1H), 2.78 (appt t, $J = 11.0$ Hz, 1H), 2.64 (appt q, $J = 6.5$ Hz, 1H), 2.55 (dd, $J = 12.0, 4.5$ Hz, 1H), 2.41 (t, $J = 7.0$ Hz, 2H), 2.35 (dd, $J = 12.0, 7.0$ Hz, 1H), 2.18 (ddd, $J = 14.0, 8.0, 8.0$ Hz, 1H), 2.16–2.10 (m, 1H), 2.08 (br d, $J = 7.0$ Hz, 1H), 1.78–1.72 (m, 1H), 1.70–1.63 (m, 1H), 1.53 (d, $J = 14.0$ Hz, 1H), 1.26–

¹² Dess, D. B.; Martin, J. C. *J. Org. Chem.* **1983**, *48*, 4155–4156.

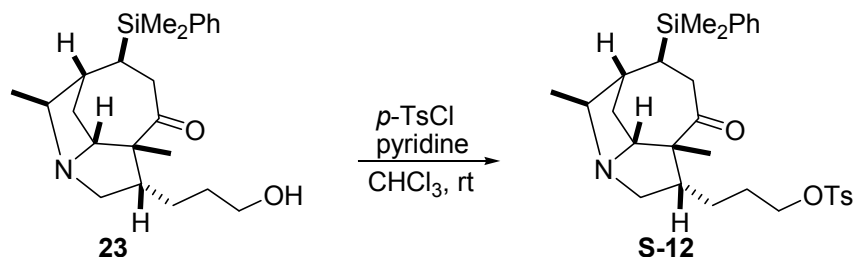
1.20 (m, 1H), 1.08 (d, $J = 6.5$ Hz, 3H), 1.05 (s, 3H), 0.355 (s, 3H), 0.346 (s, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 213.8 (C), 201.5 (C), 137.7 (C), 134.0 (CH), 129.1 (CH), 127.8 (CH), 75.2 (CH), 74.2 (CH), 60.5 (CH_2), 60.0 (C), 52.3 (CH), 44.6 (CH), 43.0 (CH_2), 41.0 (CH_2), 31.9 (CH), 29.5 (CH_2), 24.9 ($\text{CH}_3 + \text{CH}_2$), 20.9 (CH_3), -3.1 (CH_3), -3.2 (CH_3), missing carbon attributed to accidental equivalence; IR (film) 3070, 2956, 2929, 2865, 2721, 1725, 1683, 1457, 1445, 1428, 1389, 1372, 1320, 1250, 1221, 1150, 1111, 1081 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{23}\text{H}_{34}\text{NO}_2\text{Si}$ [(M+H) $^+$]: 384.2359; found: 384.2366.



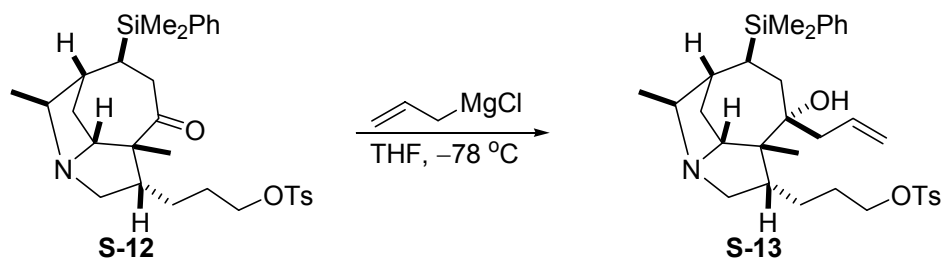
(3*S*,3*aR*,6*S*,7*R*,8*aR*,9*S*,*Z*)-Octahydro-3-(4-iodobut-3-enyl)-3*a*,9-dimethyl-6-

(dimethyl(phenyl)silyl)-1,7-methanocyclohepta[*b*]pyrrol-4(*5H*)-one (24). A solution of sodium hexamethyldisilazide (0.16 mL, 0.16 mmol, 1.0 M in THF) was added dropwise to a suspension of iodomethyltriphenylphosphonium iodide (0.104 g, 0.196 mmol) in THF (1.5 mL). The suspension was cooled to $-78\text{ }^\circ\text{C}$ and HMPA (0.3 mL) was added, followed by a solution of aldehyde **S-11** (0.030 g, 0.078 mmol) in THF (1.5 mL). The mixture was stirred for 15 min at $-78\text{ }^\circ\text{C}$ and then allowed to warm to rt over 30 min. Water (0.75 mL) was added and the mixture was extracted with Et_2O (4×5 mL). The combined organic extracts were dried over Na_2SO_4 , filtered, and concentrated in vacuo. The residue was purified by silica gel flash column chromatography (1:19 acetone:hexanes) to give **24** (0.024 g, 60%) as a pale yellow oil: $[\alpha]_{589}^{25} +3.5$, $[\alpha]_{577}^{25} +4.7$, $[\alpha]_{546}^{25} +5.8$, $[\alpha]_{435}^{25} +31$ ($c = 0.64$, CH_2Cl_2); ^1H NMR (500 MHz, CDCl_3) δ 7.55 (m, 2H), 7.38 (m, 3H), 6.20 (d, $J = 7.3$ Hz, 1H), 6.08 (dd, $J = 14.0, 6.9$ Hz, 1H), 3.75 (dd, $J = 8.8, 8.8$ Hz, 1H), 3.34 (d, $J = 8.0$ Hz, 1H), 2.82 (dd, $J = 10.6, 10.6$ Hz, 1H), 2.68 (dd, $J = 12.9, 6.3$ Hz, 1H), 2.53 (dd, $J = 11.9, 3.8$ Hz, 1H), 2.39 (dd, $J = 11.9, 7.2$ Hz, 1H), 2.25–2.01 (m, 5H), 1.59–1.48 (m, 2H), 1.36 (m, 1H), 1.24 (m, 1H), 1.10 (d, $J = 6.5$ Hz, 3H), 1.01 (s, 3H), -0.35 (s, 3H), -0.36 (s, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 214.0 (C),

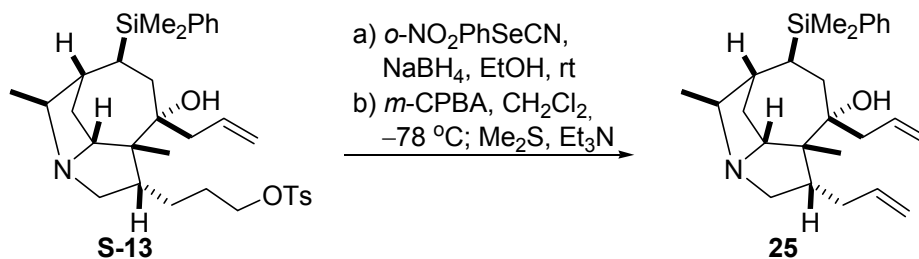
140.4 (CH), 137.9 (C), 134.1 (CH), 129.2 (CH), 127.9 (CH), 83.2 (CH), 75.0 (CH), 74.0 (CH), 60.9 (CH₂), 60.3 (C), 52.4 (CH), 44.8 (CH), 41.3 (CH₂), 34.1 (CH₂), 32.0 (CH), 29.8 (CH₂), 26.9 (CH₂), 25.0 (CH₃), 21.0 (CH₃), -2.9 (CH₃), -3.1 (CH₃); IR (film): 2931, 1684 cm⁻¹; HRMS (ESI) *m/z* calcd for C₂₄H₃₅INOSi [(M+H)⁺]: 508.1533; found: 508.1526.



Tosylate S-12. Pyridine (17 μ L, 0.22 mmol) and *p*-toluenesulfonyl chloride (0.021 g, 0.11 mmol) were added sequentially to a solution of alcohol **23** (0.021 g, 0.054 mmol) in chloroform (0.5 mL) at rt. The resultant red solution was stirred for 17 h at rt. Saturated aqueous NaHCO₃ (1 mL) was added and the mixture was extracted with CH₂Cl₂ (4 x 2 mL). The combined organic extracts were dried over Na₂SO₄, filtered, and concentrated in vacuo. The residue was dissolved in heptane (3 mL) and concentrated in vacuo (x 2) to remove pyridine by azeotropic distillation. The residue was purified by silica gel flash column chromatography (1:10:90 NH₄OH (aq):MeOH:CH₂Cl₂) to give **S-12** (0.025 g, 84%) as a pink oil: $[\alpha]^{24}_{589} +24$, $[\alpha]^{24}_{577} +25$, $[\alpha]^{24}_{546} +33$, $[\alpha]^{24}_{435} +91$ ($c = 0.99$, CH₂Cl₂); ¹H NMR (500 MHz, CDCl₃) δ 7.76 (d, $J = 8.3$, 2H), 7.51 (m, 2H), 7.42–7.35 (m, 3H), 7.33 (d, $J = 8.2$, 2H), 3.96 (dd, $J = 11.3, 4.7$, 2H), 3.75 (dd, $J = 9.4, 9.4$, 1H), 3.52 (br s, 1H), 2.76–2.67 (m, 2H), 2.52 (dd, $J = 11.9, 5.6$, 1H), 2.44 (s, 3H), 2.28 (dd, $J = 12.0, 6.6$, 1H), 2.22 (m, 1H), 2.16 (d, $J = 7.7$, 1H), 2.07 (ddd, $J = 16.1, 8.6, 8.6$, 1H), 1.64–1.51 (m, 3H), 1.51–1.40 (m, 2H), 1.2 (m, 1H), 1.16 (d, $J = 6.5$ Hz, 3H), 1.03 (s, 3H), 0.35 (s, 3H), 0.34 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 213.3 (C), 144.9 (C), 134.0 (CH), 133.0 (C), 130.03 (CH), 129.95 (CH), 129.5 (C), 128.0 (CH), 127.9 (CH), 75.3 (CH₂), 70.1 (CH₂), 59.7 (CH₂), 59.1 (C), 51.6 (CH), 44.6 (CH), 40.2 (CH₂), 31.7 (CH₃), 28.8 (CH), 28.4 (CH₂), 24.1 (CH₂), 23.2 (CH), 21.7 (CH₃), 20.3 (CH₃), -3.4 (CH₃), -3.7 (CH₃), missing carbon attributed to accidental equivalence; IR (film): 2957, 1685 cm⁻¹; HRMS (ESI) *m/z* calcd for C₃₀H₄₂NO₄SSi [(M+H)⁺]: 540.2604; found: 540.2606.

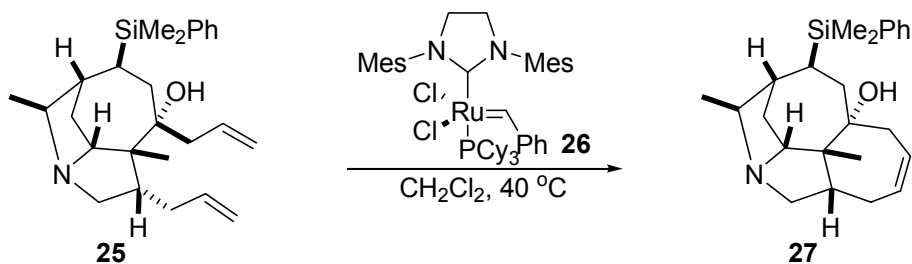


Allyl addition product S-13. A solution of allylmagnesium chloride (87 μL , 0.17 mmol, 2.0 M in THF) was added to a solution of tosylate **S-12** (0.023 g, 0.043 mmol) in THF (1.5 mL) at $-78\text{ }^\circ\text{C}$. After 15 min at $-78\text{ }^\circ\text{C}$, additional allylmagnesium chloride solution (43 μL , 0.086 mmol, 2.0 M in THF) was added. The reaction was maintained 20 min at $-78\text{ }^\circ\text{C}$, and then water (1 mL) was added and the mixture was allowed to warm to rt. The mixture was extracted with Et_2O (5 x 3 mL). The combined extracts were dried over Na_2SO_4 , filtered, and concentrated in vacuo. The residue was purified by silica gel flash column chromatography (gradient: 1:10:90 NH_4OH (aq): MeOH : CH_2Cl_2 to 2:20:80 NH_4OH (aq): MeOH : CH_2Cl_2) to give **S-13** (0.020 g, 78%) as a colorless oil: $[\alpha]_{589}^{24} -20$, $[\alpha]_{577}^{24} -10$, $[\alpha]_{546}^{24} +3.4$, $[\alpha]_{435}^{24} +11$ ($c = 0.47$, CH_2Cl_2); ^1H NMR (500 MHz, CDCl_3) δ 7.78 (d, $J = 8.2$ Hz, 2H), 7.46 (d, $J = 5.5$ Hz, 2H), 7.43–7.32 (m, 5H), 5.59 (dddd, $J = 16.8, 9.0, 9.0, 9.0$ Hz, 1H), 5.31 (d, $J = 9.9$ Hz, 1H), 5.16 (d, $J = 17.1$ Hz, 1H), 4.03 (dd, $J = 11.9, 5.9$ Hz, 2H), 3.59–3.39 (m, 3H), 3.00 (m, 1H), 2.48 (m, 1H), 2.45 (s, 3H), 2.13–1.99 (m, 3H), 1.98–1.79 (m, 3H), 1.70–1.40 (m, 5H), 1.33–1.12 (m, 4H), 0.93 (s, 3H), 0.29 (s, 3H), 0.28 (s, 3H), missing H attributed to deuterium exchange with solvent; ^{13}C NMR (125 MHz, CDCl_3): δ 144.9 (C), 133.8 (CH), 133.1 (C), 132.1 (CH), 130.0 (CH), 129.4 (CH), 128.0 (CH), 127.9 (CH), 126.0 (C), 123.5 (CH_2), 80.1 (CH), 76.7 (C), 75.6 (CH), 70.6 (CH_2), 58.0 (CH_2), 53.4 (CH), 49.3 (C), 44.2 (CH_2), 43.6 (CH), 31.9 (CH_2), 29.8 (CH), 28.6 (CH_2), 27.4 (CH_3), 26.4 (CH_2), 25.1 (CH_2), 21.7 (CH_3), 21.4 (CH_3), -4.5 (CH_3), -4.7 (CH_3); IR (film): 3417, 2954 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{33}\text{H}_{48}\text{NO}_4\text{SSi}$ $[(\text{M}+\text{H})^+]$: 582.3073; found: 582.3083.

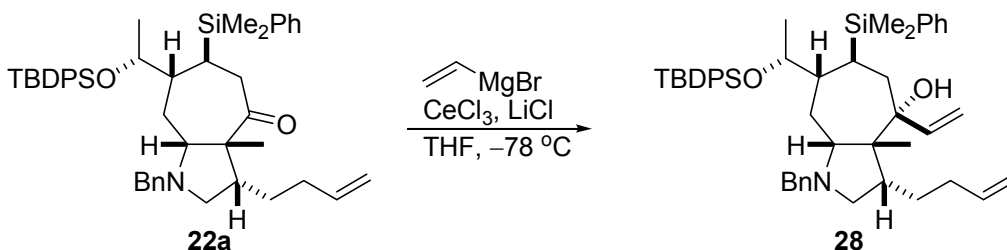


Diene 25. Sodium borohydride (4.7 mg, 0.12 mmol) was added to a suspension of 2-nitrophenyl selenocyanate (0.028 g, 0.12 mmol) in ethanol (0.5 mL) at 0 °C. After 10 min at 0 °C, a solution of tosylate **S-13** (0.024 g, 0.041 mmol) in ethanol (1 mL) was added and the suspension was allowed to warm to rt. The mixture was stirred at rt for 14 h, and then water (1 mL) was added to the deep red solution. The mixture was extracted with CH₂Cl₂ (4 x 3 mL) and the combined extracts were dried over Na₂SO₄, filtered, and concentrated in vacuo. The residue was purified by silica gel flash column chromatography (1:10:90 NH₄OH (aq):MeOH:CH₂Cl₂) to give the selenide (0.018 g, 69%) as a yellow oil, which was immediately used in the subsequent elimination step. The selenide (8.5 mg, 0.014 mmol) was dissolved in CH₂Cl₂ (0.7 mL) and the resultant yellow solution was cooled to -78 °C. A solution of *m*-chloroperoxybenzoic acid (3.8 mg, 0.017 mmol) in CH₂Cl₂ (0.2 mL) was added dropwise by syringe. The solution was maintained for 1.5 h at -78 °C, then dimethyl sulfide (51 μL, 0.70 mmol) was added, followed by triethylamine (48 μL, 0.35 mmol). The solution was allowed to warm to rt and stirred at rt for 17 h. Saturated aqueous sodium bicarbonate (1 mL) was added, and the mixture was extracted with CH₂Cl₂ (4 x 3 mL). The combined organic extracts were dried over Na₂SO₄, filtered, and concentrated in vacuo. The residue was purified by silica gel flash column chromatography (1:10:90 NH₄OH (aq):MeOH:CH₂Cl₂) to give diene **25** (4.6 mg, 81%) as a colorless oil: $[\alpha]_D^{24}$ ₅₈₉ +4.6, $[\alpha]_D^{24}$ ₅₇₇ +4.9, $[\alpha]_D^{24}$ ₅₄₆ +4.7, $[\alpha]_D^{24}$ ₄₃₅ +7.5 (*c* = 0.53, CH₂Cl₂); ¹H NMR (500 MHz, CDCl₃) δ 7.48 (d, *J* = 3.9 Hz, 2H), 7.36 (m, 3H), 5.74 (m, 1H), 5.63 (m, 1H), 5.25 (d, *J* = 9.9 Hz, 1H), 5.12 (d, *J* = 17.1 Hz, 1H), 5.05 (d, *J* = 17.0 Hz, 1H), 4.96 (d, *J* = 10.1 Hz, 1H), 3.37 (m, 1H), 3.28–3.15 (m, 2H), 2.91 (dd, *J* = 10.4 Hz, 10.4, 1H), 2.59–2.50 (m, 2H), 2.09 (dd, *J* = 13.0, 6.4 Hz, 1H), 2.01–1.90 (m, 3H), 1.64–1.53 (m, 2H), 1.46 (m, 1H), 1.37 (m, 1H), 1.28–1.23 (m, 2H), 1.09 (d, *J* = 15.7 Hz, 3H), 0.92 (s, 3H), 0.29 (s, 3H), 0.27 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 138.4 (CH), 137.8 (C), 133.8 (CH), 133.0 (CH), 129.1 (CH), 127.9 (CH), 122.4 (CH₂), 115.4

(CH₂), 80.8 (CH), 77.3 (C), 75.1 (CH), 59.1 (CH₂), 54.0 (CH), 48.9 (C), 44.5 (CH₂), 43.4 (CH), 33.9 (CH₂), 31.8 (CH₂), 27.6 (CH), 26.8 (CH₂), 23.8 (CH₃), 21.6 (CH₃), -4.4 (CH₃), -4.6 (CH₃); IR (film): 3320, 2954 cm⁻¹; HRMS (ESI) *m/z* calcd for C₂₆H₄₀NOSi [(M+H)⁺]: 410.2879; found: 410.2877.

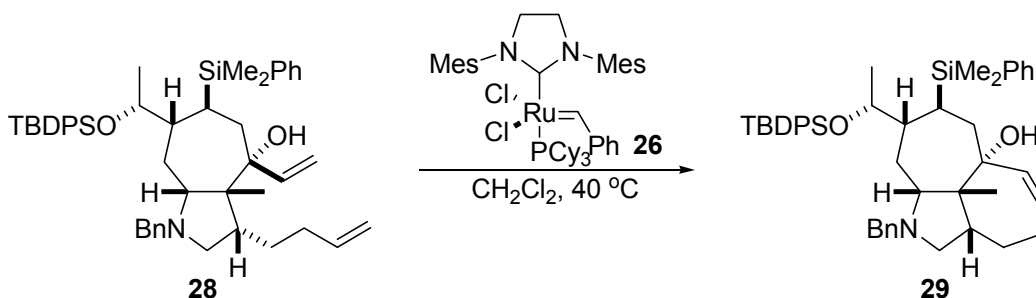


Tetracyclic alcohol 27. The Grubbs second generation catalyst (**26**) (benzylidene[1,3-bis(2,4,6-trimethylphenyl)-2-imidazolidinylidene]dichloro(tricyclohexylphosphine)ruthenium) (3.5 mg, 4.1 μmol) was added to a solution of diene **25** (0.017 g, 0.041 mmol) in CH₂Cl₂ (1.5 mL), and the resultant brown solution was heated at reflux for 3 h. The solution was allowed to cool to rt and then concentrated in vacuo. The residue was purified by silica gel flash column chromatography (1:10:90 NH₄OH (aq):MeOH:CH₂Cl₂) to give **27** (4.6 mg, 81%) as a purple oil: ¹H NMR (500 MHz, CDCl₃) δ 7.45 (m, 2H), 7.38 (m, 3H), 5.67 (m, 1H), 5.47 (m, 1H), 4.09 (d, *J* = 6.7 Hz, 1H), 3.99 (dd, *J* = 9.2, 9.2 Hz, 1H), 3.71 (s, 1H), 3.10 (dd, *J* = 10.3, 10.3 Hz, 1H), 2.60 (s, 1H), 2.52 (d, *J* = 19.5 Hz, 1H), 2.41–2.27 (m, 2H), 2.21–2.13 (m, 2H), 2.05 (d, *J* = 19.8 Hz, 1H), 1.97 (s, 1H), 1.84 (dd, *J* = 14.1, 14.1 Hz, 1H), 1.59 (d, *J* = 13.2 Hz, 1H), 1.53 (dd, *J* = 13.1, 3.6 Hz, 1H), 1.39 (d, *J* = 7.0 Hz, 3H), 1.37 (dd, *J* = 15.0, 3.5 Hz, 1H), 1.14 (s, 3H), 0.32 (s, 3H), 0.31 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 136.7 (C), 133.8 (CH), 129.6 (CH), 128.1 (CH), 128.0 (CH), 125.0 (CH), 79.0 (CH), 78.0 (C), 76.4 (CH), 60.1 (CH₂), 49.4 (C), 48.3 (CH), 47.5 (CH₂), 44.2 (CH), 35.9 (CH₂), 28.8 (CH₂), 27.1 (CH), 25.8 (CH₂), 24.7 (CH₃), 19.5 (CH₃), -4.2 (CH₃), -4.9 (CH₃); IR (film): 3320, 2954 cm⁻¹; HRMS (ESI) *m/z* calcd for C₂₄H₃₆NOSi [(M+H)⁺]: 382.2566; found: 382.2563.



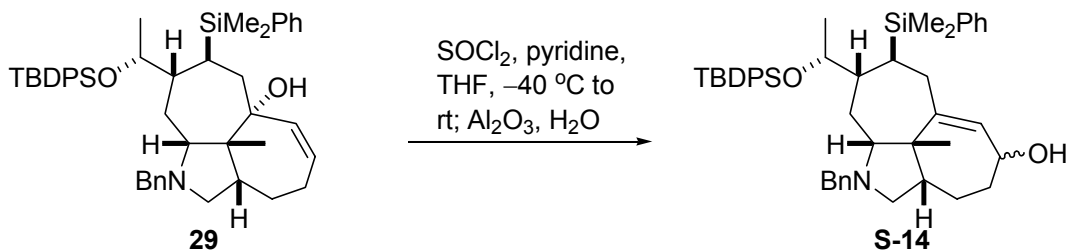
(3*S*,3*aR*,4*R*,6*S*,7*R*,8*aR*)-1-Benzyl-3-(but-3-enyl)-7-((*R*)-1-(*tert*-butyldiphenylsilyloxy)ethyl)-6-(dimethyl(phenyl)silyl)-3*a*-methyl-4-vinyl-decahydrocyclohepta[*b*]pyrrol-4-ol (28**).** Lithium chloride (0.199 g, 4.70 mmol) and anhydrous cerium (III) trichloride (0.579 g, 2.35 mmol) were added to a round-bottomed flask in a glove box. The flask was removed from the glove box, THF (3 mL) was added, and the suspension was stirred vigorously for 2.5 h at rt, producing a near colorless solution. This solution was added to ketone **22a** (0.171 g, 0.235 mmol) by cannula with a THF (2 mL) rinse. The solution was stirred for 2.5 h at rt, then cooled to $-78\text{ }^{\circ}\text{C}$. A solution of vinylmagnesium bromide (1.7 mL, 2.4 mmol, 1.4 M in THF) was quickly added. After 20 min at $-78\text{ }^{\circ}\text{C}$, methanol (2 mL) was added. The cold bath was removed, and 10% aqueous AcOH (6 mL) and Et₂O (20 mL) were added sequentially. After the solution had warmed to rt, the layers were separated. The aqueous layer was extracted with Et₂O (30 mL). The combined organic layers were washed sequentially with saturated aqueous sodium bicarbonate (30 mL) and brine (30 mL). The organic portion was dried over Na₂SO₄, filtered, and concentrated in vacuo. The residue was purified by silica gel flash column chromatography (gradient: 1:19 acetone:hexanes to 1:9 acetone:hexanes) to give **28** (0.129 g, 72%) as a colorless foam: $[\alpha]_{589}^{23} -14$, $[\alpha]_{577}^{23} -16$, $[\alpha]_{546}^{23} -17$, $[\alpha]_{435}^{23} -32$ ($c = 1.2$, CH₂Cl₂); ¹H NMR (500 MHz, CDCl₃) δ 7.67 (d, $J = 7.8$ Hz, 2H), 7.61 (d, $J = 7.2$ Hz, 2H), 7.49–7.44 (m, 2H), 7.44–7.30 (m, 9H), 7.22 (m, 1H), 7.11 (m, 4H), 6.04 (dd, $J = 17.0, 10.7$ Hz, 1H), 5.73 (m, 1H), 5.24 (d, $J = 16.9$ Hz, 1H), 4.94 (d, $J = 10.2$ Hz, 1H), 4.92 (d, $J = 9.1$ Hz, 1H), 4.86 (d, $J = 10.7$ Hz, 1H), 4.24 (d, $J = 13.0$ Hz, 1H), 3.94 (dd, $J = 5.6, 3.8$ Hz, 1H), 3.27 (d, $J = 13.3$ Hz, 1H), 2.84 (dd, $J = 13.0, 7.9$ Hz, 1H), 2.65 (m, 2H), 2.51 (dd, $J = 9.1, 9.1$ Hz, 1H), 2.14 (dd, $J = 8.2, 8.2$ Hz, 1H), 1.94 (m, 2H), 1.78–1.55 (m, 6H), 1.35 (ddd, $J = 12.9, 12.9, 4.3$ Hz, 1H), 1.17 (s, 3H), 1.04 (s, 9H), 0.94 (m, 1H), 0.52 (d, $J = 5.9$ Hz, 3H), 0.01 (s, 3H), -0.06 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 146.3 (CH), 139.7 (C), 138.8 (C), 138.7 (CH), 135.9 (CH), 135.9

(CH), 134.6 (C), 134.5 (C), 133.6 (CH), 129.6 (CH), 129.5 (CH), 128.5 (CH), 128.4 (CH), 128.3 (CH), 127.6 (CH), 127.5 (CH), 127.0 (CH), 114.5 (CH₂), 109.8 (CH₂), 80.5 (C), 73.2 (CH), 71.5 (CH), 57.3 (CH₂), 56.4 (CH₂), 51.5 (CH), 50.5 (CH), 42.1 (CH₂), 40.9 (CH), 36.6 (C), 33.9 (CH₂), 30.9 (CH₂), 28.4 (CH₂), 27.0 (CH₃), 25.5 (CH₃), 23.3 (CH), 19.2 (C), 16.1 (CH₃), -1.4 (CH₃), -4.9 (CH₃); IR (film): 3270, 2930, 2359 cm⁻¹; HRMS (ESI) *m/z* calcd for C₄₉H₆₆NO₂Si₂ [(M+H)⁺]: 756.4632; found: 756.4639.

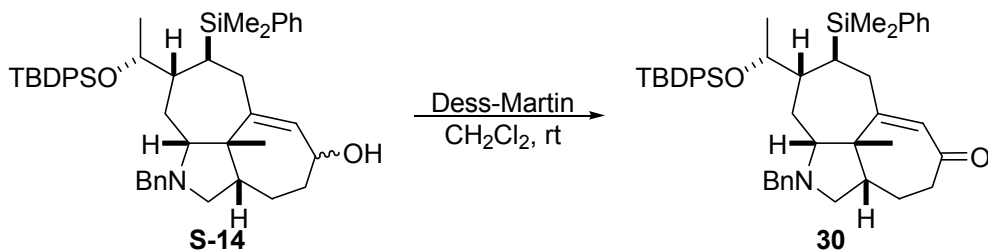


Alcohol 29. The Grubbs second generation catalyst (**26**) (benzylidene[1,3-bis(2,4,6-trimethylphenyl)-2-imidazolidinylidene] dichloro(tricyclohexylphosphine)ruthenium) (8.5 mg, 10 μ mol) was added to a solution of diene **28** (0.151 g, 0.200 mmol) in CH₂Cl₂ (20 mL). The resultant brown solution was heated at reflux for 2.5 h. The solution was then cooled to rt and concentrated in vacuo. The residue was purified by silica gel flash column chromatography (gradient: 1:19 acetone:hexanes to 1:9 acetone:hexanes) to give **29** (0.139 g, 96%) as a brown oil: $[\alpha]_{589}^{23}$ -22, $[\alpha]_{577}^{24}$ -23, $[\alpha]_{546}^{24}$ -26 (*c* = 0.84, CH₂Cl₂); ¹H NMR (500 MHz, CDCl₃) δ 7.68 (d, *J* = 6.8 Hz, 2H), 7.62 (d, *J* = 6.8 Hz, 2H), 7.48–7.35 (m, 8H), 7.30 (dd, *J* = 7.4, 7.4 Hz, 2H), 7.26–7.21 (m, 2H), 7.17 (m, 4H), 5.54 (m, 1H), 5.37 (d, *J* = 11.8 Hz, 1H), 4.24 (d, *J* = 13.3 Hz, 1H), 3.97 (dd, *J* = 5.8, 3.3 Hz, 1H), 2.99 (d, *J* = 13.3 Hz, 1H), 2.94 (m, 1H), 2.68 (dd, *J* = 12.4, 5.1 Hz, 1H), 2.59 (d, *J* = 8.9 Hz, 1H), 2.42–2.29 (m, 2H), 2.21–2.03 (m, 2H), 1.89–1.71 (m, 3H), 1.68–1.62 (m, 1H), 1.58 (dd, *J* = 15.0, 4.1 Hz, 1H), 1.49 (m, 1H), 1.35–1.25 (m, 2H), 1.08 (s, 3H), 1.05 (s, 9H), 0.61 (d, *J* = 5.9 Hz, 3H), 0.02 (s, 3H), -0.04 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 140.4 (CH), 140.2 (C), 139.9 (C), 136.1 (CH), 136.0 (CH), 134.7 (C), 134.6 (C), 133.6 (CH), 131.4 (CH), 129.61 (CH), 129.57 (CH), 128.5 (CH), 128.4 (CH), 128.2 (CH), 127.7 (CH), 127.6 (CH), 126.6 (CH), 79.1 (C), 77.6 (CH), 71.7 (CH), 61.0 (CH₂), 58.4 (CH₂), 54.2 (C), 50.2 (CH), 43.3 (CH₂), 42.8 (CH), 32.0 (CH₂), 31.1 (CH₃), 30.6

(CH₂), 29.4 (CH₂), 27.2 (CH), 27.1 (CH₃), 19.3 (C), 16.5 (CH₃), -2.3 (CH₃), -3.8 (CH₃), missing carbon attributed to accidental equivalence; IR (film): 3440, 2929 cm⁻¹; HRMS (ESI) *m/z* calcd for C₄₇H₆₂NO₂Si₂ [(M+H)⁺]: 728.4319; found: 728.4333.

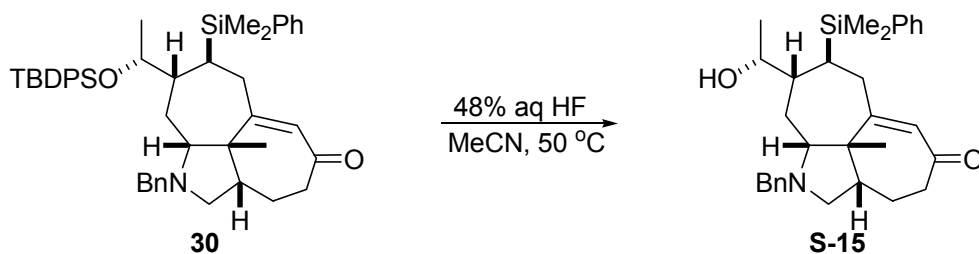


Alcohol S-14. Pyridine (34 μ L, 0.42 mmol) and thionyl chloride (24 μ L, 0.34 mmol) were added sequentially to a solution of allylic alcohol **29** (0.122 g, 0.168 mmol) in THF (4 mL) at -40 °C. After 30 min at -40 °C, aluminum oxide (2 g), water (2 mL), and THF (6 mL) were added, and the mixture was allowed to warm to rt. The suspension was stirred vigorously for 18 h at rt, then filtered through celite, washing with CH₂Cl₂ (50 mL). The filtrate was concentrated in vacuo and the residue was purified by silica gel flash column chromatography (gradient: 1:19 acetone:hexanes to 1:9 acetone:hexanes) to give alcohol **S-14** (0.081 g, 66%) as a colorless foam and a 1.1:1 mixture of diastereomers. This compound was not characterized because it was obtained as a mixture. A copy of the ¹H NMR spectrum of the mixture is included in the spectral section for reference purposes. Satisfactory low-resolution mass spectral data were obtained: LRMS (ESI) *m/z* for C₄₇H₆₂NO₂Si₂ [(M+H)⁺]: 728.4.



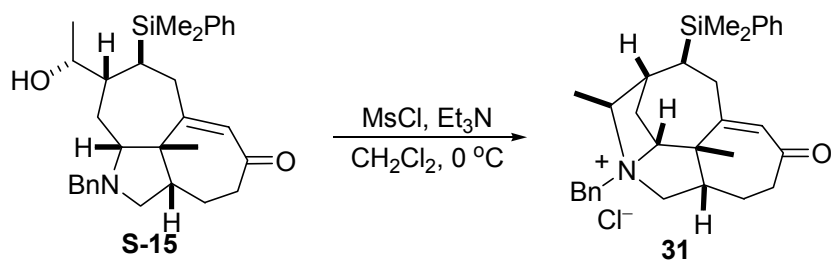
Enone 30. Pyridine (19 μ L, 0.24 mmol) and Dess–Martin periodinane (44 mg, 0.10 mmol) were added sequentially to a solution of alcohols **S-14** (0.069 g, 0.095 mmol) in water-saturated CH₂Cl₂ (5 mL). The resultant suspension was stirred for 1 h at rt. A solution of sodium thiosulfate (4 g) in 80% saturated aqueous sodium bicarbonate (20

mL) was then added. The layers were separated, and the aqueous portion was extracted with CH_2Cl_2 (3 x 30 mL). The combined organic extracts were dried over Na_2SO_4 , filtered, and concentrated in vacuo. The residue was purified by silica gel flash column chromatography (gradient: 1:19 acetone:hexanes to 1:9 acetone:hexanes) to give **30** (0.047 g, 68%) as a colorless oil: $[\alpha]^{23}_{589} +13$, $[\alpha]^{23}_{577} +15$, $[\alpha]^{23}_{546} +19$, $[\alpha]^{23}_{435} +65$ ($c = 0.70$, CH_2Cl_2); ^1H NMR (500 MHz, CDCl_3) δ 7.62 (d, $J = 7.5$ Hz, 2H), 7.59 (d, $J = 7.7$ Hz, 2H), 7.46–7.16 (m, 16H), 5.69 (s, 1H), 3.96 (m, 1H), 3.82 (d, $J = 13.8$ Hz, 1H), 3.72 (d, $J = 13.6$ Hz, 1H), 3.09 (dd, $J = 8.1, 8.1$ Hz, 1H), 2.79 (d, $J = 10.0$ Hz, 1H), 2.67–2.59 (m, 2H), 2.50 (dd, $J = 8.9, 5.0$ Hz, 1H), 2.44 (m, 1H), 2.30 (d, $J = 12.8$ Hz, 1H), 2.12 (m, 1H), 2.02 (m, 2H), 1.72 (dd, $J = 9.6, 9.6$ Hz, 1H), 1.64 (d, $J = 13.9$ Hz, 1H), 1.56 (br s, 1H), 1.34 (s, 3H), 1.04 (s, 9H), 0.86 (dd, $J = 11.7, 11.7$ Hz, 1H), 0.73 (d, $J = 16.1$ Hz, 3H), 0.05 (s, 3H), 0.00 (s, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 204.8 (C), 166.9 (C), 139.6 (C), 138.5 (C), 135.94 (CH), 125.86 (CH), 134.5 (C), 134.3 (C), 133.4 (CH), 129.63 (CH), 129.59 (CH), 129.1 (CH), 128.9 (CH), 128.3 (CH), 128.2 (CH), 127.9 (CH), 127.6 (CH), 127.5 (CH), 126.7 (CH), 73.5 (CH), 70.6 (CH), 58.2 (CH_2), 56.3 (C), 55.9 (CH_2), 49.5 (CH), 49.1 (CH), 43.0 (CH_2), 39.2 (CH_2), 35.2 (CH_2), 32.9 (CH_2), 29.4 (CH_2), 27.4 (CH), 27.0 (CH_3), 19.1 (C), 16.9 (CH_3), -3.3 (CH_3), -3.4 (CH_3); IR (film) 2927, 1651 cm^{-1} ; HRMS (ESI) m/z calcd for $\text{C}_{47}\text{H}_{60}\text{NO}_2\text{Si}_2$ $[(\text{M}+\text{H})^+]$: 726.4163; found: 726.4157.



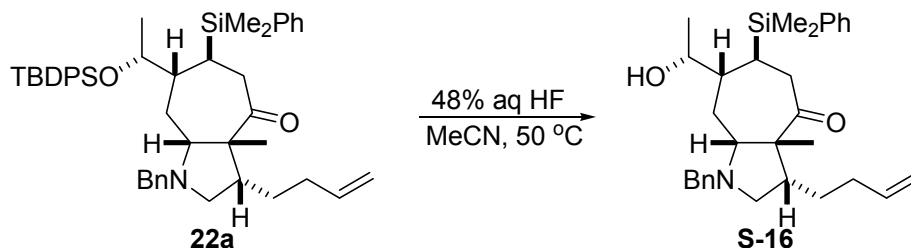
Alcohol S-15. Hydrogen fluoride (0.4 mL, 48% aq) was added to a solution of silyl ether **30** (0.021 g, 0.028 mmol) in MeCN (1.6 mL) in a Nalgene vial and the solution was heated at 50 °C for 17.5 h. Additional hydrogen fluoride (0.4 mL, 48% aq) was added and the solution was maintained for another 9 h at 50 °C. The solution was then cooled to rt and poured into a stirring, biphasic mixture of Et₂O (70 mL) and aqueous 1 N NaOH solution (60 mL). The mixture was stirred for 20 min at rt, then the layers were separated.

The aqueous layer was extracted with Et₂O (2 x 70 mL). The combined organic extracts were dried over Na₂SO₄, filtered, and concentrated in vacuo. The residue was purified by silica gel flash column chromatography (gradient: 1:19 acetone:hexanes to 1:9 acetone:hexanes) to give **S-15** (0.011 g, 80%) as a colorless foam, along with recovered starting material (**30**) (1 mg, 5%): $[\alpha]^{23}_{589} +2.16$, $[\alpha]^{23}_{577} +3.98$, $[\alpha]^{23}_{546} +7.03$ ($c = 1.14$, CH₂Cl₂); ¹H NMR (500 MHz, CDCl₃) δ 7.48 (m, 2H), 7.43–7.22 (m, 8H), 5.79 (s, 1H), 3.94 (m, 1H), 3.79 (d, $J = 13.3$ Hz, 1H), 3.65 (d, $J = 13.3$ Hz, 1H), 3.11 (dd, $J = 7.1, 7.1$ Hz, 1H), 2.72–2.61 (m, 2H), 2.52 (m, 1H), 2.47 (m, 1H), 2.38 (d, $J = 12.7$ Hz, 1H), 2.19 (d, $J = 12.7$ Hz, 1H), 2.14–2.00 (m, 3H), 1.66 (m, 1H), 1.48 (dd, $J = 11.5, 11.5$ Hz, 1H), 1.35 (s, 3H), 1.25 (m, 1H), 0.94 (dd, $J = 11.8, 11.8$ Hz, 1H), 0.84 (d, $J = 6.3$ Hz, 3H), 0.36 (s, 3H), 0.34 (s, 3H), missing H attributed to deuterium exchange with solvent; ¹³C NMR (125 MHz, CDCl₃) δ 204.7 (C), 166.5 (C), 139.4 (C), 138.5 (C), 133.4 (CH), 129.3 (CH), 129.2 (CH), 128.6 (CH), 128.2 (CH), 128.0 (CH), 126.8 (CH), 72.5 (CH), 69.0 (CH), 58.8 (CH₂), 56.2 (C), 55.9 (CH₂), 49.3 (CH), 49.0 (CH), 42.9 (CH₂), 39.3 (CH₂), 34.7 (CH), 32.8 (CH₃), 28.5 (CH₂), 27.4 (CH₂), 16.3 (CH₃), –2.8 (CH₃), –3.1 (CH₃); IR (film): 3429, 2924, 1637 cm⁻¹; HRMS (ESI) m/z calcd for C₃₁H₄₂NO₂Si [(M+H)⁺]: 488.2985; found: 488.2977.



Benzylammonium salt 31. Triethylamine (4 μL , 0.03 mmole) and methanesulfonyl chloride (2 μL , 0.02 mmole) were added sequentially to a solution of alcohol **S-15** (9.1 mg, 0.019 mmole) in CH₂Cl₂ (1 mL) at 0 °C. The cold bath was removed and the solution was stirred at rt for 24 h. Additional triethylamine (5 μL) and methanesulfonyl chloride (3 μL) were added and the solution was stirred for 24 h at rt. The reaction mixture was loaded directly onto a silica gel column that had been pretreated with sodium bromide as

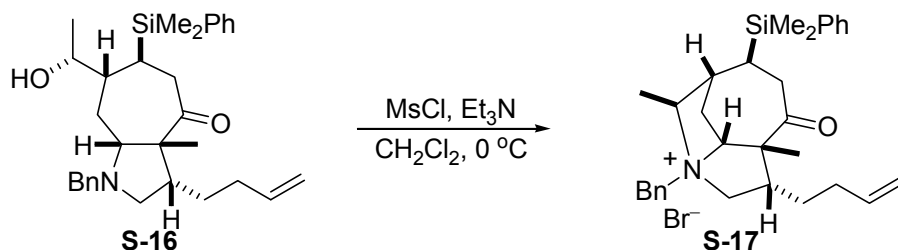
per the literature method.¹³ The product was eluted with a 9:1 mixture of CH₂Cl₂:MeOH and the product-containing fractions were concentrated in vacuo. The residue was dissolved in CH₂Cl₂ (20 mL) and washed with a 1:1 mixture of brine:saturated NaHCO₃ solution (10 mL), dried over Na₂SO₄, filtered, and concentrated in vacuo to give **31** as a colorless solid: $[\alpha]_{589}^{24} +55$, $[\alpha]_{577}^{24} +58$, $[\alpha]_{546}^{24} +68$, $[\alpha]_{435}^{24} +153$, $[\alpha]_{405}^{24} +204$ ($c = 0.65$, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.77 (d, $J = 6.5$ Hz, 2H), 7.52-7.50 (m, 2H), 7.48-7.42 (m, 6H), 6.00 (s, 1H), 5.61 (d, $J = 12.7$ Hz, 1H), 4.66 (dd, $J = 13.6, 9.1$ Hz, 1H), 4.19 (d, $J = 12.7$ Hz, 1H), 3.99 (dd, $J = 13.6, 13.6$ Hz, 1H), 3.78 (m, 1H), 3.58 (d, $J = 6.5$ Hz, 1H), 2.76 (m, 1H), 2.54 (m, 2H), 2.42 (m, 2H), 2.31 (dd, $J = 13.9, 4.3$ Hz, 1H), 1.97 (m, 1H), 1.85 (d, $J = 7.0$ Hz, 3H), 1.81 (m, 2H), 1.62 (m, 1H), 1.12 (m, 1H), 1.10 (s, 3H), 0.40 (s, 3H), 0.39 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 199.8 (C), 155.2 (C), 135.1 (C), 133.7 (CH), 132.9 (CH), 132.8 (CH), 130.8 (CH), 130.1 (CH), 129.6 (CH), 128.4 (CH), 127.9 (C), 87.1 (CH), 81.3 (CH), 62.3 (CH₂), 57.6 (CH₂), 50.7 (C), 47.3 (CH), 41.9 (CH), 37.7 (CH₂), 35.0 (CH₂), 33.6 (CH), 25.4 (CH₃), 24.0 (CH₂), 18.7 (CH₂), 14.8 (CH₃), -4.8 (CH₃), -4.9 (CH₃); IR (film) 2953, 2924, 1648 cm⁻¹; HRMS (ESI) m/z calcd for C₃₁H₄₀NOSi [(M)⁺]: 470.2879; found: 470.2877.



(3*S*,3*aR*,6*S*,7*R*,8*aR*)-1-Benzyl-3-(but-3-enyl)-6-(dimethyl(phenyl)silyl)-7-((*R*)-1-hydroxyethyl)-3*a*-methyl-octahydrocyclohepta[*b*]pyrrol-4(*5H*)-one (S-16). Hydrogen fluoride (0.3 mL, 48% aq) was added to a solution of silyl ether **22a** (0.031 g, 0.043 mmol) in MeCN (2.5 mL) in a Nalgene vial at rt. The solution was then heated at 50 °C for 24 h. Additional hydrogen fluoride (0.4 mL, 48% aq) was added and the solution was maintained for 21 h at 50 °C. The solution was then cooled to rt and added to a stirring, biphasic mixture of Et₂O (70 mL) and aqueous 1 N NaOH solution (60 mL). The mixture was stirred for 20 min at rt, then the layers were separated and the aqueous layer was

¹³ Bluhm, L. H.; Li, T. *Tetrahedron Lett.* **1998**, *39*, 3623–3626.

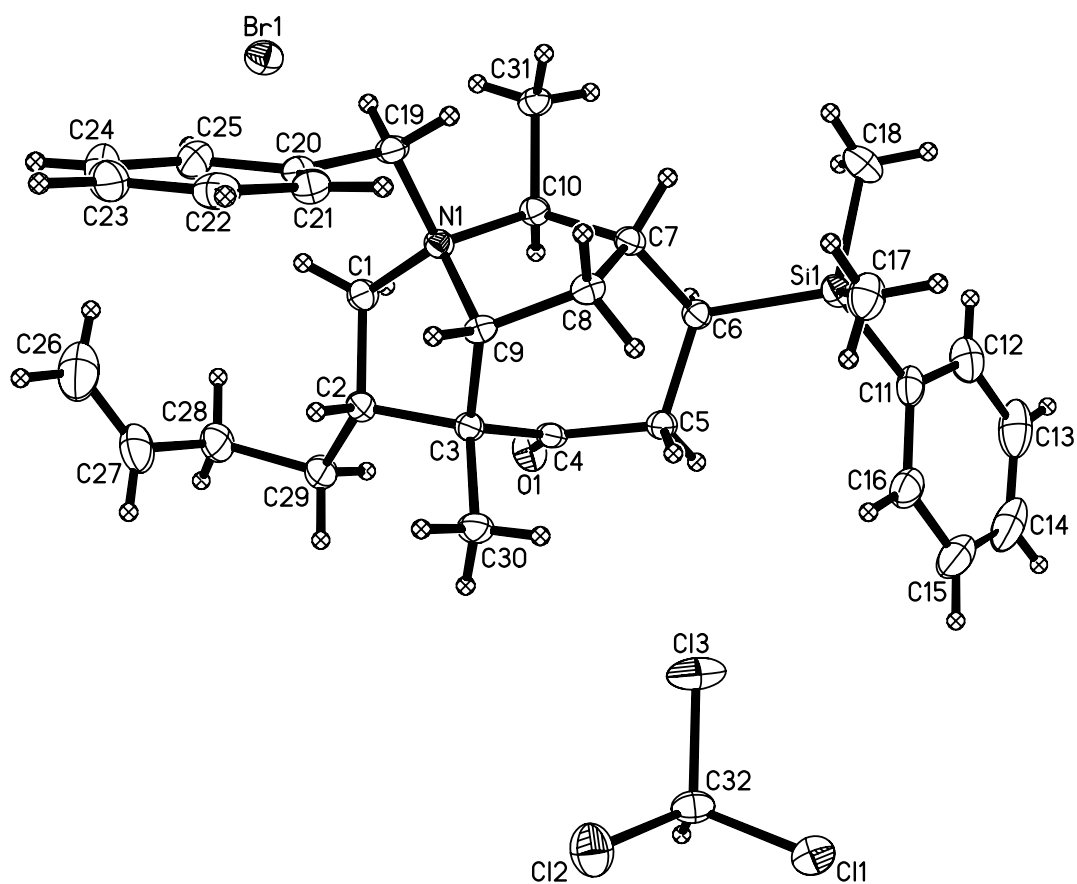
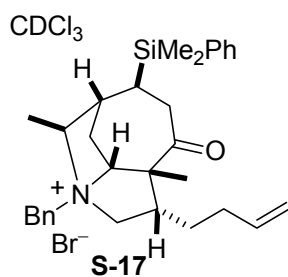
extracted with Et₂O (2 x 20 mL). The combined organic extracts were dried over Na₂SO₄, filtered, and concentrated in vacuo. The residue was purified by silica gel flash column chromatography (5:1 hexanes:acetone) to give **S-16** (15.5 mg, 74%) as a colorless oil: $[\alpha]^{24}_{589} -68$, $[\alpha]^{24}_{577} -75$, $[\alpha]^{24}_{546} -86$, $[\alpha]^{24}_{435} -158$, $[\alpha]^{24}_{405} -190$ ($c = 0.23$, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.53-7.51 (m, 2H), 7.38-7.29 (m, 8H), 5.70-5.61 (m, 1H), 4.95-4.90 (m, 2H), 4.10 (d, $J = 13.2$ Hz, 1H), 3.93-3.90 (m, 1H), 3.55 (d, $J = 13.2$ Hz, 1H), 3.14 (dd, $J = 11.3, 4.8$ Hz, 1H), 3.01 (dd, $J = 11.6, 8.8$ Hz, 1H), 2.84-2.80 (m, 2H), 2.53 (dd, $J = 11.3, 6.1$ Hz, 1H), 1.97-1.93 (m, 2H), 1.86-1.71 (m, 4H), 1.67-1.65 (m, 1H), 1.47-1.41 (m, 1H), 1.02-0.99 (m, 1H), 1.01 (s, 3H), 0.88 (d, $J = 6.4$ Hz, 3H), 0.88-0.85 (m, 1H), 0.38 (s, 3H), 0.35 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 213.8 (C), 138.3 (C), 138.0 (CH), 137.1 (C), 134.6 (CH), 129.9 (CH), 129.2 (CH), 128.7 (CH), 127.8 (CH), 115.2 (CH₂), 73.5 (CH), 71.7 (CH), 62.0 (C), 61.0 (CH₂), 56.6 (CH₂), 47.5 (CH), 43.6 (CH₂), 42.4 (CH), 33.2 (CH₂), 32.9 (CH₂), 28.6 (CH₂), 24.2 (CH₃), 21.8 (CH), 21.0 (CH₃), -3.1 (CH₃), -4.0 (CH₃), missing carbon attributed to accidental equivalence; IR (film) 2965, 1691, 1455 cm⁻¹; HRMS (ESI) m/z calcd for C₃₁H₄₄NO₂Si [(M+H)⁺]: 490.3141; found: 490.3135.



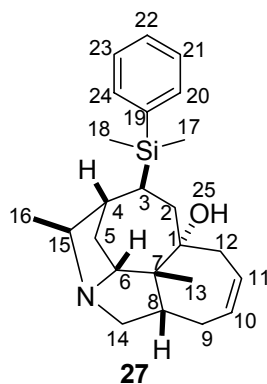
(3*S*,3*aR*,6*S*,7*R*,8*aR*,9*S*)-Octahydro-1-benzyl-3-(but-3-enyl)-3*a*,9-dimethyl-6-(dimethyl(phenyl)silyl)-1,7-methanocyclohepta[*b*]pyrrolium-4(5*H*)-one bromide (S-17**).** Triethylamine (7 μ L, 0.05 mmole) and methanesulfonyl chloride (3 μ L, 0.04 mmole) were added sequentially to a solution of alcohol **S-16** (15 mg, 0.031 mmole) in CH₂Cl₂ (1 mL) at 0 °C. The solution was stirred at 0 °C for 30 min, then diluted with CH₂Cl₂ (3 mL) and saturated aqueous NaHCO₃ solution (3 mL). The layers were separated, and the aqueous layer was extracted with CH₂Cl₂ (3 x 1 mL). The combined organic extracts were concentrated in vacuo and the residue was purified on a silica gel column that had been pretreated with sodium bromide as per the literature method.¹³ The product was

eluted with a 9:1 mixture of CH₂Cl₂:MeOH and the product-containing fractions were concentrated in vacuo. The residue was dissolved in CH₂Cl₂, washed with a 1:1 mixture of dilute aqueous NaBr solution:saturated NaHCO₃ solution, dried over Na₂SO₄, filtered, and concentrated in vacuo to give **S-17** (15.0 mg, 89%) as a colorless solid. Slow diffusion of diethyl ether into a solution of **S-17** in CDCl₃ (0.2 mL) provided crystals suitable for X-ray analysis: $[\alpha]^{24}_{589} +27$, $[\alpha]^{24}_{577} +31$, $[\alpha]^{24}_{546} +38$, $[\alpha]^{24}_{435} +111$, $[\alpha]^{24}_{405} +167$ ($c = 0.41$, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 7.72 (d, $J = 7.0$ Hz, 2H), 7.57-7.55 (m, 2H), 7.44-7.43 (m, 3H), 7.41-7.35 (m, 3H), 5.58-5.50 (m, 1H), 5.02 (d, $J = 13.0$ Hz, 1H), 4.99 (d, $J = 13.0$ Hz, 1H), 4.93-4.89 (m, 2H), 4.45 (dd, $J = 13.0, 9.0$ Hz, 1H), 3.50 (d, $J = 6.7$ Hz, 1H), 3.32-3.25 (m, 2H), 3.14-3.08 (m, 1H), 3.00-2.97 (m, 1H), 2.59 (dd, $J = 12.2, 12.2$ Hz, 1H), 2.27 (dd, $J = 11.9, 5.1$ Hz, 1H), 2.17-2.09 (m, 1H), 2.00-1.93 (m, 1H), 1.80-1.70 (m, 5H), 1.62-1.56 (m, 1H), 1.09 (dd, $J = 12.5, 5.0$ Hz, 1H), 1.05-1.01 (m, 1H), 0.99 (s, 3H), 0.43 (s, 3H), 0.39 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 213.4 (C), 137.2 (CH), 135.1 (C), 134.3 (CH), 133.2 (CH), 130.7 (CH), 130.1 (CH), 129.6 (CH), 128.5 (CH), 128.3 (C), 116.5 (CH₂), 87.9 (CH), 79.6 (CH), 66.1 (CH₂), 58.5 (CH₂), 56.7 (C), 49.0 (CH), 42.4 (CH), 37.8 (CH₂), 33.5 (CH₂), 30.7 (CH), 25.9 (CH₂), 24.5 (CH₂), 20.0 (CH₃), 15.0 (CH₃), -4.56 (CH₃), -4.60 (CH₃); IR (film) 2923, 1698, 1454 cm⁻¹; HRMS (ESI) m/z calcd for C₃₁H₄₂NOSi [(M)⁺]: 472.3036; found: 472.3019.

X-ray structure of S-17¹⁴



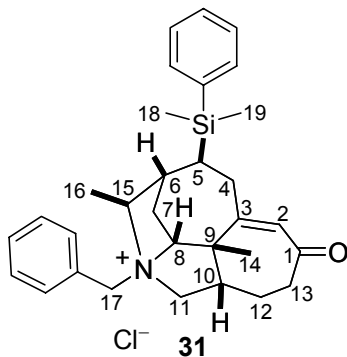
¹⁴ The thermal ellipsoid plot is shown at the 50% probability level.

**Table S1.** NMR data for **27** in CDCl₃.

Atom	¹ H shift (ppm)	¹ H mult.	<i>J</i> value(s) (Hz)	¹³ C shift (ppm)	¹³ C mult.	HMBC	COSY	NOESY
1				78.0	C			
2a	1.84	dd	14.1,14.1	35.9	CH ₂	3,7	2b,3	2b,5b,12a,13
2b	1.37	dd	15.0,3.5	35.9	CH ₂	1,3,4,7,12	2a,3	3,12a,12b
3	1.53	dd	13.1,3.6	27.1	CH	4,5,10	2a,2b	2b,15
4	2.21-2.13	m		44.2	CH	2,3,5	5a	5b,15,16
5a	2.21-2.13	m		25.8	CH ₂	6,7	4,5b	5b,6
5b	1.59	d	13.2	25.8	CH ₂	3,4,6,15	5a	2a,4,5a,6,13
6	3.71	s		79.0	CH	1,4,15	5a,5b	5a,5b,8,13
7				49.4	C			
8	2.41-2.27	m		48.3	CH	1,7,9	9a,9b	6,13,14a
9a	2.41-2.27	m		28.8	CH ₂	8	8,9b	9b,10
9b	2.60	s		28.8	CH ₂	4	8,9a	9a,10,14b
10	5.67	m		125.0	CH		9a,11	9a,9b,11
11	5.48	m		128.0	CH		10,12a,12b	10,12a,12b
12a	2.52	d	19.5	47.5	CH ₂		12b	2a,11,12b,13

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12b	2.05	d	19.8	47.5	CH ₂		12a	2b,11,12a
13	1.14	s		24.7	CH ₃	1,6,7,8		2a,5b,6,8,12a
14a	3.99	dd	9.2,9.2	60.1	CH ₂		8,14b	8,14b
14b	3.10	dd	10.3,10.3	60.1	CH ₂	8,9,15	8,14a	9b,14a,15
15	4.09	d	6.7	76.4	CH	3,4,14	4,16	3,4,14b,16
16	1.39	d	7.0	19.5	CH ₃	4,15	15	4,15
17	0.31	s		-4.9	CH ₃			2a,2b,3,4,5a,5b,20,24
18	0.32	s		-4.2	CH ₃			2a,2b,3,4,5a,5b,20,24
19				136.7	C	20,21,23,24		
20	7.45	m		133.8	CH	19,21,22	21	2a,3,4
21	7.42- 7.33	m		128.1	CH	19,20,22	20,22	
22	7.42- 7.33	m		129.6	CH	20,21,23,24	21,23	
23	7.42- 7.33	m		128.1	CH	19,22,24	22,24	
24	7.45	m		133.8	CH	19,22,23	23	2a,3,4
25	1.97	s			OH			

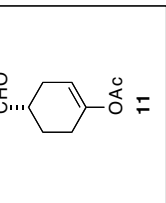
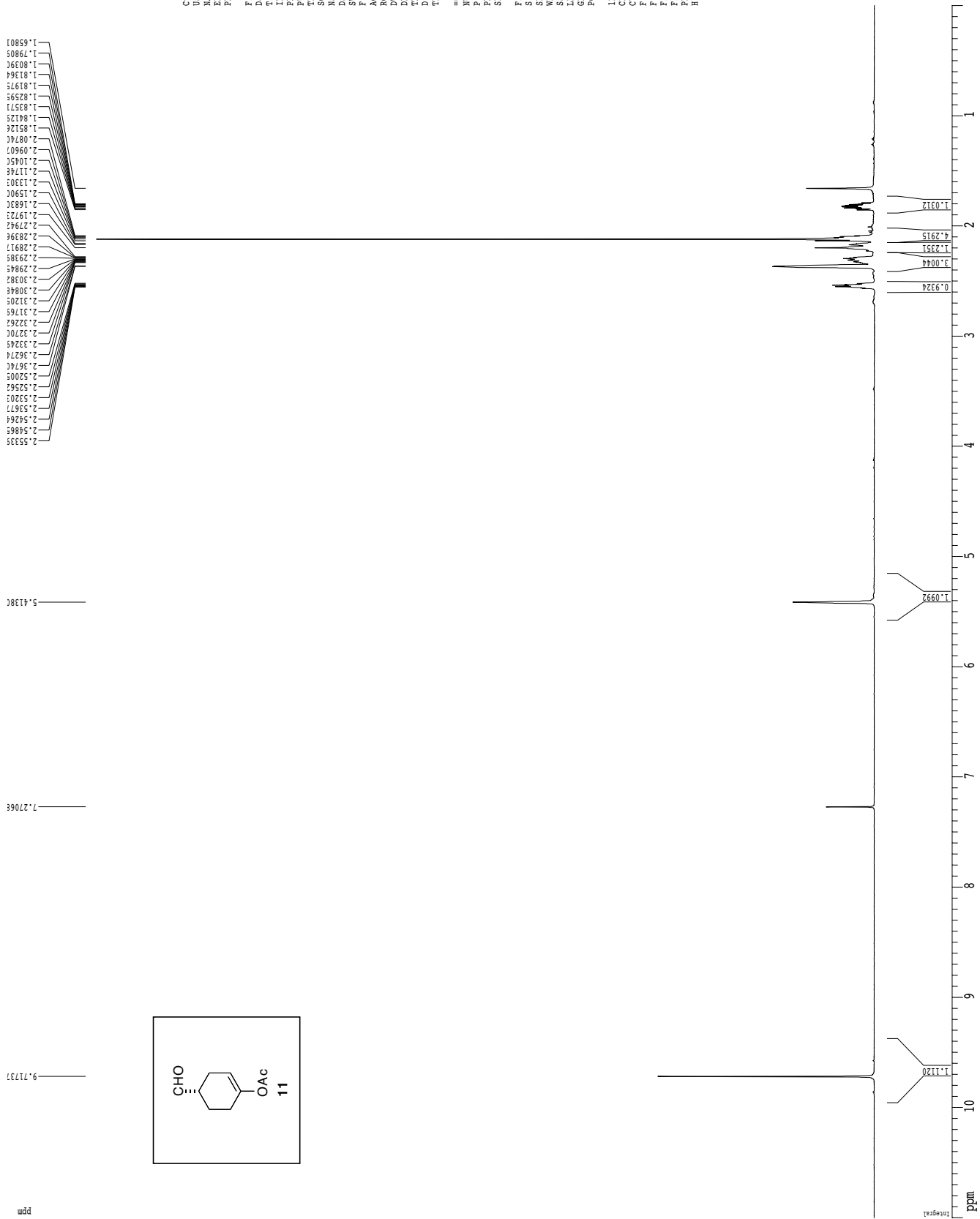
**Table S2.** NMR data for **31** in CDCl₃.

Atom	¹ H shift (ppm)	¹ H mult.	<i>J</i> value(s) (Hz)	¹³ C shift (ppm)	¹³ C mult.	HMBC	COSY
1				199.8	C		
2	6.00	s		132.8	CH	1, 3, 9, 4	4a,4b,13b
3				155.2	C		
4a	2.42	m		35.0	CH ₂	2,3,5,6,9	2,4b,5
4b	2.31	dd	13.9, 4.3	35.0	CH ₂	2,3,5,6,9	2,4a,5
5	1.12	m		33.6	CH	3,4,6,7,15	4a,4b
6	2.54	m		41.9	CH	4,7,8,16	7a,7b,15
7a	2.42	m		24.0	CH ₂	5,6,8,9	6,7b,8
7b	1.81	m		24.0	CH ₂	5,6,9	6,7a
8	3.58	d	6.5	81.3	CH	3,6,7,9,10,14,15,17	7a
9				50.7	C		
10	1.62	m		47.3	CH	3	11a,11b,12a,12b
11a	4.66	dd	13.6,9.1	62.3	CH ₂	8,9,10,12,15,17	10,11b
11b	3.99	dd	13.6,13.6	62.3	CH ₂	10,12,15,17	10,11a
12a	1.97	m		18.7	CH ₂	9	10,12b,13a,13b
12b	1.81	m		18.7	CH ₂	1,9,10,13	10,12a,13a,13b

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13a	2.76	m		37.7	CH ₂	1,10,12	12a,12b,13b
13b	2.54	m		37.7	CH ₂	1,10,12	12a,12b,13a
14	1.10	s		25.4	CH ₃	3,8,9,10	
15	3.78	m		87.1	CH	5,6,11,16,17	6,16
16	1.85	d	7.0	14.8	CH ₃	6,15	15
17a	5.61	d	12.7	57.6	CH ₂	8,11,15	17b
17b	4.19	d	12.7	57.6	CH ₂	8,11,15	17a
18	0.40	s		-4.8	CH ₃		
19	0.39	s		-4.9	CH ₃		

¹H spectrum



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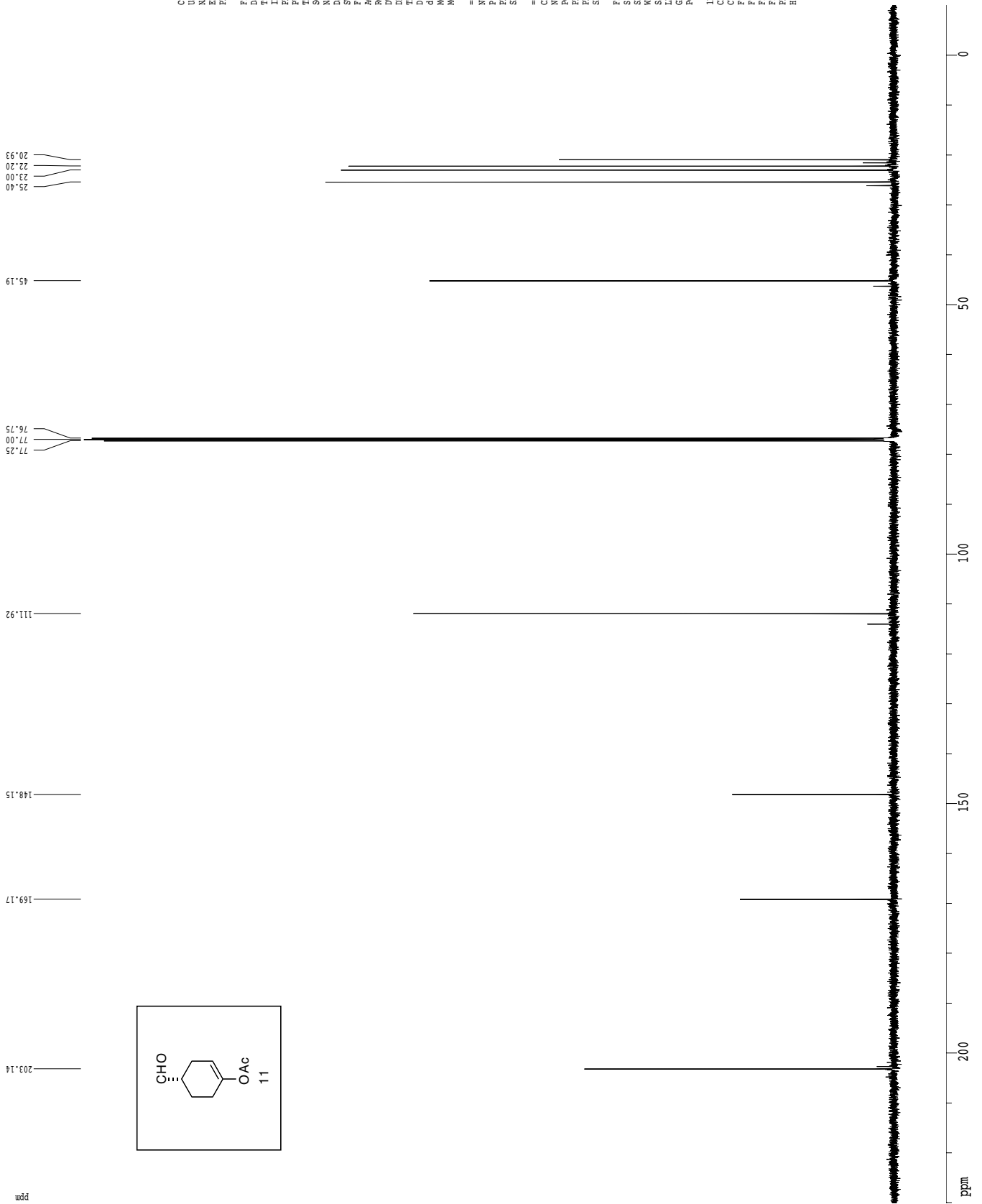
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13C spectrum with 1H decoupling



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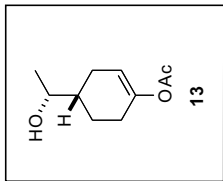
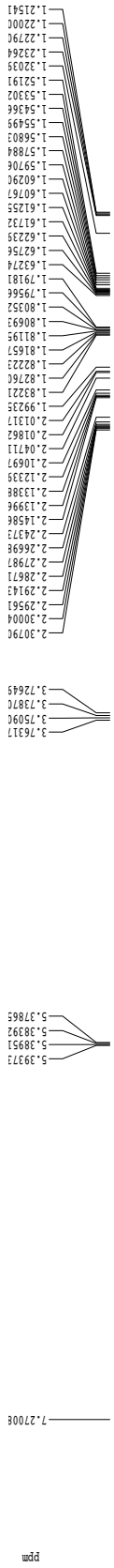
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¹H spectrum



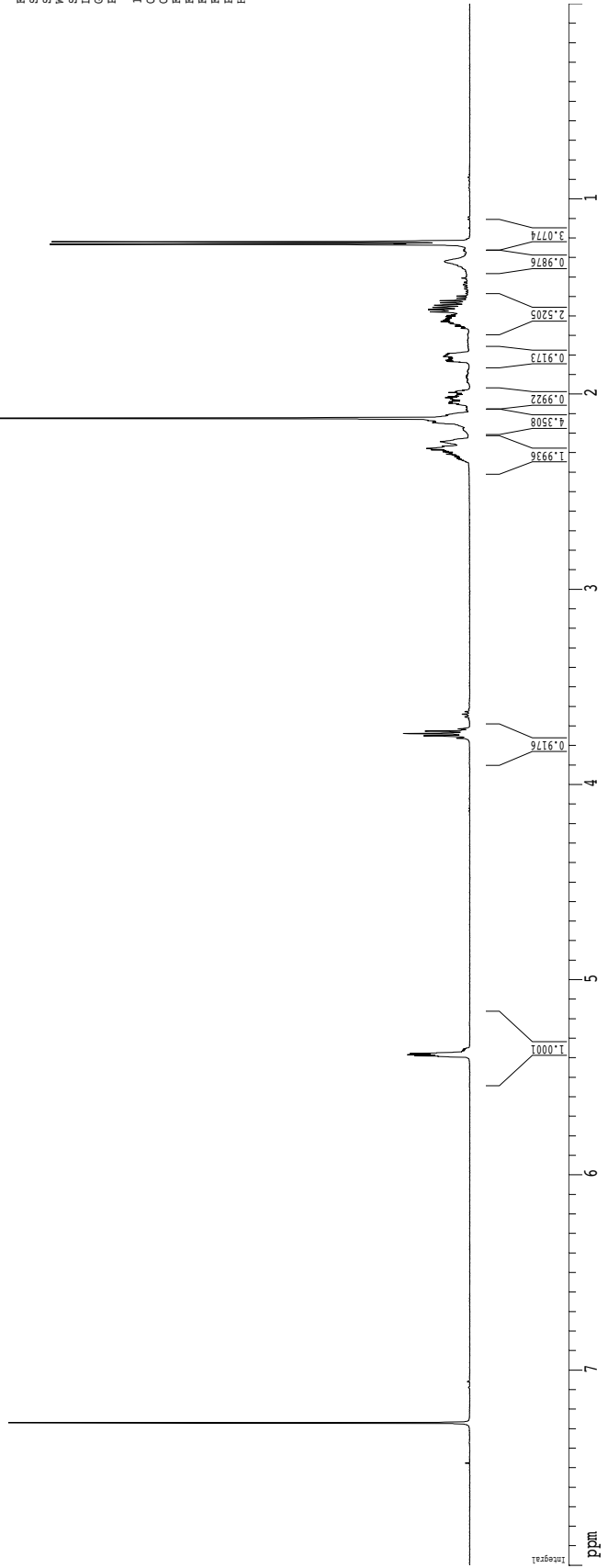
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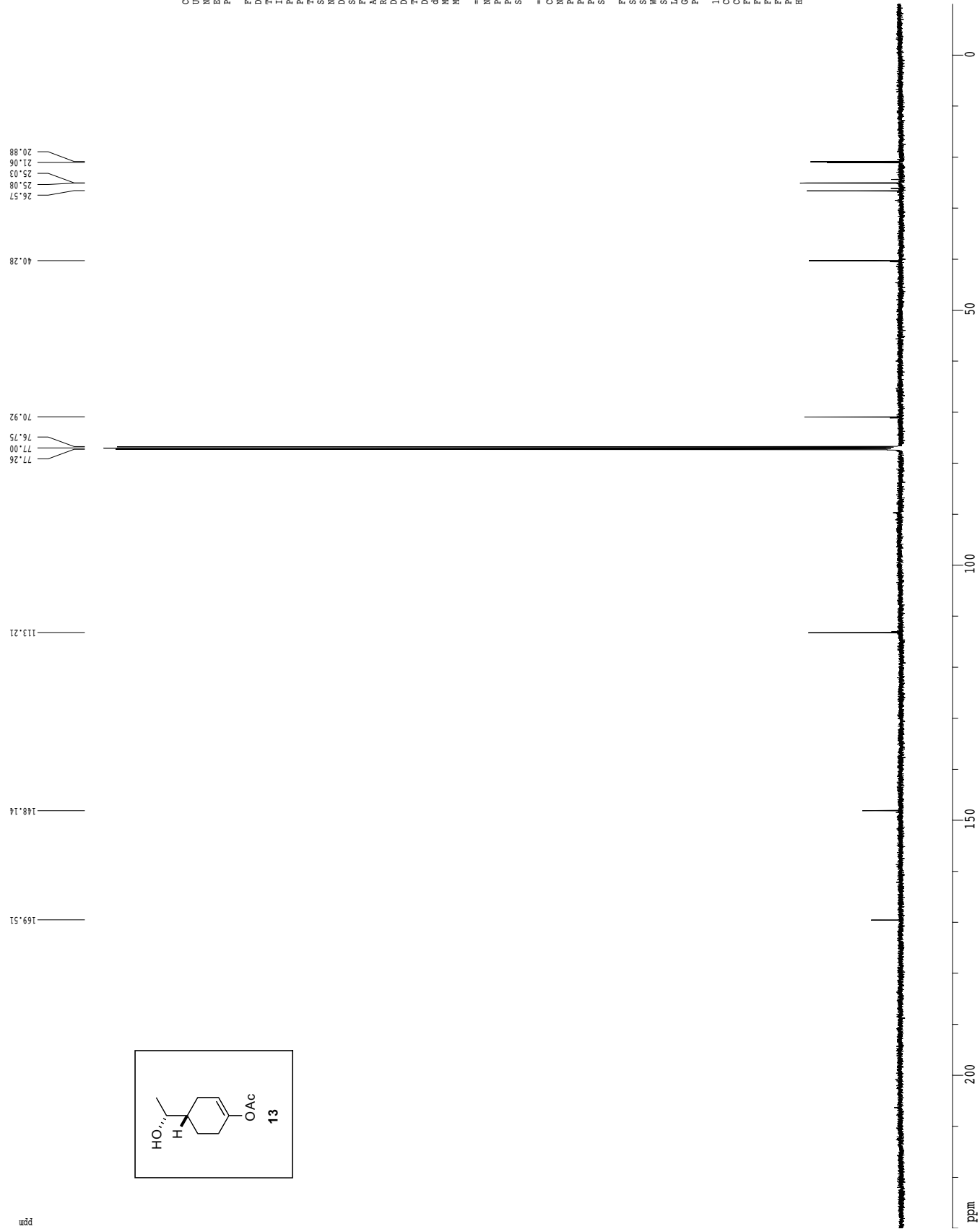
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 F1 8.000 ppm
 F2 4001.76 Hz
 F3 0.000 ppm
 F4 0.000 Hz
 F5 0.000 Hz
 HGCM 175.51581 Hz/cm



13C spectrum with 1H decoupling



Current Data Parameters
 USER travis
 NAME TD-2-119-2
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060928
 Time 15:29
 INSTRUM cryo500
 PROBDW 5 mm CPPTCI 1H-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 424
 DS 2
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813940 sec
 RG 18390.4
 DW 16.500 usec
 DE 6.00 usec
 TE 298.0 K
 D1 1.0000000 sec
 D2 0.1000000 sec
 MCREST 0.0300000 sec
 MCWRR 0.0150000 sec

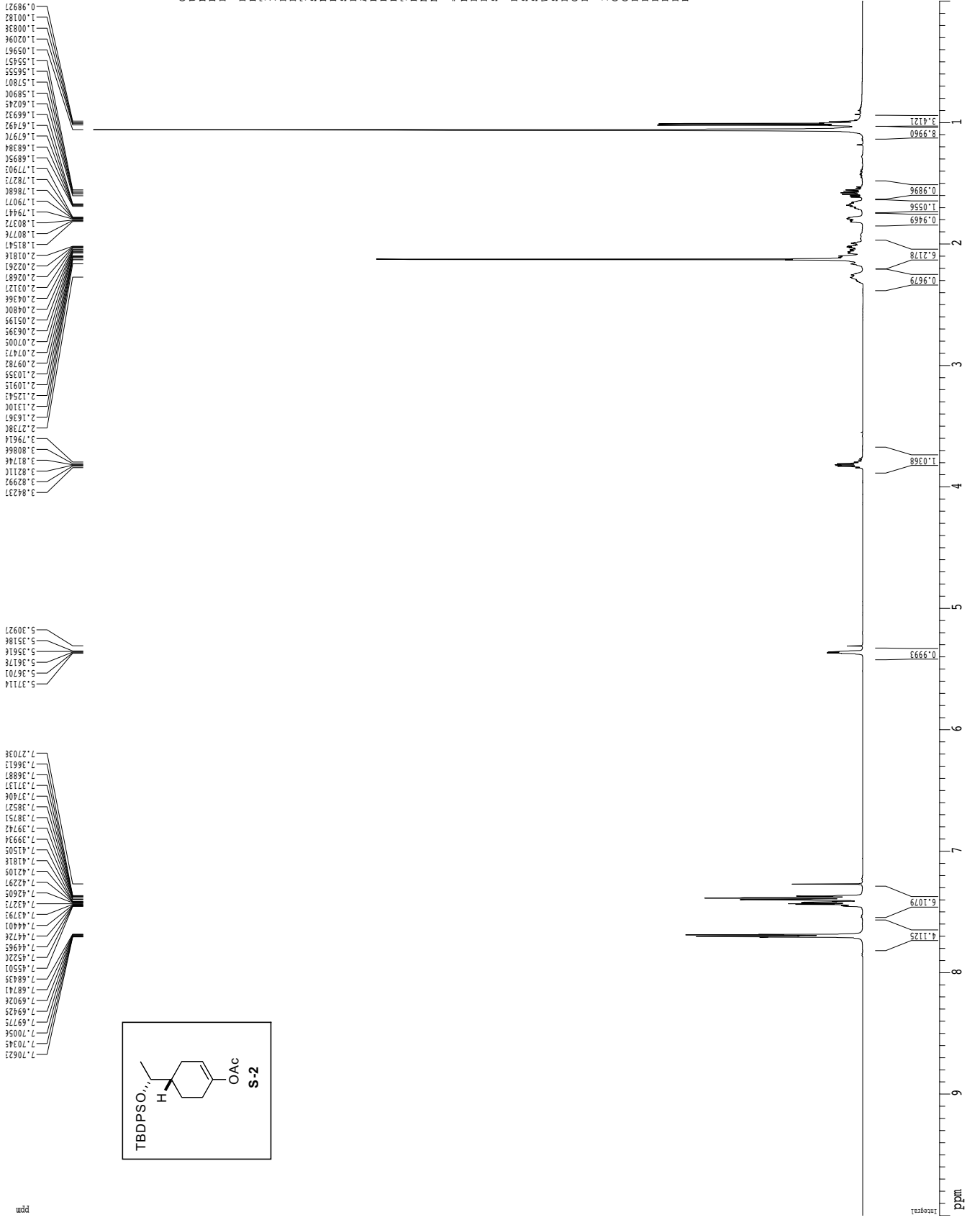
==== CHANNEL f1 =====
 NUC1 13C
 P1 15.00 usec
 PL1 -1.00 dB
 SFO1 125.7942348 MHz

==== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 1.60 dB
 PL12 23.54 dB
 SFO2 500.2225011 MHz

F2 - Processing parameters
 SI 65536
 SF 125.7804424 MHz
 RMW 24.00 MHz
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CI 15.65 cm
 F1 20.00 ppm
 F2 2839.50 ppm
 F3 10.00 ppm
 F4 -1257.80 Hz
 PPM0N 10.52632 ppm/cm
 HZCN 1324.00462 Hz/cm

1H spectrum



Current Data Parameters
 USER Travis
 NMR TD-410-2
 EXNO 1
 PROCNO 1

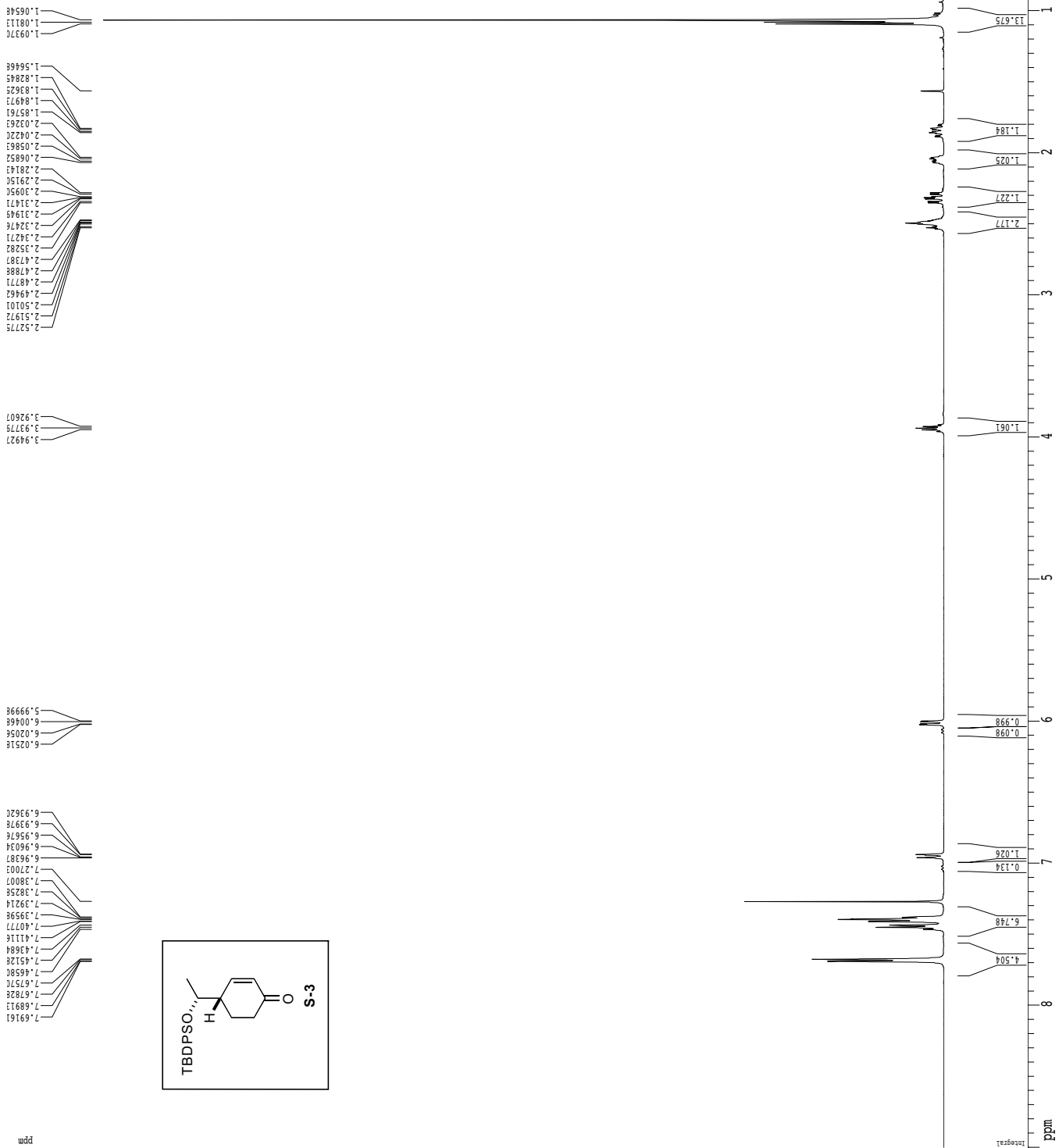
F2 - Acquisition Parameters
 Date_ 20070106
 Time 8.54
 INSTRUM crysol00
 PROBHD 5 mm CPCLP1
 PULPROG zgpg30
 TD 81728
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098943 Hz
 AQ 3.093918 sec
 RG 4.5
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.1000000 sec
 ACQST 0.0000000 sec
 ACWRK 0.0150000 sec

==== CHANNEL f1 =====
 NUC1 1H
 P1 8.00 usec
 PL1 1.60 dB
 SFO1 500.2255015 MHz

F2 - Processing parameters
 SI 65536
 SF 500.220263 MHz
 WTM EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 4.00

1D NMR plot parameters
 CX 22.80 cm
 CY 15.00 cm
 FLIP 10.000 ppm
 F1 5002.20 Hz
 F2 0.000 ppm
 F3 0.00 Hz
 FWHM 0.4860 ppm/cm
 HZCM 213.39476 Hz/cm

1H spectrum



Current Data Parameters
 USER travis
 NAME TD-2-211-1
 EXPNO 1
 PROCNO 1

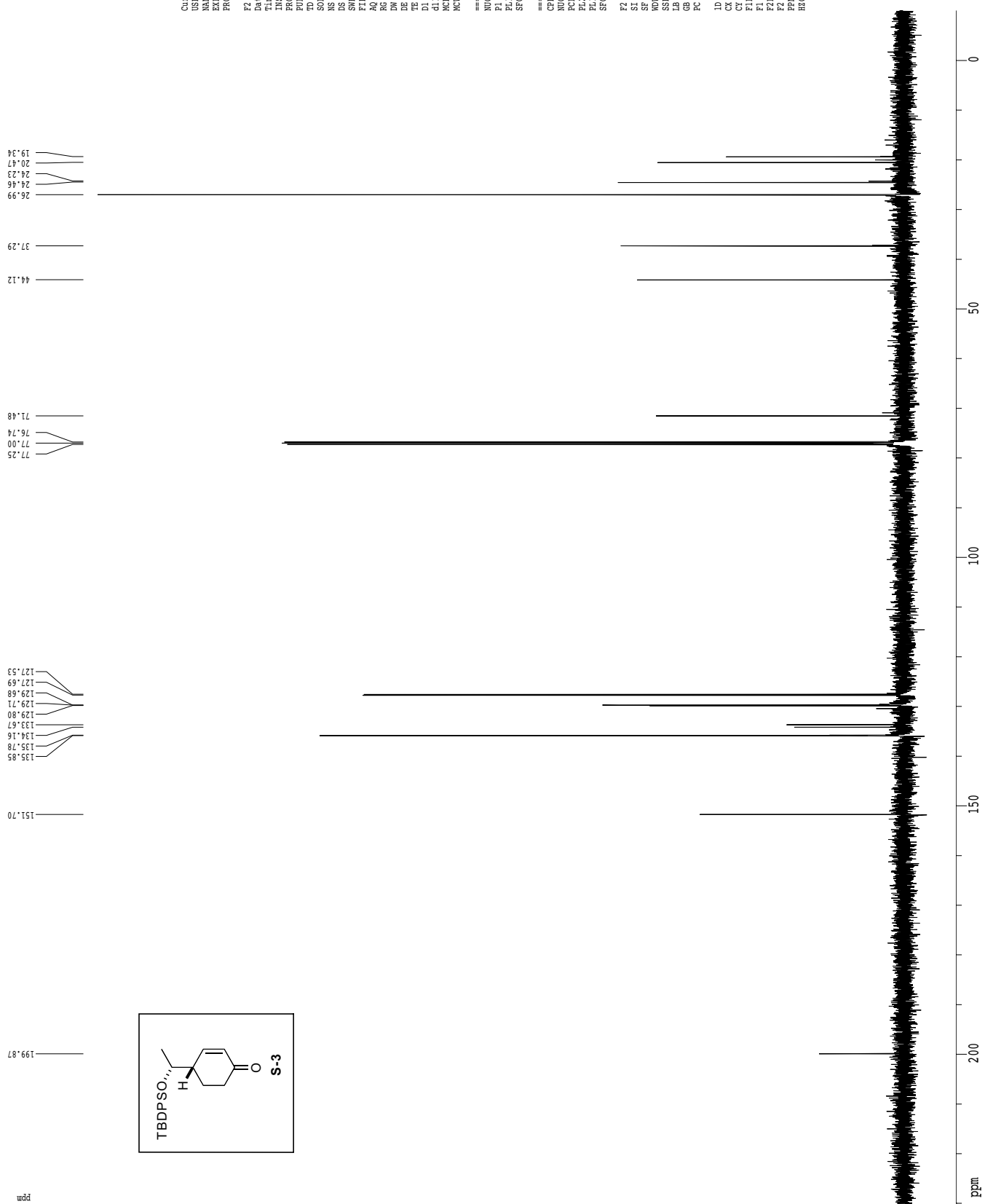
F2 - Acquisition Parameters
 Date_ 20070102
 Time 12:52
 INSTRUM ccp500
 PROBRD 5 mm CPCL1 H1-
 PULPROG zg30
 TD 81728
 SOLVENT CDCl3
 NS 8
 DS 8
 SWH 8013.872 Hz
 FIDRES 0.068043 Hz
 AQ 5.0948774 sec
 RG 5
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.1000000 sec
 d11 0.0500000 sec
 ACQST 0.0500000 sec
 ACWR 0.03500000 sec

***** CHANNEL f1 *****
 NUC1 1H
 P1 8.00 usec
 PL1 1.60 dB
 SFO1 500.225015 MHz

F2 - Processing parameters
 SI 65536
 SF 500.2200266 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 4.00

ID NMR F2.c Parameters
 CX 22.80 cm
 CY 15.00 cm
 FIP 9.000 ppm
 F1 4501.88 Hz
 F2 0.000 ppm
 F3 0.00 Hz
 HPCW 0.784848 ppm/cm
 HZCX 197.45528 Hz/cm

13C spectrum with 1H decoupling



```

Current Data Parameters
USER          travis
NAME          td-2-125-2
EXPNO         1
PROCNO        1

F2 - Acquisition Parameters
Date_         20061002
Time          15:17
PROBHD        gpcp13
PULPROG       zgpg30
PROBHD        5 mm broadbnd
PULPROG       zgpg30
TD            65536
SOLVENT       CDCl3
NS            128
DS            4
SWH           30303.031 Hz
FIDRES        0.462388 Hz
AQ            1.0813940 sec
RG            375.5
WDW           EM
SSB           0
GB            0
PC            16.500 usec
TE            298.0 K
D1            1.00000000 sec
d11           0.03000000 sec
d12           0.00000000 sec
d13           0.00000000 sec
d14           0.00000000 sec
d15           0.00000000 sec
d16           0.00000000 sec
d17           0.00000000 sec
d18           0.00000000 sec
d19           0.00000000 sec
d20           0.00000000 sec
d21           0.00000000 sec
d22           0.00000000 sec
d23           0.00000000 sec
d24           0.00000000 sec
d25           0.00000000 sec
d26           0.00000000 sec
d27           0.00000000 sec
d28           0.00000000 sec
d29           0.00000000 sec
d30           0.00000000 sec
d31           0.00000000 sec
d32           0.00000000 sec
d33           0.00000000 sec
d34           0.00000000 sec
d35           0.00000000 sec
d36           0.00000000 sec
d37           0.00000000 sec
d38           0.00000000 sec
d39           0.00000000 sec
d40           0.00000000 sec
d41           0.00000000 sec
d42           0.00000000 sec
d43           0.00000000 sec
d44           0.00000000 sec
d45           0.00000000 sec
d46           0.00000000 sec
d47           0.00000000 sec
d48           0.00000000 sec
d49           0.00000000 sec
d50           0.00000000 sec
d51           0.00000000 sec
d52           0.00000000 sec
d53           0.00000000 sec
d54           0.00000000 sec
d55           0.00000000 sec
d56           0.00000000 sec
d57           0.00000000 sec
d58           0.00000000 sec
d59           0.00000000 sec
d60           0.00000000 sec
d61           0.00000000 sec
d62           0.00000000 sec
d63           0.00000000 sec
d64           0.00000000 sec
d65           0.00000000 sec
d66           0.00000000 sec
d67           0.00000000 sec
d68           0.00000000 sec
d69           0.00000000 sec
d70           0.00000000 sec
d71           0.00000000 sec
d72           0.00000000 sec
d73           0.00000000 sec
d74           0.00000000 sec
d75           0.00000000 sec
d76           0.00000000 sec
d77           0.00000000 sec
d78           0.00000000 sec
d79           0.00000000 sec
d80           0.00000000 sec
d81           0.00000000 sec
d82           0.00000000 sec
d83           0.00000000 sec
d84           0.00000000 sec
d85           0.00000000 sec
d86           0.00000000 sec
d87           0.00000000 sec
d88           0.00000000 sec
d89           0.00000000 sec
d90           0.00000000 sec
d91           0.00000000 sec
d92           0.00000000 sec
d93           0.00000000 sec
d94           0.00000000 sec
d95           0.00000000 sec
d96           0.00000000 sec
d97           0.00000000 sec
d98           0.00000000 sec
d99           0.00000000 sec
d100          0.00000000 sec

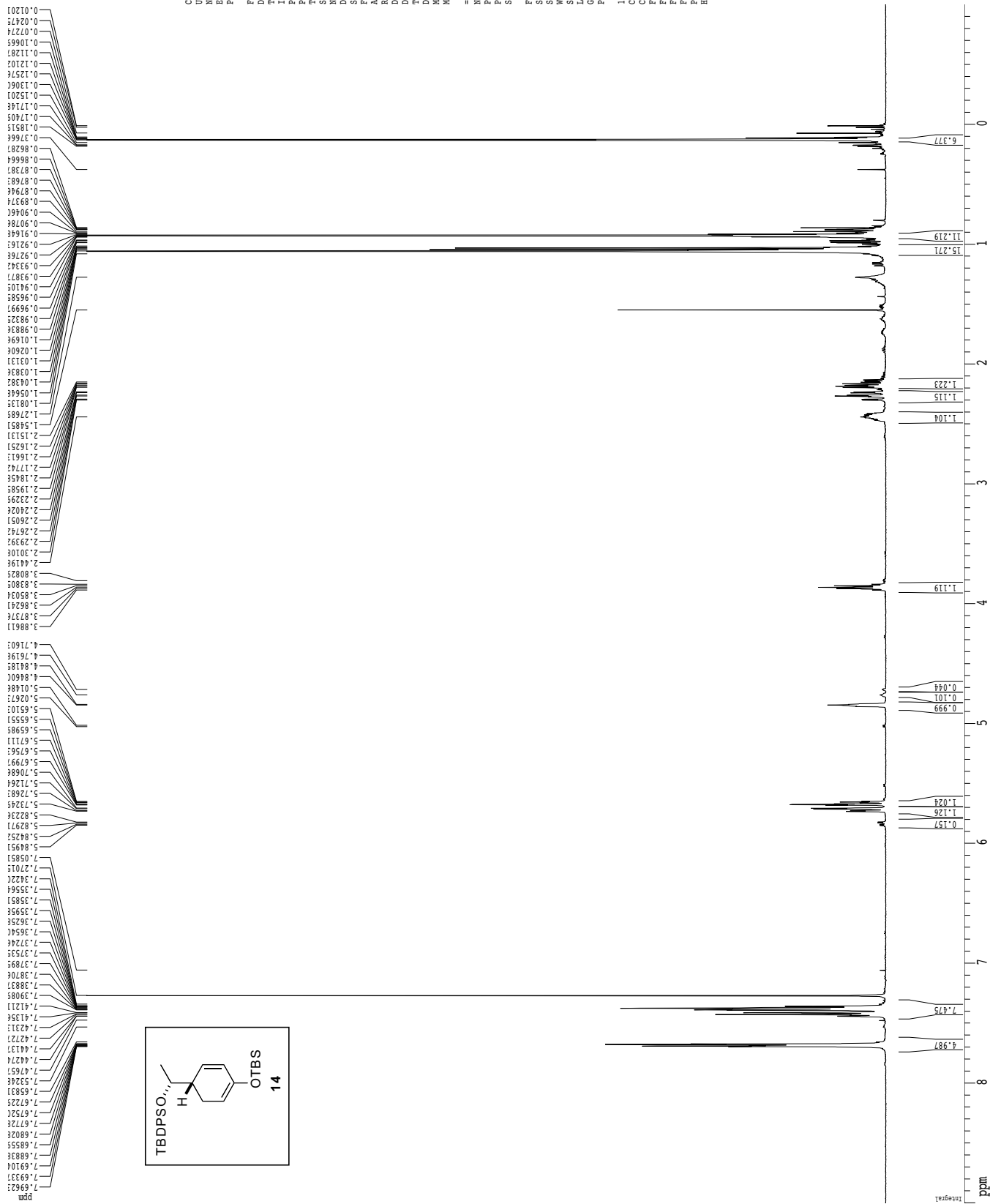
===== CHANNEL f1 =====
NUC1          13C
P1            7.08 usec
PL1           0.00 dB
SFO1          125.7213258 MHz

===== CHANNEL f2 =====
CPDPRG2       waltz16
NUC2          1H
PCPD2         80.00 usec
PL2           1.00 dB
PL12          1.00 dB
PL13          1.00 dB
SFO2          499.9324997 MHz

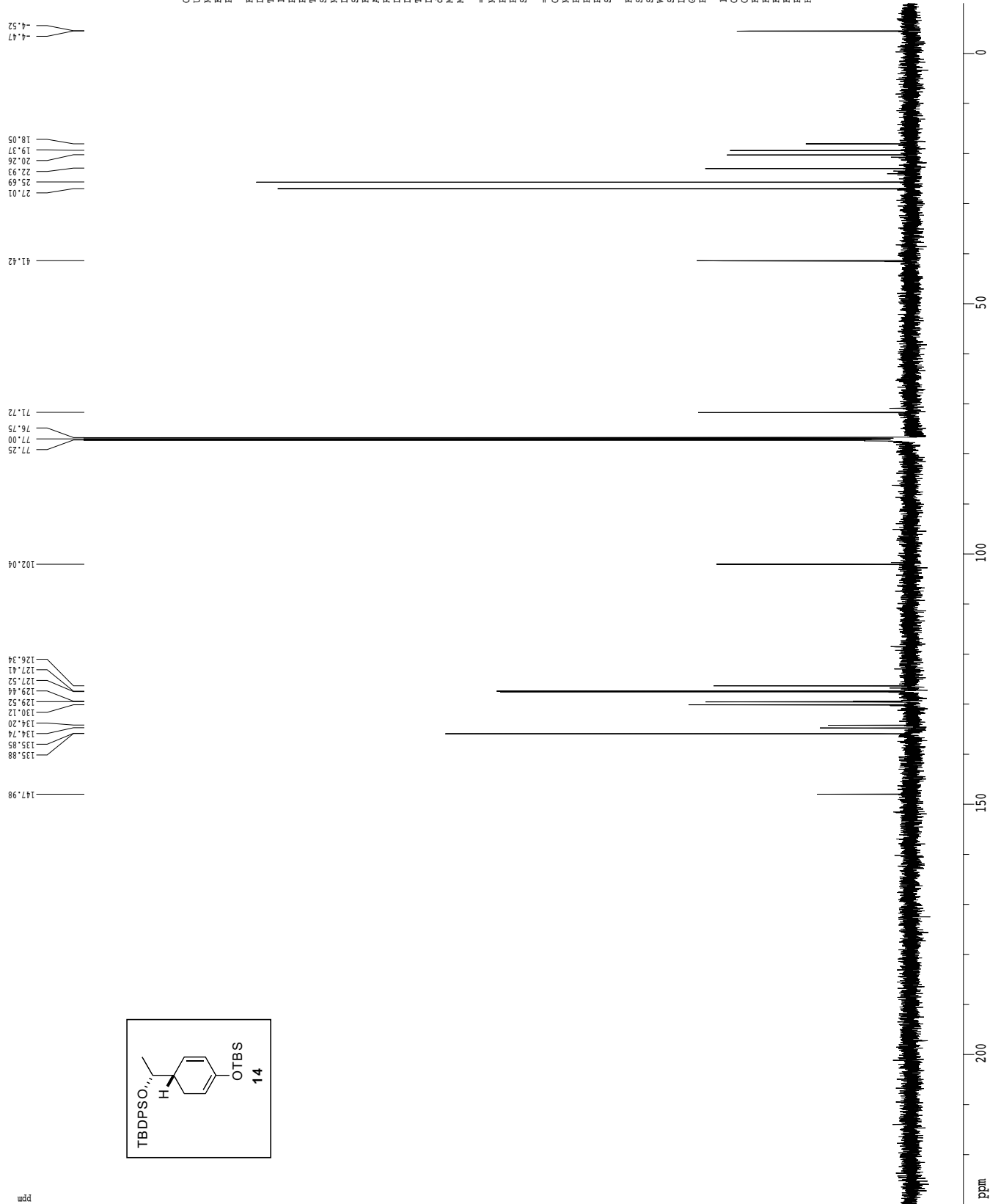
F2 - Processing parameters
SI            65536
SF            125.7075075 MHz
WDW           EM
SSB           0
LB            1.00 Hz
GB            0
PC            2.00

ID NMR plot parameters
CX            22.80 cm
CY            15.65 cm
FIDP          238.000 PPM
F1            28912.73 Hz
F2P           -10.000 PPM
F2            1257.18 Hz
FREQCY        125.7075075 PPM/cm
HZ CM        1323.25934 Hz/cm
    
```

¹H spectrum



¹³C spectrum with ¹H decoupling



```

Current Data Parameters
USER      travis
NAME      TD-2-180-2-2
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     20061121
Time      15.25
INSTRUM   cryo500
PROBHD    5 mm CPXI 1H-
PULPROG   zgpg30
TD         65418
SOLVENT   CDCl3
NS         247
DS         4
SWH        30303.031 Hz
FIDRES     0.463222 Hz
AQ         1.0784470 sec
RG         11585.2
DE         16.500 usec
TE         296.0 K
NUC1       13C
NUC2       1H
AQ1        0.6300000 sec
NUC1PROG   zgpg30
MORF       0.40000000 sec
MORF2      0.41500000 sec

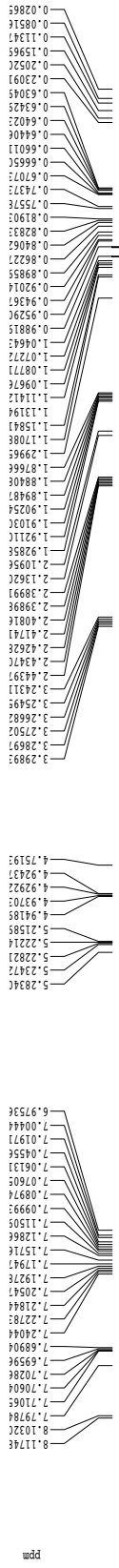
===== CHANNEL f1 =====
NUC1       13C
P1         15.00 usec
PL1        -1.00 dB
SFO1       125.7942348 MHz

===== CHANNEL f2 =====
CDEPRG2    waltz16
NUC2       1H
PCPD2      100.00 usec
PL2         1.60 dB
PL12       23.54 dB
SFO2       500.2225011 MHz

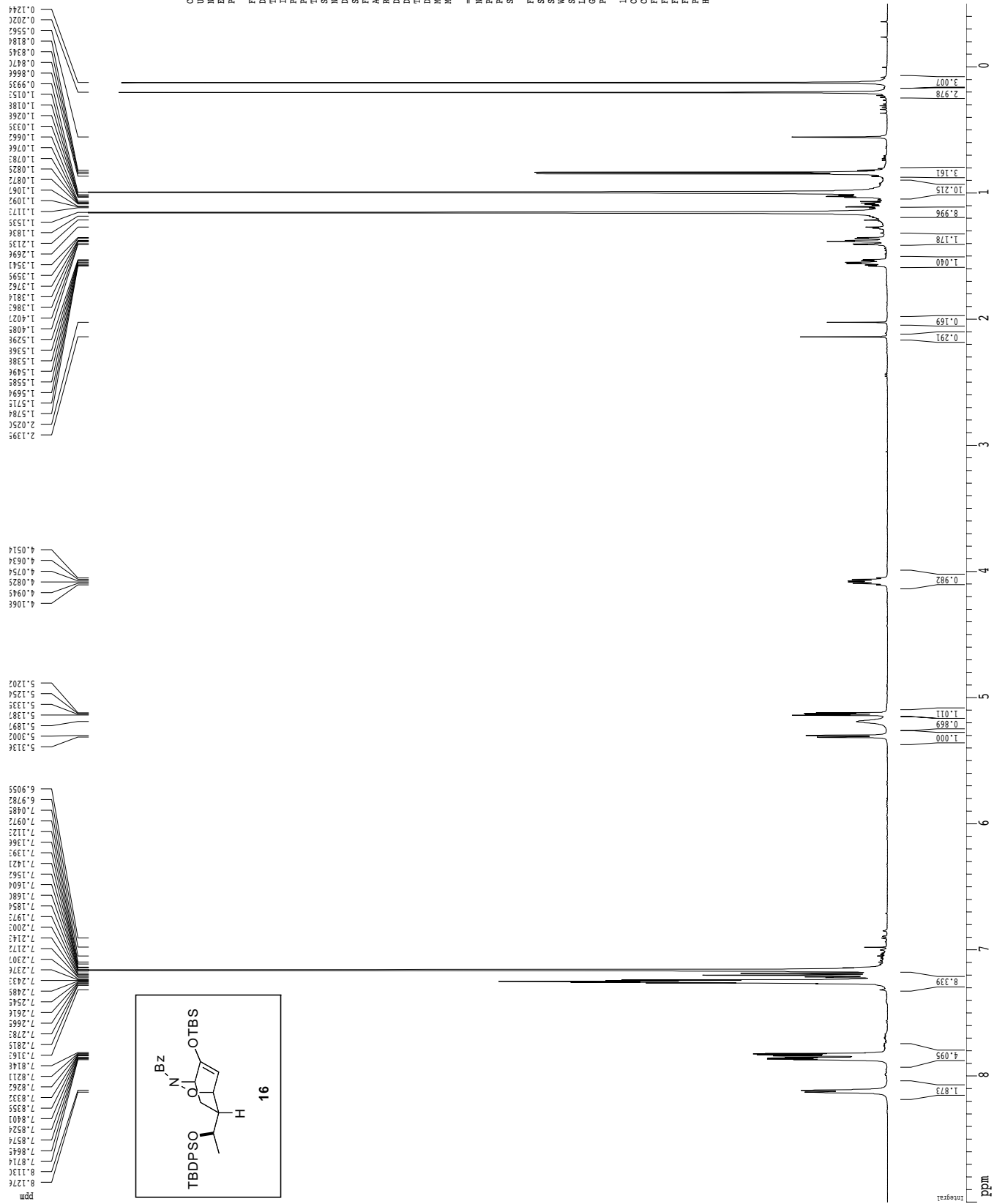
F2 - Processing parameters
SI         65536
SF         125.7884272 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00

JD NMR plot parameters
CX         2.00 cm
CY         2.00 cm
FT1        230.000 ppm
F1         28929.50 Hz
F2         -10.000 ppm
PP4MCH     10.52632 ppm/cm
HZCM       1324.00632 Hz/cm
    
```

1H spectrum



1H spectrum



Current Data Parameters
 USER travis
 NAME TD-2-06-4
 EXPNO 1
 PROCNO 1

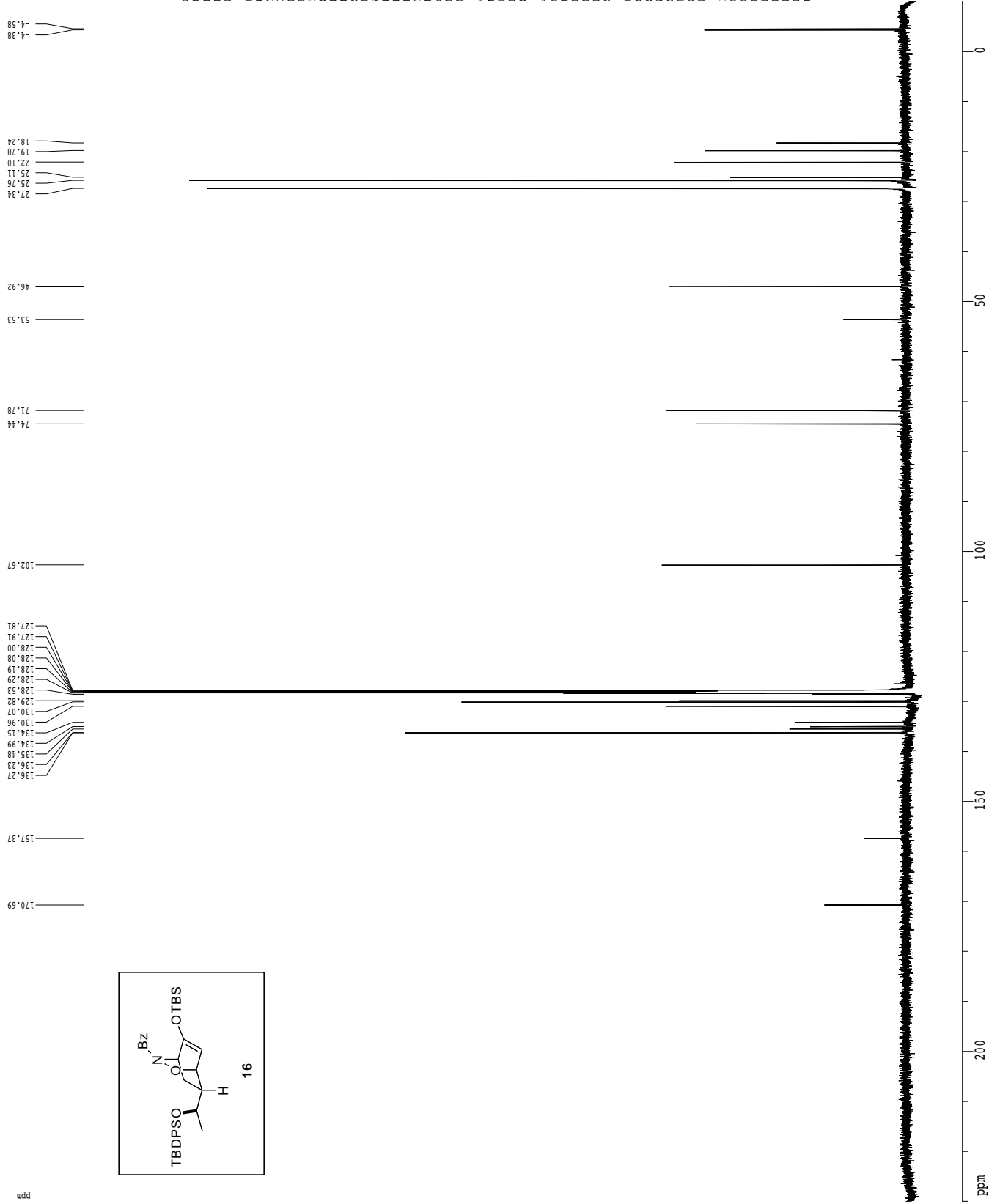
F2 - Acquisition Parameters
 Date_ 20070130
 Time 16.32
 INSTRUM cryo500
 PROBRD 5 mm CPFI 1H-
 PULPROG zgpg30
 TD 81728
 SOLVENT C6D6
 NS 2
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.0998774 sec
 RG 4
 DM 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D 0.10000000 sec
 ACQRES 0.00000000 sec
 MCKEY 0.00000000 sec
 MCKEY 0.01500000 sec

***** CHANNEL f1 *****
 NUC1 1H
 P1 8.00 usec
 PL1 0.00 dB
 SFO1 500.223913 MHz

F2 - Processing parameters
 SI 65536
 SF 500.220000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 4.00

1D NMR plot parameters
 CX 22.80 cm
 CY 30.00 cm
 F1 9.000 ppm
 F2 4501.98 Hz
 F3 0.500 ppm
 F4 -25.000 ppm
 FPMCM 0.41647 ppm/cm
 BPCCM 208.42500 Hz/cm

¹³C spectrum with ¹H decoupling



Current Data Parameters
 USER travis
 NAME TD-2-206-4
 EXPTNO 4
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20070130
 Time 16:37
 INSTRUM spect
 PROBD 5 mm CPCT-1H
 PULPROG zgpg30
 TD 65418
 SOLVENT C6D6
 NS 241
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.463222 Hz
 AQ 1.0794470 sec
 RG 325
 DW 16.500 usec
 DE 6.00 usec
 TE 323.0 K
 D1 1.4000000 sec
 d11 0.4300000 sec
 MCREST 0.0000000 sec
 MCPRK 0.0150000 sec

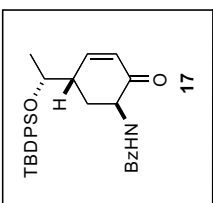
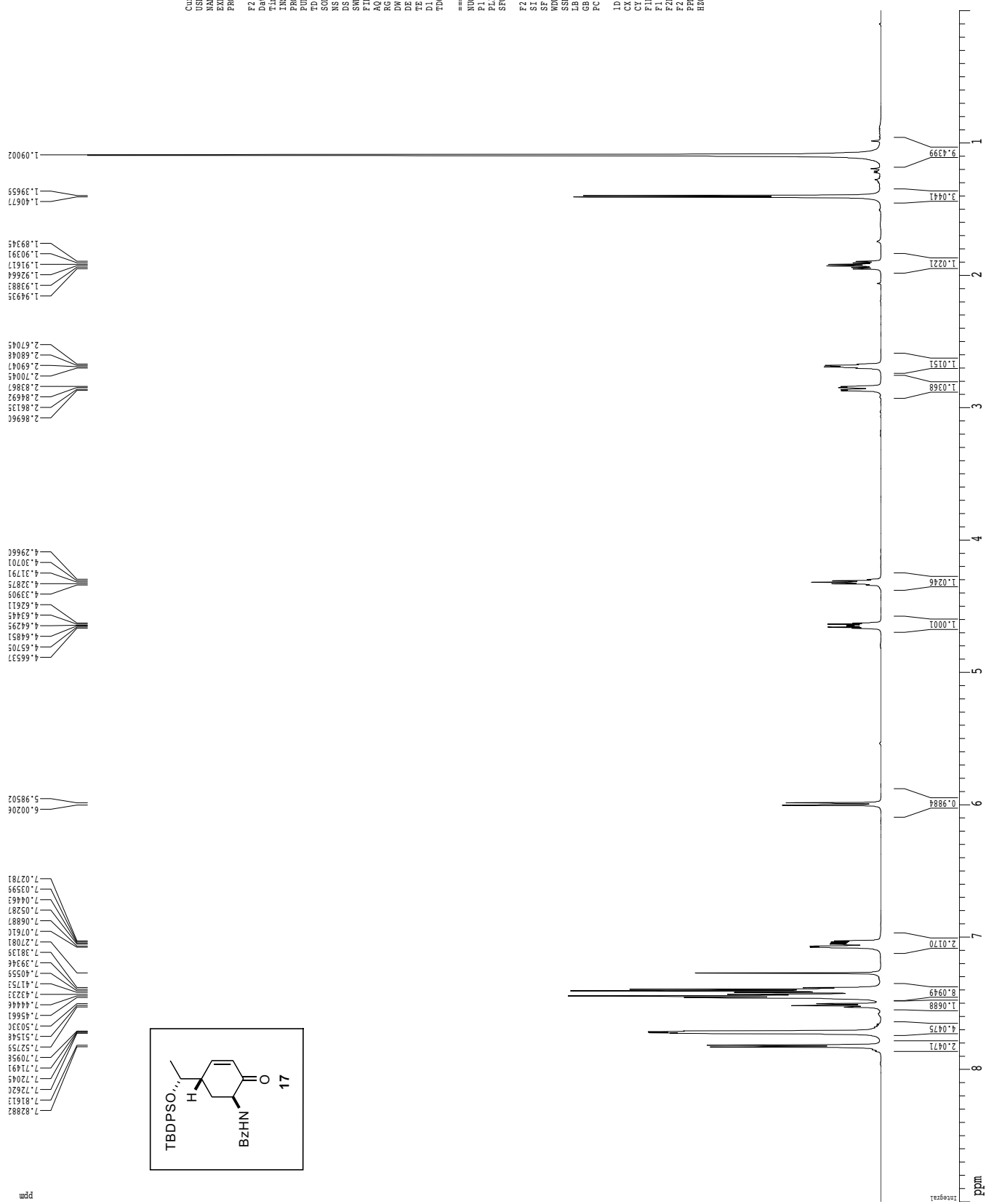
===== CHANNEL f1 =====
 NUC1 ¹³C
 P1 15.00 usec
 PL1 -1.00 dB
 SFO1 125.7942548 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 ¹H
 PCDP2 100.00 usec
 PL2 0.00 dB
 PL12 23.50 dB
 SFO2 500.2255111 MHz

F2 - Processing parameters
 SI 65536
 SF 125.7603744 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CY 150.00 cm
 F1P 230.000 ppm
 F2P -10.000 ppm
 F3P 0.000 Hz
 BRANCH 1.05432 /cm
 HECM 1324.00391 Hz/cm

1H spectrum



Current Data Parameters
 USER travis
 NAME TD-2-231-1
 EXPNO 11
 PROCNO 1

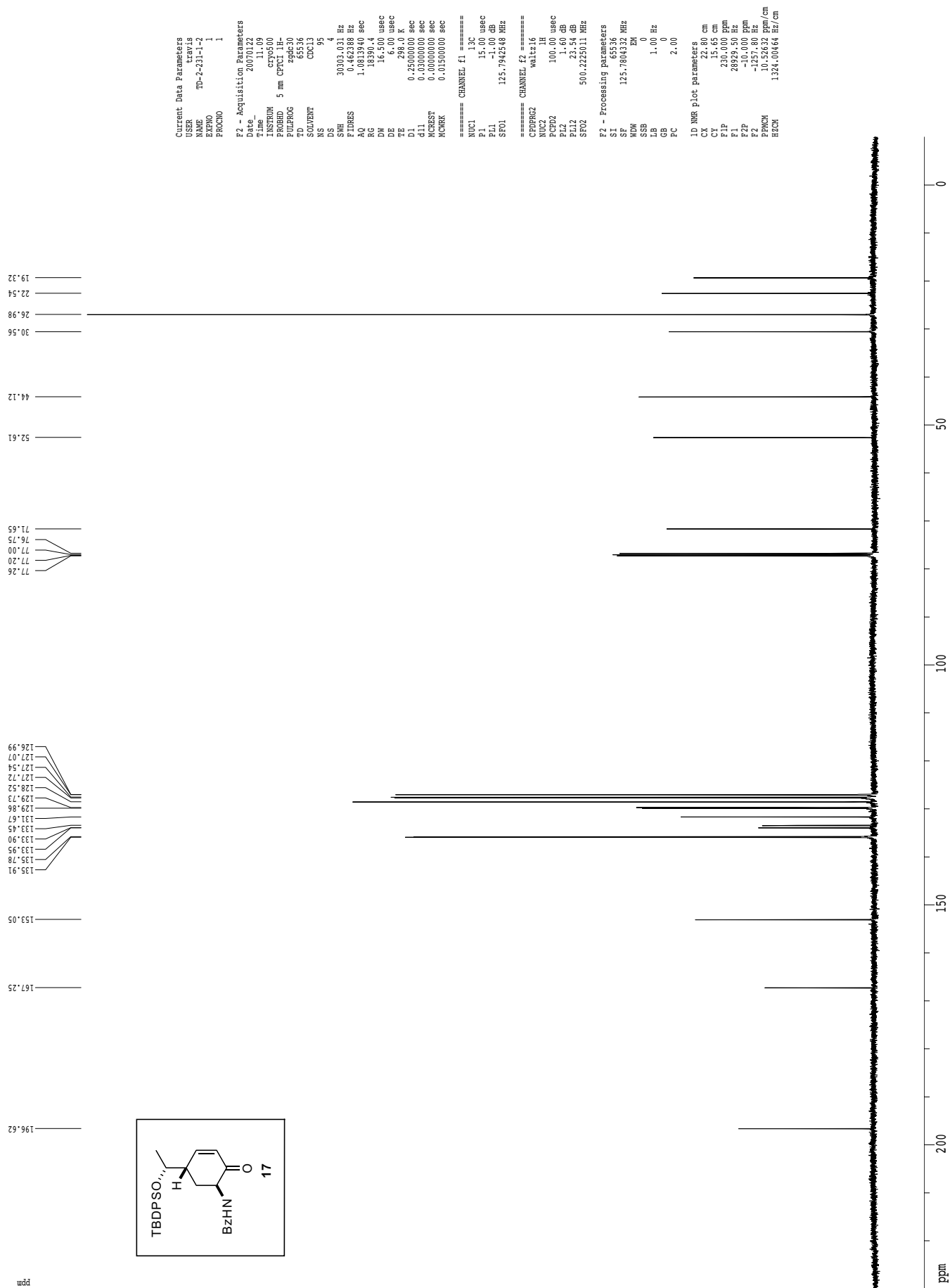
 F2 - Acquisition Parameters
 Date_ 20070122
 Time 10.32
 INSTRUM av600
 PROBRD 5 mm TBI H/13
 PULPROG zg30
 TD 47936
 CQCLP1 8
 NS 8
 DS 2
 SWH 9615.385 Hz
 FIDRES 0.081178 Hz
 AQ 5.0928259 sec
 RG 32
 DW 52.000 usec
 DE 6.00 usec
 TE 293.2 K
 C1 1
 D1 0.10000000 sec
 D10 1

 ===== CHANNEL f1 =====
 NUC1 1H
 P1 8.00 usec
 PL1 -1.00 dB
 SFO1 600.1342009 MHz

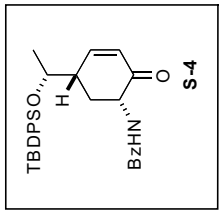
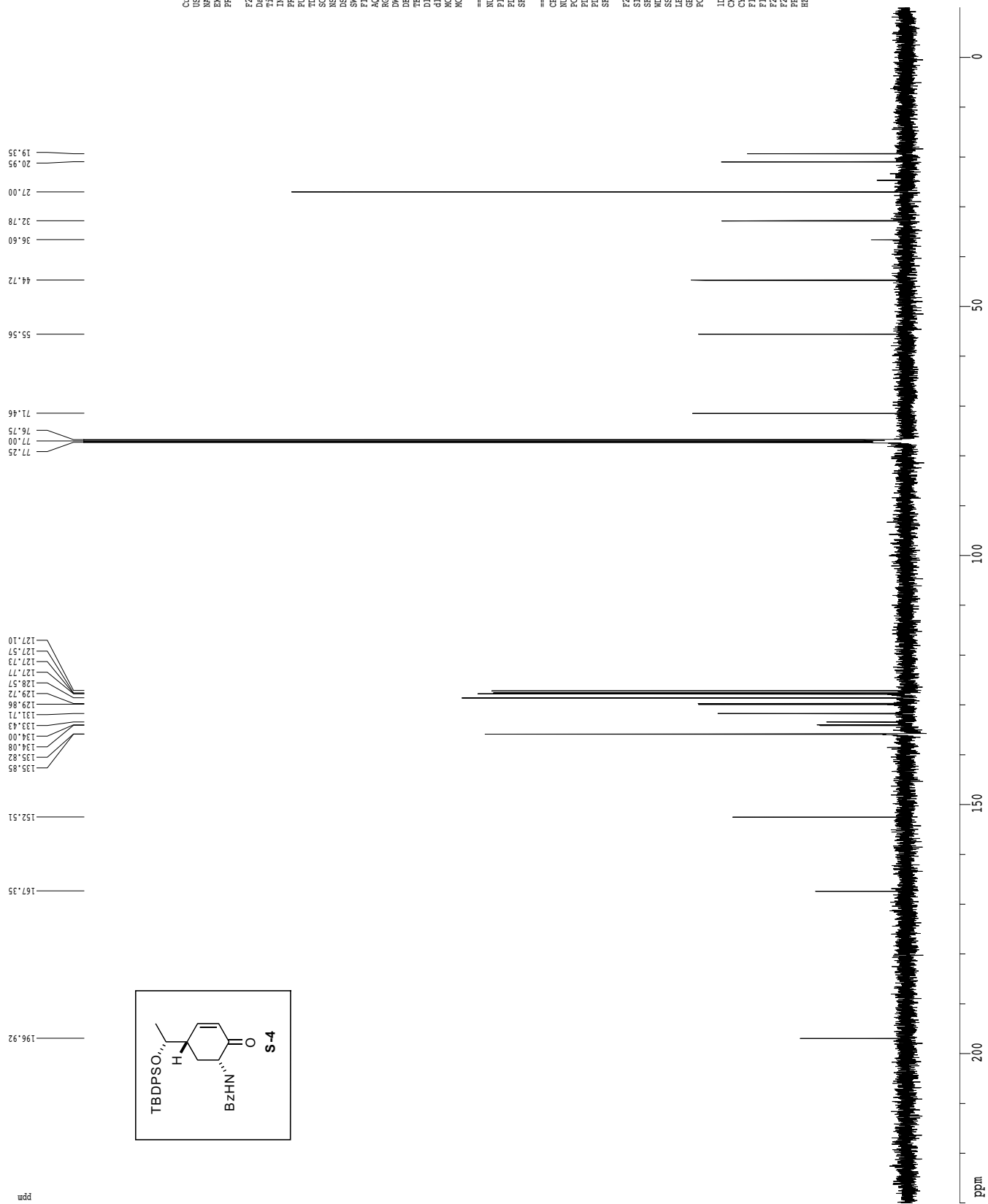
 F2 - Processing parameters
 SI 65536
 SF 600.1300282 MHz
 WDM EM 0
 LB 0.30 Hz
 GB 0
 EC 1.00

 D0 NMR plot parameters
 XZ 0.00 cm
 CZ 0.00 cm
 F1P 8.000 PPM
 F1 5401.17 Hz
 F2P 0.000 PPM
 F2 0.00 Hz
 FPMCH 0.39474 PPM/cm
 HZCM 236.89343 Hz/cm

¹³C spectrum with ¹H decoupling



¹³C spectrum with ¹H decoupling



```

Current Data Parameters
=====
USER          TD-2-192-3-2
NAME          TD-2-192-3-2
EXPNO        3
PROCNO       1

F2 - Acquisition Parameters
=====
Date_         20061206
Time         10:48
INSTRUM      cryo500
PROBHD       5 mm CPYAC
PULPROG      zgpg30
SFO1         500.225011 MHz
TD           65418
SOLVENT      CDCl3
NS           201
DS           4
SWH          30303.031 Hz
FIDRES      0.463222 Hz
AQ           1.079470 sec
RG           133.98
AQ           1.500 usec
DE           6.00 usec
TE           298.0 K
D1           0.2500000 sec
d11          0.0300000 sec
MCRETST     0.0000000 sec
MORPK       0.0150000 sec

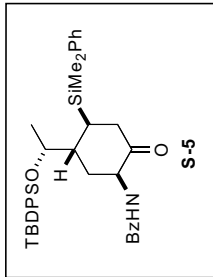
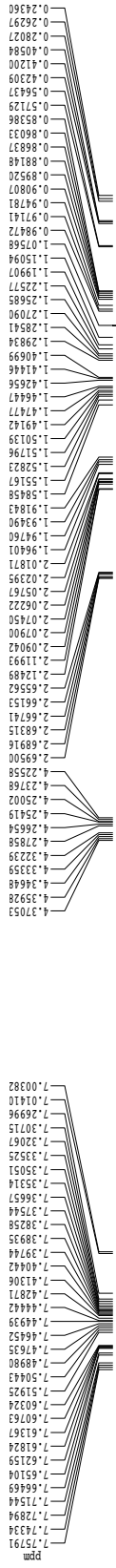
===== CHANNEL f1 =====
NUC1         13C
P1           15.00 usec
PL1          -1.00 dB
SFO1        125.7942548 MHz

===== CHANNEL f2 =====
CPDPRG2     waltz16
NUC2         1H
PCPD2       100.00 usec
PL2         1.60 dB
SFO2        500.225011 MHz

F2 - Processing parameters
=====
SI           65536
SF          125.7804382 MHz
WDW         EM
SSB         0
LB          1.00 Hz
GB          0
PC          2.00

ID NMR plot parameters
=====
CX          22.80 cm
CY          45.00 cm
F1P        230.000 ppm
F1         28929.50 Hz
F2P        -10.000 ppm
F2         -1257.80 Hz
FREQCH     10.3652 ppm/cm
HSCN       1324.00452 Hz/cm
    
```

¹H spectrum



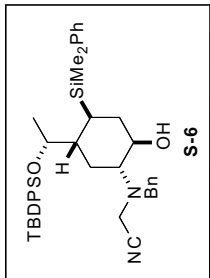
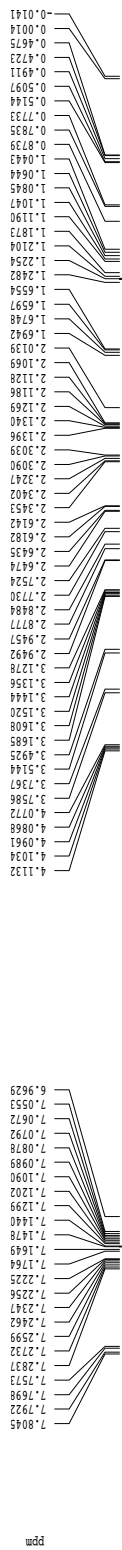
```

Current Data Parameters
=====
Date_ 20070227
Time_ 10:02
USER TD-2-255-5
EXPNO 5
PROCNO 1

F2 - Acquisition Parameters
=====
Date_ 20070227
Time_ 10:02
USER TD-2-255-5
EXPNO 5
PROCNO 1
PROBHD 5 mm CPYCI 1H-
PULPROG zg30
TD 81728
SOLVENT CDCl3
NS 8
DS 2
AQ 5.0988714 sec
RG 18
FIDRES 0.013572 Hz
AQRES 0.013572 Hz
SFO1 500.225015 MHz
NUC1 1H
P1 8.00 usec
PL1 1.60 dB
SFO2 500.225015 MHz
F2 - Processing parameters
=====
SI 45856
SF 500.220246 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 4.00

ID: NMR plot parameters
=====
CX 22.80 cm
CY 30.00 cm
FIP 9.000 ppm
F1 4501.98 Hz
F2 -1.000 ppm
FZ -500.22 Hz
PPMCH 0.43860 ppm/cm
HZCM 219.39419 Hz/cm
    
```


¹H spectrum



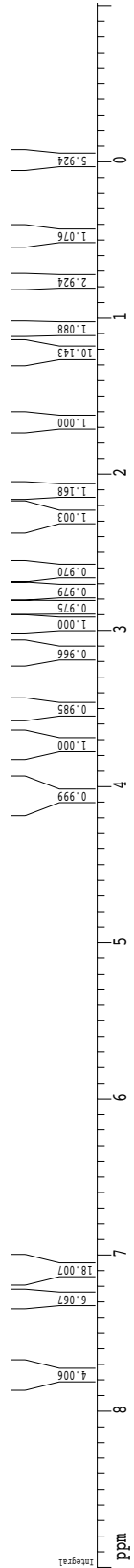
Current Data Parameters
 USER travis
 NAME TD-2-29-2-1
 EXVNO 101
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20070726
 Time_ 14:53
 INSTRUM av600
 PROBRD 5 mm TBI JH/13
 PULPROG zg30
 TD 97538
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 9615.74 Hz
 FWHM 0.08433 Hz
 AQ 5.0928259 sec
 RG 28.5
 DM 52.000 usec
 DE 6.00 usec
 TE 298.1 K
 D1 0.1000000 sec
 TDO 1

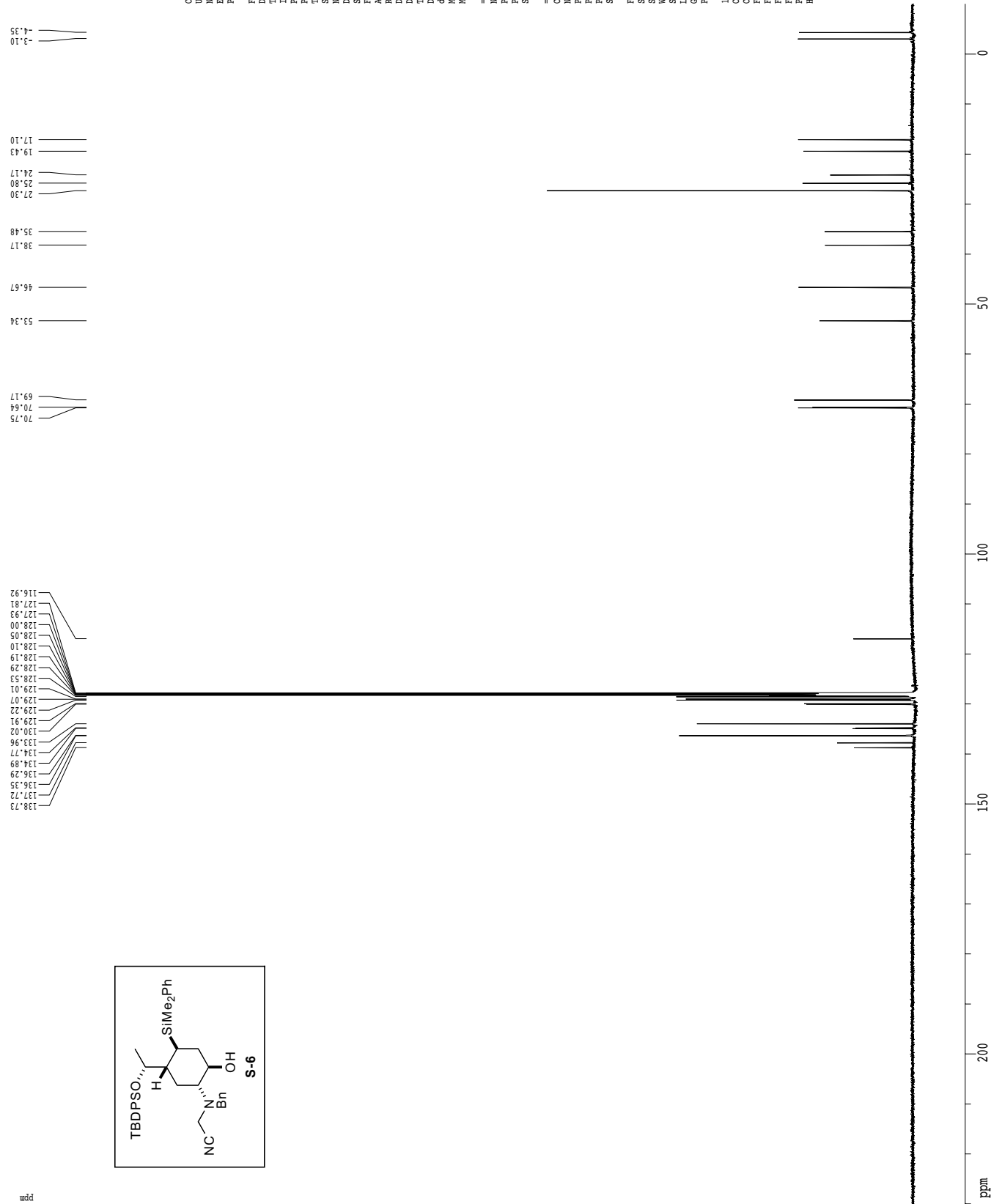
===== CHANNEL f1 =====
 NUC1 ¹H
 P1 8.00 usec
 PL1 -1.00 dB
 SFO1 600.1342009 MHz

F2 - Processing parameters
 SI 65536
 SF 600.1300040 MHz
 EQ
 ZG 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 22.80 cm
 CY 30.00 cm
 FIP 9.000 ppm
 F1 5401.17 Hz
 F2 5401.17 Hz
 PPM -610.78 ppm
 PPMCM 0.43937 ppm/cm
 HZCM 263.68207 Hz/cm



13C spectrum with 1H decoupling



Current Data Parameters
 USER travis
 NAME TD-2-249-2
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20070206
 Time_ 14.05
 INSTRUM ctyo500
 PROBRD 5 mm CPFI 1H-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 213
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813940 sec
 RG 3251
 DW 16.500 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.5000000 sec
 d11 0.0000000 sec
 ACQRES 0.0000000 sec
 NUC1 13C
 NUC2 1H
 SFO1 125.7942548 MHz
 SFO2 500.2225011 MHz

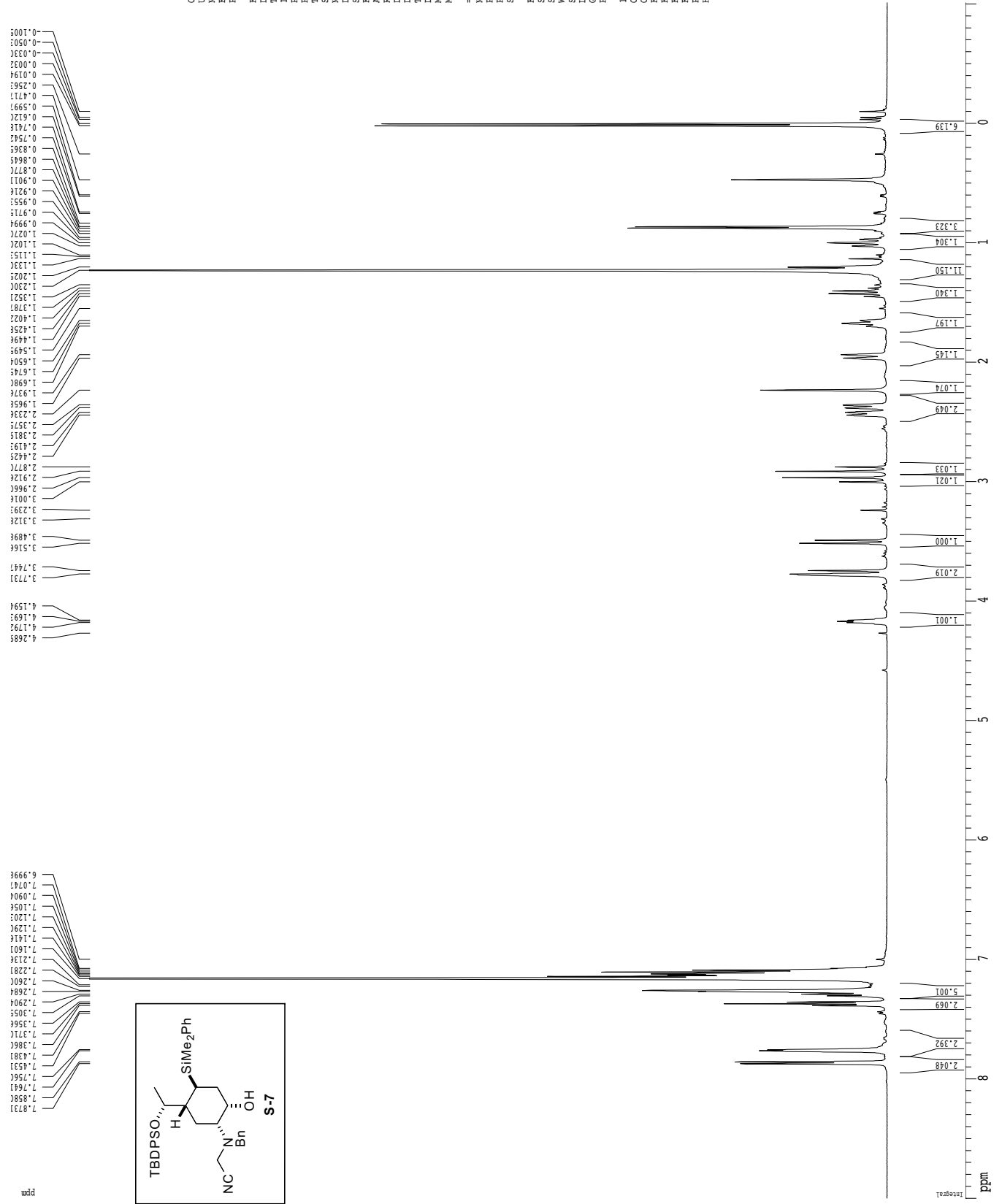
==== CHANNEL f1 =====
 NUC1 13C
 P1 15.00 usec
 PL1 -1.00 dB
 SFO1 125.7942548 MHz

==== CHANNEL f2 =====
 NUC2 1H
 P2 100.00 usec
 PL2 1.60 dB
 SFO2 500.2225011 MHz

F2 - Processing parameters
 SI 65536
 SF 125.7803656 MHz
 RM 0
 GB 0
 LB 1.00 Hz
 PC 2.00

ID NMR plot parameters
 CC 22.80 cm
 CD 24.00 cm
 CL 24.00 cm
 FI 28929.49 Hz
 F2 -10.000 ppm
 F2 -1257.80 Hz
 FPMCM 10.52632 ppm/cm
 HZCM 1324.00403 Hz/cm

1H spectrum



Current Data Parameters
 USER travis
 NAME TD-3-102-2
 EXTNO 1
 PROCNO 1

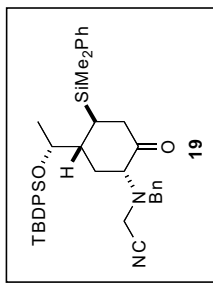
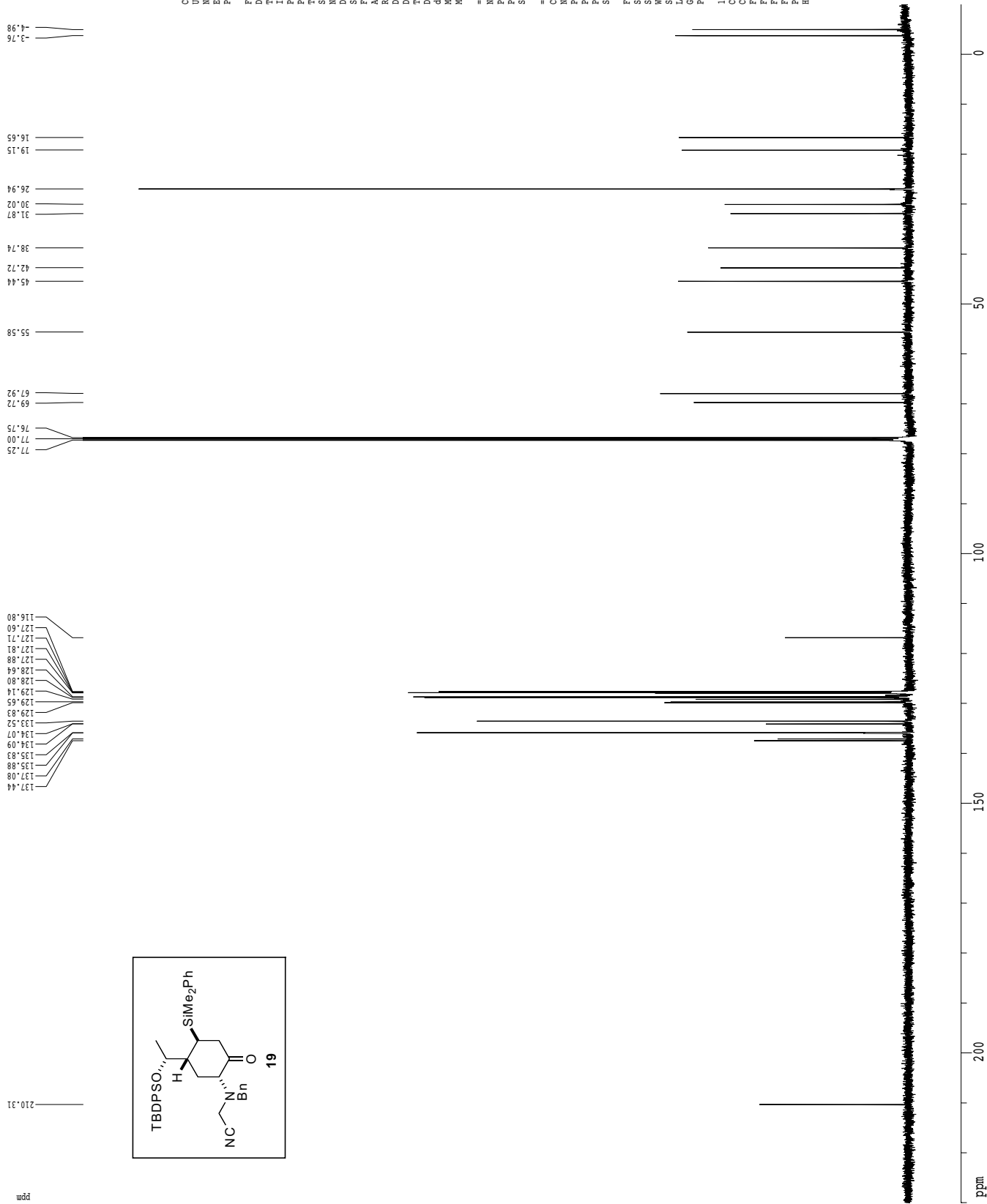
F2 - Acquisition Parameters
 Date_ 20070709
 Time_ 14:54
 INSTRUM cxts500
 PROBRD 5 mm CPYCI 1H-
 PULPROG zg30
 TD 81728
 SOLVENT C6D6
 NS 8
 DS 2
 SWH 8012.80 Hz
 FWHM 0.048803 Hz
 AQ 5.0988774 sec
 RG 5.7
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.1000000 sec
 ACQST 0.0100000 sec
 PCWAK 0.0130000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 8.00 usec
 PL1 1.60 dB
 SFO1 500.2250015 MHz

F2 - Processing parameters
 SI 65536
 SF 500.2260000 MHz
 WDW EM
 SSF 0
 SSB 0
 LB 0.30 Hz
 GB 0
 CB 4.00

ID: NMR Plot parameters
 CX 22.80 cm
 CY 30.00 cm
 FIP 9.000 ppm
 F1 4501.98 Hz
 F2 -1.009 ppm
 F2 -504.87 Hz
 PPMXN 0.43900 ppm/cm
 HZCMX 213.39680 Hz/cm

¹³C spectrum with ¹H decoupling



```

Current Data Parameters
USER          ctavis
NAME         TD-3-286-1
EXPNO        4
PROCNO       1

F2 - Acquisition Parameters
Date_        20080314
Time         17.03
INSTRUM      cryo500
PROBHD       5 mm CPCT 1H-
PULPROG      zgpg30
GAMMA        13
SOLVENT      CDCl3
NS           245
DS           4
SWH          30303.031 Hz
FIDRES       0.462388 Hz
AQ           1.0813940 sec
RG           13004
DM           16.500 usec
DE           2.00 usec
TE           300.2 K
D1           0.5000000 sec
d11          0.0300000 sec
MCHEST       0.0000000 sec
MCPRK        0.0150000 sec

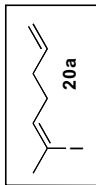
===== CHANNEL f1 =====
NUC1          13C
P1           14.75 usec
PL1          0.00 dB
SFO1         125.7945248 MHz

===== CHANNEL f2 =====
CPDPRG2      waltz16
NUC2          1H
PCPD2        100.00 usec
PCPD2        1.60 dB
PL2          24.80 dB
SFO2         500.2250111 MHz

F2 - Processing parameters
SI           65536
SF           125.7804309 MHz
WDW          EM
SSB          0
LB           1.00 Hz
GB           0
EC           2.00

ID_NMR Plot parameters
CY          32.80 cm
CY          30.00 cm
F1P         230.000 ppm
F2P         28929.50 Hz
F2P         -10.000 ppm
F2          -1257.80 Hz
PPMXX       10.52632 ppm/cm
HZCX        1324.00464 Hz/cm
    
```


¹H spectrum



Current Data Parameter
 USER mellis
 NAME ME-II-68-ch
 EXPNO 1
 PROCNO 1

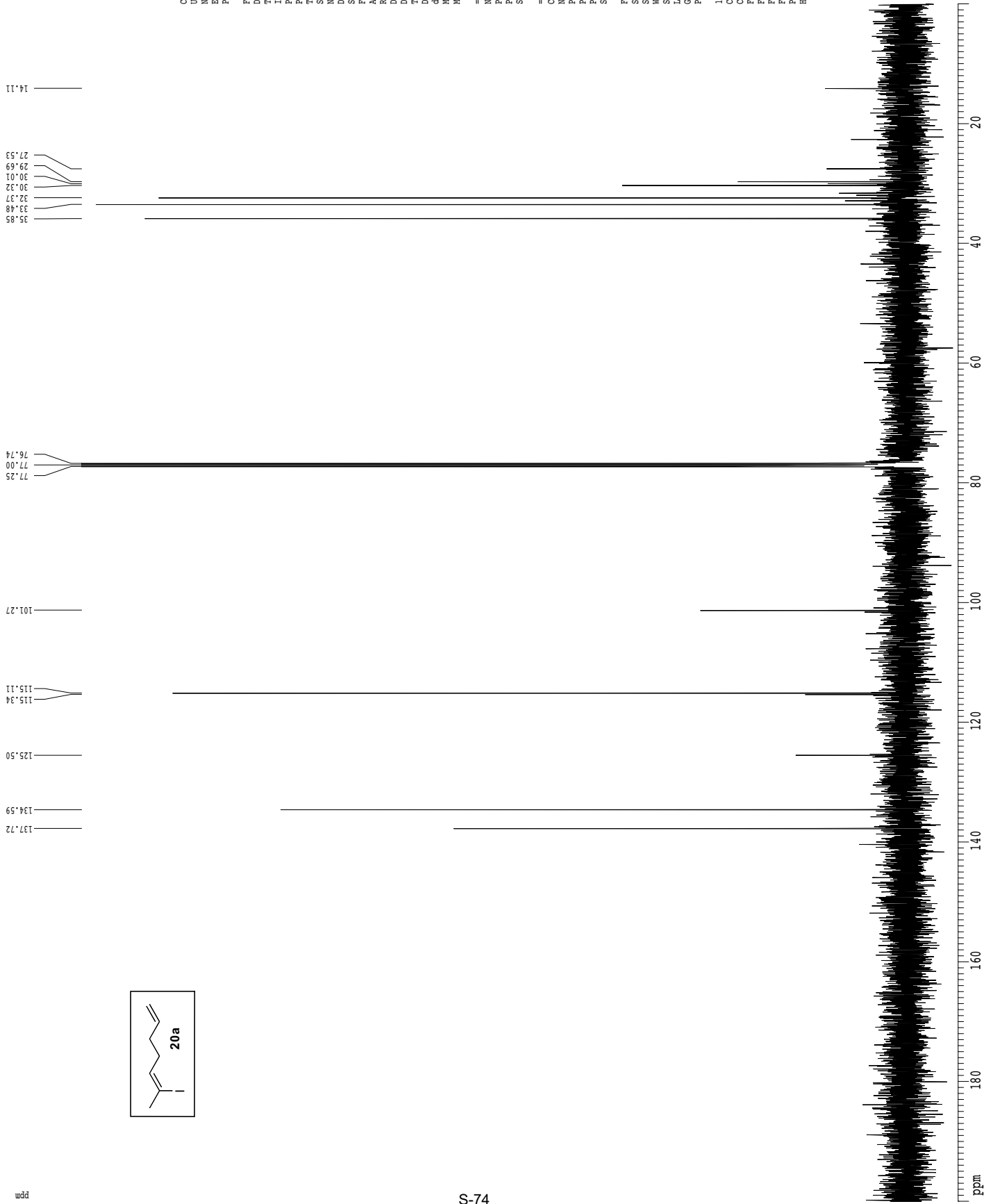
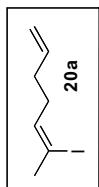
F2 - Acquisition Parameters
 Date_ 20090511
 Time 10.31
 INSTRUM gn500
 PULPROG 5 mm broadband
 FIDRES 81.728
 TD 65536
 SOLVENT DMSO
 NS 16
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.099874 sec
 RG 128
 DM 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.100000 sec
 d11 0.050000 sec
 ACQRES 0.0150000 sec
 MCPRK 0.0150000 sec

==== CHANNEL f1 =====
 NUC1 ¹H
 P1 12.00 usec
 PL1 -3.00 dB
 SFO1 499.723480 MHz

F2 - Processing parameter
 SI 65536
 SF 499.7200262 MHz
 EQ
 ASB 0
 GB 0.20 Hz
 GR 0
 GA 0
 GB 0
 GC 1.00

1D NMR plot parameters
 CX 22.80 cm
 CY 15.00 cm
 CZ 8.000 ppm
 F1P 3997.76 Hz
 F2P 0.000 ppm
 F3P 0.000 Hz
 PPRCM -0.35088 ppm/cm
 HZCM 175.34036 Hz/cm

¹³C spectrum with ¹H decoupling



Current Data Parameters
 USER mellis
 NAME ME-II-68-ch
 EXENO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20090511
 Time 10.38
 INSTRUM gn500
 PROBD 5 mm broadband
 PULPROG zgpg30
 TD 65536
 SOLVENT CCL3
 NS 331
 DS 4
 SWH 30303.031 Hz
 SF 0.462388 Hz
 FIDRES 1.0813846 sec
 RG 6502
 DM 16.500 usec
 DE 4.50 usec
 TE 298.0 K
 D1 0.25000000 sec
 d11 0.03000000 sec
 ACQRES 0.00000000 sec
 MCPRK 0.01500000 sec

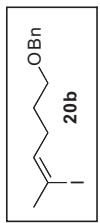
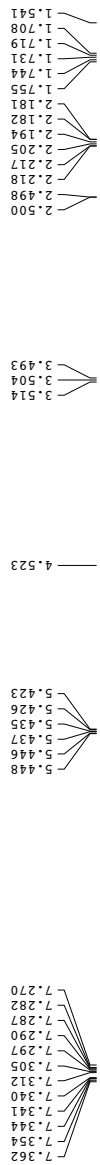
===== CHANNEL f1 =====
 NUC1 ¹³C
 P1 7.08 usec
 PL1 0.00 dB
 SFO1 125.668160 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 ¹H
 P2 80.00 usec
 PL2 -3.00 dB
 PL12 14.70 dB
 SFO2 499.7224986 MHz

F2 - Processing parameter
 S1 65536
 SF 125.6547018 MHz
 EM
 WDM 0
 LB 1.00 Hz
 GB 0
 FC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CY 15.65 cm
 F1P 200.000 ppm
 F2P 25130.94 Hz
 F2 0.000 ppm
 F2 0.00 Hz
 FWHM 8.77193 ppm/cm
 HZCM 1102.23425 Hz/cm

1H spectrum



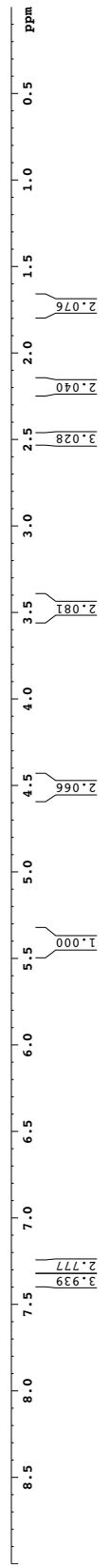
```

Current Data Parameters
USER      travis
NAME      TD 4 51 4
EXPNO     101
PROCNO    1

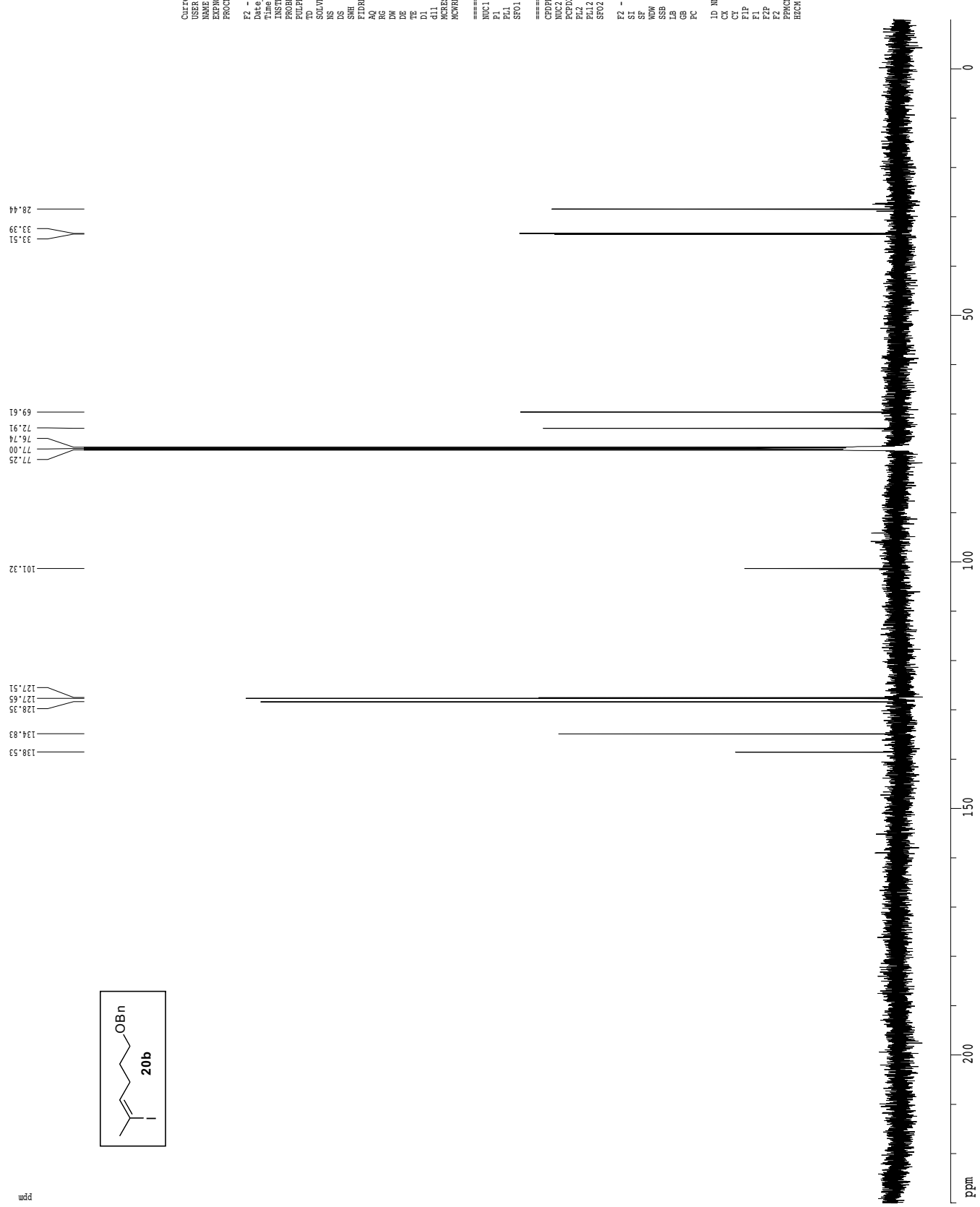
F2 Acquisition Parameters:
Date_     20080326
Time_     12.00
INSTRUM   av600
PROBHD    5 mm BBO BB 1H
PULPROG   zg30
TD         97938
SOLVENT   CDCl3
NS         40
DS         4
SWH        9615.385 Hz
FIDRES     0.098178 Hz
AQ         5.0928259 sec
RG         812
DM         52.000 us
DE         6.00 us
TE         298.0 K
D1         0.10000000 sec
TD0        1

===== CHANNEL f1 =====
NUC1       1H
P1         9.00 us
PL1        5.30 dB
SFO1       600.1342009 MHz

F2 Processing parameters
SI         6556
SF         600.1300274 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00
    
```



¹³C spectrum with ¹H decoupling



```

Current Data Parameters
Name      TD-1-18-1
EXPNO    2
PROCNO   1

F2 - Acquisition Parameters
Date_    20070815
Time     14.05
INSTRUM  crysov0
PROBHD   5 mm CP131
PULPROG  zgpg30
TD        65536
SOLVENT  CDCl3
NS        189
DS        4
SWH       30303.021 Hz
FIDRES    0.462388 Hz
AQ        1.0813940 sec
RG        10221.3
RW        16.00 usec
DE        6.00 usec
TE        298.0 K
D1        1.5000000 sec
d11       0.0300000 sec
MCREST    0.0000000 sec
MCWPRK    0.0150000 sec

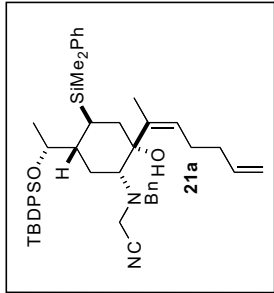
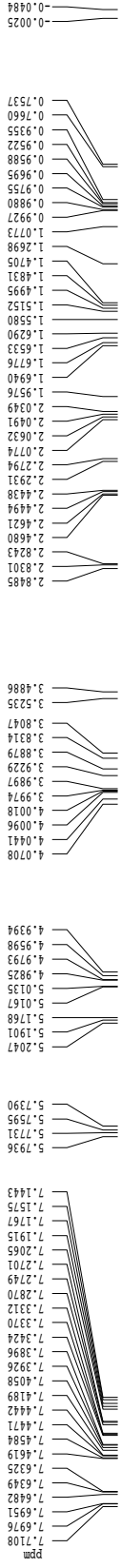
===== CHANNEL f1 =====
NUC1      13C
P1        18.00 usec
PL1       -1.00 dB
SFO1      125.7942548 MHz

===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2      1H
PCPD2     100.00 usec
PL2       1.60 dB
PL12      21.54 dB
SFO2      500.222011 MHz

F2 - Processing Parameters
SI        65536
SF        125.7804282 MHz
WDW       EM
SSB       0
LB        1.00 Hz
GB        0
PC        2.00

1D NMR plot parameters
CX        22.80 cm
CY        100.00 cm
F1P       230.000 ppm
F2P       -10.000 ppm
F2        -1257.80 Hz
PROCNO    10158632 ppm/cm
RG        1324.00452 Hz/cm
    
```

¹H spectrum



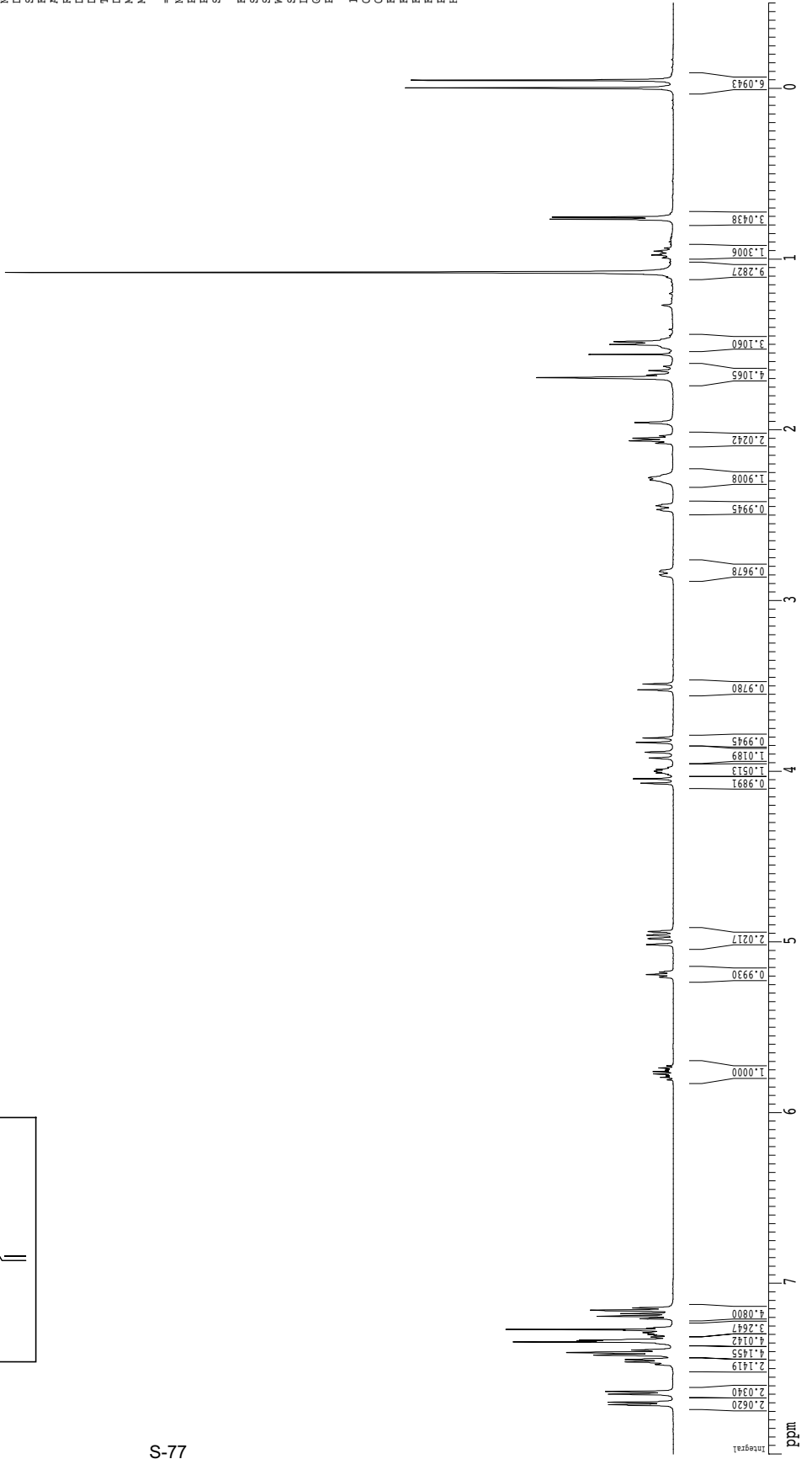
Current Data Parameter
 USER mellis
 NAME kh-1-124
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20090306
 Time 11:04
 INSTRUM cryo500
 PROBRD 5 mm CPCL1 H-
 PULPROG zgpg30
 F4 8178
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.098874 sec
 RG 10.1
 DM 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.1000000 sec
 ACQRES 0.0000000 sec
 SCANS 0.0130000 sec

==== CHANNEL f1 =====
 NUC1 ¹H
 P1 7.50 usec
 PL1 1.60 dB
 SF01 500.2235015 MHz

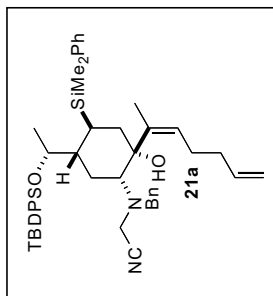
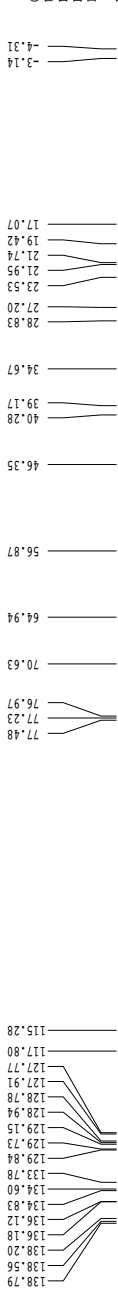
F2 - Processing parameter
 S1 65536
 SF 500.2200269 MHz
 EN
 WDW EM
 SSB 0
 CB 0
 GB 0
 PC 4.00

1D NMR plot parameters
 CX 22.80 cm
 CY 10.51 cm
 F1P 8.000 ppm
 F2 4001.76 Hz
 F2P -0.500 ppm
 F2 -250.11 Hz
 FWHM 0.37281 ppm/cm
 HZCX 186.48555 Hz/cm



13C spectrum with 1H decoupling

wdd



```

Current Data Parameters
USER      mellis
NAME      Kh-1-123d
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     20090306
Time      11.08
INSTRUM   cryo500
PROBHD    5 mm CPCTC 1H-
PULPROG   SpinEcho90gp-prd
TD         65536
SOLVENT   CDCl3
NS         1024
DS         16
SWH        30803.031 Hz
FIDRES     0.462388 Hz
AQ         1.0813940 sec
RG         7288.2
DM         16.500 usec
DE         6.00 usec
TE         298.0 K
D1         0.1500000 sec
d11        0.0100000 sec
d16        0.0002000 sec
d17        0.0001960 sec
MCREST    0.0000000 sec
MCWRK     0.0150000 sec
F2         29.70 usec

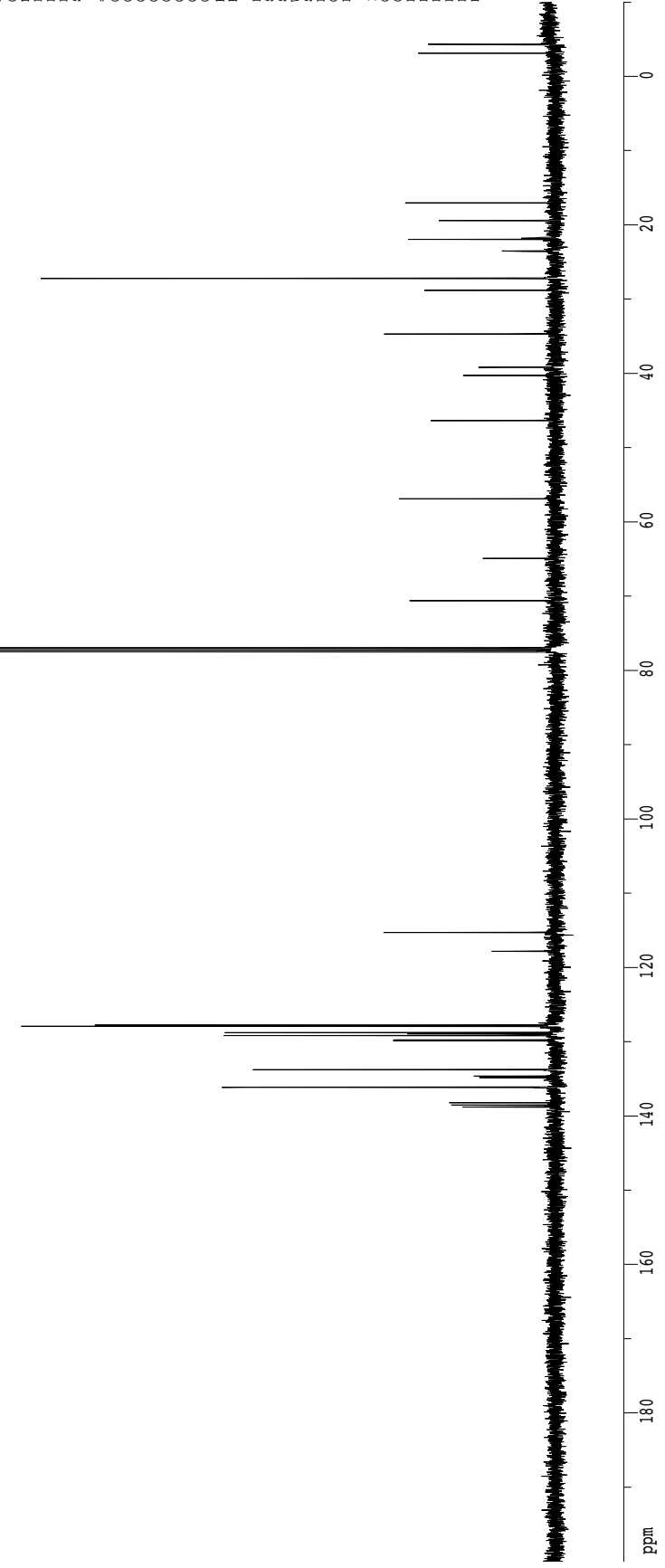
===== CHANNEL f1 =====
NUC1       13C
P1         14.85 usec
PL1        0.00 usec
P12        200.00 usec
PL2        12.00 dB
PL11       -1.00 dB
SFO1       125.7942548 MHz
SFO2       3.60 dB
SFO3       3.60 dB
SFO4       3.60 dB
SFO5       3.60 dB
SFO6       3.60 dB
SFO7       3.60 dB
SFO8       3.60 dB
SFO9       3.60 dB
SFO10      3.60 dB
SFO11      3.60 dB
SFO12      3.60 dB
SFO13      3.60 dB
SFO14      3.60 dB
SFO15      3.60 dB
SFO16      3.60 dB
SFO17      3.60 dB
SFO18      3.60 dB
SFO19      3.60 dB
SFO20      3.60 dB
SFO21      3.60 dB
SFO22      3.60 dB
SFO23      3.60 dB
SFO24      3.60 dB
SFO25      3.60 dB
SFO26      3.60 dB
SFO27      3.60 dB
SFO28      3.60 dB
SFO29      3.60 dB
SFO30      3.60 dB
SFO31      3.60 dB
SFO32      3.60 dB
SFO33      3.60 dB
SFO34      3.60 dB
SFO35      3.60 dB
SFO36      3.60 dB
SFO37      3.60 dB
SFO38      3.60 dB
SFO39      3.60 dB
SFO40      3.60 dB
SFO41      3.60 dB
SFO42      3.60 dB
SFO43      3.60 dB
SFO44      3.60 dB
SFO45      3.60 dB
SFO46      3.60 dB
SFO47      3.60 dB
SFO48      3.60 dB
SFO49      3.60 dB
SFO50      3.60 dB
SFO51      3.60 dB
SFO52      3.60 dB
SFO53      3.60 dB
SFO54      3.60 dB
SFO55      3.60 dB
SFO56      3.60 dB
SFO57      3.60 dB
SFO58      3.60 dB
SFO59      3.60 dB
SFO60      3.60 dB
SFO61      3.60 dB
SFO62      3.60 dB
SFO63      3.60 dB
SFO64      3.60 dB
SFO65      3.60 dB
SFO66      3.60 dB
SFO67      3.60 dB
SFO68      3.60 dB
SFO69      3.60 dB
SFO70      3.60 dB
SFO71      3.60 dB
SFO72      3.60 dB
SFO73      3.60 dB
SFO74      3.60 dB
SFO75      3.60 dB
SFO76      3.60 dB
SFO77      3.60 dB
SFO78      3.60 dB
SFO79      3.60 dB
SFO80      3.60 dB
SFO81      3.60 dB
SFO82      3.60 dB
SFO83      3.60 dB
SFO84      3.60 dB
SFO85      3.60 dB
SFO86      3.60 dB
SFO87      3.60 dB
SFO88      3.60 dB
SFO89      3.60 dB
SFO90      3.60 dB
SFO91      3.60 dB
SFO92      3.60 dB
SFO93      3.60 dB
SFO94      3.60 dB
SFO95      3.60 dB
SFO96      3.60 dB
SFO97      3.60 dB
SFO98      3.60 dB
SFO99      3.60 dB
SFO100     3.60 dB

===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2       1H
P2         100.00 usec
PL2        0.00 dB
PL12       24.60 dB
SFO2       500.2225011 MHz

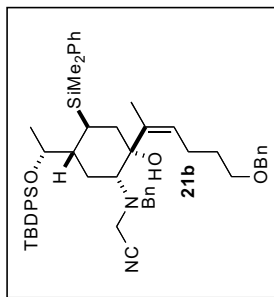
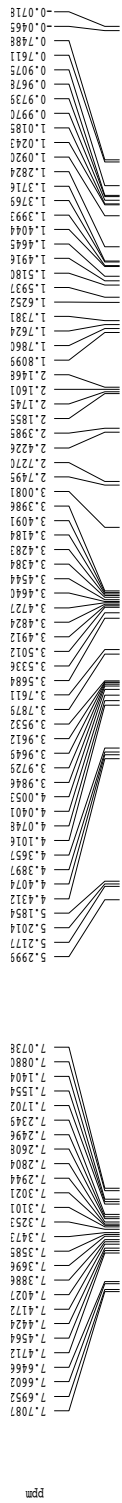
===== GRADIENT CHANNEL =====
GPMAM1     SINE.100
GPMAM2     SINE.100
GPR1       0.00 %
GPR2       0.00 %
GPR3       0.00 %
GPR4       0.00 %
GPR5       0.00 %
GPR6       0.00 %
GPR7       0.00 %
GPR8       0.00 %
GPR9       0.00 %
GPR10      0.00 %
GPR11      0.00 %
GPR12      0.00 %
GPR13      0.00 %
GPR14      0.00 %
GPR15      0.00 %
GPR16      0.00 %
GPR17      0.00 %
GPR18      0.00 %
GPR19      0.00 %
GPR20      0.00 %
GPR21      0.00 %
GPR22      0.00 %
GPR23      0.00 %
GPR24      0.00 %
GPR25      0.00 %
GPR26      0.00 %
GPR27      0.00 %
GPR28      0.00 %
GPR29      0.00 %
GPR30      0.00 %
GPR31      0.00 %
GPR32      0.00 %
GPR33      0.00 %
GPR34      0.00 %
GPR35      0.00 %
GPR36      0.00 %
GPR37      0.00 %
GPR38      0.00 %
GPR39      0.00 %
GPR40      0.00 %
GPR41      0.00 %
GPR42      0.00 %
GPR43      0.00 %
GPR44      0.00 %
GPR45      0.00 %
GPR46      0.00 %
GPR47      0.00 %
GPR48      0.00 %
GPR49      0.00 %
GPR50      0.00 %
GPR51      0.00 %
GPR52      0.00 %
GPR53      0.00 %
GPR54      0.00 %
GPR55      0.00 %
GPR56      0.00 %
GPR57      0.00 %
GPR58      0.00 %
GPR59      0.00 %
GPR60      0.00 %
GPR61      0.00 %
GPR62      0.00 %
GPR63      0.00 %
GPR64      0.00 %
GPR65      0.00 %
GPR66      0.00 %
GPR67      0.00 %
GPR68      0.00 %
GPR69      0.00 %
GPR70      0.00 %
GPR71      0.00 %
GPR72      0.00 %
GPR73      0.00 %
GPR74      0.00 %
GPR75      0.00 %
GPR76      0.00 %
GPR77      0.00 %
GPR78      0.00 %
GPR79      0.00 %
GPR80      0.00 %
GPR81      0.00 %
GPR82      0.00 %
GPR83      0.00 %
GPR84      0.00 %
GPR85      0.00 %
GPR86      0.00 %
GPR87      0.00 %
GPR88      0.00 %
GPR89      0.00 %
GPR90      0.00 %
GPR91      0.00 %
GPR92      0.00 %
GPR93      0.00 %
GPR94      0.00 %
GPR95      0.00 %
GPR96      0.00 %
GPR97      0.00 %
GPR98      0.00 %
GPR99      0.00 %
GPR100     0.00 %

F2 - Processing parameters
SI         65536
SF         125.7804000 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00

ID NMR plot parameters
CX         72.51 cm
CY         1.50 cm
CZ         1.50 cm
F1P        200.000 ppm
F1         25156.08 Hz
F2P        -10.000 ppm
F2         -1257.80 Hz
PPMCK      9.21053 ppm/cm
HZCK       1158.50366 Hz/cm
    
```

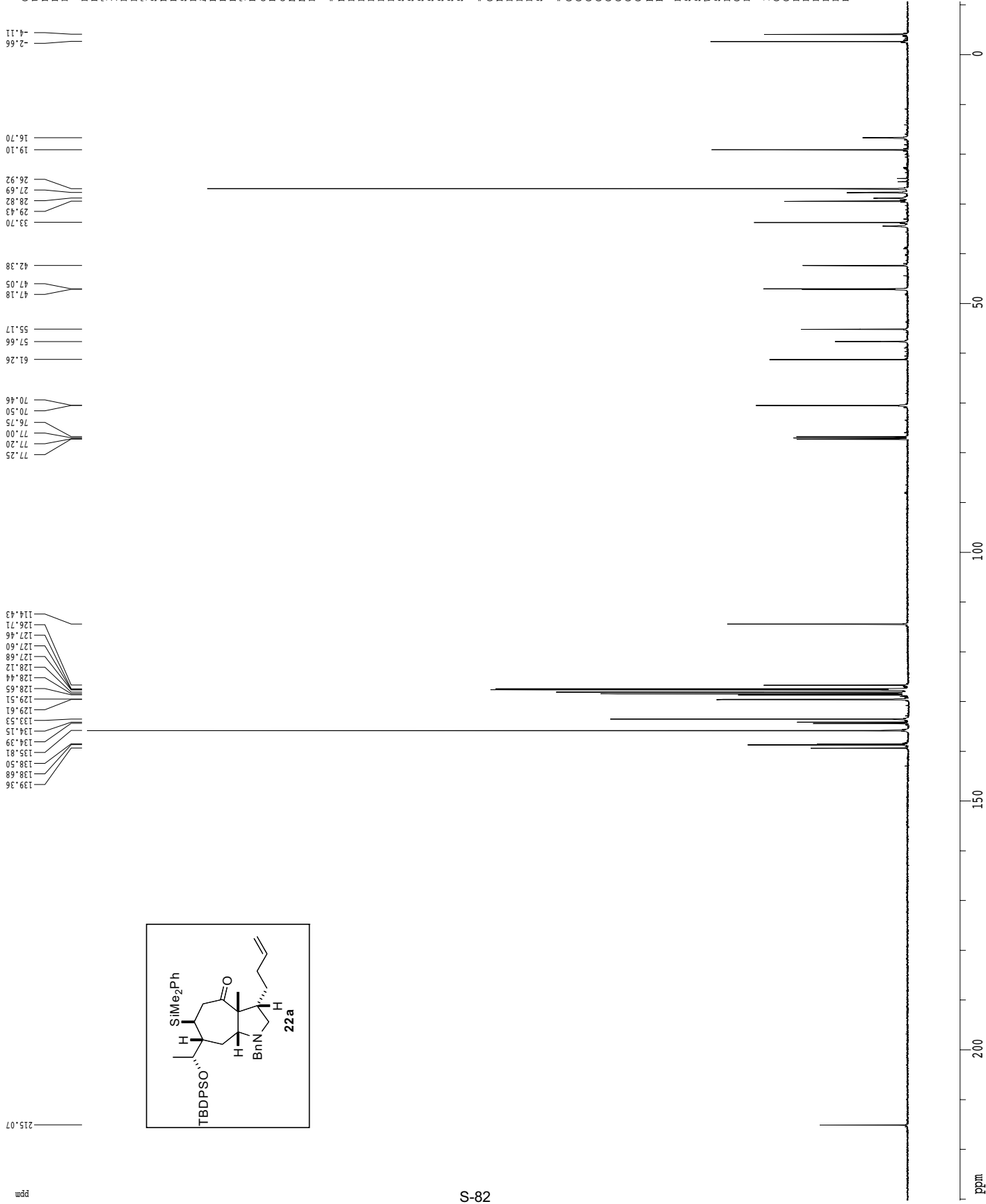


1H spectrum



Current Data Parameters
 USER travis
 NAME TD-3-155-3
 PROCNO 1
 F2 - Acquisition Parameters
 Date 20070829
 Time 14.14
 INSTRUM cryo500
 PROBD 5 mm CPXI H-
 PULPROG zgpg30
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.096043 Hz
 AQ 5.099774 sec
 RG 316
 DE 6.00 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCWRR 0.01580000 sec
 ===== CHANNEL f1 =====
 NUCL1 1H
 P1 8.00 usec
 PL1 1.60 dB
 SFO1 500.2235015 MHz
 F2 - Processing parameters
 SI 65536
 SF 500.2200303 MHz
 SW 4000
 GB 0.30 Hz
 CB 0
 PC 4.00
 LD NMR PLOT parameters
 CX 22.80 cm
 CY 15.00 cm
 CZ 4.00 cm
 FL 4501.98 Hz
 FR 0.00000000 Hz
 F2P -1.070 ppm
 F2 -535.21 Hz
 PPMCK 0.44166 ppm/cm
 HZCK 220.82926 Hz/cm

13C spectrum with 1H decoupling



```

Current Data Parameters
USER          mellis
NAME          MP-1--22
EXPNO        2
PROCNO       1

F2 - Acquisition Parameters
Date_        20081108
Time         11.26
INSTRUM      cryo500
PROBHD       5 mm CPYCT 1H-
PULPROG      Spinechop93gp.prd
TD           65536
SOLVENT      CDCl3
NS           2048
DS           4
SWH           30303.031 Hz
FIDRES       0.462388 Hz
AQ           1.0814105 sec
RG           7298.2
DM           16.500 usec
DE           6.00 usec
TE           300.2 K
D1           0.28000000 sec
d11          0.03000000 sec
d16          0.00020000 sec
d17          0.00019600 sec
MCWREST      0.00000000 sec
MCWREK       0.01500000 sec
P2           29.70 usec

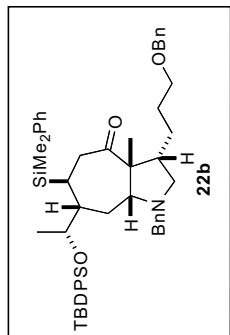
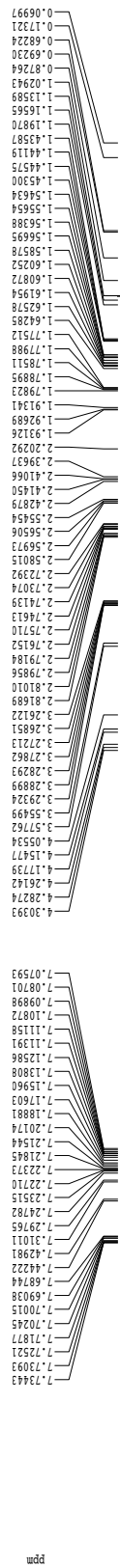
===== CHANNEL f1 =====
NUC1         13C
P1           14.85 usec
P11          500.00 usec
P12          2000.00 usec
PL0          120.00 dB
PL1          -1.00 dB
SFO1         125.7942548 MHz
SF1          3.60 dB
SFO2         0.0000000 MHz
SF2          3.60 dB
SFO3         0.0000000 MHz
SF3          3.60 dB
SFO4         0.0000000 MHz
SF4          3.60 dB
SFO5         0.0000000 MHz
SF5          3.60 dB
SFO6         0.0000000 MHz
SF6          3.60 dB
SFO7         0.0000000 MHz
SF7          3.60 dB
SFO8         0.0000000 MHz
SF8          3.60 dB
SFO9         0.0000000 MHz
SF9          3.60 dB
SFO10        0.0000000 MHz
SF10         3.60 dB
SFO11        0.0000000 MHz
SF11         3.60 dB
SFO12        0.0000000 MHz
SF12         3.60 dB
SFO13        0.0000000 MHz
SF13         3.60 dB
SFO14        0.0000000 MHz
SF14         3.60 dB
SFO15        0.0000000 MHz
SF15         3.60 dB
SFO16        0.0000000 MHz
SF16         3.60 dB
SFO17        0.0000000 MHz
SF17         3.60 dB
SFO18        0.0000000 MHz
SF18         3.60 dB
SFO19        0.0000000 MHz
SF19         3.60 dB
SFO20        0.0000000 MHz
SF20         3.60 dB

===== CHANNEL f2 =====
CPDPRG2      waltz16
NUC2         1H
P2           100.00 usec
P21          2000.00 usec
P22          2000.00 usec
PL2          120.00 dB
PL12         24.60 dB
SFO2         500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM1       SINE.100
GPNAM2       SINE.100
GPR1         0.00 %
GPR2         0.00 %
GPR3         0.00 %
GPR4         0.00 %
GPR5         0.00 %
GPR6         0.00 %
GPR7         0.00 %
GPR8         0.00 %
GPR9         0.00 %
GPR10        0.00 %
GPR11        0.00 %
GPR12        0.00 %
GPR13        0.00 %
GPR14        0.00 %
GPR15        0.00 %
GPR16        0.00 %
GPR17        0.00 %
GPR18        0.00 %
GPR19        0.00 %
GPR20        0.00 %

F2 - Processing parameters
SI           32768
SF           125.760462 MHz
WDW          EM
SSB          0
LB           1.00 Hz
GB           0
PC           2.00

ID NMR plot parameters
C1           12.65 cm
C2           12.65 cm
F1P          230.244 ppm
F1           28960.18 Hz
F2P          -10.676 ppm
F2           -1342.85 Hz
FPMCH        10.56667 ppm/cm
H2CN         1329.08044 Hz/cm
    
```



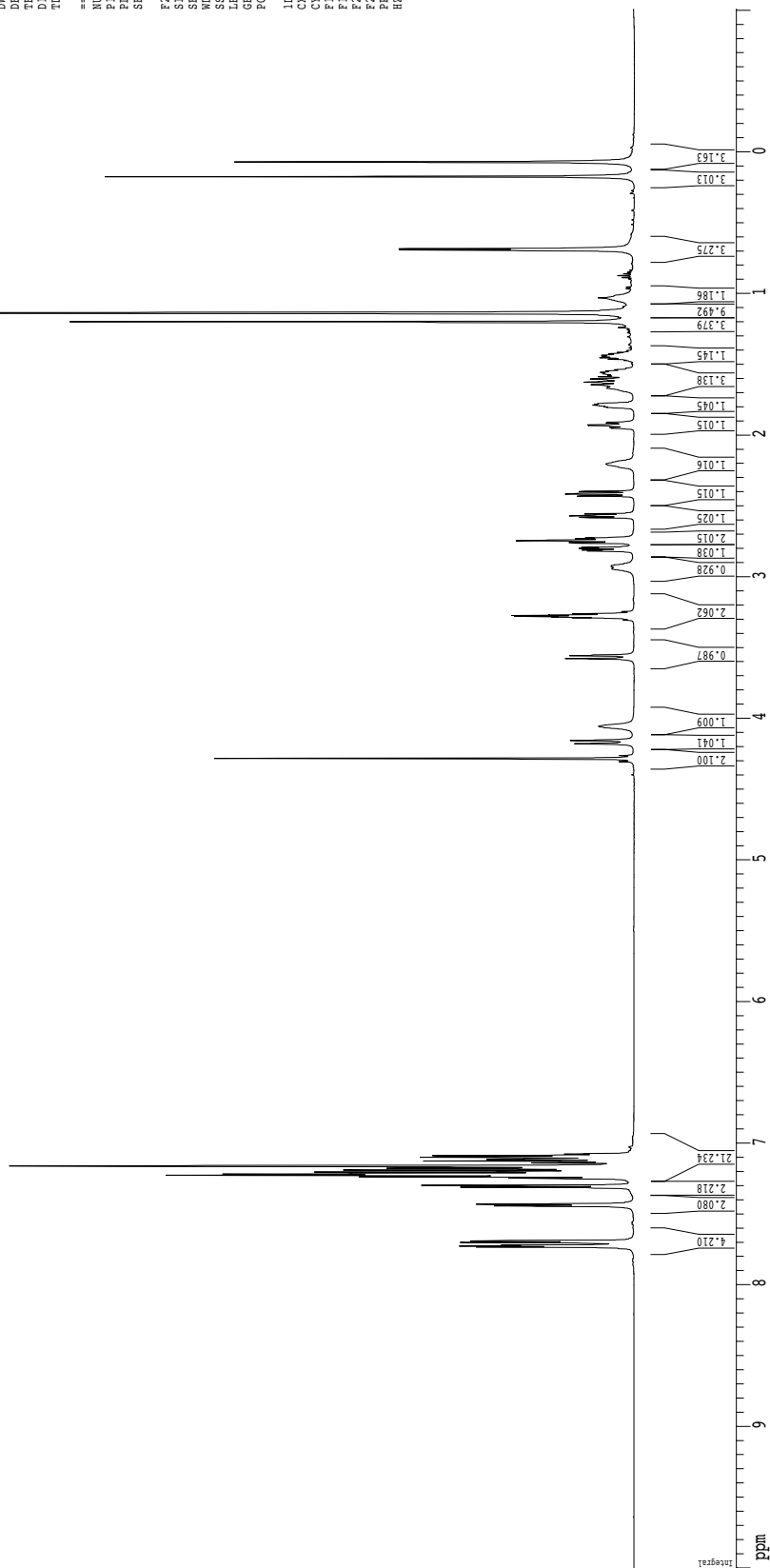
Current Data Parameters
 USER travis
 NAME TD-3-161-2
 EXNO 101
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20070901
 Time 10.39
 Date_ 09/01/07
 PROBNM 5 mm F31 1H/13
 PULPROG zgpg30
 TD 97938
 SOLVENT C6D6
 NS 8
 DS 2
 SWH 9615.385 Hz
 FIDRES 0.099178 Hz
 AQ 5.0928295 sec
 RG 655
 DW 52.000 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.10000000 sec
 TD0 1

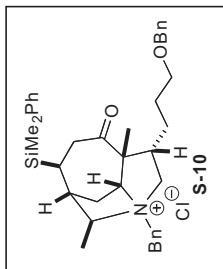
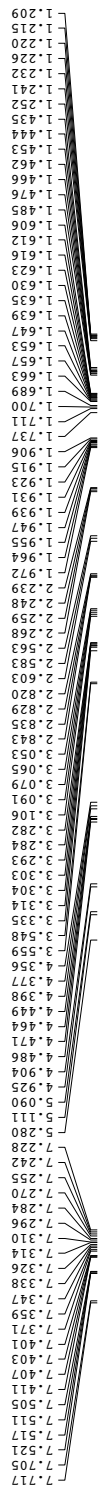
===== CHANNEL f1 =====
 NUCL1 1H
 P1 8.00 usec
 PL1 0.00 dB
 SFO1 600.1342009 MHz

F2 - Processing parameters
 SI 65536
 SF 600.1299978 MHz
 WDM EN
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 22.80 cm
 CY 30.00 cm
 FIP 10.000 ppm
 F1 6001.30 Hz
 F2P -1.007 ppm
 F2 -604.53 Hz
 FREQZ 0.00000000 Hz/cm
 HZCX 289.72961 Hz/cm



¹H spectrum



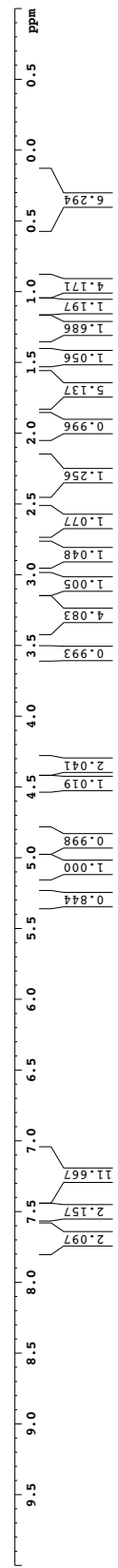
```

Current Data Parameters
USER      travis
NAME      TD 4 43 1
EXPNO     101
PROCNO    1

F2 Acquisition Parameter:
Date_     20080308
Time      16.38
INSTRUM   av600
PROBHD    5 mm BBO BB 1H
PULPROG   zg30
TD         97938
SOLVENT   CDCl3
NS         8
DS         4
SFO1      9615.385 Hz
FIDRES    0.098178 Hz
AQ         5.0928259 se
RG         128
DW         52.000 us
DE         6.00 us
TE         298.0 K
D1         0.10000000 se
TDO        1

===== CHANNEL f1 =====
NUC1       1H
P1         9.00 us
PL1        5.30 dB
SFO1      600.1342009 MH

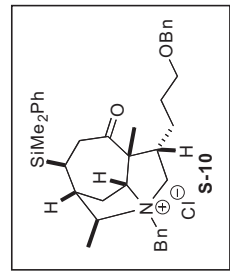
F2 Processing parameters
SI         65536
SF         600.1300283 MH
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00
    
```



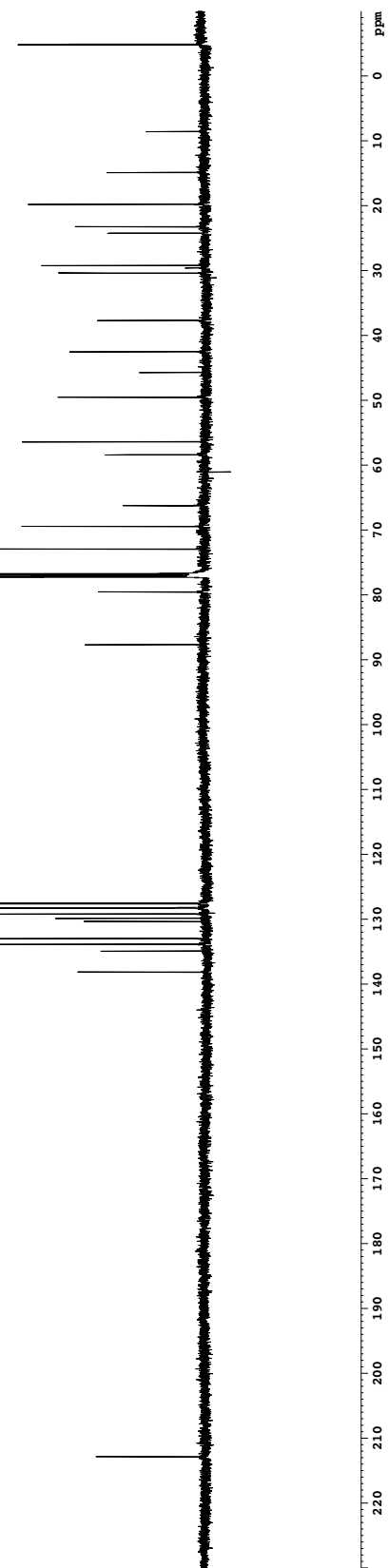
¹³C spectrum with ¹H decoupling

138.175
134.947
133.897
133.006
130.362
129.884
129.222
128.364
128.303
128.265
127.657
127.561

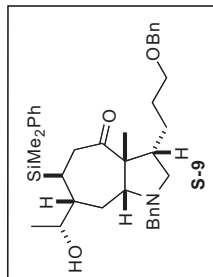
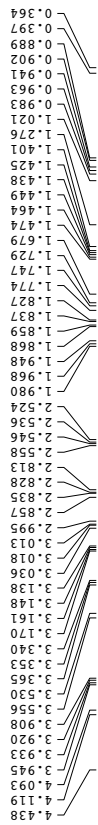
87.698
79.557
77.255
77.207
77.001
76.746
72.948
69.454
66.269
58.438
56.438
49.556
45.751
42.554
37.722
30.402
29.611
29.254
24.243
23.251
19.824
14.906
8.585
4.787
4.873



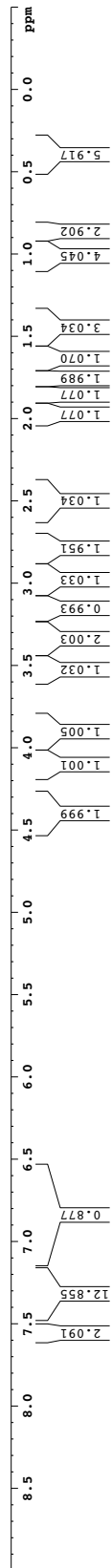
Current Data Parameters
 NSMR trav15
 EXPR TD 3 300 2
 PROCNO 1
 F2 Acquisition Parameters:
 Date_ 20080204
 Time 12.49
 INSTRUM cryo500
 PROBD 5 mm CPYCI 1H
 TOUFRQG zgpg30
 SOLVENT CDCl3
 NS 130
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813940 sec
 RG 11585.2
 DW 16.500 usec
 DE 660.0 usec
 TE 298.0 K
 D1 1.0000000 sec
 d11 0.0300000 sec
 MCREST 0.0000000 sec
 MCWRK 0.0150000 sec
 ===== CHANNEL f1 =====
 NUC1 ¹³C
 P1 15.00 usec
 PL1 0.00 dB
 SFO1 125.7942548 MHz
 ===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 ¹H
 PCPD2 100.00 usec
 PL2 1.60 dB
 PL12 23.54 dB
 SFO2 500.2225011 MHz
 F2 Processing Parameters
 SI 65536
 SF 125.7804346 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 FC 2.00



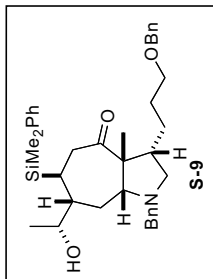
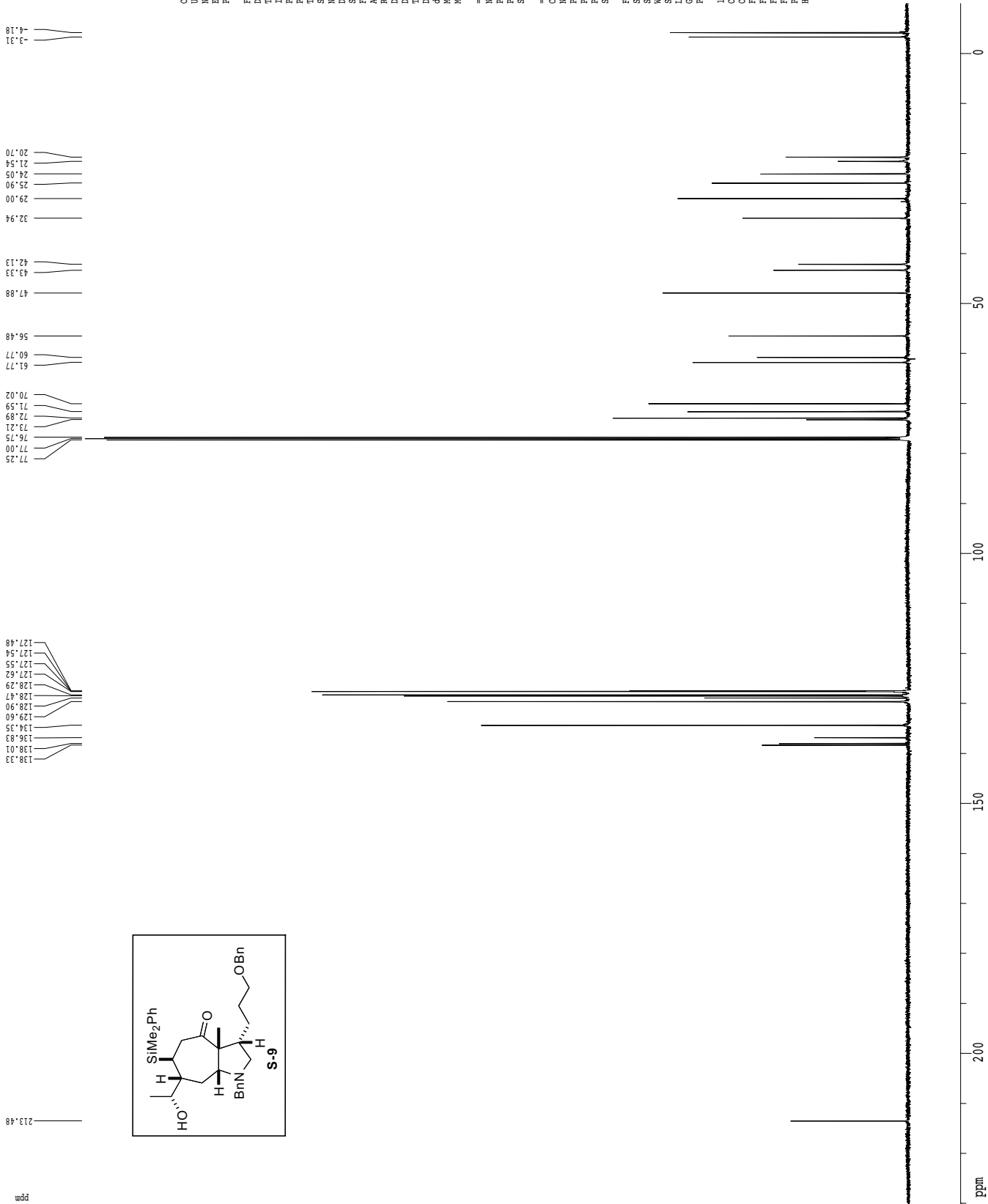
1H spectrum



Current Data Parameters
 USER travis
 NAME TD 4 33 2
 PROCNO 1
 F2 Acquisition Parameters:
 Date 20080308
 Time 9.38
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H
 PULPROG zg30
 TD 81728
 SOLVENT CDCl3
 NS 2
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.0988774 sec
 RG 3.2
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 TT 0.1000000 sec
 MCREST 0.0000000 sec
 MCWRK 0.01500000 sec
 ===== CHANNEL f1 =====
 NUC1 1H
 P1 7.38 usec
 PL1 1.60 dB
 SF01 500.2235015 MHz
 F2 Processing Parameters
 SI 65536
 SF 500.2200251 MHz
 EM 0
 WDW 0
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 4.00



13C spectrum with 1H decoupling



```

Current Data Parameters
USER      travis
NAME      TD-4-31-2
EXPNO     7
PROCNO    1

F2 - Acquisition Parameters
Date_     20080308
Time      9.39
INSTRUM   cryo500
PROBHD    5 mm CPCT 1H-
PULPROG   zgpg30
GAMES    65316
SOLVENT   CDCl3
NS         255
DS         4
SWH        30303.01 Hz
FIDRES     0.462388 Hz
AQ         1.0813940 sec
RG         8192
DM         16,500 usec
DE         6.00 usec
TE         300.2 K
DT         1.6000000 sec
d11        0.63000000 sec
MCREST     0.60000000 sec
MCWRK     0.61500000 sec

===== CHANNEL f1 =====
NUC1       13C
P1         14.75 usec
PL1        -1.00 dB
SFO1       125.7942348 MHz

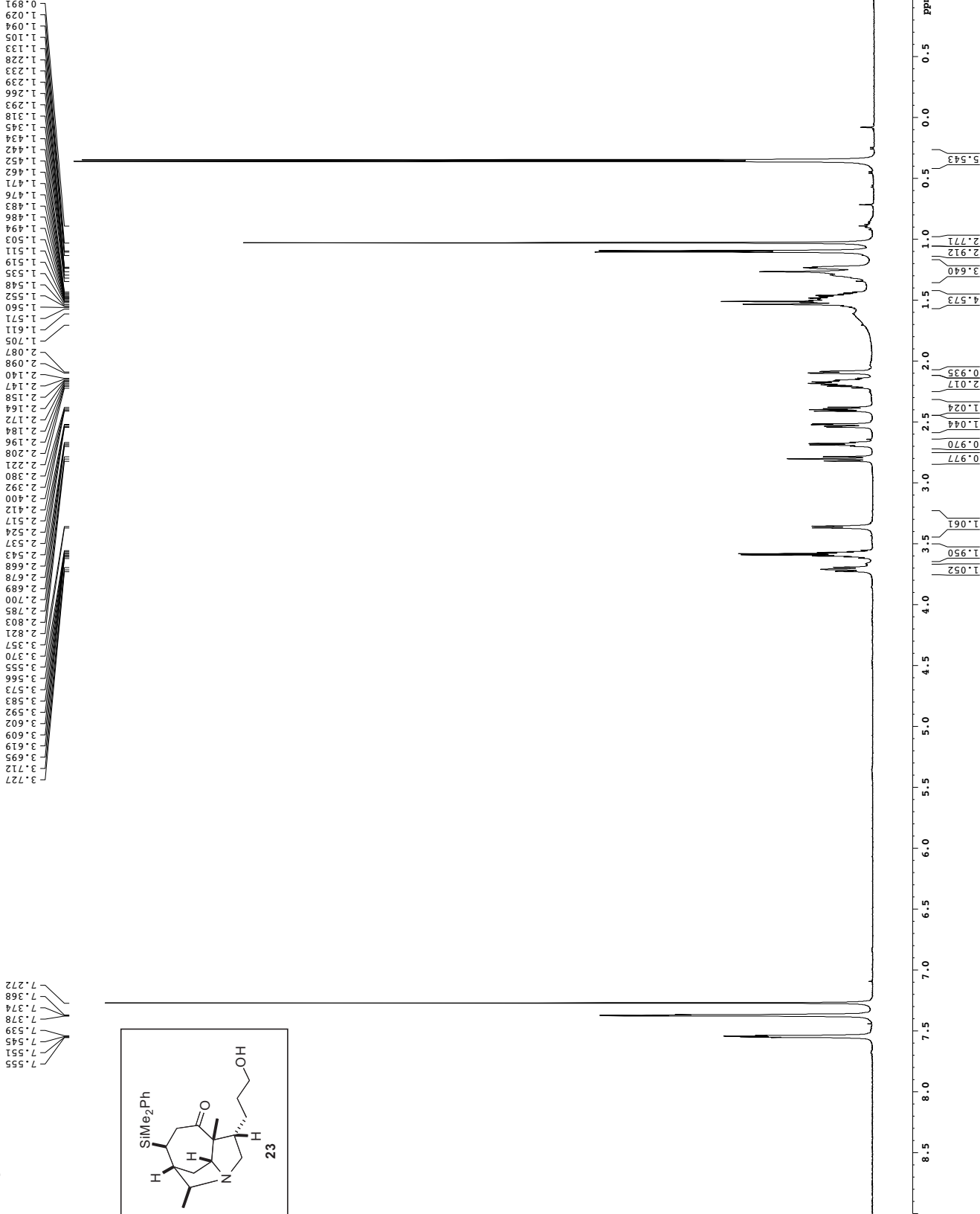
===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2       1H
PCPD2     100.00 usec
PL2        1.60 dB
PL12       24.80 dB
SFO2       500.2225011 MHz

F2 - Processing parameters
SI         655.556
SF         125.7804356 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00

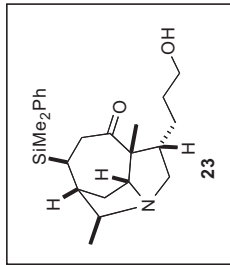
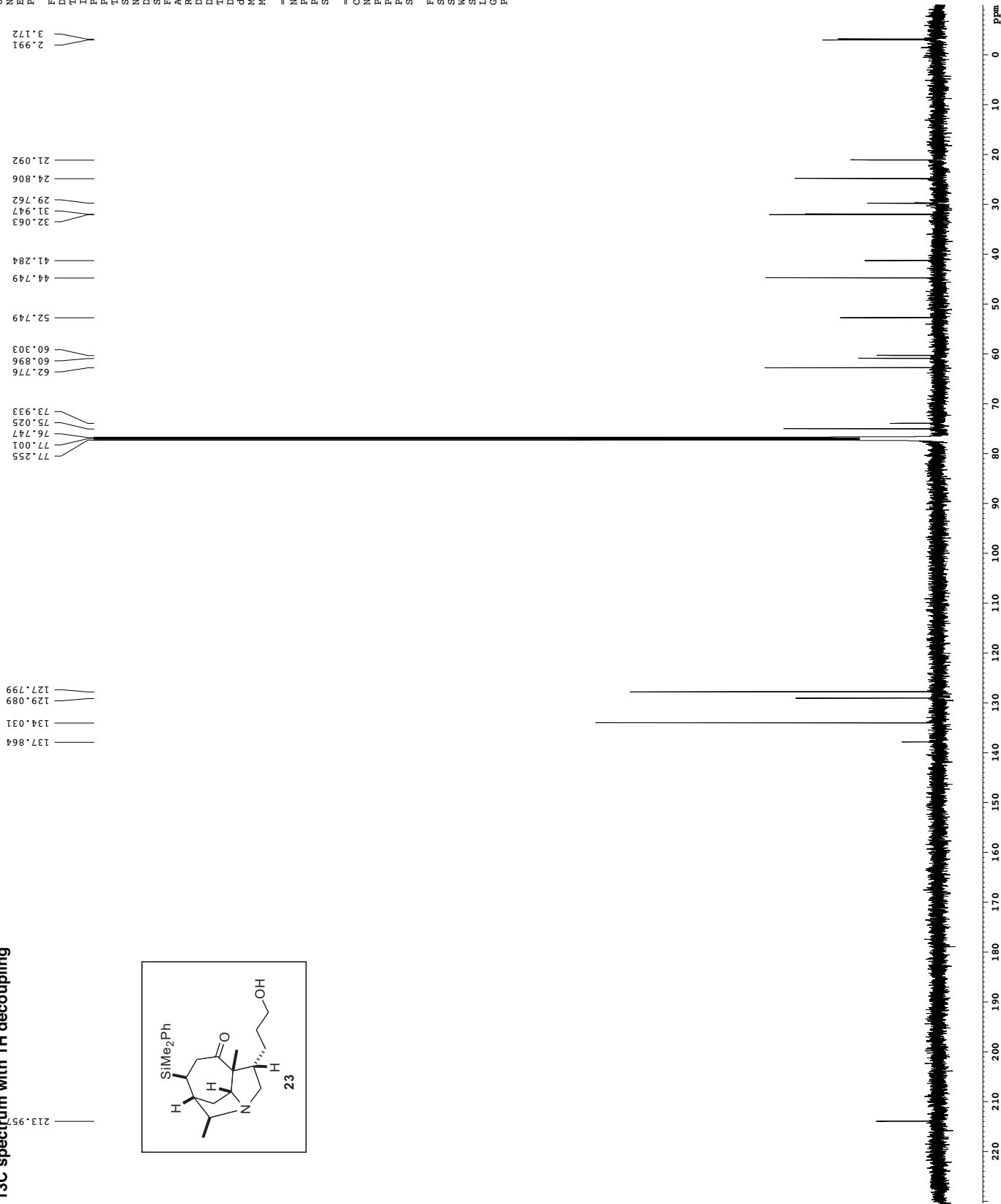
ID_NMR     plot parameters
CX         125.80 cm
CY         15.65 cm
F1P        230.000 ppm
F2P        28929.50 Hz
F3P        -10.000 ppm
F4P        -1257.80 Hz
PP4CM      10.52632 ppm/cm
HZCM       1324.00464 Hz/cm
    
```


1H spectrum

Current Data Parameters
 USER: gms
 TD: 4 203
 EXPTNO: 101
 PROCNO: 1
 F2 Acquisition Parameter:
 Date_ 20080211
 Time_ 20.48
 INSTRUM av600
 PULPROG zgpg30
 TD 65536
 FIDRES 0.096178 Hz
 AQ 5.0926259 sec
 RG 256
 DW 52.600 usec
 DE 6.000 usec
 TE 288.0 K
 D1 0.10000000 sec
 TD0 1
 ===== CHANNEL f1 =====
 NUC1 1H
 P1 8.00 usec
 PL1 0.00 dB
 SFO1 600.1342009 MHz
 F2 Processing parameters
 SI 65536
 SF 600.1300282 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



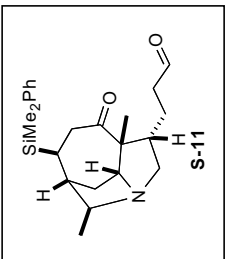
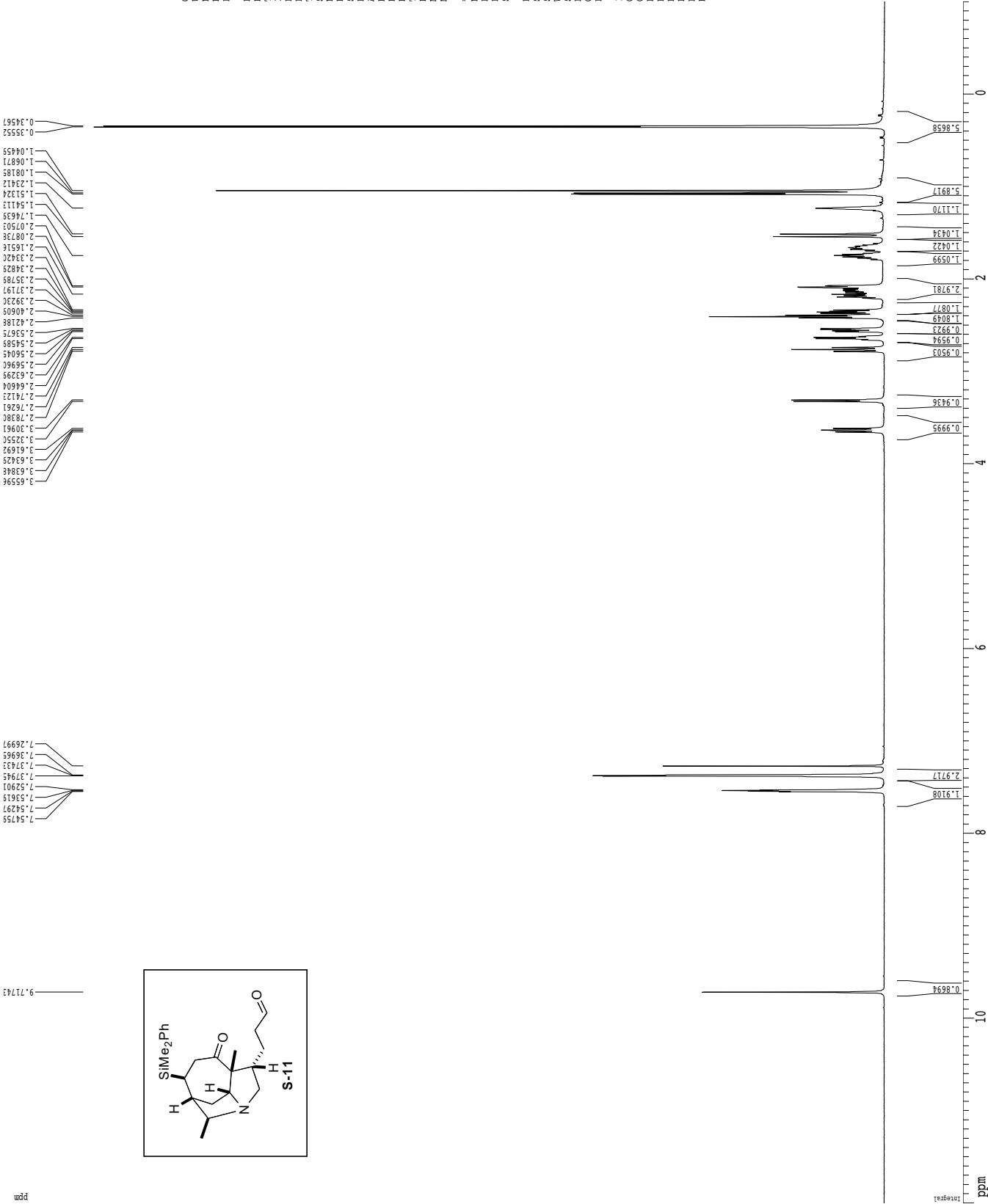
¹³C spectrum with ¹H decoupling



```

Current Data Parameters
USER      travis
NAME      TD 4 20 3
EXPNO     5
PROCNO    1
=====
F2 Acquisition Parameters:
Date_     20080211
Time_     21.30
INSTRUM   cryo500
PROBHD    5 mm CPTCI IH
PULPROG   zgpg30
TD         65418
SOLVENT   CDCl3
NS         794
DS         4
SWH        30303.031 Hz
FIDRES     0.463222 Hz
AQ         1.0794470 sec
RG         11585.2
DE         16.500 us
TE         298.0 K
D1         1.0000000 sec
d11        0.0000000 sec
d12        0.0000000 sec
d13        0.0000000 sec
d14        0.0000000 sec
d15        0.0000000 sec
d16        0.0000000 sec
d17        0.0000000 sec
d18        0.0000000 sec
d19        0.0000000 sec
d20        0.0000000 sec
d21        0.0000000 sec
d22        0.0000000 sec
d23        0.0000000 sec
d24        0.0000000 sec
d25        0.0000000 sec
d26        0.0000000 sec
d27        0.0000000 sec
d28        0.0000000 sec
d29        0.0000000 sec
d30        0.0000000 sec
===== CHANNEL f1 =====
NUC1       13C
P1         14.75 us
PL1        1.00 dB
SFO1       125.7942548 MHz
===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2       1H
PCPD2     100.00 us
PL2        1.60 dB
PL12       24.80 dB
SFO2       500.2225011 MHz
===== Processing parameters =====
SI         65536
SF         125.7604376 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00
  
```

1H spectrum



Current Data Parameters
 USER travis
 NAME TD-4-36-3
 EXPNO 1
 PROCNO 1

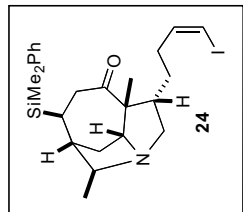
F2 - Acquisition Parameters
 Date_ 20080228
 Time_ 19.04
 INSTRUM cryo500
 PROBRD 5 mm CPXI H-
 PULPROG zgpg30
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.0998774 sec
 RG 5
 DW 62.400 usec
 DE 100.000 usec
 TE 298.0 K
 D1 0.1000000 sec
 MCREST 0.0000000 sec
 MCWRK 0.0150000 sec

***** CHANNEL f1 *****
 NUCL1 1H
 P1 7.00 usec
 PL1 1.60 dB
 SFO1 500.225015 MHz

F2 - Processing parameters
 SI 65536
 SF 500.220258 MHz
 WDW EM
 SS 0
 GB 0.30 Hz
 CB 0
 PC 4.00

1D NMR plot parameters
 CX 22.80 cm
 CT 15.00 cm
 FL 15.00 ppm
 FT 409.64 Hz
 FZ 0.00000000
 F2 -510.22 Hz
 PPMCK 0.57018 ppm/cm
 HZCKN 285.21320 Hz/cm

¹H spectrum



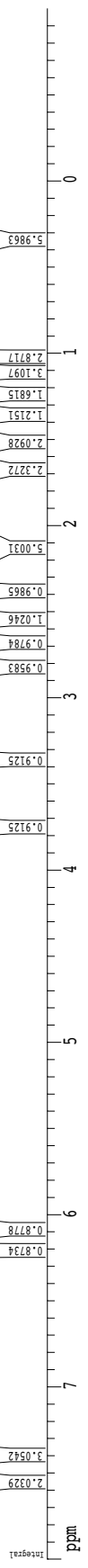
Current Data Parameter:
 USER mellis
 NAME ME-1-204-ch
 EXNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20080602
 Time 14.02
 INSTRUM cryo500
 PROBRD 5 mm CPYCI 1H
 PULPROG zgpg30
 TD 81728
 SFO1 500.1350000 MHz
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.099398 sec
 RG 4.5
 DM 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.1000000 sec
 d11 0.1000000 sec
 ACQST 0.1000000 sec
 ACWRK 0.0150000 sec

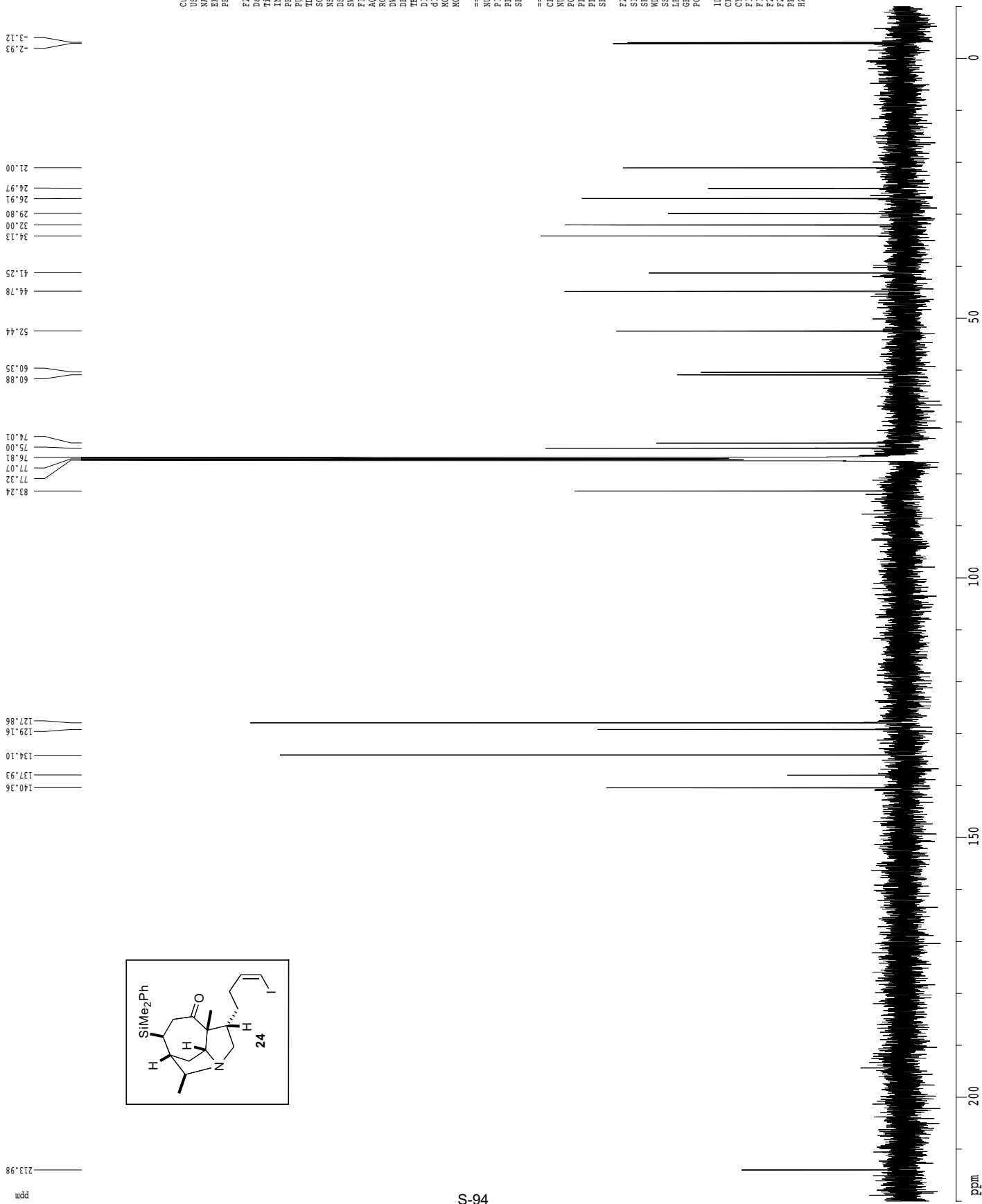
==== CHANNEL f1 =====
 NUC1 1H
 P1 7.38 usec
 PL1 1.60 dB
 SFO1 500.2235015 MHz

F2 - Processing parameter
 SI 65536
 SF 500.2200261 MHz
 EQ
 ASB 0
 GB 0
 GR 0
 SC 4.00

1D NMR plot parameters
 CX 22.80 cm
 CY 15.00 cm
 F1P 8.000 ppm
 F1 4001.76 Hz
 F2P -1.000 ppm
 F2 -500.22 Hz
 FREQC 0.39878 ppm/cm
 HZCCH 197.4528 Hz/cm



13C spectrum with 1H decoupling



```

Current Data Parameters
USER      mellis
NAME      ME-1-204-ch
EXNO      3
PROCNO    1

F2 - Acquisition Parameters
Date_     20080602
Time      14.17
INSTRUM   cryo500
PROBHD    5 mm CPTCI IH-
PULPROG   zgpg30
TD         65418
SOLVENT   CDCl3
NS         1024
DS         4
SWH        30303.031 Hz
FIDRES     0.463222 Hz
AQ         1.0794635 sec
RG         11585.2
DM         16.500 usec
DE         6.00 usec
TE         298.0 K
d1         0.25000000 sec
d11        0.03000000 sec
ACQRES    0.00000000 sec
MCPRK     0.01500000 sec

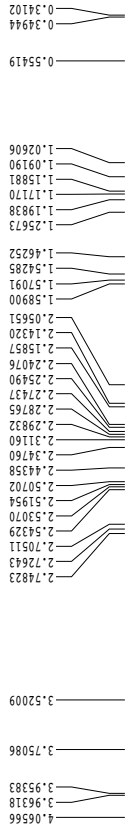
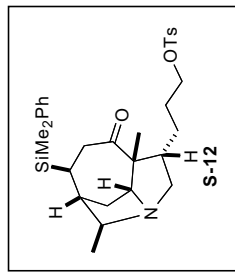
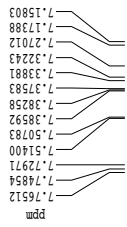
===== CHANNEL f1 =====
NUC1       13C
P1         14.75 usec
PL         -1.00 dB
SFO1      125.7942546 MHz

===== CHANNEL f2 =====
CPOPRG2    waltz16
NUC2        1H
PCPD2      100.00 usec
PL2        1.60 dB
PL12       24.80 dB
SFO2      500.2225011 MHz

F2 - Processing parameter
SI         65536
SF         125.7804190 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00

ID_NMR plot parameters
CX         22.80 cm
CY         162.98 cm
ETP        220.000 ppm
F1P        27671.69 Hz
F2P        -10.000 ppm
F2         -1257.80 Hz
PPMCH     10.08772 ppm/cm
HZCM      1268.83765 Hz/cm
    
```

¹H spectrum



Current Data Parameter
 USER mellis
 NAME ME-1-258-ch
 EXPTNO 1
 PROCNO 1

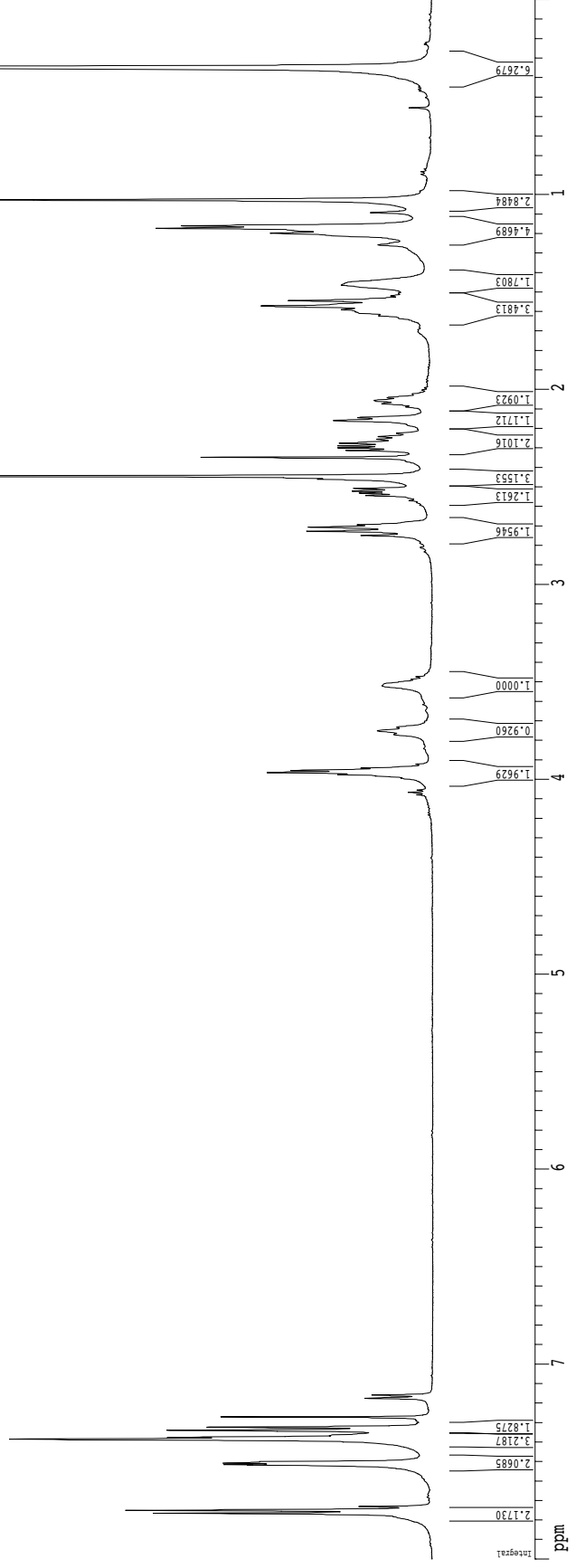
F2 - Acquisition Parameters
 Date_ 20080710
 Time 13:39
 INSTRUM crys00
 PROBRD 5 mm CPCL1 H-1
 PULPROG zgpg30
 TD 65536
 SFO1 500.136000 MHz
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.099398 sec
 RG 4.5
 DM 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.100000 sec
 d11 0.100000 sec
 ACQST 0.0150000 sec
 ACWRK 0.0150000 sec

==== CHANNEL f1 =====
 NUC1 ¹H
 P1 7.38 usec
 PL1 1.60 dB
 SFO1 500.136015 MHz

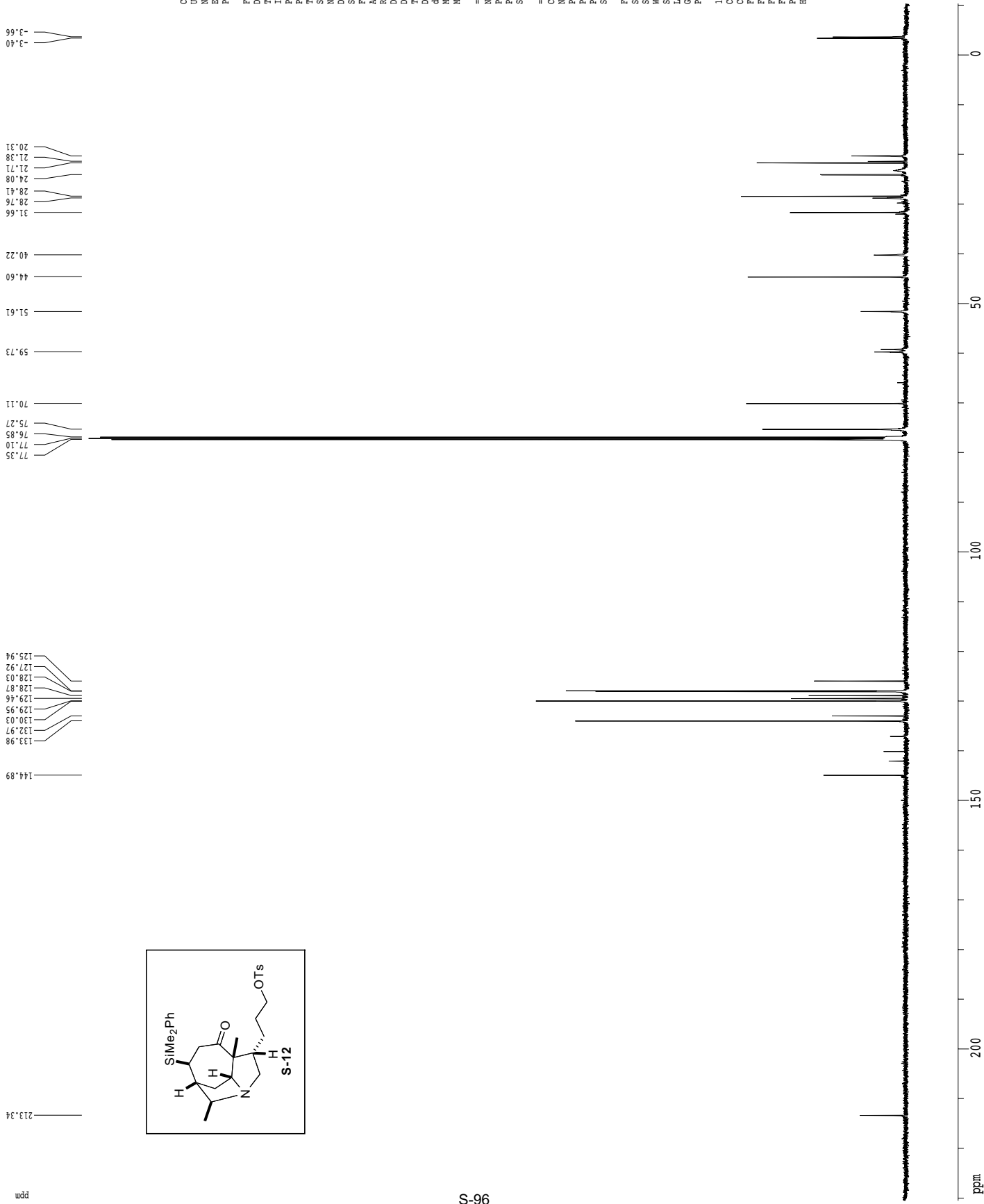
F2 - Processing parameter
 SI 65536
 SF 500.136015 MHz
 EQ
 SSB 0
 CB 0.20 Hz
 GB 0
 PC 0
 SC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CY 15.00 cm
 TP 8.000 ppm
 F1 4001.76 Hz
 F2 0.000 ppm
 F3 0.000 Hz
 PPRCM -0.35088 ppm/cm
 HZCM 175.51581 Hz/cm

56-S



13C spectrum with 1H decoupling



```

Current Data Parameters
USER      mellis
NAME      ME-1-258-ch
EXNO      3
PROCNO    1

F2 - Acquisition Parameters
Date_     20080710
Time      13:53
INSTRUM   cryo500
PROBHD    5 mm CPXI 1H-
PULPROG   zgpg30
TD         65418
SOLVENT    CDCl3
NS         824
DS         4
SWH        30303.031 Hz
FIDRES     0.463222 Hz
AQ         1.0794635 sec
RG         11585.2
DM         16.500 usec
DE         6.00 usec
TE         298.0 K
D1         0.25000000 sec
d11        0.03000000 sec
ACQRES    0.00000000 sec
MCPRK     0.01500000 sec

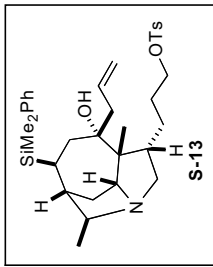
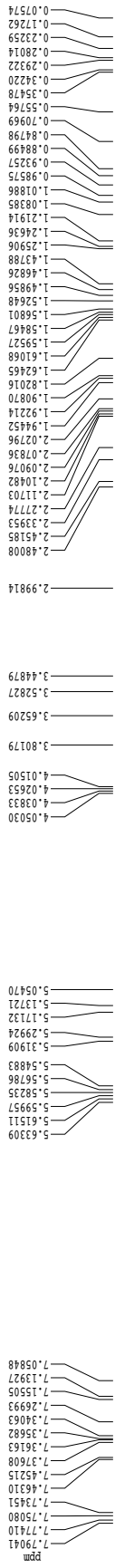
===== CHANNEL f1 =====
NUC1       13C
P1         14.75 usec
PL         -1.00 dB
SFO1       125.7942346 MHz

===== CHANNEL f2 =====
CPOBRC2    waltz16
NUC2        1H
PCPD2      100.00 usec
PL2         1.60 dB
PL12        24.80 dB
SFO2        500.2225011 MHz

F2 - Processing parameter
SI         65536
SF         125.7804190 MHz
WDW        EM
SSB         0
LB         1.00 Hz
GB         0
PC         2.00

ID_NMR plot parameters
CX         22.80 cm
CY         15.65 cm
FLP        230.637 ppm
F1         29009.68 Hz
F2         -10.287 ppm
F3         -1293.96 Hz
FPMCH      10.56688 ppm/cm
HZCM       1329.10706 Hz/cm
    
```


¹H spectrum



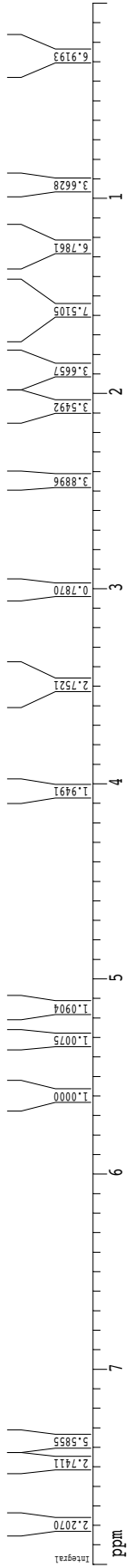
Current Data Parameter:
 USER mellis
 NAME NE-1-259-ch
 EXENO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20080714
 Time 10:25
 INSTRUM cryo500
 PROBRD 5 mm CPCL1
 PULPROG zgpg30
 TD 81728
 SFO1 500.136261 MHz
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.099398 sec
 RG 5.7
 DM 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.100000 sec
 d11 0.100000 sec
 ACQST 0.10150000 sec
 ACWRK 0.10150000 sec

==== CHANNEL f1 =====
 NUC1 ¹H
 P1 7.38 usec
 PL1 1.60 dB
 SFO1 500.2235015 MHz

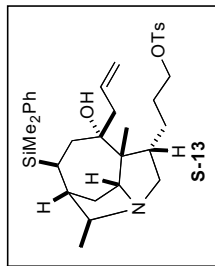
F2 - Processing parameter
 SI 65536
 SF 500.2200255 MHz
 EQ
 SSB 0
 GB 0.30 Hz
 GR 0
 ZA 4.00

1D NMR plot parameters
 CX 22.80 cm
 CY 15.00 cm
 FID 8.000 ppm
 F1 4001.76 Hz
 F2 0.000 ppm
 F3 0.000 Hz
 FWHM 0.35088 ppm/cm
 HZCN 175.51581 Hz/cm



13C spectrum with 1H decoupling

ppm



```

Current Data Parameters
USER      mellis
NAME      ME-1-253-ch
EXNO      3
PROCNO    1

F2 - Acquisition Parameters
Date_     20080714
Time      10.33
INSTRUM   cryo500
PROBHD    5 mm CPYCI IH-
PULPROG   zgpg30
TD         65418
SOLVENT    CDCl3
NS         1370
DS         4
SWH        30303.031 Hz
FIDRES     0.463222 Hz
AQ         1.0794635 sec
RG         11585.2
DM         16.500 usec
DE         6.00 usec
TE         298.0 K
d11        0.25000000 sec
d12        0.03000000 sec
d13        0.03000000 sec
d14        0.03000000 sec
d15        0.03000000 sec
d16        0.03000000 sec
d17        0.03000000 sec
d18        0.03000000 sec
d19        0.03000000 sec
d20        0.03000000 sec
d21        0.03000000 sec
d22        0.03000000 sec
d23        0.03000000 sec
d24        0.03000000 sec
d25        0.03000000 sec
d26        0.03000000 sec
d27        0.03000000 sec
d28        0.03000000 sec
d29        0.03000000 sec
d30        0.03000000 sec
===== CHANNEL f1 =====
NUC1       13C
P1         14.75 usec
PL1        -1.00 dB
SFO1       125.7942546 MHz

===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2       1H
PCPD2      100.00 usec
PL2        1.60 dB
PL12       24.80 dB
SFO2       500.2225011 MHz

F2 - Processing parameter
SI         65536
SF         125.7804190 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00

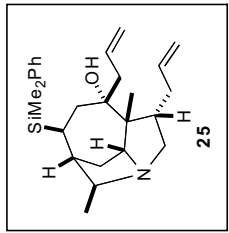
ID_NMR plot parameters
CX         22.80 cm
CY         143.06 cm
ETP        200.000 ppm
F1P        25156.08 Hz
F2P        -10.000 ppm
F3P        -1.257.80 Hz
FPMCH      9.21053 ppm/cm
HZCM       1158.50378 Hz/cm
    
```

77.227
77.073
76.819



ppm

¹H spectrum



Current Data Parameter:
 USER mellis
 NAME NE-1-266-ch
 EXPTNO 1
 PROCNO 1

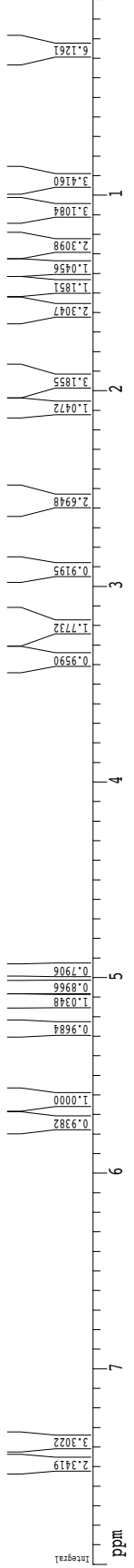
F2 - Acquisition Parameters
 Date_ 20080726
 Time 13.03
 INSTRUM cryo500
 PROBRD 5 mm CPCL1 JN
 PULPROG zgpg30
 TD 81728
 SFO1 500.135000 MHz
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.099398 sec
 RG 5
 DM 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.100000 sec
 d11 0.100000 sec
 ACQST 0.100000 sec
 ACWRK 0.0150000 sec

==== CHANNEL f1 =====
 NUC1 ¹H
 P1 7.38 usec
 PL1 1.60 dB
 SFO1 500.2235015 MHz

F2 - Processing parameter
 SI 65536
 SF 500.2200256 MHz
 EQ
 SSB 0
 CB 0.00 Hz
 GB 0
 DB 0
 SC 4.00

1D NMR plot parameters
 CX 22.80 cm
 CY 15.00 cm
 FID 8.000 ppm
 F1 4001.76 Hz
 F2 0.000 ppm
 F3 0.000 Hz
 FWHM 0.35088 ppm/cm
 HZCN 175.51581 Hz/cm

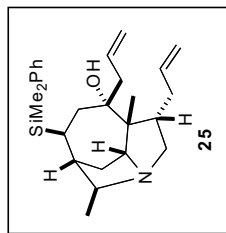
66-S



Integral

¹³C spectrum with ¹H decoupling

ppm



```

Current Data Parameters
USER      mellis
NAME      ME-1-266-ch
EXNO      3
PROCNO    1

F2 - Acquisition Parameters
Date_     20080726
Time      13.12
INSTRUM   cryo500
PROBHD    5 mm CPXI 1H-
PULPROG   zgpg30
TD         65418
SOLVENT   CDCl3
NS         1024
DS         4
SWH        30303.031 Hz
FIDRES     0.463222 Hz
AQ         1.0794635 sec
RG         11585.2
DM         16.500 usec
DE         6.00 usec
TE         298.0 K
d1         0.25000000 sec
d11        0.03000000 sec
ACQRES    0.00000000 sec
MCPRK     0.01500000 sec

===== CHANNEL f1 =====
NUC1       13C
P1         14.75 usec
PL         -1.00 dB
SFO1      125.7942546 MHz

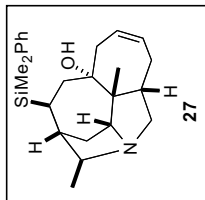
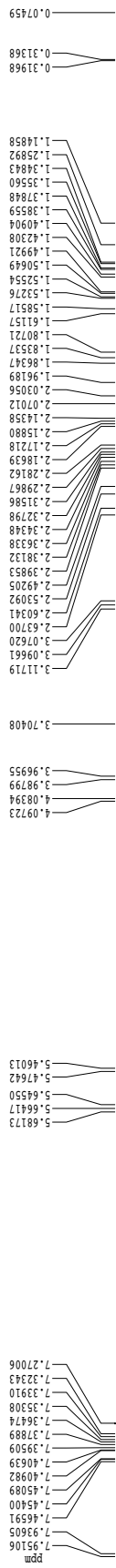
===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2        1H
P2         100.00 usec
PL2        1.60 dB
PL12       24.80 dB
SFO2      500.2225011 MHz

F2 - Processing parameter
SI         65536
SF         125.7804190 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00

ID_NMR plot parameters
CX         22.80 cm
CY         73.99 cm
ETP        200.000 ppm
F1P        25156.08 Hz
F2P        -10.000 ppm
F2         -1257.80 Hz
FPMCH      9.21053 ppm/cm
HZCM       1158.50378 Hz/cm
    
```



¹H spectrum



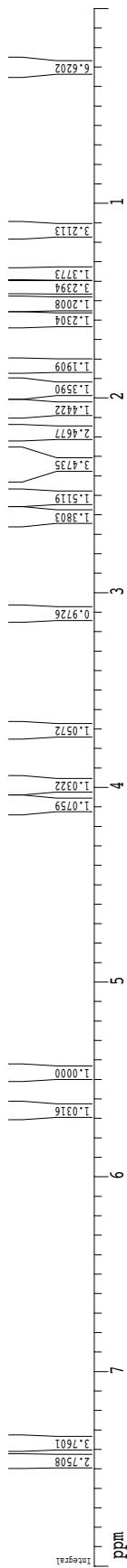
Current Data Parameters
 USER melis
 NAME ME-I-235-old
 EXPERNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date 20080916
 Time 14.31
 INSTRUM cryo500
 PROBRD 5 mm CPTCI IH-
 PULPROG zg30
 TD 81728
 SFO1 500.136261 MHz
 SOLVENT CDCl3
 NS 2
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.0988774 sec
 RG 7.1
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.1000000 sec
 MCREST 0.0000000 sec
 MCWRR 0.0150000 sec

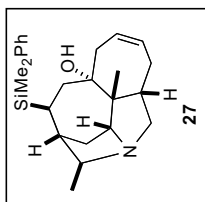
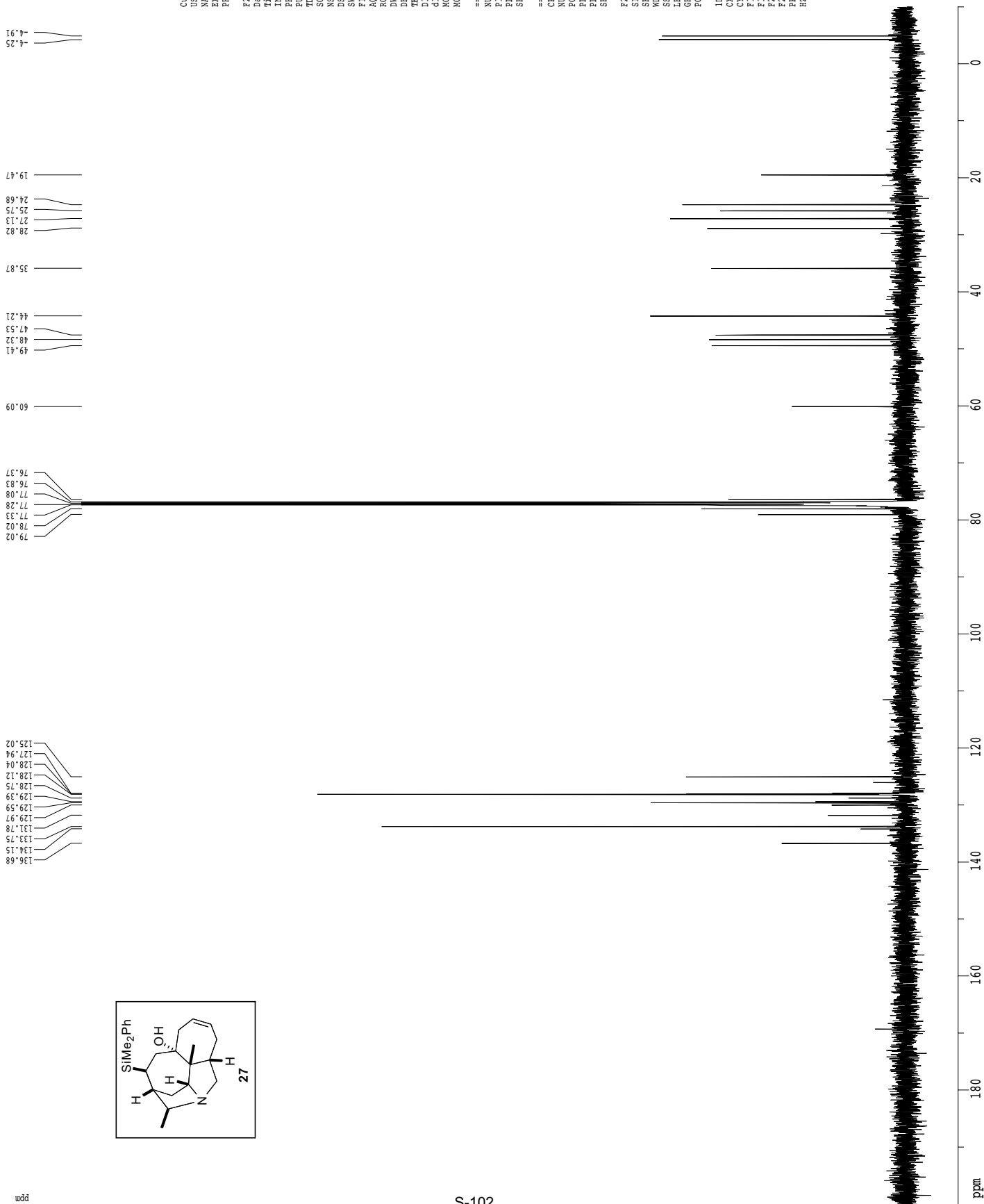
===== CHANNEL f1 =====
 NUC1 ¹H
 P1 7.38 usec
 PL1 1.60 dB
 SFO1 500.2235015 MHz

F2 - Processing parameters
 S1 65536
 SF 500.2200261 MHz
 EM
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 4.00

ID NMR Plot parameters
 CX 22.80 cm
 CY 36.94 cm
 F1P 8.000 ppm
 F1F 4001.76 Hz
 F2P 0.000 ppm
 F2F 0.00 Hz
 PPMCH 0.35888 ppm/cm
 HZCN 175.51981 Hz/cm



13C spectrum with 1H decoupling



```

Current Data Parameters
USER      mellis
NAME      ME-1-295-04d
EXNO      3
PROCNO    1

F2 - Acquisition Parameters
Date_     20080916
Time      14.35
INSTRUM   cryo500
PROBHD    5 mm CP1CI 1H-
PULPROG   zgpg30
TD         65418
SOLVENT    CDCl3
NS         726
DS         4
SWH        30303.031 Hz
FIDRES     0.463222 Hz
AQ         1.0794476 sec
RG         9195.2
DM         16.500 usec
DE         6.00 usec
TE         298.0 K
d11        0.25000000 sec
d12        0.03000000 sec
d13        0.00000000 sec
d14        0.00000000 sec
d15        0.01500000 sec
MCPRK     0.01500000 sec

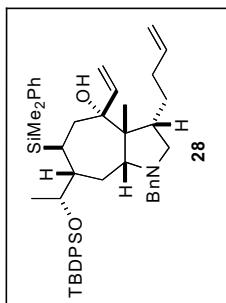
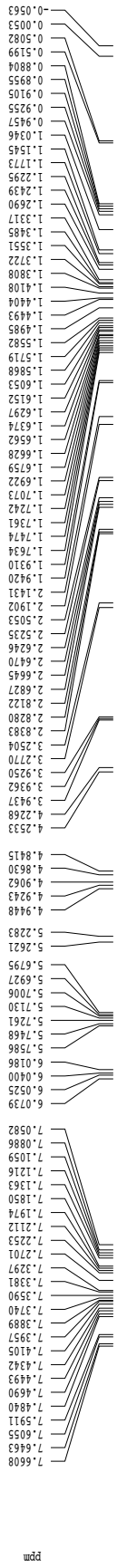
===== CHANNEL f1 =====
NUC1       13C
P1         14.75 usec
PL1        -1.00 dB
SFO1       125.7942546 MHz

===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2        1H
P2         100.00 usec
PL2         1.60 dB
PL12       24.80 dB
SFO2       500.2225011 MHz

F2 - Processing parameter
SI         65536
SF         125.7804190 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00

ID NMR plot parameters
CX         272.80 cm
CY         145.47 cm
ETP        200.000 ppm
F1P        25156.08 Hz
F2P        -10.000 ppm
F2         -1257.80 Hz
FPMCH      9.21053 ppm/cm
HZCM       1158.50378 Hz/cm
    
```

¹H spectrum



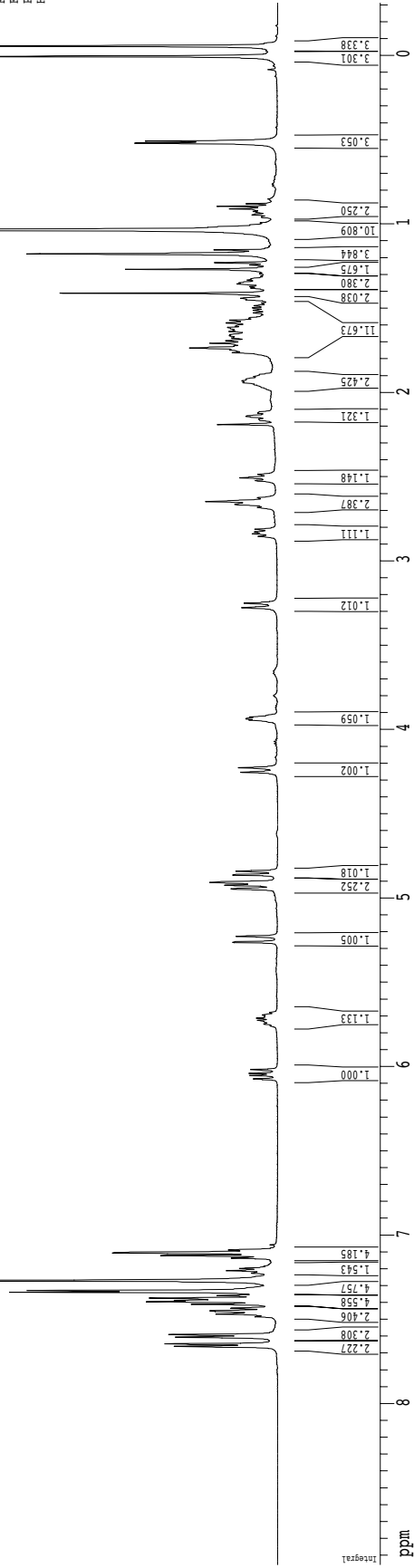
Current Data Parameter
 USER mellis
 NAME ME-II-24
 EXNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20081107
 Time 16:04
 INSTRUM cryo500
 PROBD 5 mm CPTCI JN
 PULPROG zgpg30
 TD 81728
 SFO1 500.1350000
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.099398 sec
 RG 4.5
 DM 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.1000000 sec
 d11 0.1000000 sec
 ACQST 0.10150000 sec
 ACWRK 0.01500000 sec

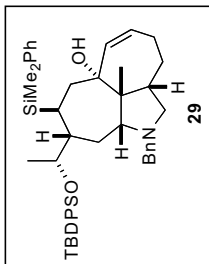
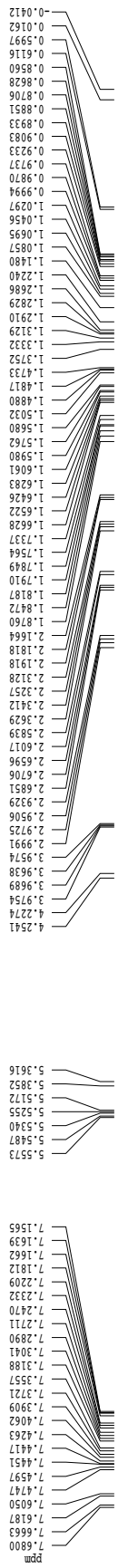
==== CHANNEL f1 =====
 NUC1 ¹H
 P1 7.50 usec
 PL1 1.60 dB
 SFO1 500.22235015 MHz

F2 - Processing parameter
 SI 65536
 SF 500.2200246 MHz
 EQ
 SSB 0
 GB 0.20 Hz
 CB 0
 SC 4.00

1D NMR plot parameters
 CX 22.80 cm
 CY 15.00 cm
 F1P 8.855 ppm
 F2P 4479.51 Hz
 F2 -0.309 ppm
 F2 -154.62 Hz
 F2PCMC -0.40632 ppm/cm
 HZCM 203.25117 Hz/cm



¹H spectrum



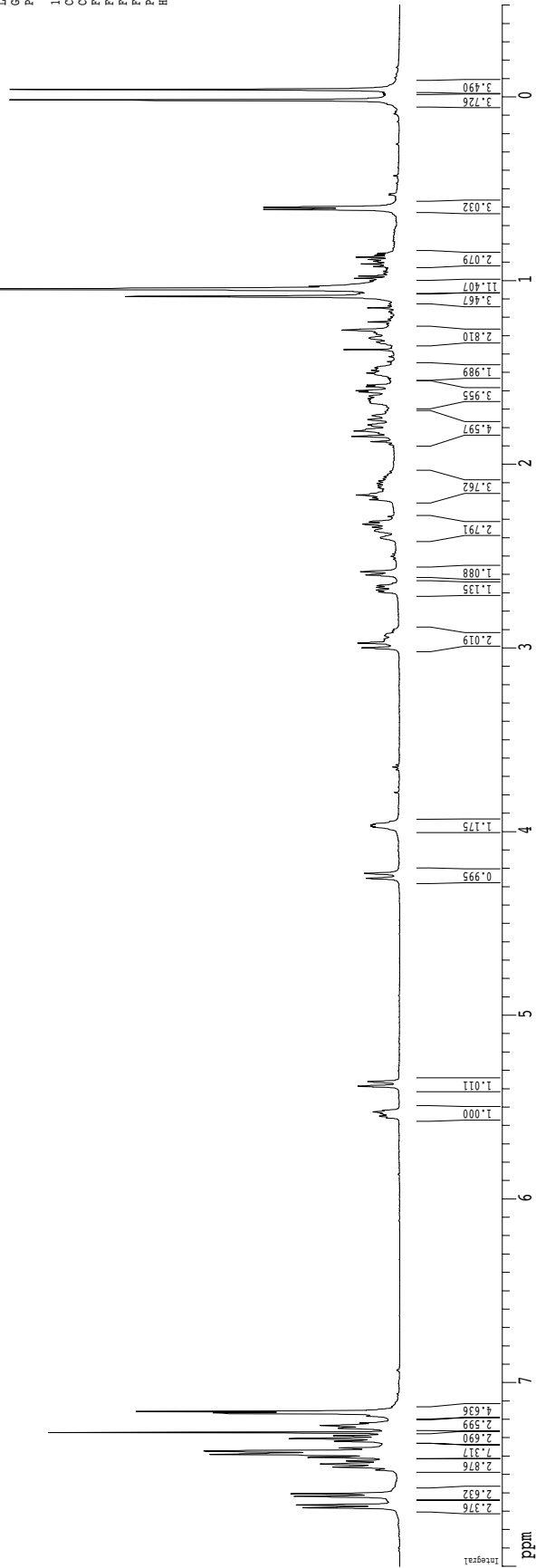
```

Current Data Parameter
USER          mellis
NAME          ME-II-25
EXFNO        1
PROCNO        1

F2 - Acquisition Parameters
Date_         2008117
Time          8.15
INSTRUM      cry500
PROBHD       5 mm CPTCI
PULPROG      zgpg30
TD           81728
SOLVENT      CDCl3
NS           8
DS           2
SWH          8012.820 Hz
FIDRES      0.098043 Hz
AQ          5.099398 sec
RG          11.3
DM          62.400 usec
DE          6.00 usec
TE          298.0 K
D1          0.1000000 sec
d11         0.0000000 sec
d12         0.0000000 sec
d13         0.0150000 sec
===== CHANNEL f1 =====
NUC1         1H
P1          7.50 usec
PL1         1.60 dB
SFO1        500.2225015 MHz

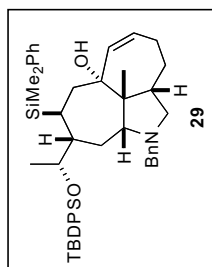
F2 - Processing parameter
SI          65536
SF          500.2200239 MHz
EN
GB          0
GB1         0
GB2         0
GB3         0
PC         4.00

ID NMR plot parameters
CX          22.80 cm
CY          15.00 cm
FTF         8.000 ppm
F1F         4001.76 Hz
F2F         -0.500 ppm
F2          -250.11 Hz
PPHMC      0.37281 ppm/cm
HZHC       186.48555 Hz/cm
    
```



13C spectrum with 1H decoupling

140.44
139.85
136.07
136.01
134.72
134.61
133.63
131.42
129.61
129.57
128.55
128.15
127.65
127.57
126.55



79.07
77.56
77.52
77.27
77.06
76.81
71.73
61.04
58.40
54.24
50.19
43.29
42.76
32.03
31.13
30.62
29.43
27.20
27.09

Current Data Parameters
 USER mellis
 NAME MP-1-25
 EXPNO 2
 PROCNO 1
 F2 - Acquisition Parameters
 Date_ 20081117
 Time 8.23
 INSTRUM cryo500
 PROBHD 5 mm CPCTC 1H-
 PULPROG SpinEcho90gp-prd
 TD 6556
 SFO2 125.754258 MHz
 SOLVENT CDCl3
 NS 1016
 DS 4
 SHW 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0814105 sec
 RQ 7298.2
 DM 16.500 usec
 DE 6.00 usec
 TE 300.2 K
 D1 0.25000000 sec
 D11 0.03000000 sec
 D17 0.00020000 sec
 d17 0.00019600 sec
 MCREST 0.00000000 sec
 MCRBK 0.01500000 sec
 P2 29.70 usec

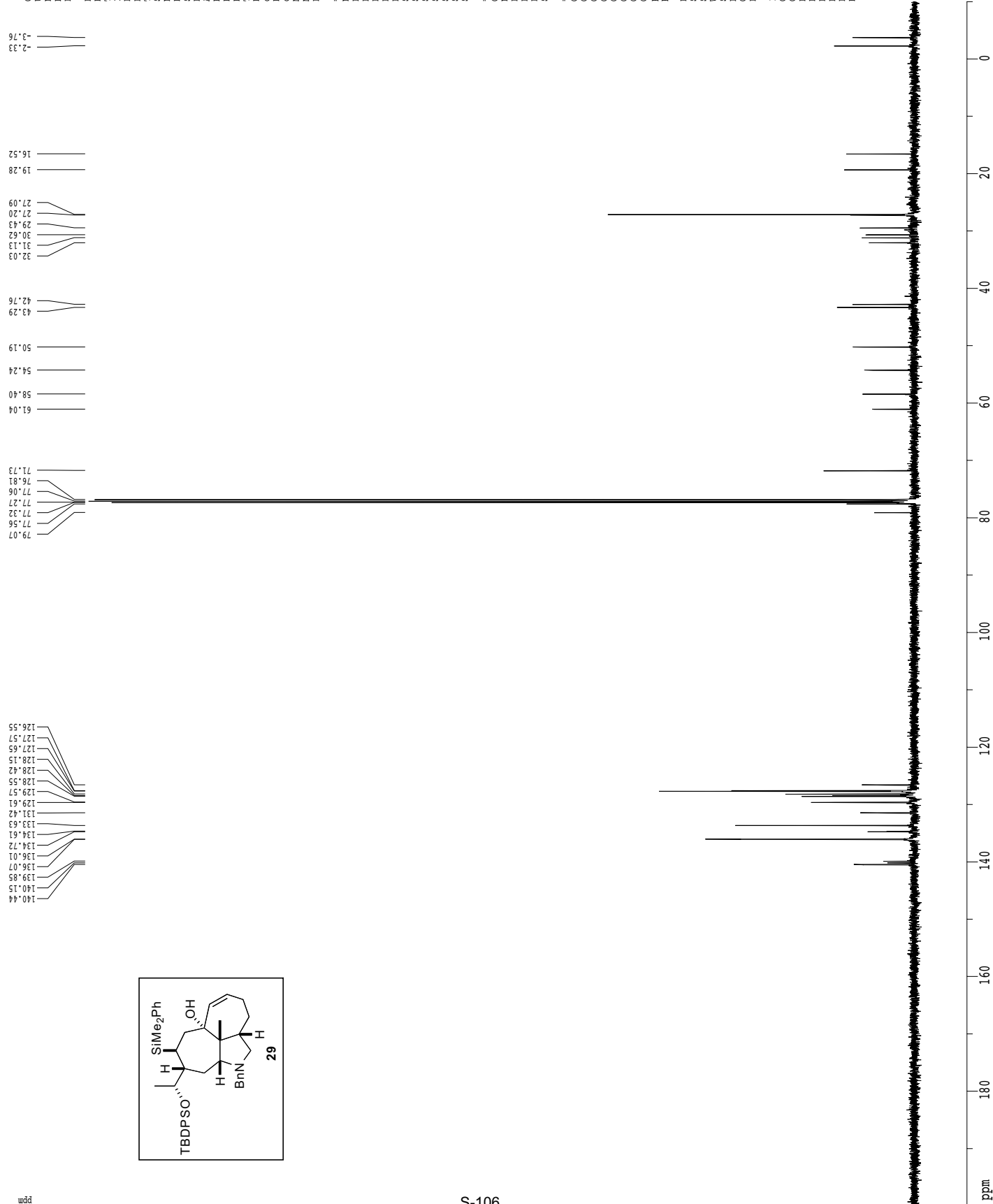
===== CHANNEL f1 =====
 NUC1 13C
 P1 14.85 usec
 P11 500.00 usec
 P12 2000.00 usec
 PL0 120.00 dB
 PL1 -1.00 dB
 SFO1 125.794258 MHz
 SF1 3.60 dB
 SP1 3.60 dB
 SP11 3.60 dB
 SP12 3.60 dB
 SPNAM1 Cp660.5
 SPPOFF1 Cp660cm.4
 SFOFF1 0.00 Hz
 SPOFF2 0.00 Hz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 P2 100.00 usec
 P21 60.00 usec
 P22 24.60 dB
 SFO2 500.2225011 MHz

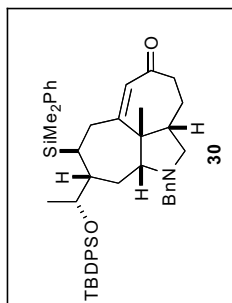
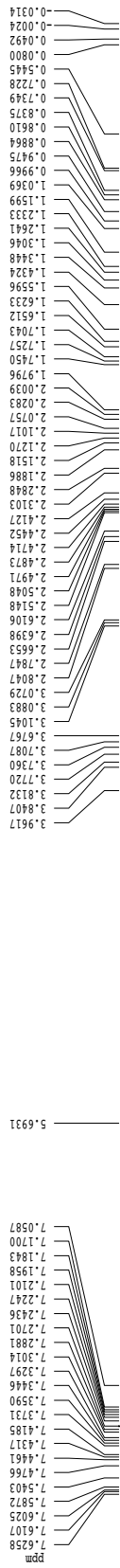
===== GRADIENT CHANNEL =====
 GPNAM1 SINE.100
 GPNAM2 SINE.100
 GPX1 0.00 %
 GPX2 0.00 %
 GPY1 0.00 %
 GPY2 0.00 %
 GPZ1 30.00 %
 GPZ2 50.00 %
 P15 500.00 usec
 P16 1000.00 usec

F2 - Processing parameters
 SI 32768
 SF 125.7804310 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 F1 125.754258 MHz
 F2 125.754258 MHz
 F1P 200.000 ppm
 F2P 25156.08 Hz
 F2 -10.000 ppm
 F2 -1257.80 Hz
 FPMCH 9.21053 ppm/cm
 HZCM 1158.50378 Hz/cm



1H spectrum



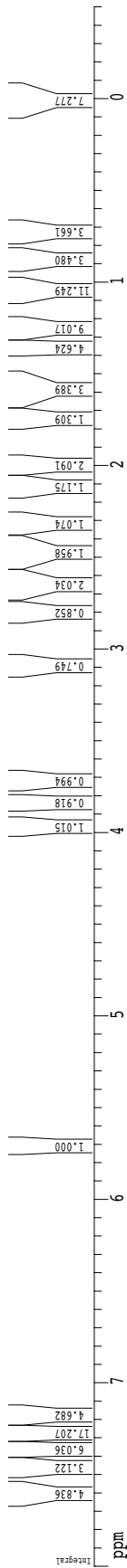
Current Data Parameters
 USER melis
 NAME ME-II-51
 EXPERNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 2009110
 Time 13.14
 INSTRUM cryo500
 PROBRD 5 mm CPTCI IH-
 PULPROG zg30
 TD 8128
 SFO1 500.136250 MHz
 SOLVENT CDCl3
 NS 2
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.0988774 sec
 RG 4.5
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.1000000 sec
 MCREST 0.0000000 sec
 MCWRR 0.0150000 sec

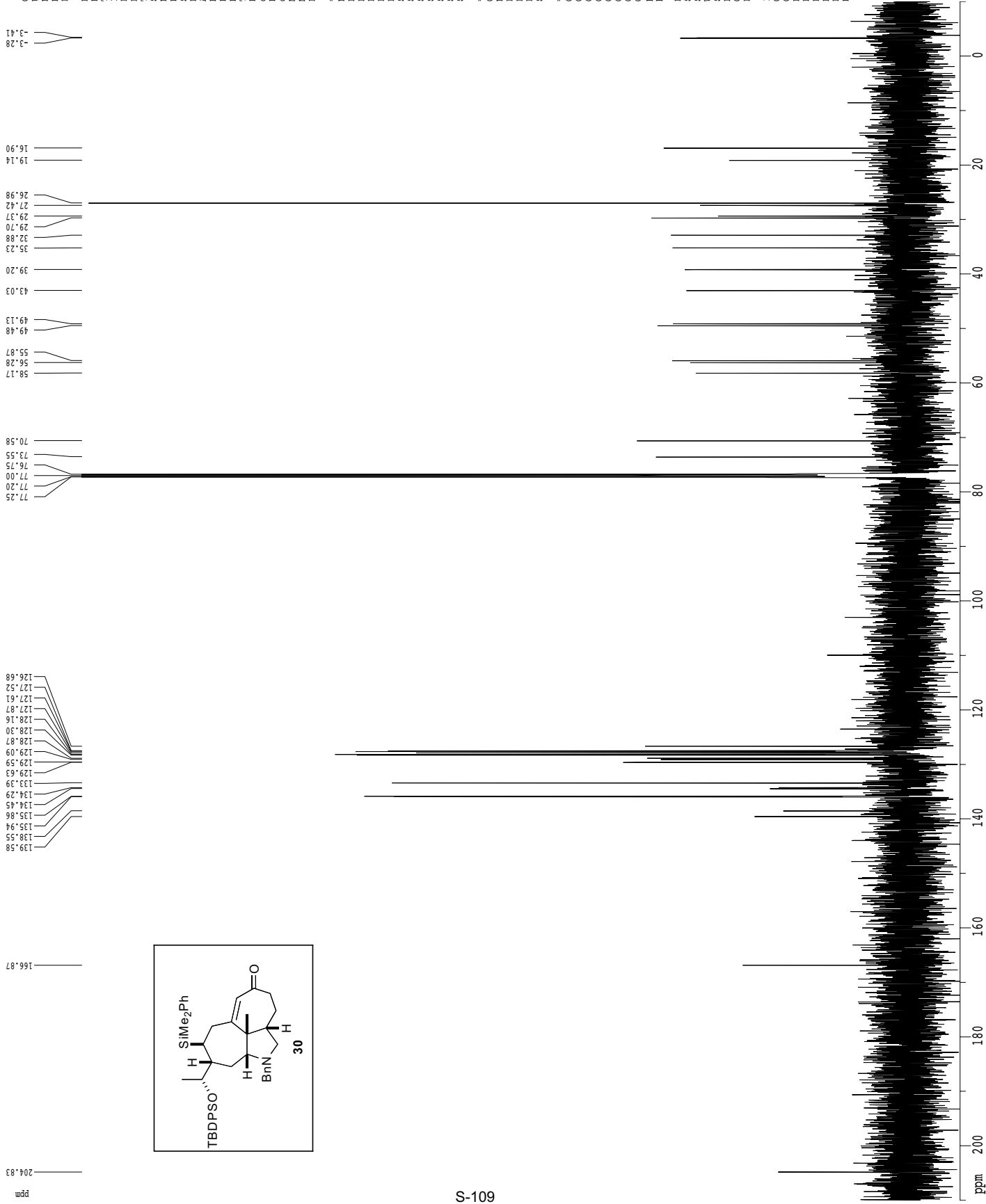
===== CHANNEL f1 =====
 NUC1 1H
 P1 7.50 usec
 PL1 1.60 dB
 SFO1 500.2235015 MHz

F2 - Processing parameter
 S1 65536
 SF 500.2200258 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 4.00

ID_NMR Plot parameters
 CX 22.80 cm
 CY 15.00 cm
 F1P 8.000 ppm
 F2P 4001.76 Hz
 F2 -250.11 Hz
 PPMCH 0.37281 ppm/cm
 HZCN 186.48555 Hz/cm



13C spectrum with 1H decoupling



```

Current Data Parameters
USER      mellis
NAME      MP-11-51
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     20090110
Time      13.23
INSTRUM   cryo500
PROBHD    5 mm CPCTC 1H-
PULPROG   Spinechop90gp.prd
TD         6556
SOLVENT   CDCl3
NS         16
DS         4
SHW        30303.031 Hz
FIDRES     0.462388 Hz
AQ          1.0813940 sec
RG          91.95.2
DM          16.500 usec
DE          6.00 usec
TE          300.2 K
D1          0.28000000 sec
d11         0.03000000 sec
d16         0.00020000 sec
d17         0.00019600 sec
MCREST     0.00000000 sec
MCWREST    0.01500000 sec
P2          29.70 usec

===== CHANNEL f1 =====
NUC1       13C
P1         14.85 usec
P2         500.00 usec
P3         2000.00 usec
PL0        120.00 dB
PL1        -1.00 dB
SFO1       125.7942548 MHz
SF         125.7600000 MHz
SFO2        3.60 dB
SFO3        3.60 dB
SFO4        3.60 dB
SFO5        3.60 dB
SFO6        3.60 dB
SFO7        3.60 dB
SFO8        3.60 dB
SFO9        3.60 dB
SFO10       3.60 dB
SFO11       3.60 dB
SFO12       3.60 dB
SFO13       3.60 dB
SFO14       3.60 dB
SFO15       3.60 dB
SFO16       3.60 dB
SFO17       3.60 dB
SFO18       3.60 dB
SFO19       3.60 dB
SFO20       3.60 dB
SFO21       3.60 dB
SFO22       3.60 dB
SFO23       3.60 dB
SFO24       3.60 dB
SFO25       3.60 dB
SFO26       3.60 dB
SFO27       3.60 dB
SFO28       3.60 dB
SFO29       3.60 dB
SFO30       3.60 dB
SFO31       3.60 dB
SFO32       3.60 dB
SFO33       3.60 dB
SFO34       3.60 dB
SFO35       3.60 dB
SFO36       3.60 dB
SFO37       3.60 dB
SFO38       3.60 dB
SFO39       3.60 dB
SFO40       3.60 dB
SFO41       3.60 dB
SFO42       3.60 dB
SFO43       3.60 dB
SFO44       3.60 dB
SFO45       3.60 dB
SFO46       3.60 dB
SFO47       3.60 dB
SFO48       3.60 dB
SFO49       3.60 dB
SFO50       3.60 dB
SFO51       3.60 dB
SFO52       3.60 dB
SFO53       3.60 dB
SFO54       3.60 dB
SFO55       3.60 dB
SFO56       3.60 dB
SFO57       3.60 dB
SFO58       3.60 dB
SFO59       3.60 dB
SFO60       3.60 dB
SFO61       3.60 dB
SFO62       3.60 dB
SFO63       3.60 dB
SFO64       3.60 dB
SFO65       3.60 dB
SFO66       3.60 dB
SFO67       3.60 dB
SFO68       3.60 dB
SFO69       3.60 dB
SFO70       3.60 dB
SFO71       3.60 dB
SFO72       3.60 dB
SFO73       3.60 dB
SFO74       3.60 dB
SFO75       3.60 dB
SFO76       3.60 dB
SFO77       3.60 dB
SFO78       3.60 dB
SFO79       3.60 dB
SFO80       3.60 dB
SFO81       3.60 dB
SFO82       3.60 dB
SFO83       3.60 dB
SFO84       3.60 dB
SFO85       3.60 dB
SFO86       3.60 dB
SFO87       3.60 dB
SFO88       3.60 dB
SFO89       3.60 dB
SFO90       3.60 dB
SFO91       3.60 dB
SFO92       3.60 dB
SFO93       3.60 dB
SFO94       3.60 dB
SFO95       3.60 dB
SFO96       3.60 dB
SFO97       3.60 dB
SFO98       3.60 dB
SFO99       3.60 dB
SFO100      3.60 dB

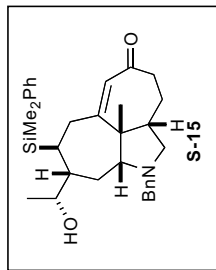
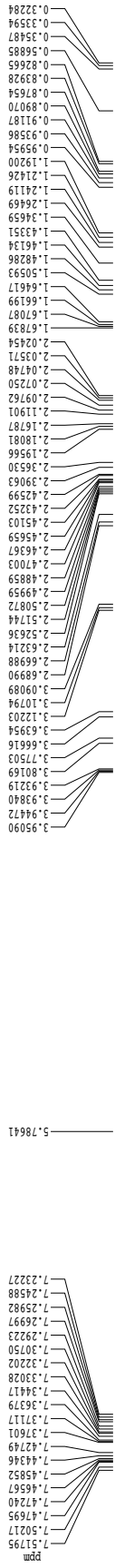
===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2       1H
P2         100.00 usec
P3         60.00 usec
PL0        24.60 dB
PL1        24.60 dB
SFO1       500.2225011 MHz

===== GRADIENT CHANNEL =====
GPMAM1     SINE.100
GPMAM2     SINE.100
GPR1       0.00 %
GPR2       0.00 %
GPR3       0.00 %
GPR4       0.00 %
GPR5       0.00 %
GPR6       0.00 %
GPR7       0.00 %
GPR8       0.00 %
GPR9       0.00 %
GPR10      0.00 %
GPR11      0.00 %
GPR12      0.00 %
GPR13      0.00 %
GPR14      0.00 %
GPR15      0.00 %
GPR16      0.00 %
GPR17      0.00 %
GPR18      0.00 %
GPR19      0.00 %
GPR20      0.00 %
GPR21      0.00 %
GPR22      0.00 %
GPR23      0.00 %
GPR24      0.00 %
GPR25      0.00 %
GPR26      0.00 %
GPR27      0.00 %
GPR28      0.00 %
GPR29      0.00 %
GPR30      0.00 %
GPR31      0.00 %
GPR32      0.00 %
GPR33      0.00 %
GPR34      0.00 %
GPR35      0.00 %
GPR36      0.00 %
GPR37      0.00 %
GPR38      0.00 %
GPR39      0.00 %
GPR40      0.00 %
GPR41      0.00 %
GPR42      0.00 %
GPR43      0.00 %
GPR44      0.00 %
GPR45      0.00 %
GPR46      0.00 %
GPR47      0.00 %
GPR48      0.00 %
GPR49      0.00 %
GPR50      0.00 %
GPR51      0.00 %
GPR52      0.00 %
GPR53      0.00 %
GPR54      0.00 %
GPR55      0.00 %
GPR56      0.00 %
GPR57      0.00 %
GPR58      0.00 %
GPR59      0.00 %
GPR60      0.00 %
GPR61      0.00 %
GPR62      0.00 %
GPR63      0.00 %
GPR64      0.00 %
GPR65      0.00 %
GPR66      0.00 %
GPR67      0.00 %
GPR68      0.00 %
GPR69      0.00 %
GPR70      0.00 %
GPR71      0.00 %
GPR72      0.00 %
GPR73      0.00 %
GPR74      0.00 %
GPR75      0.00 %
GPR76      0.00 %
GPR77      0.00 %
GPR78      0.00 %
GPR79      0.00 %
GPR80      0.00 %
GPR81      0.00 %
GPR82      0.00 %
GPR83      0.00 %
GPR84      0.00 %
GPR85      0.00 %
GPR86      0.00 %
GPR87      0.00 %
GPR88      0.00 %
GPR89      0.00 %
GPR90      0.00 %
GPR91      0.00 %
GPR92      0.00 %
GPR93      0.00 %
GPR94      0.00 %
GPR95      0.00 %
GPR96      0.00 %
GPR97      0.00 %
GPR98      0.00 %
GPR99      0.00 %
GPR100     0.00 %

F2 - Processing parameters
SI         32768
SF         125.7600000 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00

1D NMR plot parameters
F2         125.7600000 MHz
CY         12.65 cm
FIDRES     0.462388 Hz
F1         210.000 ppm
F2P        -10.000 ppm
F2         -1257.80 Hz
FPMCH      9.64912 ppm/cm
HZCN       1213.67078 Hz/cm
    
```

¹H spectrum



S-110

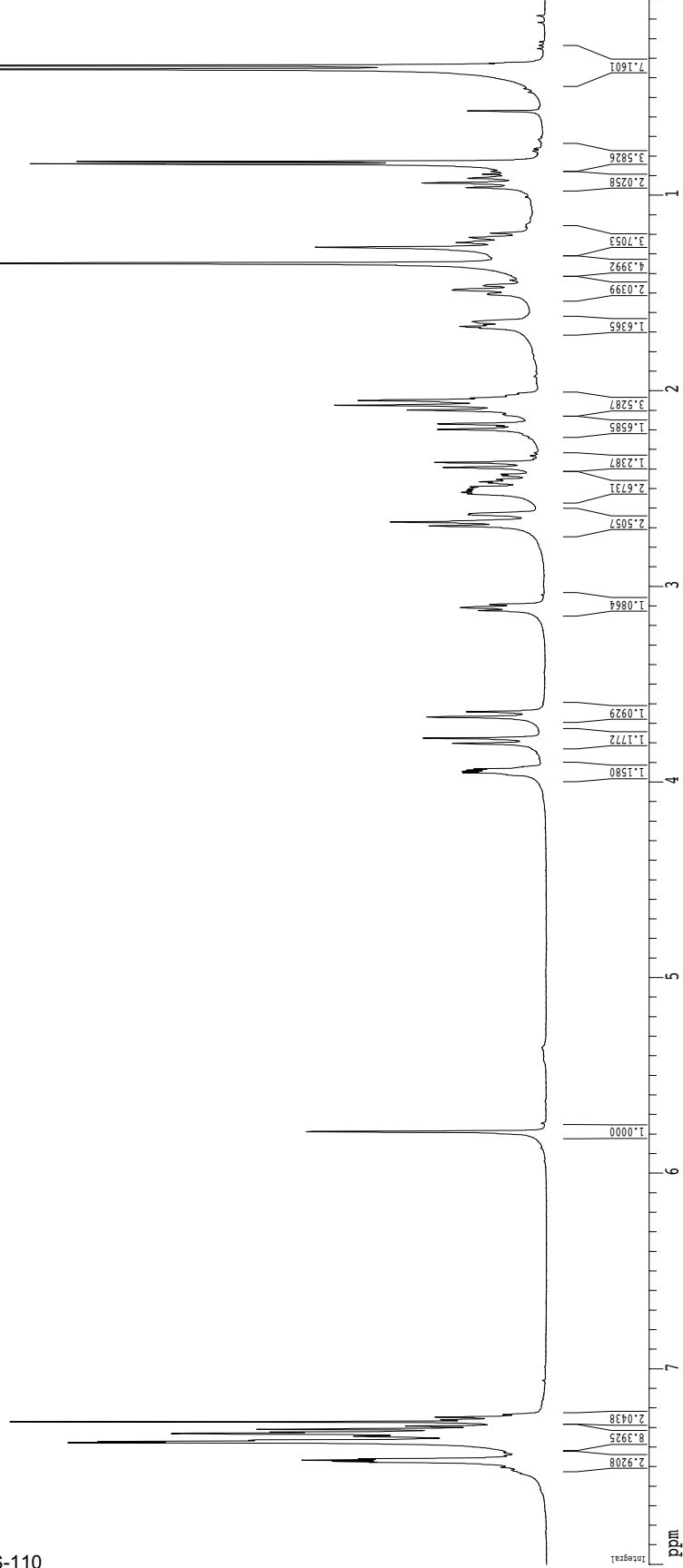
Current Data Parameter
 USER mellis
 NAME ME-II-100-ch
 EXNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20090305
 Time_ 11:27
 INSTRUM cryo500
 PROBRD 5 mm CPCL1
 PULPROG zgpg30
 TD 81728
 SFO1 500.136268
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.099398 sec
 RG 6.3
 DM 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.100000 sec
 dPRST 0.000000 sec
 ACQPRK 0.0150000 sec

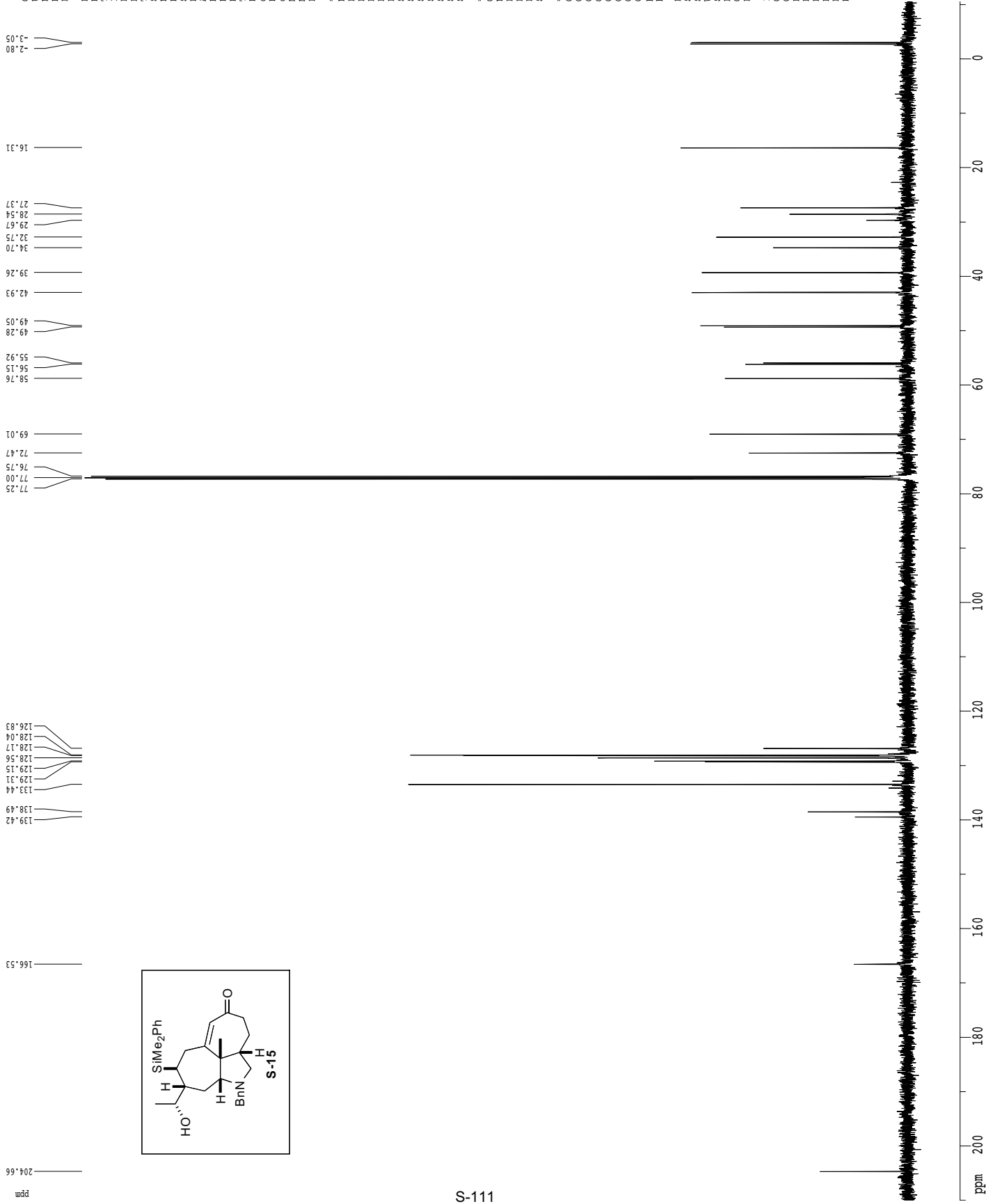
==== CHANNEL f1 =====
 NUC1 ¹H
 P1 7.50 usec
 PL1 1.60 dB
 SFO1 500.2235015 MHz

F2 - Processing parameter
 SI 65536
 SF 500.2200268 MHz
 EQ
 SSB 0
 GB 0.20 Hz
 GR 0
 GC 4.00

1D NMR plot parameters
 CX 22.80 cm
 CY 15.00 cm
 FID 8.000 ppm
 F1 4001.76 Hz
 F2 0.000 ppm
 F3 0.000 Hz
 FWHM 0.35088 ppm/cm
 HZCN 175.51581 Hz/cm



13C spectrum with 1H decoupling



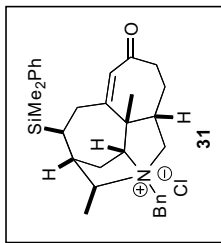
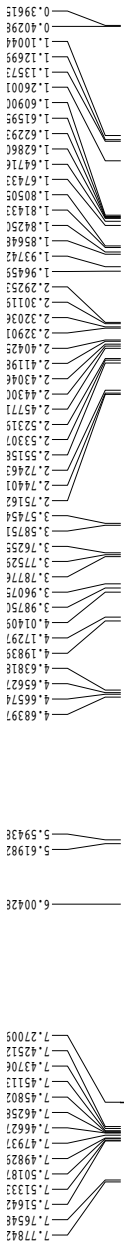
Current Data Parameters
 USER mellis
 NAME ME-II-100-ch
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date 20090305
 Time 11:31
 INSTRUM cryo500
 PROBHD 5 mm CPCTC 1H-
 PULPROG Spinechop90gp-prd
 TD 6556
 SFO1 125.794258 MHz
 SOLVENT CDCl3
 NS 716
 DS 2
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0814105 sec
 RG 4096
 DW 16.500 usec
 DE 6.00 usec
 TE 300.2 K
 D1 0.28000000 sec
 d11 0.03000000 sec
 D16 0.00020000 sec
 d17 0.00019600 sec
 MCREST 0.00000000 sec
 MCREST 0.01500000 sec
 P2 29.70 usec

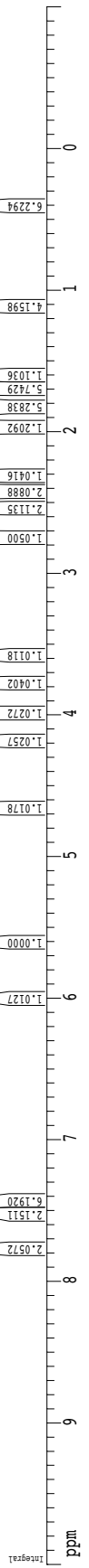
===== CHANNEL f1 =====
 NUC1 13C
 P1 14.85 usec
 P11 500.00 usec
 P12 2000.00 usec
 PL0 120.00 dB
 PL1 -1.00 dB
 SFO1 125.794258 MHz
 SF1 3.60 dB
 SP1 3.60 dB
 SP11 3.60 dB
 SP12 3.60 dB
 SP13 3.60 dB
 SP14 3.60 dB
 SP15 3.60 dB
 SP16 3.60 dB
 SP17 3.60 dB
 SP18 3.60 dB
 SP19 3.60 dB
 SP20 3.60 dB
 SP21 3.60 dB
 SP22 3.60 dB
 SP23 3.60 dB
 SP24 3.60 dB
 SP25 3.60 dB
 SP26 3.60 dB
 SP27 3.60 dB
 SP28 3.60 dB
 SP29 3.60 dB
 SP30 3.60 dB
 SP31 3.60 dB
 SP32 3.60 dB
 SP33 3.60 dB
 SP34 3.60 dB
 SP35 3.60 dB
 SP36 3.60 dB
 SP37 3.60 dB
 SP38 3.60 dB
 SP39 3.60 dB
 SP40 3.60 dB
 SP41 3.60 dB
 SP42 3.60 dB
 SP43 3.60 dB
 SP44 3.60 dB
 SP45 3.60 dB
 SP46 3.60 dB
 SP47 3.60 dB
 SP48 3.60 dB
 SP49 3.60 dB
 SP50 3.60 dB
 SP51 3.60 dB
 SP52 3.60 dB
 SP53 3.60 dB
 SP54 3.60 dB
 SP55 3.60 dB
 SP56 3.60 dB
 SP57 3.60 dB
 SP58 3.60 dB
 SP59 3.60 dB
 SP60 3.60 dB
 SP61 3.60 dB
 SP62 3.60 dB
 SP63 3.60 dB
 SP64 3.60 dB
 SP65 3.60 dB
 SP66 3.60 dB
 SP67 3.60 dB
 SP68 3.60 dB
 SP69 3.60 dB
 SP70 3.60 dB
 SP71 3.60 dB
 SP72 3.60 dB
 SP73 3.60 dB
 SP74 3.60 dB
 SP75 3.60 dB
 SP76 3.60 dB
 SP77 3.60 dB
 SP78 3.60 dB
 SP79 3.60 dB
 SP80 3.60 dB
 SP81 3.60 dB
 SP82 3.60 dB
 SP83 3.60 dB
 SP84 3.60 dB
 SP85 3.60 dB
 SP86 3.60 dB
 SP87 3.60 dB
 SP88 3.60 dB
 SP89 3.60 dB
 SP90 3.60 dB
 SP91 3.60 dB
 SP92 3.60 dB
 SP93 3.60 dB
 SP94 3.60 dB
 SP95 3.60 dB
 SP96 3.60 dB
 SP97 3.60 dB
 SP98 3.60 dB
 SP99 3.60 dB
 SP100 3.60 dB
 SFOF1 Cmp6cm.4
 SFOF2 Cmp6cm.4
 SFOF3 0.00 Hz
 SFOF4 0.00 Hz
 SFOF5 0.00 Hz
 SFOF6 0.00 Hz
 SFOF7 0.00 Hz
 SFOF8 0.00 Hz
 SFOF9 0.00 Hz
 SFOF10 0.00 Hz
 SFOF11 0.00 Hz
 SFOF12 0.00 Hz
 SFOF13 0.00 Hz
 SFOF14 0.00 Hz
 SFOF15 0.00 Hz
 SFOF16 0.00 Hz
 SFOF17 0.00 Hz
 SFOF18 0.00 Hz
 SFOF19 0.00 Hz
 SFOF20 0.00 Hz
 SFOF21 0.00 Hz
 SFOF22 0.00 Hz
 SFOF23 0.00 Hz
 SFOF24 0.00 Hz
 SFOF25 0.00 Hz
 SFOF26 0.00 Hz
 SFOF27 0.00 Hz
 SFOF28 0.00 Hz
 SFOF29 0.00 Hz
 SFOF30 0.00 Hz
 SFOF31 0.00 Hz
 SFOF32 0.00 Hz
 SFOF33 0.00 Hz
 SFOF34 0.00 Hz
 SFOF35 0.00 Hz
 SFOF36 0.00 Hz
 SFOF37 0.00 Hz
 SFOF38 0.00 Hz
 SFOF39 0.00 Hz
 SFOF40 0.00 Hz
 SFOF41 0.00 Hz
 SFOF42 0.00 Hz
 SFOF43 0.00 Hz
 SFOF44 0.00 Hz
 SFOF45 0.00 Hz
 SFOF46 0.00 Hz
 SFOF47 0.00 Hz
 SFOF48 0.00 Hz
 SFOF49 0.00 Hz
 SFOF50 0.00 Hz
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 SFOF70 0.00 Hz
 SFOF71 0.00 Hz
 SFOF72 0.00 Hz
 SFOF73 0.00 Hz
 SFOF74 0.00 Hz
 SFOF75 0.00 Hz
 SFOF76 0.00 Hz
 SFOF77 0.00 Hz
 SFOF78 0.00 Hz
 SFOF79 0.00 Hz
 SFOF80 0.00 Hz
 SFOF81 0.00 Hz
 SFOF82 0.00 Hz
 SFOF83 0.00 Hz
 SFOF84 0.00 Hz
 SFOF85 0.00 Hz
 SFOF86 0.00 Hz
 SFOF87 0.00 Hz
 SFOF88 0.00 Hz
 SFOF89 0.00 Hz
 SFOF90 0.00 Hz
 SFOF91 0.00 Hz
 SFOF92 0.00 Hz
 SFOF93 0.00 Hz
 SFOF94 0.00 Hz
 SFOF95 0.00 Hz
 SFOF96 0.00 Hz
 SFOF97 0.00 Hz
 SFOF98 0.00 Hz
 SFOF99 0.00 Hz
 SFOF100 0.00 Hz

¹H spectrum

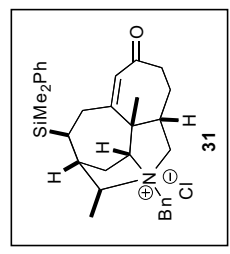
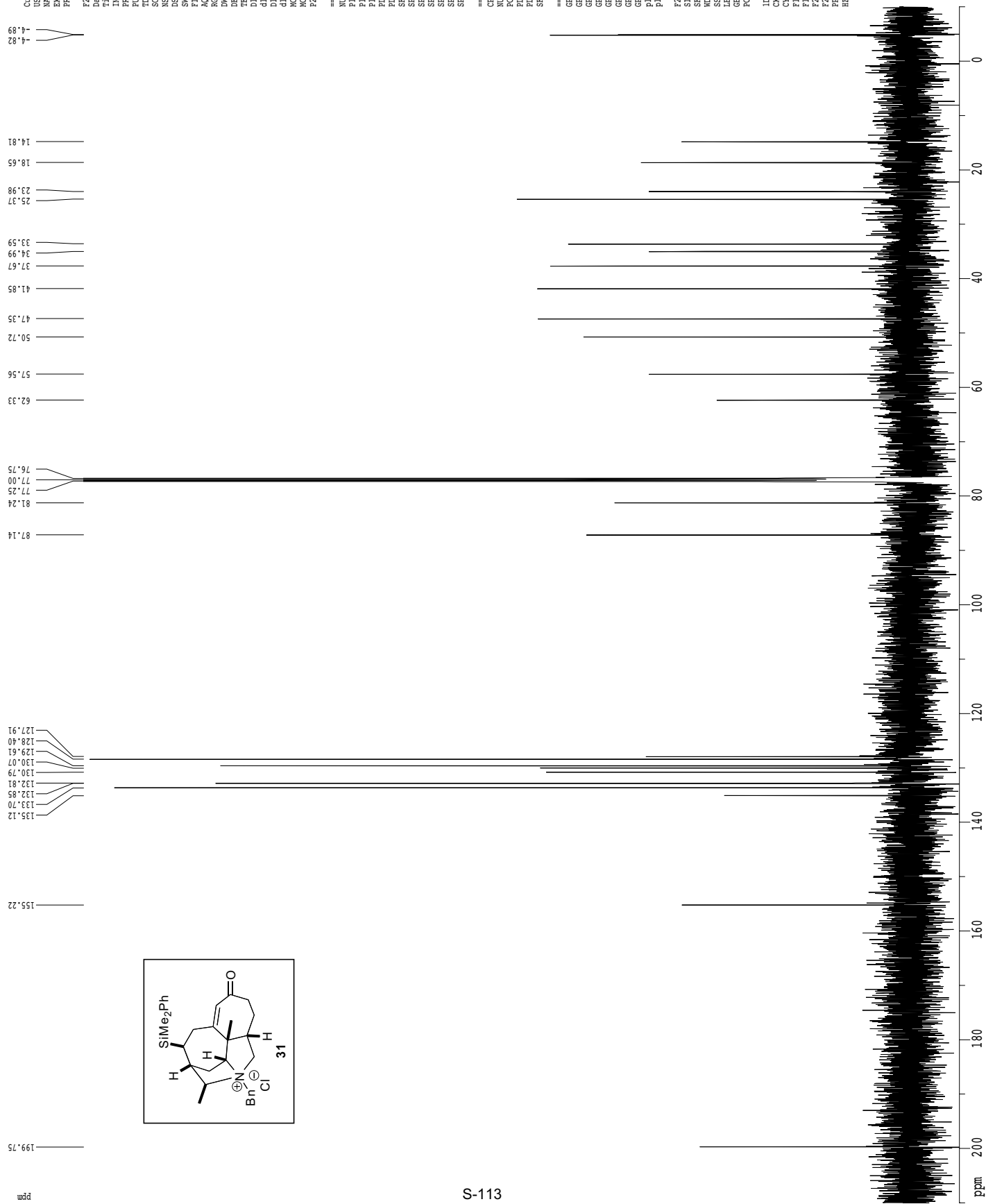
ppm



Current Data Parameters
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 SAMPLE jfm-1-170 ff 4
 EXPTNO 1
 PROCNO 1
 F2 - Acquisition Parameters
 Date 20/09/09
 Time 15.50
 INSTRUM cryo500
 PROBRD 5 mm CPCL1.H
 PULPROG zg30
 TD 81728
 SOLVENT CDCl3
 NS 51
 DS 2
 SWH 8012.822 Hz
 FIDRES 0.189603 Hz
 AQ 5.0398774 sec
 RG 5
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCWRR 0.01500000 sec
 ===== CHANNEL f1 =====
 NUC1 1H
 P1 7.20 usec
 PL 0.00 dB
 SFO1 500.233015 MHz
 F2 - Processing parameters
 SI 65536
 SF 500.220261 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 4.00
 ID NMR plot parameters
 CX 22.80 cm
 CY 15.00 cm
 CZ 10.00 cm
 F1 5002.20 Hz
 F2 -1.000 ppm
 F2 -500.22 Hz
 PPMCH 0.48246 ppm/cm
 HZCM 241.33423 Hz/cm



¹³C spectrum with ¹H decoupling



Current Data Parameters
 USER manlin
 NAME jmc-1-170 fr 4
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20090609
 Time_ 16:31
 Date_ 20090609
 Time_ 16:31
 PULPROG zgpg30
 PROGRAM 5 mm CPMAS JH
 PULPROG Spinechop30pp.prd
 TD 65536
 SOLVENT CMC13
 NS 2429
 DS 16
 SWH 30003.031 Hz
 FIDRES 0.48268 Hz
 AQ 1.00132 sec
 RG 3298.2 sec
 DM 16.500 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.25000000 sec
 d11 0.03000000 sec
 D16 0.00020000 sec
 F1 0.00000000 sec
 F2 0.00000000 sec
 MCHRG 0.01500000 sec
 MCHRG 0.01500000 sec
 F2 29.70 usec

==== CHANNEL f1 =====
 NUC1 ¹³C
 P1 14.85 usec
 PL1 500.00 usec
 PL2 500.00 usec
 PL3 500.00 usec
 PL4 500.00 usec
 PL5 500.00 usec
 PL6 500.00 usec
 PL7 500.00 usec
 PL8 500.00 usec
 PL9 500.00 usec
 PL10 500.00 usec
 PL11 500.00 usec
 PL12 500.00 usec
 SF01 125.7942548 MHz
 SF1 3.60 dB
 SF2 3.60 dB
 SFRAM1 Cnp60, 0.5, 20.1
 SFRAM2 Cnp60comp.4
 SFOFF1 0.00 Hz
 SFOFF2 0.00 Hz

==== CHANNEL f2 =====
 CPMRG2 waltz16
 NUC2 ¹H
 PCDP2 100.00 usec
 PL2 1.60 dB
 PL3 1.60 dB
 SF02 500.2225011 MHz

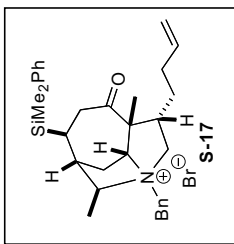
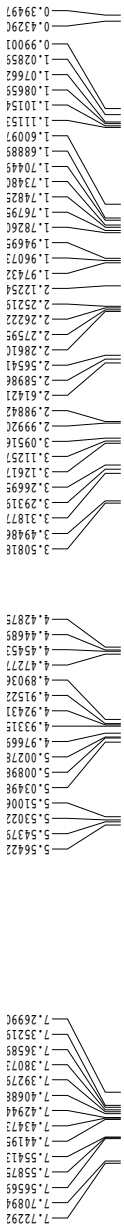
==== GRADIENT CHANNEL =====
 GPMAM1 SINE.100
 GPMAM2 SINE.100
 GPK1 0.00 %
 GPK2 0.00 %
 GPK3 0.00 %
 GPK4 0.00 %
 GPK5 0.00 %
 GPK6 0.00 %
 GPK7 0.00 %
 GPK8 0.00 %
 GPK9 0.00 %
 GPK10 0.00 %
 GPK11 0.00 %
 GPK12 0.00 %
 GPK13 0.00 %
 GPK14 0.00 %
 GPK15 0.00 %
 GPK16 0.00 %
 GPK17 0.00 %
 GPK18 0.00 %
 GPK19 0.00 %
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 GPK37 0.00 %
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 GPK42 0.00 %
 GPK43 0.00 %
 GPK44 0.00 %
 GPK45 0.00 %
 GPK46 0.00 %
 GPK47 0.00 %
 GPK48 0.00 %
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 GPK57 0.00 %
 GPK58 0.00 %
 GPK59 0.00 %
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 GPK62 0.00 %
 GPK63 0.00 %
 GPK64 0.00 %
 GPK65 0.00 %
 GPK66 0.00 %
 GPK67 0.00 %
 GPK68 0.00 %
 GPK69 0.00 %
 GPK70 0.00 %
 GPK71 0.00 %
 GPK72 0.00 %
 GPK73 0.00 %
 GPK74 0.00 %
 GPK75 0.00 %
 GPK76 0.00 %
 GPK77 0.00 %
 GPK78 0.00 %
 GPK79 0.00 %
 GPK80 0.00 %
 GPK81 0.00 %
 GPK82 0.00 %
 GPK83 0.00 %
 GPK84 0.00 %
 GPK85 0.00 %
 GPK86 0.00 %
 GPK87 0.00 %
 GPK88 0.00 %
 GPK89 0.00 %
 GPK90 0.00 %
 GPK91 0.00 %
 GPK92 0.00 %
 GPK93 0.00 %
 GPK94 0.00 %
 GPK95 0.00 %
 GPK96 0.00 %
 GPK97 0.00 %
 GPK98 0.00 %
 GPK99 0.00 %
 GPK100 0.00 %

F2 - Processing parameters
 SI 65536
 SF 125.7804277 MHz
 WDW EM
 SSB 0
 GB 0
 PC 2.00

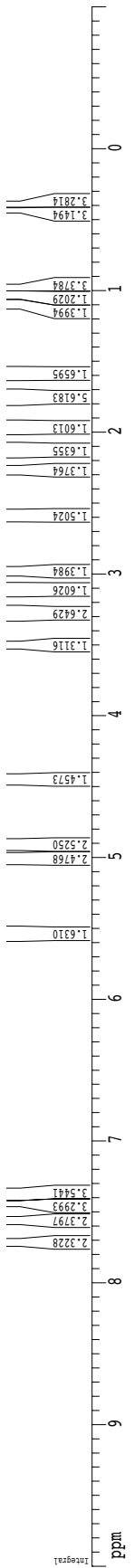
ID NMR plot parameters
 CX 22.80 cm
 CY 15.65 cm
 FIP 210.000 ppm
 F1 2643.839 Hz
 F2 125780.477 MHz
 PPMCM 9.64812 ppm/cm
 HzCM 1213.67078 Hz/cm

¹H spectrum

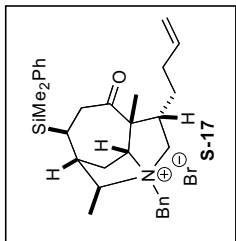
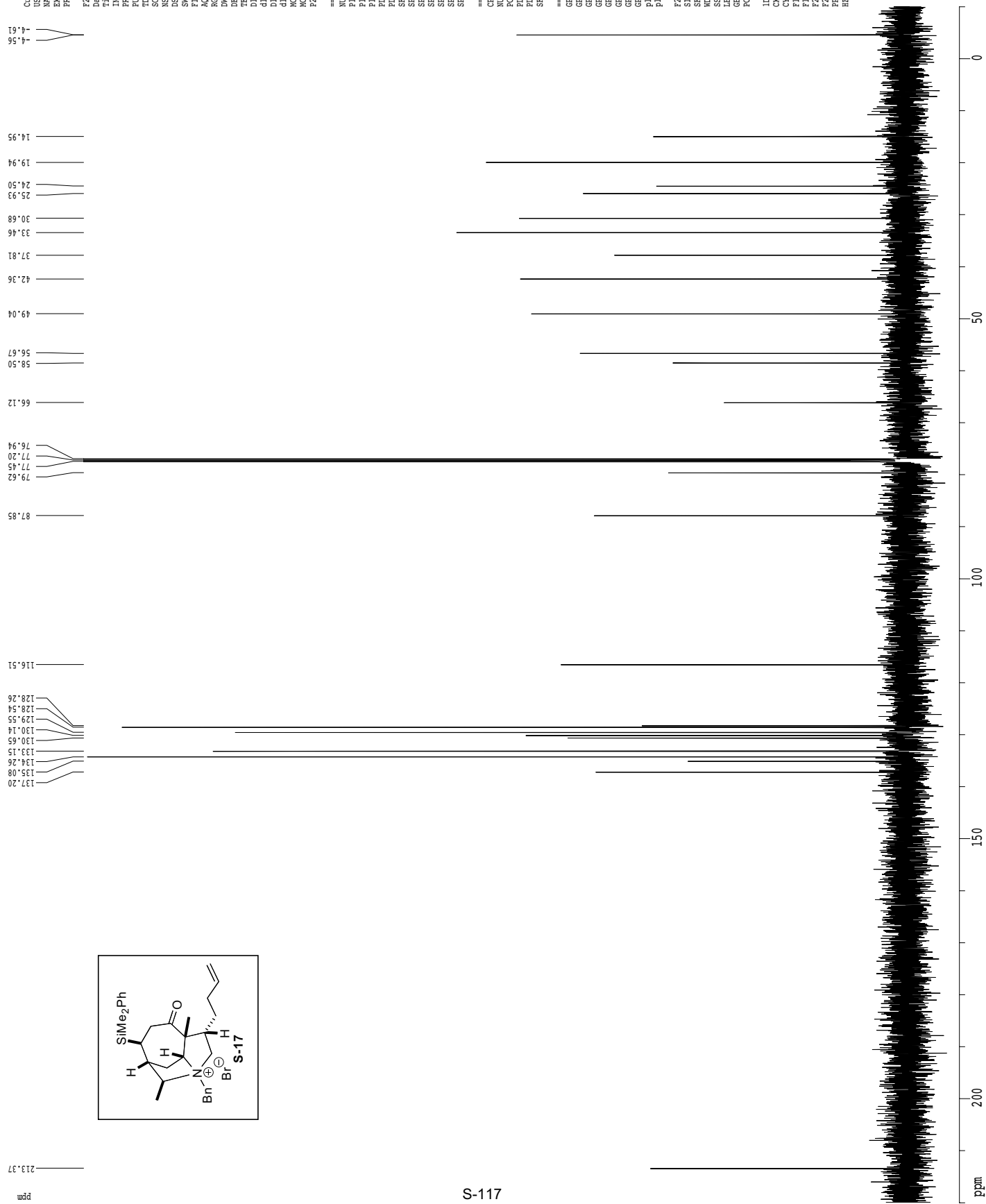
ppm



Current Data Parameters
 USER manlin
 SAMPLE Jrm-1-200
 EXPTNO 1
 PROCNO 1
 F2 - Acquisition Parameters
 Date 20/09/04
 Time 7.22
 INSTRUM cryo500
 PROBHD 5 mm CPCL1 IH-
 PULPROG zg30
 TD 81728
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 8013.822 Hz
 FIDRES 0.189603 Hz
 AQ 5.0398774 sec
 RG 6.3
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCWRR 0.01500000 sec
 ===== CHANNEL f1 =====
 NUC1 ¹H
 P1 7.20 usec
 PL 0.00 dB
 SFO1 500.2335015 MHz
 F2 - Processing parameters
 SI 65536
 SF 500.220263 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 4.00
 ID NMR plot parameters
 CX 22.80 cm
 CY 15.00 cm
 CZ 10.00 cm
 F1 5002.20 Hz
 F2 -1.000 ppm
 F2 -500.22 Hz
 PPMCH 0.48246 ppm/cm
 HZCM 241.33423 Hz/cm



13C spectrum with 1H decoupling



Current Data Parameters
 USER manlin
 NAME jtm-120
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 2009024
 Time_ 12:27
 Time2_ 12:27
 PULPROG Spinechop30pp.prd
 PROBRD 5 mm CPYCI 1H
 TD 65536
 SOLVENT CDCl3
 NS 329
 DS 16
 SWH 30003.031 Hz
 FIDRES 0.48398 Hz
 AQ 1.03132 sec
 RG 3298.2 sec
 DM 16.500 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.25000000 sec
 d11 0.03000000 sec
 D16 0.00020000 sec
 F1 0.00000000 sec
 F2 0.00000000 sec
 MCHRG 0.01500000 sec
 MCHRR 0.01500000 sec
 P2 31.00 usec

==== CHANNEL f1 =====
 NUC1 13C
 P1 15.50 usec
 PL1 500.00 usec
 F1 125.760404 MHz
 PL2 1.60 dB
 P11 -1.00 dB
 P12 -1.00 dB
 SF01 125.7942548 MHz
 SF1 3.20 dB
 SF2 3.20 dB
 SFRM1 Ctp60, 0.520.1
 SFRM2 Ctp60comp.4
 SFOF1 0.00 Hz
 SFOF2 0.00 Hz

==== CHANNEL f2 =====
 CDPGR2 waltz16
 NUC2 1H
 PCDP2 100.00 usec
 PL2 1.60 dB
 PL12 24.60 dB
 SF02 500.2225011 MHz

==== GRADIENT CHANNEL =====
 GPM1 SINE.100
 GPM2 SINE.100
 GPK1 0.00 %
 GPK2 0.00 %
 GFL1 0.00 %
 GFL2 0.00 %
 GFL3 30.00 %
 GFL4 30.00 %
 P16 500.00 usec
 P17 1000.00 usec

F2 - Processing parameters
 SI 65536
 SF 125.760404 MHz
 WDW EM
 SSB 0
 GB 0
 PC 2.00

ID NMR plot parameters
 CX 22.80 cm
 CY 15.65 cm
 FIP 270.000 ppm
 F1 27671.69 Hz
 F2 27671.69 Hz
 F3 -1257.80 Hz
 PPMCM 10.0872 ppm/cm
 RECM 1268.83740 Hz/cm