

Supplementary material

Figure S1. Alignment of SERK proteins

Amino acid sequence alignment of deduced *N. attenuata* NaBAK1, NaSERK1 (partial), *N. benthamiana* NbSERK3/BAK1 (partial), and other SERK proteins. NCBI (GenBank) accession numbers of *SERK* genes and SERK proteins included in this alignment are: *N. attenuata*: NaBAK1 (HM639279), NaSERK1 (HM639280); *N. benthamiana*: NbSERK3/BAK1 (CK291393); *Arabidopsis thaliana*: AtSERK1 (NP_177328), AtSERK2 (NP_174683), AtSERK3 (NP_567920), AtSERK4 (NP_178999), AtSERK5 (NP_179000); *Solanum tuberosum*: StSERK1 (ABO14172); *Medicago truncatula*: MtSERK1 (AAN64294); *Oryza sativa*: OsSERK1 (BAD86793); *Zea mays*: ZmSERK1 (CAC37638); ZmSERK2 (CAC37639). Amino acids in white background are different from the consensus sequence.

Figure S2. Morphology of EV and NaBAK1-VIGS plants

N. attenuata plants were inoculated with *Agrobacterium* carrying pTV00 empty vector or pTV-NaBAK1 to generate EV and NaBAK1-silenced (NaBAK1-VIGS) plants, respectively. (A) Plants at early elongation stage. (B) Plants at flowering stage.

Figure S3. NaBAK1 positively regulates the W+OS- and W+W-induced accumulation of jasmonic acid (JA) and JA-isoleucine (JA-Ile).

Leaves of EV and NaBAK1-VIGS plants were wounded with a pattern wheel and 20 μ l of water (W+W) or *M. sexta* oral secretions (W+OS) were immediately applied to each leaf. Individual leaves from 5 replicate plants were harvested at the indicated times. Asterisks represent significantly different levels between EV and NaBAK1-VIGS plants at indicated times (unpaired *t*-test; *, $p < 0.05$; **, $p < 0.01$; ***, $p < 0.001$; N = 5). (A) Mean (\pm SE) JA concentrations in EV and NaBAK1-VIGS plants. (B) Mean (\pm SE) JA-Ile concentrations in EV and NaBAK1-VIGS

plants.

Figure S4. EV and NaBAK1-VIGS plants have the same levels of SA and ethylene production after W+OS induction.

Leaves of EV and NaBAK1-VIGS plants were wounded with a pattern wheel and 20 μ l of water (W+W), *M. sexta* oral secretions (W+OS), or FAC A solution (W+FAC) were immediately applied to each leaf. Individual leaves from 5 replicate plants were harvested after 1.5 h.

Asterisks represent significantly different levels between EV and NaBAK1-VIGS plants at indicated times (unpaired *t*-test; *, $p < 0.05$; **, $p < 0.01$; ***, $p < 0.001$; N = 5). (A) Mean SA concentrations (\pm SE) in EV and NaBAK1-VIGS plants. Leaves of EV and NaBAK1-VIGS plants were wounded with a pattern wheel and treated with 20 μ l of *M. sexta* OS (W+OS). (B) Ethylene emission (\pm SE) was measured using a photoacoustic spectrometer 5 h after W+OS treatment (N = 5).

Figure S5. Transcript accumulation of JA biosynthetic genes in EV and NaBAK1-VIGS plants

Mean transcript levels (\pm SE) of *NaLOX3* (A), *NaAOS* (B), *NaAOC* (C), *NaOPR3* (D), and *NaACX1* (E) were measured with qPCR. Asterisks represent significantly different levels between EV and NaBAK1-VIGS plants at indicated times (unpaired *t*-test; *, $p < 0.05$; **, $p < 0.01$; ***, $p < 0.001$; N = 5).

Figure S6. EV and NaBAK1-VIGS plants have similar levels of DTGs in response to W+OS and MeJA treatment

(A) Leaves from EV and NaBAK1-VIGS plants were left untreated (control) or wounded with a pattern wheel and 20 μ l of *M. sexta* oral secretions were immediately applied to each leaf (W+OS). Samples were harvested three days after treatment, and DTG content were determined.

(B) EV and NaBAK1-VIGS leaves were treated with 20 μl of lanolin or lanolin containing MeJA ($7.5 \mu\text{g } \mu\text{l}^{-1}$). Samples were harvested three days after treatment, and DTG content were determined. Error bars represent standard errors; different letters indicate significant differences (two-way ANOVA, Fisher's PLSD test; $p < 0.05$; $N = 5$).

Figure S7. *M. sexta* has similar growth rates on EV and NaBAK1-VIGS plants

Each EV and NaBAK1-VIGS plant was infested with one *M. sexta* neonate. The larval mass was subsequently recorded on day 6, 8, and 11 ($N = 25$).

MtSERK1	----MEET--KFCALAFICAFFLLLHPLWLVSANMEGDALHNLRTNLQD---PNNVLQ	50
OsSERK1	----MAHR---WAVWAVLLLRLLVP--AARVLANMEGDALHSLRTNLVD---PNNVLQ	47
ZmSERK1	----MAASL---RWWWSAVVFSVVVG--VIFVVANTEGDALYSLRQSLKD---NNNVLQ	47
StSERK1	----MVKVMEKDAVVVSLVWVLLLVVHHLKLIYANMEGDALHSLRVNLQD---PNNVLQ	52
NaSERK1	-----	0
AtSERK1	----MES----SYVVFILLSLILLPNHSLWLASANLEGDALHILRVTLVD---PNNVLQ	48
AtSERK2	----MGRKKFEAFGFVCLISLLLLFN-SLWLASNNMEGDALHSLRANLVD---PNNVLQ	51
ZmSERK2	----MAASASAGRWWAVVLAVAVLLG--PGQVVANTEGDALYSLRQSLKD---ANNVLQ	50
AtSERK3/BAK1	----MERRLMIPCFFWLILVLDLVLRLR---VSENAEGDALSAKNSLAD---PNKVLQ	47
NaSERK3/BAK1	----MDQWILG--ILGFVSAFLCLIGLLLVESANIEGDALNALKTNLAD---PNNVLQ	50
NbSERK3/BAK1	-----	0
AtSERK4/BKK1	MTSSKMEQRSLL-CFLYLLLLFNFTLR---VAGNAEGDALTQKNSLSSGDPANNVLQ	54
AtSERK5	----MEHGSSR-GFIWLLLELDFVSR---VTGKTQVDALIALRSSLSSGDHTNNILQ	49
MtSERK1	SWDPTLVNPTWFHVTCNNDNSVIRVDLGNAA LSGTLVPQLGQLKNLQYLELYSNNITGP	110
OsSERK1	SWDPTLVNPTWFHVTCNNDNSVIRVDLGNAA LSGTLVPQLGQLKNLQYLELYSNNISGT	107
ZmSERK1	SWDPTLVNPTWFHVTCNNDNSVIRVDLGNAA LSGELVPQLGQLKNLQYLELYSNNISGP	107
StSERK1	SWDPTLVNPTWFHVTCNNDNSVIRVDLGNAA LSGTLVPQLGQLKNLQYLELYSNNISGL	112
NaSERK1	-----	0
AtSERK1	SWDPTLVNPTWFHVTCNNDNSVIRVDLGNAA LSGHLVPELGLKKNLQYLELYSNNITGP	108
AtSERK2	SWDPTLVNPTWFHVTCNNDNSVIRVDLGNAA LSGQLVPQLGQLKNLQYLELYSNNITGP	111
ZmSERK2	SWDPTLVNPTWFHVTCNNDNSVIRVDLGNAA LSGVLVPQLGQLKNLQYLELYSNNISGT	110
AtSERK3/BAK1	SWDATLVNPTWFHVTCNNDNSVIRVDLGNAA LSGQLVLPQLGQLKNLQYLELYSNNITGT	107
NaSERK3/BAK1	SWDPTLVNPTWFHVTCNNDNSVIRVDLGNAA LSGQLVPQLGQLKNLQYLELYSNNISGR	110
NbSERK3/BAK1	-----	0
AtSERK4/BKK1	SWDATLVNPTWFHVTCNNDNSVIRVDLGNAA LSGKLVPELGLKNLQYLELYSNNITGE	114
AtSERK5	SWNATLVNPTWFHVTCNNDNSVIRVDLGNAA LSGELVPQLGQLKNLQYLELYSNNITGE	109
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OsSERK1	IPSELGNTNLVSLDLYLNRFNGPI PDSLGKLLKLRFLRLNNSLSGSIPIKSLTALITALQ	167
ZmSERK1	IPPELGNTNLVSLDLYLNRFNGPI PDITLGLSKLRFLRLNNSLSGQIPIKTLTNINTLQ	167
StSERK1	IPSDLGNTNLVSLDLYLNRFNGPI PDSLGKLSKLRFLRLNNSLTGNIIPMSLTNISSLQ	172
NaSERK1	-----	0
AtSERK1	IPSNLGNTNLVSLDLYLNRFNGPI PELGLKLSKLRFLRLNNSLTGSIIPMSLTNITTLQ	168
AtSERK2	VPSDLGNTNLVSLDLYLNRFNGPI PDSLGKLEKLRFLRLNNSLTGPIIPMSLTNIMTLQ	171
ZmSERK2	IPPELGNTNLVSLDLYLNRFNGPI PDSLGKLVKLRFLRLNNSLVGPIIPMSLTNISTLQ	170
AtSERK3/BAK1	IPEQLGNTNLVSLDLYLNRFNGPI PSTLGLKLRFLRLNNSLSGEIIPMSLTAVITLQ	167
NaSERK3/BAK1	IPPELGNTNLVSLDLYLNRFNGPI PDITLGLKLRFLRLNNSLVGRIIPMLTIVISLQ	170
NbSERK3/BAK1	-----	0
AtSERK4/BKK1	IPEELGNTNLVSLDLYLNRFNGPI PSISGLKLGKLRFLRLNNSLSGEIIPMLTISVQ-LQ	173
AtSERK5	IPEELGNTNLVSLDLYLNRFNGPI PSISGLKLGKLRFLRLNNSLSGEIIPMSLTALP-LD	168

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 ZmSERK1 VLDLSNNNLSGVEPSSGFSLSLFTPISFANNPNLCGPSTTKPCPGAPPFSPPPPYNPPAP- 226
 StSERK1 VLDLSNNRSLGVVPDNGSFSLSLFTPISFANNLLDLCGPVTGRPCPGSPPFSPPPPFVPPPI 232
 NaSERK1 ----- 0
 AtSERK1 VLDLSNNRSLGSVVPDNGSFSLSLFTPISFANNLLDLCGPVTSHPCPGSPPFSPPPPFVPPPI 228
 AtSERK2 VLDLSNNRSLGSVVPDNGSFSLSLFTPISFANNLLDLCGPVTSRCPGSPPFSPPPPFVPPPI 231
 ZmSERK2 VLDLSNNNLSGQVPESTGFSLSLFTPISFANNPNLCGPSTSKPCPGAPPFSPPPPFNPPSP 230
 AtSERK3/BAK1 VLDLSNNELTGDIPVNGSFSLSLFTPISFANTKLTP-----LP-----ASPPPFISPTPP 215
 NaSERK3/BAK1 VLDLSNNNLTGFEVNGSFSLSLFTPISFANNP-----LDIPPAAPPPFISPTPTS 219
 NbSERK3/BAK1 ----- 0
 AtSERK4/BKK1 VLDTSNRSLSGDIPVNGSFSLSLFTPISFANNSLTD-----LP-----PPPTSTSTPTTP 221
 AtSERK5 VLDTSNRSLSGDIPVNGSFSLSLFTPISFANNKLR-----PPPASPSPS-- 210

MtSERK1 SPGSGGATGAIAGGVAAGAALLFAAPAIAFAWRRRKQEEFFDVPAAEDPEVHLGQLK 290
 OsSERK1 QSPGSSSTGAIAGGVAAGAALLFAAPAIAFAWRRRKQEEFFDVPAAEDPEVHLGQLK 287
 ZmSERK1 TSSKGVSSSTGAIAGGVAAGTALLFAAPAIAFAWRRRKQEEFFDVPAAEDPEVHLGQLK 286
 StSERK1 SPGNGATGAIAGGVAAGAALLFAAPAIAFAWRRRKQEEFFDVPAAEDPEVHLGQLK 292
 NaSERK1 ----- 0
 AtSERK1 STPSGYGITGAIAGGVAAGAALLFAAPAIAFAWRRRKQLDFFDVPAAEDPEVHLGQLK 288
 AtSERK2 PTPGGYSATGAIAGGVAAGAALLFAAPAIAFAWRRRKQEEFFDVPAAEDPEVHLGQLK 291
 ZmSERK2 TQSTGASSTGAIAGGVAAGAALLFAAPAIAFAWRRRKQEEFFDVPAAEDPEVHLGQLK 290
 AtSERK3/BAK1 SPAGSNRITGAIAGGVAAGAALLFAAPAIAFAWRRRKQDHFDDVPAAEDPEVHLGQLK 275
 NaSERK3/BAK1 SSGVGNISATGAIAGGVAAGAALLFAAPAIAFAWRRRKQDHFDDVPAAEDPEVHLGQLK 279
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 AtSERK4/BKK1 PPSGG-QMTAIAAGGVAAGAALLFAAPAIAFAWRRRKQDHFDDVPAAEDPEVHLGQLK 280
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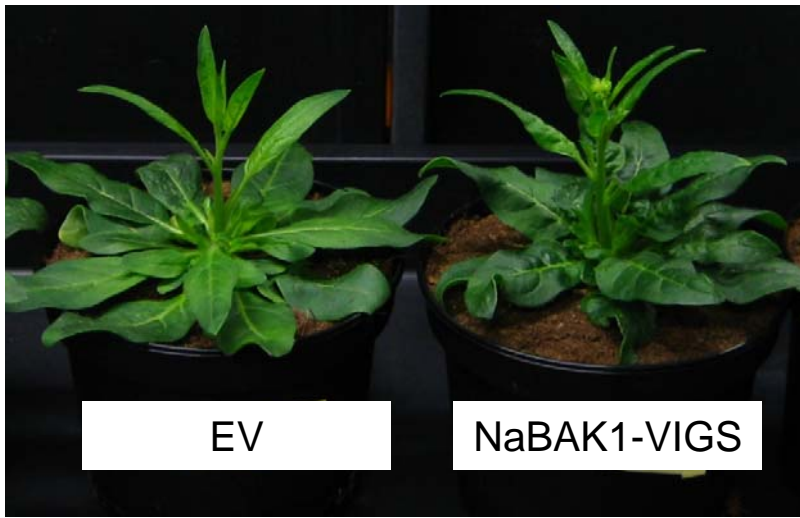
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NbSERK3/BAK1	-----	0
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ZmSERK1	IGHIAPEYLSHGKSSSEKTDVFGYGYMELLEITGQRAFDLARLANDDDVMLLDWVKGLLKE	526
StSERK1	IGHIAPEYLSHGKSSSEKTDVFGYGYMELLEITGQRAFDLARLANDDDVMLLDWVKGLLKE	532
NaSERK1	IGHIAPEYLSHGKSSSEKTDVFGYGYMELLEITGQRAFDLARLANDDDVMLLDWVKGLLKE	148
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ZmSERK2	IGHIAPEYLSHGKSSSEKTDVFGYGYMELLEITGQRAFDLARLANDDDVMLLDWVKGLLKE	530
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NaSERK3/BAK1	IGHIAPEYLSHGKSSSEKTDVFGYGYMELLEITGQRAFDLARLANDDDVMLLDWVKGLLKE	519
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AtSERK5	IGHIAPEYLSHGKSSSEKTDVFGYGYMELLEITGQRAFDLARLANDDDVMLLDWVKGLLKE	501

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OsSERK1	KKLEMLVDPDLQSNYLDVEVESLIQVALLCTQSSPMERPKMSEVVRMLEGDGLAERWEW	587
ZmSERK1	KKLEMLVDPDLQGRYVDQEVESLIQVALLCTQSSPMERPKMSEVVRMLEGDGLAERWEW	586
StSERK1	KKLEMLVDPDLQNKYVEAEVEQLIQVALLCTQSSPMERPKMSEVVRMLEGDGLAERWEW	592
NaSERK1	KKLEMLVDPDLQNKYVEAEVEQLIQVALLCTQSSPMERPKMSEVVRMLEGDGLAERWEW	208
AtSERK1	KKLEMLVDPDLQINYEVEVELEQLIQVALLCTQSSPMERPKMSEVVRMLEGDGLAERWEW	588
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NbSERK3/BAK1	KKYEMLVDADLQGNYEVEVEQLIQVALLCTQSSPMERPKMSEVVRMLEGDGLAERWEW	101
AtSERK4/BKK1	KKLESILVDAELEGKYVEVEVEQLIQVALLCTQSSPMERPKMSEVVRMLEGDGLAERWEW	580
AtSERK5	KKLESILVDAELEGKYVEVEVEQLIQVALLCTQSSPMERPKMSEVVRMLEGDGLAERWEW	561
MtSERK1	QKGEVLRQEVVELAPHPNS--DWIV--DSTENLHAVELSGPR	627
OsSERK1	QKIEVVRQEVVELGPHNS--EWIV--DSTENLHAVELSGPR	624
ZmSERK1	QKVEVVRQEVVELAPRHN---DWIV--DSTYNLRAVELSGPR	622
StSERK1	QKVEVLRQEVVELAPHPNS--DWIV--DSTENLHAVELSGPR	629
NaSERK1	QKVEVLRQEVVELAPHPNS--DWIV--DSTENLHAVELSGPR	245
AtSERK1	QKVEVLRQEVVELAPHPNS--DWIV--DSTENLHAVELSGPR	625
AtSERK2	QKVEVLRQEVVELSSHPTS--DWIL--DSTENLHAVELSGPR	628
ZmSERK2	QKVEVVRQEVVELAPLRN---DWIV--DSTYNLRAVELSGPR	626
AtSERK3/BAK1	QKEEMVRQDFNYPTHHPAVSGWIIIGDSTSQIENEYESGPR	615
NaSERK3/BAK1	QKEEMVRQDYP-AHHPHT--DWIIGDSTYNLRPEELSGPR	616
NbSERK3/BAK1	QKEEMVRQDYP-AHHPHT--DWIIGDSTYNLRPEELSGPR	138
AtSERK4/BKK1	QKEEMPIHDFNYQAYPHAGTDWLIPIYSNLIENDYESGPR	620
AtSERK5	QKEEMPIHDFNYQAYPHAGTDWLIPIYSNLIENDYESGPR	601

Figure S1

A



B

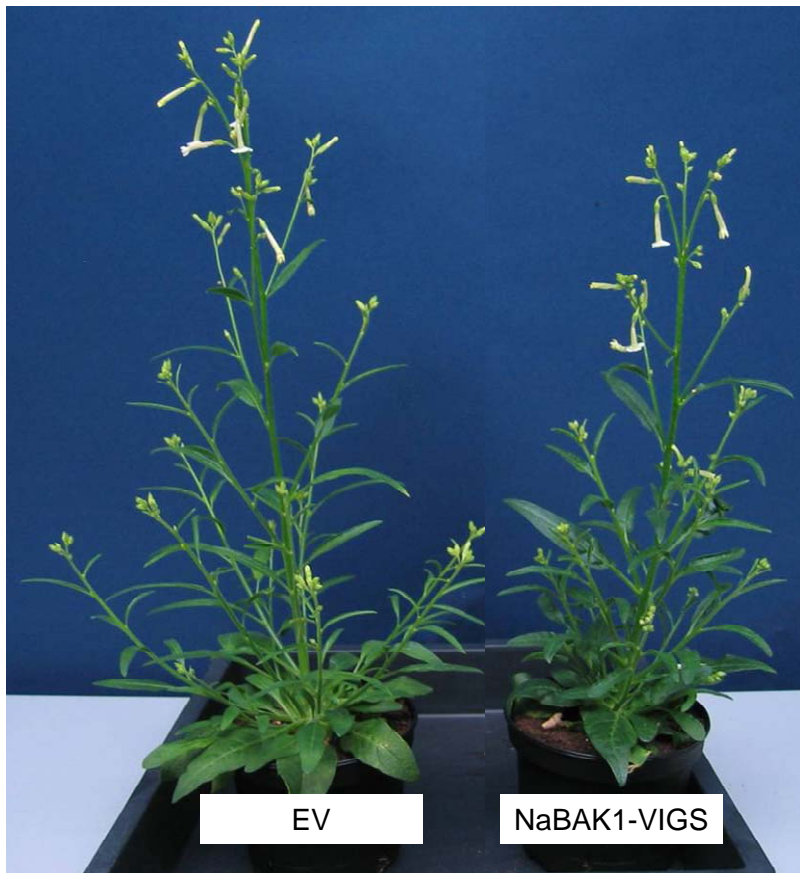


Figure S2

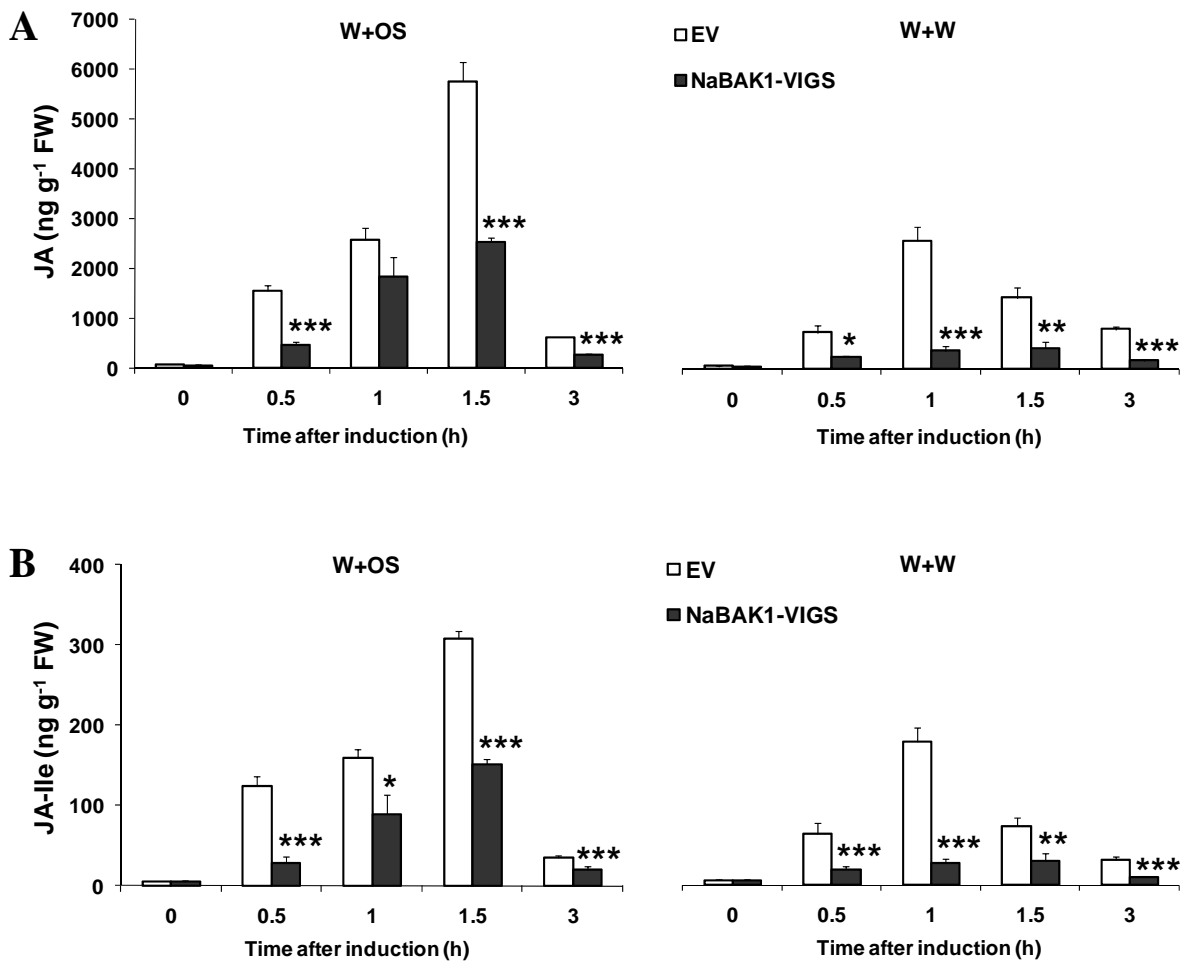


Figure S3

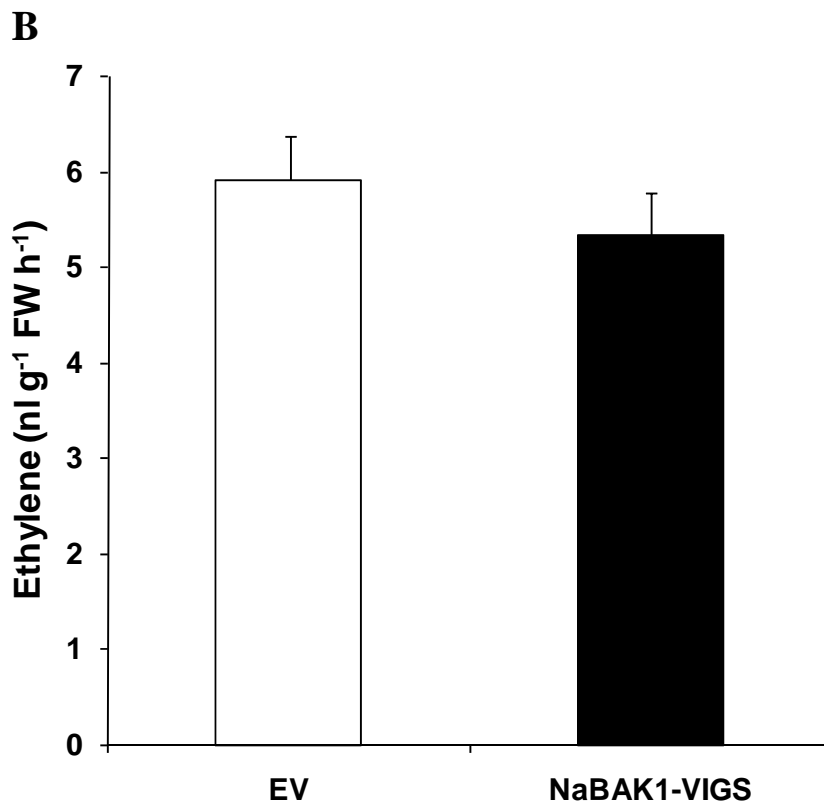
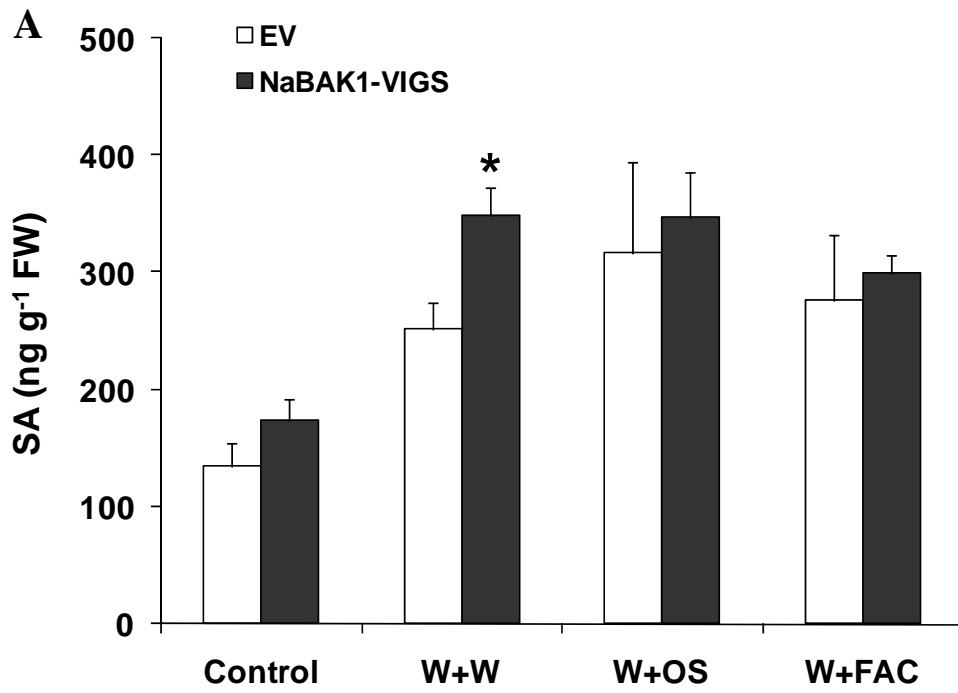
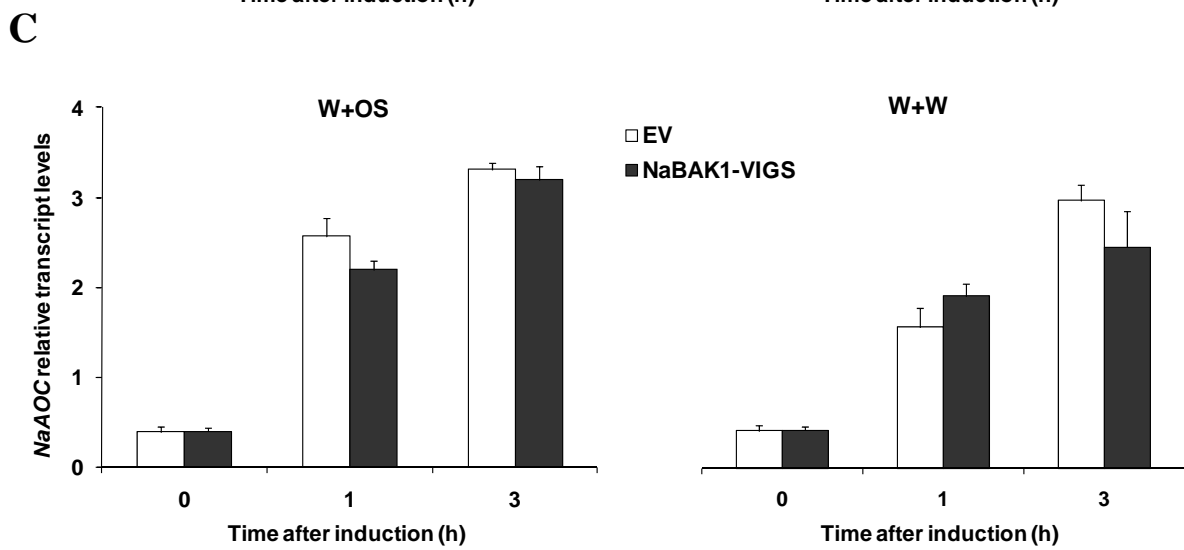
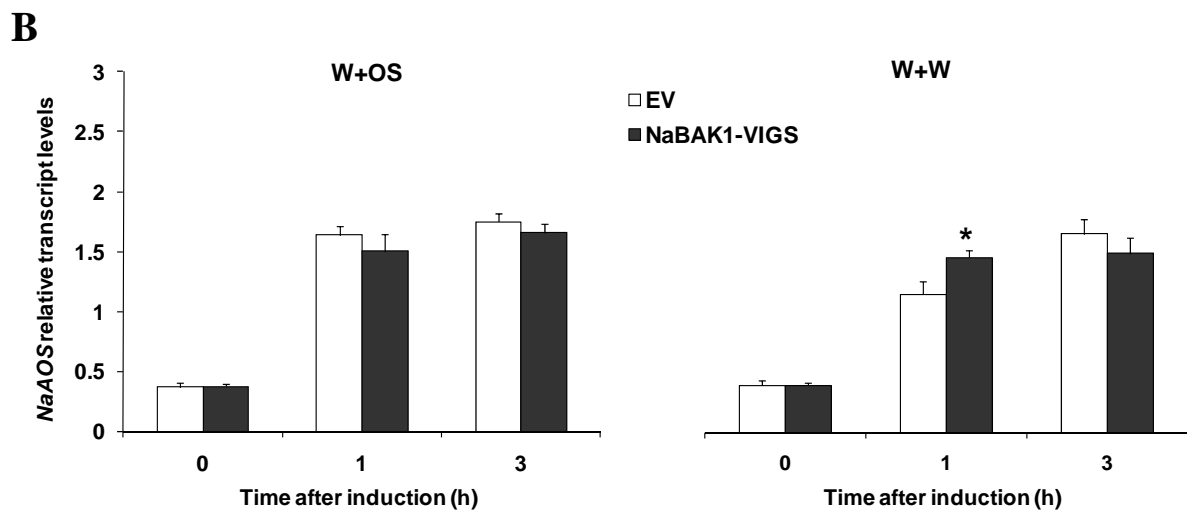
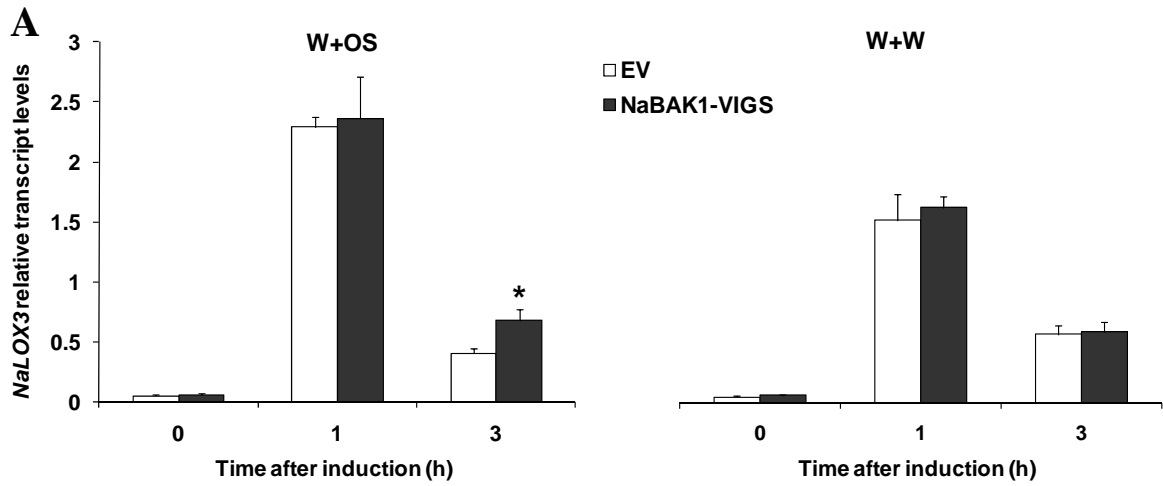


Figure S4



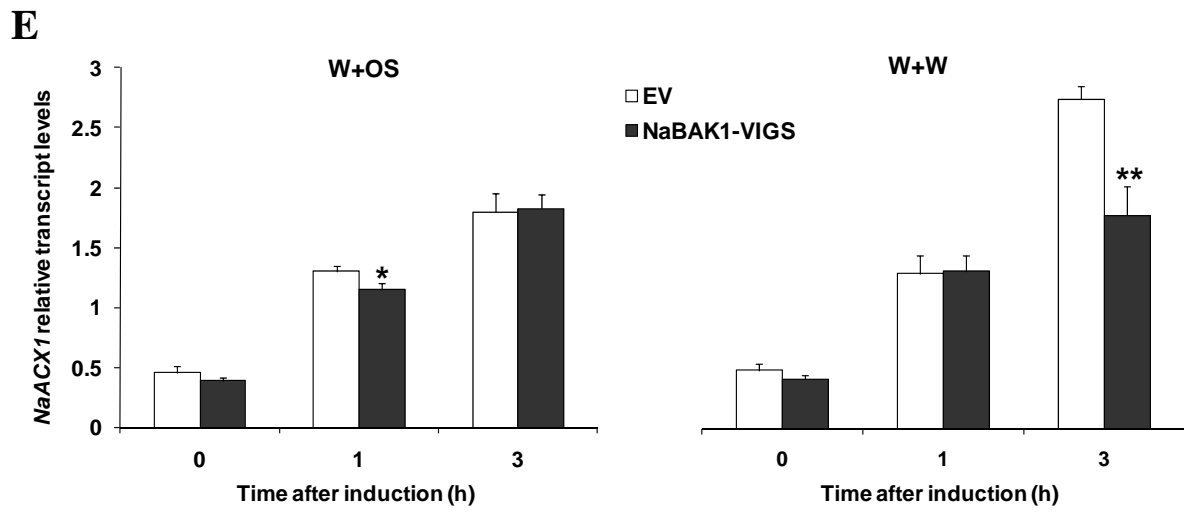
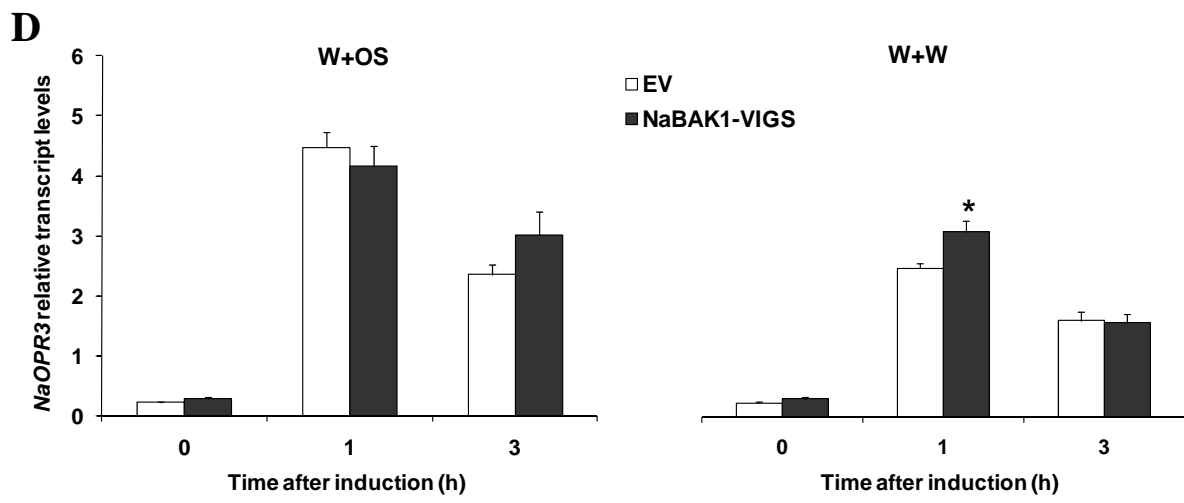


Figure S5

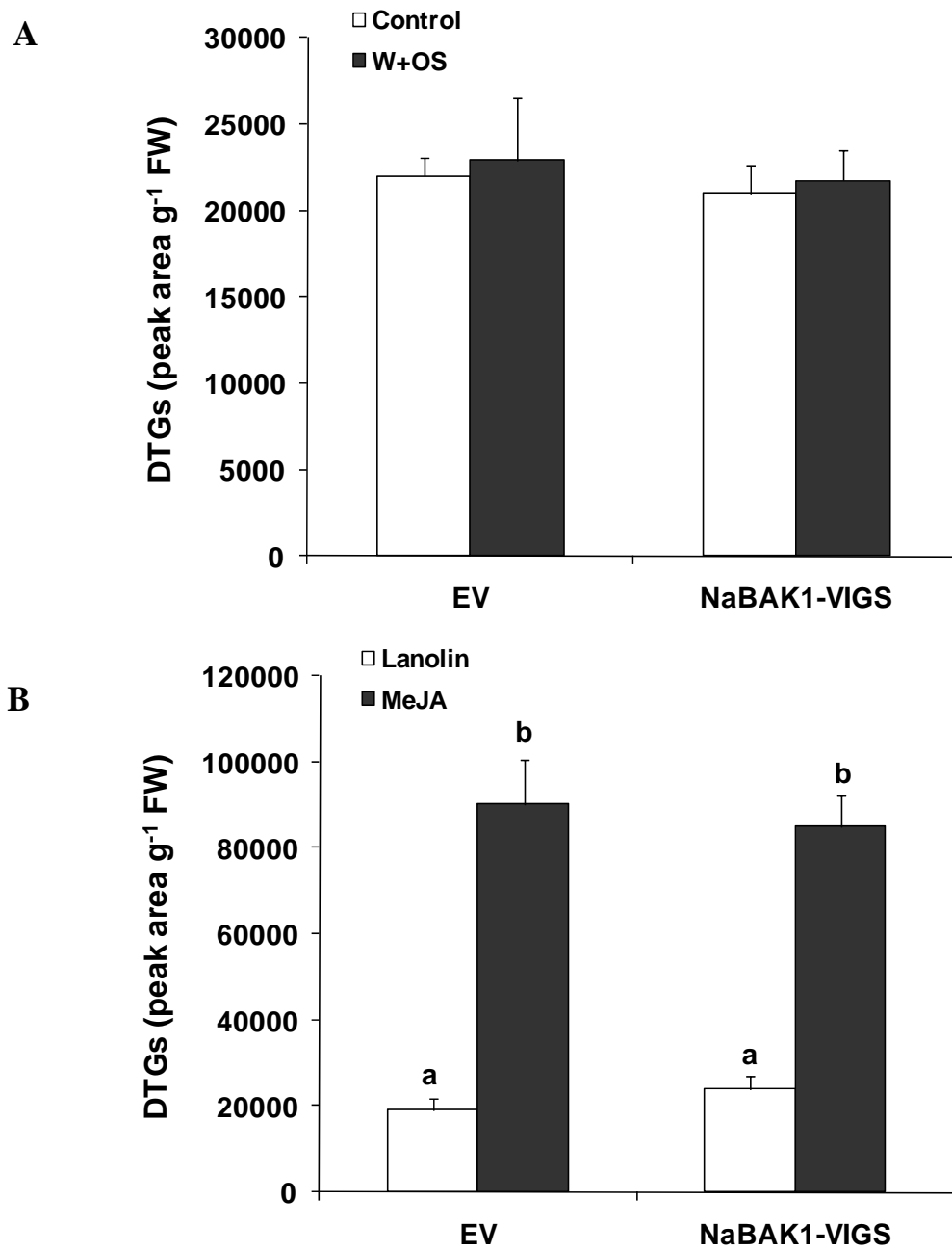


Figure S6

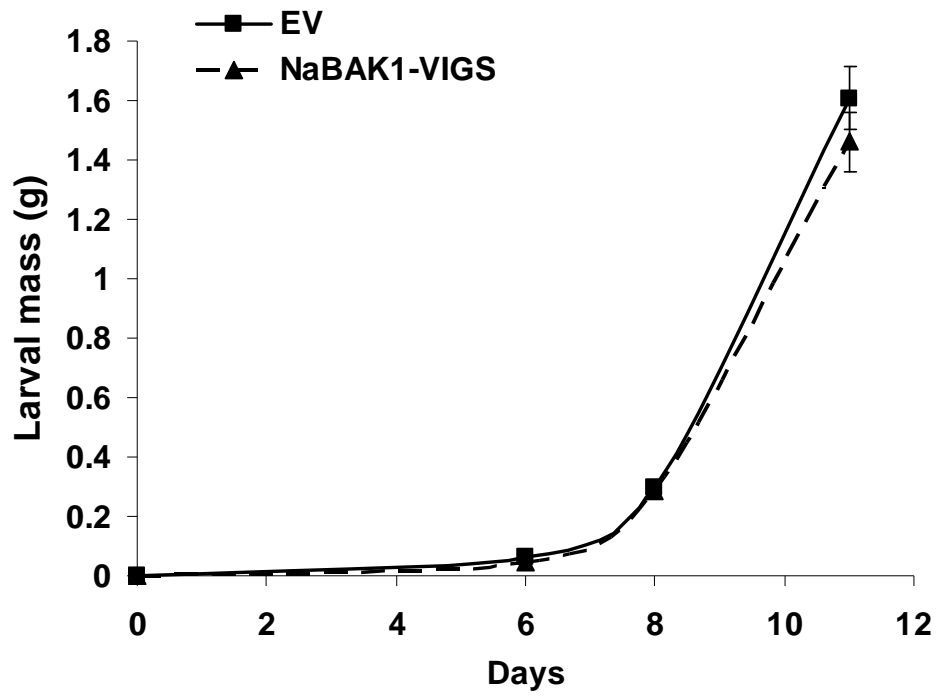


Figure S7

Table S1 Primers used for cloning of NaBAK1 and NaSERK1, and preparation of pTV-NaBAK1 construct

<i>Genes</i>	<i>Primer (5'-3')*</i>
<i>NaBAK1-1</i>	ATGGATCAATGGATATTGGGGATC
<i>NaBAK1-2</i>	GAACACCCACTATCTGATACATCCAG
<i>NaSERK1-1</i>	AATGGAAGTGTTGCATCGTGCCTG
<i>NaSERK1-2</i>	CGCCTATGTTCAACTTGTCAGGGCATAG
<i>NaBAK1-VIGS-BamHI</i>	ACGT <u>GGATCC</u> AAGTGGAGTCTGCGATAATCC
<i>NaBAK1-VIGS-SalI</i>	ACGT <u>GTCGAC</u> GATTGGGTCAAGGGACTTC

* Nucleotides underlined are *Bam*H I and *Sal* I sites

Table S2 Primer sequences used for quantitative real-time PCR (SYBR Green analysis)

<i>Genes</i>	<i>Forward Primer (5'-3')</i>	<i>Reverse Primer (5'-3')</i>
<i>NaActin2</i>	GGTCGTACCACCGGTATTGTG	GTCAAGACGGAGAATGGCATG
<i>NaBAK1</i>	TTAAGTCTTTATATTTGTATGTCAGGAA	AAAAGAAAATACATTTGTGCTTCCAC
<i>NaSERK1</i>	GAACTCTAATTTTGTGATCTTGAAAGTT	ATATAGCCACTACGCCTATGTTC
<i>NaLOX3</i>	GGCAGTGAAATTCAAAGTAAGAGC	CCCAAAATTTGAATCCACAACA
<i>NaAOS</i>	GACGGCAAGAGTTTTCCCAC	TAACCGCCGGTGAGTTCAGT
<i>NaAOC</i>	AACTACCTAACCTCTCATTTCTCA	AAGCGAAGATAGGCAGGGC
<i>NaOPR3</i>	AATGGAGTTGGAGTTTGTTT	AGGTGGTTGAAGCAGTCGTT
<i>NaACX1</i>	GAATGTCTGTTGCTTGTGCTCA	TACCGCAAAGCACCTCCAG
<i>NaTD</i>	TAAGGCATTTGATGGGAGGC	TCTCCCTGTTACGATAATGGAA
<i>NaJAR4</i>	ATGCCAGTCGGTCTAACTGAA	TGCCATTGTGGAATCCTTTTAT
<i>NaJAR6</i>	TGGAGTAAACGTTAACCCGAAA	AGAATTTGCTTGCTCAATGCCA