Glycosyl thiocarbamates as complementary building blocks for chemical glycosylation

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General remarks.

Column chromatography was performed on silica gel 60 (EM Science, 70-230 mesh), reactions were monitored by TLC on Kieselgel 60 F₂₅₄ (EM Science). The compounds were detected by examination under UV light and by charring with 10% sulfuric acid in methanol. Solvents were removed under reduced pressure at < 40 °C. CH₂Cl₂ and ClCH₂CH₂Cl were distilled from CaH₂ directly prior to application. Methanol was dried by refluxing with magnesium methoxide, distilled and stored under argon. Acetonitrile and pyridine were dried by refluxing with CaH₂ and then distilled and stored over molecular sieves (3 Å). Acetone was dried by refluxing with K₂CO₃ and then distilled and stored over molecular sieves (3 Å). KSCN was dried in vacuo at 150 °C for 24 h. Molecular sieves (3 Å or 4 Å), used for reactions, were crushed and activated in vacuo at 390 °C during 8 h in the first instance and then for 2-3 h at 390 °C directly prior to application. AgOTf was co-evaporated with toluene (3 x 10 mL) and dried *in vacuo* for 2-3 h directly prior to application. Cu(OTf)₂ was dried in vacuo for 1-2 h before using for glycosylation. Optical rotations were measured at 'Jasco P-1020' polarimeter. ¹H-n.m.r. spectra were recorded in CDCl₃ at 300 MHz, ¹³C-NMR spectra were recorded in CDCl₃ at 75 MHz (Bruker Avance). HRMS determinations were made with the use of JEOL MStation (JMS-700) Mass Spectrometer.

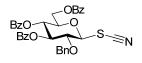
3,4,6-Tri-O-benzoyl-2-O-benzyl-α-D-glucopyranosyl bromide (12).



A solution of acetyl bromide (4 mL, 54.29 mmol) and methanol (2.1 mL, 54.29 mmol) in dichloromethane (25 mL) was added to a solution of 1,3,4,6-tetra-O-benzoyl-2-O-benzyl-D-

glucopyranose¹ (2.3 g, 3.393 mmol) in dichloromethane (15 mL) at 0 °C. The resulting reaction mixture was stirred for 2 h at 0 °C and then washed with water (1 x 15 mL), sat. aq. NaHCO₃ (15 mL), and water (3 x 15 mL). The organic layer was separated, dried with MgSO₄, concentrated in *vacuo* and dried to afford the title compound in 78% (1.7 g) yield, which was used directly in subsequent synthesis of **1a**. Spectral data of **12** was the same as reported previously.²

3,4,6-Tri-O-benzoyl-2-O-benzyl-β-D-glucopyranosyl thiocyanate (1a).

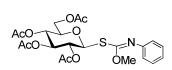


The title compound was obtained in accordance with the method reported previously.³ 18-Crown-6 (85 mg, 0.3 mmol) and KSCN (312 mg, 3.0 mmol) were added to a stirred solution of bromide 12 (690 mg, 1.0 mmol) in dry acetone (2.6 mL) and the reaction mixture was stirred under argon for 2.5 h at rt. After that, the mixture was diluted with acetone (20 mL), the solid was filtered off, rinsed with acetone (3 x 5 mL) and the combined filtrate was concentrated in vacuo. The residue was purified by column chromatography on silica gel (ethyl acetate-hexane gradient elution) to afford the title compound as a white foam in 65% (435 mg) yield. Analytical data for **1a**: $R_f = 0.48$ (ethyl acetate/hexanes, 3/7, v/v); $[\alpha]_D^{22} 21.1^\circ$ (c = 0.9, CHCl₃); ¹H n.m.r.: δ, 4.14 (dd, 1H, J_{2,3} = 9.2 Hz, H-2), 4.24 (m, 1H, J_{5,6a} = 5.0, J_{5,6b} = 2.9 Hz, H-5), 4.56 (dd, 1H, $J_{6a,6b} = 12.3$ Hz, H-6a), 4.68 (dd, H-6b), 4.83 (dd, 2H, $J^2 = 10.7$ Hz, CH₂Ph), 4.94 (d, 1H, $J_{1,2} = 10.7$ Hz, CH₂Ph), 4.94 (d, 1H, J_{1,2} = 10.7 Hz, CH₂Ph), 4.94 (d, 1H, J_{1,2} = 10.7 Hz, CH₂Ph), 4.94 (d, 1H, J_{1,2} = 10.7 Hz, CH₂Ph), 4.94 (d, 1H, J_{1,2} = 1 9.3 Hz, H-1), 5.70 (dd, 1H, J_{4.5} = 9.8 Hz, H-4), 5.93 (dd, 1H, J_{3.4} = 9.4 Hz, H-3), 7.27-8.12 (m, 25H, aromatic) ppm; ¹³C n.m.r.: δ, 62.9, 68.9, 75.4, 75.9, 77.1, 79.1, 84.3, 108.5, 128.5 (x 3), 128.6 (x 2), 128.61 (x 2), 128.7 (x 6), 128.9, 129.6, 129.9 (x 2), 130.0 (x 4), 133.3, 133.6, 136.3, 165.3, 165.6, 166.2 ppm, HR-FAB MS[M+Na]⁺ calcd for C₃₅H₂₉NO₈SNa⁺ 646.1512, found 646.1501.

O-Methyl phenylcarbamothioate.

A solution of NaOMe in methanol (1M, 22.2 mL) was added to phenyl isothiocyanate (1.7 mL, 14.7 mmol) and the resulting mixture was stirred for 15 min at rt. After that, conc. HCl was added till the pH \sim 4-5. The resulting white precipitate was filtered off and the filtrate was concentrated *in vacuo*. The crude residue containing the title compound was used directly in subsequent reactions with glycosyl bromides. The analytical data of the title compound were the same as described previously.⁴

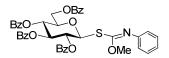
2,3,4,6-Tetra-O-acetyl-1-thio-β-D-glucopyranosyl O-methyl phenylcarbamothioate (13).



O-Methyl phenylcarbamothioate (HSNea, 61 mg, 0.364 mmol) and NaOH (9.7 mg, 0.243 mmol) were added to a stirring solution of 2,3,4,6-tetra-O-acetyl- α -D-glucopyranosyl bromide⁵ (100 mg, 0.243 mmol) in dry acetonitrile (1.2 mL) and the resulting reaction mixture was stirred for 1 h at rt. After that, the solid was filtered off and filtrate was washed with sat. aq. NaHCO₃ (5 mL) and water (3 x 5 mL). The organic layer was separated, dried with MgSO₄ and concentrated *in vacuo*. The residue was purified by column chromatography on silica gel (ethyl acetate-toluene gradient elution) to afford the title compound as a white foam in 60% (73 mg) yield. Analytical data for **13**: R_{*f*} = 0.42 (ethyl acetate/hexanes, 3/7, v/v); [α]_D²⁵ -1.9° (c = 1, CHCl₃); ¹H-n.m.r.: δ , 1.99, 2.01, 2.03, 2.09 (4s, 12H, 4 x COCH₃), 3.77 (m, 1H, J_{5,6a} = 2.3 Hz, J_{5,6b} = 4.7 Hz, H-5), 4.01 (s, 3H, OCH₃), 4.14 (dd, 1H, J_{6a,6b} = 12.4 Hz, H-6a), 4.25 (dd, 1H, H-6b), 4.99 (dd, 1H, J_{2,3} = 9.2 Hz, H-2), 5.07 (dd, 1H, J_{4,5} = 9.7 Hz, H-4), 5.22 (dd, 1H, J_{3,4} = 9.3 Hz, H-3), 5.24 (dd, 1H,

 $J_{1,2} = 10.4 \text{ Hz}, \text{H-1}, 6.83-7.27 \text{ (m, 5H, aromatic) ppm;} {}^{13}\text{C n.m.r.: } \delta, 20.7 \text{ (x 3)}, 20.9, 56.8, 62.2, 68.2, 69.9, 74.1, 76.3, 81.4, 121.5 \text{ (x 2)}, 124.4, 129.3 \text{ (x 2)}, 146.3, 154.5, 169.3, 169.5, 170.4, 170.8 ppm; \text{HR-FAB MS}[\text{M+Na}]^+ \text{ calcd for } \text{C}_{22}\text{H}_{27}\text{NO}_{10}\text{SNa}^+ 520.1253, \text{ found } 520.1263.$

2,3,4,6-Tetra-O-benzoyl-1-thio-β-D-glucopyranosyl O-methyl phenylcarbamothioate (4c).



A solution of NaOMe in methanol (1M, ~0.1 mL) was added dropwise to a solution of thiocarbamate 12 (72 mg, 0.145 mmol) in methanol (1.0 mL) till pH ~ 9 and the resulting reaction mixture was kept for 1 h at rt. After that, Dowex (H^+) was added till pH ~ 7, the resin was filtered off and washed successively with methanol (5 x 5 mL). The combined filtrate was concentrated in vacuo and dried. The residue was dissolved in pyridine (1 mL) and benzoyl chloride (0.1 mL, 0.87 mmol) was added dropwise at 0 °C under argon. The external cooling was removed, the reaction mixture was allowed to warm to rt and stirred for 4 h total. After that, methanol (1 mL) was added, the volatiles were evaporated in vacuo, and the residue was coevaporated with toluene (3 x 5 mL). The resulting residue was diluted with dichloromethane (100 mL) and washed with 1N ag. HCl (25 mL), water (25 mL), sat. ag. NaHCO₃ (25 mL), and water (3 x 25 mL). The organic layer was separated, dried with MgSO₄ and concentrated in vacuo. The residue was purified by column chromatography on silica gel (ethyl acetate - toluene gradient elution) to afford the title compound as a white foam in 83% (90 mg) yield. Analytical data for 4c: $R_f = 0.37$ (ethyl acetate/toluene, 1.5/8.5, v/v); $[\alpha]_D^{21} 0.35^\circ$ (c = 1, CHCl₃); ¹H-n.m.r.: δ, 3.90 (s, 3H, OCH₃), 4.25 (m, 1H, J_{5.6a} = 5.7 Hz, J_{5.6b} = 2.9 Hz, H-5), 4.49 (dd, 1H, J_{6a.6b} = 12.2 Hz, H-6a), 4.64 (dd, 1H, H-6b), 5.54 (dd, 1H, J_{2,3} = 9.2 Hz, H-2), 5.62 (d, 1H, J_{1,2} = 10.2 Hz, H-1), 5.64 (dd, 1H, J_{4,5} = 9.8 Hz, H-4), 5.95 (dd, J_{3,4} = 9.2 Hz, H-3), 6.72-8.06 (m, 25H, aromatic)

ppm; ¹³C-n.m.r.: δ, 56.8, 63.4, 69.6, 70.6, 74.3, 77.4, 81.9, 121.5 (x 2), 124.2, 128.5 (x 2), 128.6 (x 6), 128.7 (x 2), 128.9 (x 2), 129.1, 129.2 (x 2), 129.8, 129.9 (x 3), 130.0 (x 3), 133.4, 133.5, 133.6, 133.7, 146.3, 154.5, 165.1, 165.4, 165.9, 166.3 ppm; HR-FAB MS[M+Na]⁺ calcd for C₄₂H₃₅NO₁₀SNa⁺ 768.1879, found 768.1896.

Characterization data for glycosyl acceptors 2, 5-7

Methyl 2,3,4-tri-*O*-benzyl-α-D-glucopyranoside (2).⁶



Analytical data for **2**: ¹H n.m.r.: δ , 3.33 (s, 3H, OCH₃), 3.45-3.52 (m, 2H, H-2, 4), 3.59-3.76 (m, 3H, H-5, 6a, 6b), 3.98 (dd, 1H, J_{3,4} = 9.2 Hz, H-3), 4.53 (d, 1H, J_{1,2} = 3.5 Hz, H-1), 4.70 (dd, 2H, J² = 12.1 Hz, CH₂Ph), 4.73 (dd, 2H, J² = 11.0 Hz, CH₂Ph), 4.89 (dd, 2H, J² = 10.9 Hz, CH₂Ph), 7.21-7.34 (m, 15H, aromatic) ppm; ¹³C n.m.r.: δ , 55.12, 61.76, 70.55, 73.33, 74.93, 77.24, 79.82, 81.83, 98.01, 127.42, 127.68, 127.74, 127.76 (x 3), 127.83 (x 2), 127.92 (x 2), 128.20 (x 2), 128.27 (x 4), 137.87, 138.48 ppm.

Methyl 2,3,6-tri-*O*-benzyl-α-D-glucopyranoside (5).⁷

Analytical data for **5**: ¹H n.m.r.: δ , 3.34 (s, 3H, OCH₃), 3.52 (dd, 1H, J_{2,3} = 8.5 Hz, H-2), 3.56-3.69 (m, 4H, H-4, 5, 6a, 6b), 3.74 (dd, 1H, J_{3,4} = 9.0 Hz, H-3), 4.52 (dd, 2H, J² = 12.2 Hz, CH₂Ph), 4.59 (d, 1H, J_{1,2} = 3.7 Hz, H-1), 4.63 (dd, 2H, J² = 12.4 Hz, CH₂Ph), 4.84 (dd, 2H, J² = 11.4 Hz, CH₂Ph), 7.22-7.22 (m, 15H, aromatic) ppm; ¹³C n.m.r.: δ , 55.36, 69.57, 70.02, 70.80, 73.28, 73.69, 75.55, 79.71, 81.59, 98.32, 127.80 (x 3), 128.02, 128.14, 128.18 (x 2), 128.31 (x 2), 128.54 (x 2), 128.66 (x 2), 128.77 (x 2), 138.19, 138.24, 138.99 ppm.

Methyl 2,4,6-tri-*O*-benzyl-α-D-glucopyranoside (6).⁸



Analytical data for **6**: $R_f = 0.35$ (ethyl acetate/toluene, 2/8, v/v); $[\alpha]_D^{27} + 36^\circ$ (c = 1, CHCl₃); ¹H n.m.r.: δ , 3.29 (s, 3H, OCH₃), 3.36 (dd, 1H, J_{1,2} = 3.5 Hz, J_{2,3} = 9.6 Hz, H-2), 3.51 (dd, 1H, J_{4,5} = 9.3 Hz, H-4), 3.58-3.7 (m, 3H, H-5, 6a, 6b), 4.02 (dd, 1H, J_{3,4} = 9.1 Hz, H-3), 4.50 (dd, 2H, J² = 12.1 Hz, CH₂Ph), 4.61 (d, 1H, J_{1,2} = 3.5 Hz, H-1), 4.62 (dd, 2H, J² = 11.1 Hz, CH₂Ph), 4.64 (dd, 2H, J² = 12.1 Hz, CH₂Ph), 7.15-7.33 (m, 15H, aromatic) ppm; ¹³C n.m.r.: δ , 55.21, 68.51, 69.65, 73.11, 73.54, 73.63, 74.60, 77.29, 79.51, 97.60, 127.73, 127.76, 127.91 (x 2), 127.99 (x 2), 128.13, 128.17 (x 2), 128.42 (x 4), 128.61 (x 2), 137.94, 138.01, 138.45 ppm.

Methyl 3,4,6-tri-*O*-benzyl-α-D-glucopyranoside (7).⁹



Analytical data for 7: ¹H n.m.r.: δ , 3.43 (s, 3H, OCH₃), 3.62-3.80 (m, 6H, H-2, 3, 4, 5, 6a, 6b), 4.58 (dd, 2H, J² = 12.1 Hz, CH₂Ph), 4.67 (dd, J² = 10.8 Hz, ½ CH₂Ph), 4.81 (d, 1H, J_{1,2} = 3.2 Hz, H-1), 4.88 (m, 2H, J² = 11.2 Hz, CH₂Ph), 7.14-7.30 (m, 15H, aromatic) ppm. ¹³C n.m.r.: δ , 55.32, 68.60, 70.54, 73.03, 73.64, 75.48, 77.60, 83.38, 99.53, 127.84, 127.97 (x 2), 128.03 (x 5), 128.51 (x 4), 128.57 (x 3), 138.05, 138.30, 138.79 ppm.

General procedures for glycosylation

In the presence of MeOTf (Method A). A mixture of a glycosyl donor (0.04 mmol), glycosyl acceptor (0.032 mmol), and freshly activated molecular sieves (3 Å, 90 mg) in 1,2-dichloroethane (0.5 mL) was stirred under argon at rt for 1.5 h. MeOTf (10 μ L, 0.08 mmol) was added and the reaction mixture was stirred at rt for 22-48 h. After that, the solid was filtered off and the filtrate was washed successively with sat. aq. NaHCO₃ (5 mL) and water (3 x 5 mL). The organic layer was separated, dried with MgSO₄ and concentrated *in vacuo*. The residue was purified by column chromatography on silica gel (ethyl acetate-toluene gradient elution) to afford the corresponding disaccharide.

In the presence of AgOTf (Method B): A mixture of glycosyl donor (0.04 mmol), glycosyl acceptor (0.032 mmol), and freshly activated molecular sieves (3 Å, 90 mg) in 1,2-dichloroethane (0.5 mL) was stirred under argon for 1.5 h. AgOTf (12.3 mg, 0.048 mmol) was added and the reaction mixture was stirred for 24 h. Upon completion, the solid was filtered off and the filtrate was washed with NaHCO₃ (5 mL) and water (3 x 5 mL). The organic layer was separated, dried with MgSO₄ and concentrated *in vacuo*. The residue was purified by silica gel column chromatography (ethyl acetate-toluene gradient elution) to afford the corresponding oligosaccharide.

In the presence of $AgBF_4$ (Method C): A mixture of glycosyl donor (0.04 mmol), glycosyl acceptor (0.032 mmol), and freshly activated molecular sieves (3 Å, 90 mg) in 1,2-dichloroethane (0.5 mL) was stirred under argon for 1.5 h. AgBF₄ (9.3 mg, 0.048 mmol) was added and the reaction mixture was stirred for 5 min - 24 h. Upon completion, the solid was filtered off and the filtrate was washed with NaHCO₃ (5 mL) and water (3 x 5 mL). The organic layer was separated, dried with MgSO₄ and concentrated *in vacuo*. The residue was purified by

silica gel column chromatography (ethyl acetate-toluene gradient elution) to afford the corresponding oligosaccharide.

In the presence of $Cu(OTf)_2$ (Method D): A mixture of glycosyl donor (0.04 mmol), glycosyl acceptor (0.032 mmol), and freshly activated molecular sieves (4 Å, 90 mg) in 1,2-dichloroethane (0.5 mL) was stirred under argon for 1.5 h. Cu(OTf)₂ (17 mg, 0.048 mmol) was added and the reaction mixture was stirred for 10 min - 24 h. Upon completion, the solid was filtered off and the filtrate was washed with NaHCO₃ (5 mL) and water (3 x 5 mL). The organic layer was separated, dried with MgSO₄ and concentrated *in vacuo*. The residue was purified by silica gel column chromatography (ethyl acetate-toluene gradient elution) to afford the corresponding disaccharide.

General Procedure for Competitive Glycosylations

A mixture of glycosyl donor **4b** (47 mg, 0.0645 mmol), glycosyl donor **4c** (48 mg, 0.0645 mmol), glycosyl acceptor **2** (20 mg, 0.043 mmol), and freshly activated molecular sieves (4 Å for Cu(OTf)₂ or 3 Å for MeOTf-promoted reactions, 140 mg) in 1,2-dichloroethane (1 mL) was stirred under argon at rt for 1h. Cu(OTf)₂ (28 mg, 0.0774 mmol) or MeOTf (16 μ L, 0.129 mmol) was added and the resulting reaction mixture was stirred at rt. The reaction was monitored by TLC (ethyl acetate / toluene, 1/4, v/v); upon disappearance of the glycosyl acceptor **2** (1-1.5 h), the solid was filtered off, rinsed with CH₂Cl₂ (3 x 5 mL), and the combined filtrate was washed with sat. aq. NaHCO₃ (5 mL) and water (3 x 5 mL). The organic layer was separated, dried with MgSO₄, filtered and concentrated *in vacuo*. The residue was purified by column chromatography on silica gel (ethyl acetate – hexanes gradient elution) to afford disaccharide **8** in 88-97% yield and unreacted glycosyl donors.

Characterization data for disaccharides 8-11

Methyl 6-*O*-(2,3,4,6-tetra-*O*-benzoyl-β-D-glucopyranosyl)-2,3,4-tri-O-benzyl-α-Dglucopyranoside (8).¹⁰

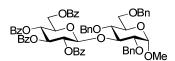
The title compound was synthesized by method D from glycosyl donor 4c (30 mg, 0.04 mmol) and glycosyl acceptor 2 (15 mg, 0.032 mmol) in 90% (30 mg) yield. Analytical data for 8 was in agreement with that reported previously.¹⁰

Methyl 4-O-(2,3,4,6-tetra-O-benzoyl-β-D-glucopyranosyl)-2,3,6-tri-O-benzyl-α-D-

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glucopyranoside (9).<sup>10</sup>
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The title compound was synthesized by method D from glycosyl donor 4c (30 mg, 0.04 mmol) and glycosyl acceptor 5 (15 mg, 0.032 mmol) in 78% (26 mg) yield. Analytical data for 9 was in agreement with that reported previously.¹⁰

Methyl 3-O-(2,3,4,6-tetra-O-benzoyl-β-D-glucopyranosyl)-2,4,6-tri-O-benzyl-α-D-

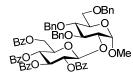


glucopyranoside (10).

The title compound was synthesized by method D from glycosyl donor **4c** (30 mg, 0.04 mmol) and glycosyl acceptor **6** (15 mg, 0.032 mmol) in 74% (25 mg) yield. Analytical data for **10**: $R_f =$

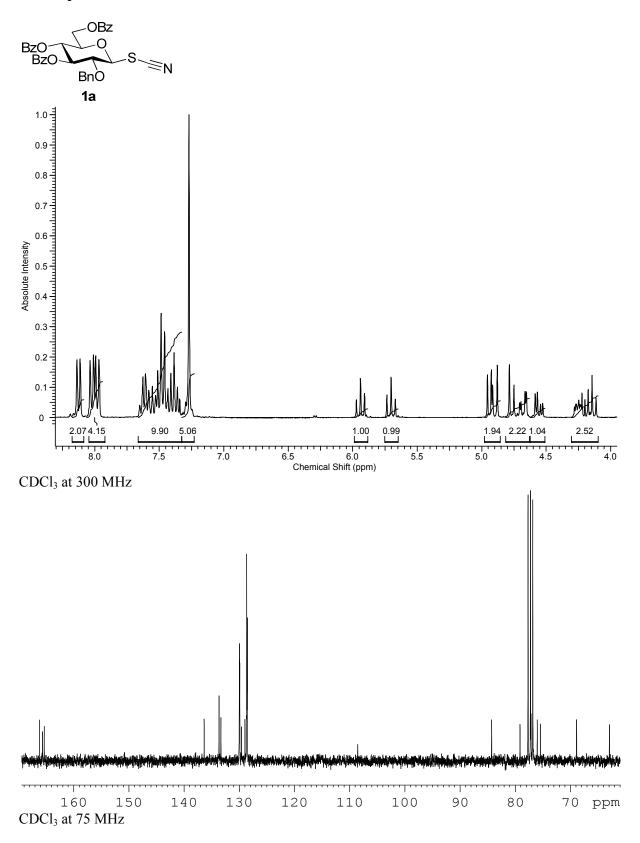
0.4 (ethyl acetate / toluene, 1/5, v/v), $[\alpha]_D^{27}$ -12.8° (c = 1, CHCl₃), ¹H-n.m.r.: δ , 3.15 (s, 3H, OCH₃), 3.25 (dd, 1H, J_{2,3} = 9.6 Hz, H-2), 3.44-3.58 (m, 4H, H-4, 5, 6a, 6b), 4.03-4.07 (m, 1H, H-5'), 4.19 (d, 1H, J_{1,2} = 3.4 Hz, H-1), 4.28 (dd, 1H, J_{3,4} = 9.7 Hz, H-3), 4.30 (dd, 2H, J² = 12.2 Hz, CH₂Ph), 4.30-4.50 (m, 2H, H-6a', 6b'), 4.70 (dd, 2H, J² = 10.8 Hz, CH₂Ph), 5.43 (d, 1H, J_{1',2'} = 8.0 Hz, H-1'), 5.57 (dd, 1H, J_{2',3'} = 9.7 Hz, H-2'), 5.63 (dd, 1H, J_{4',5'} = 9.7 Hz, H-4'), 5.86 (dd, 1H, J_{3',4'} = 9.6 Hz, H-3'), 7.0-7.94 (m, 35H, aromatic) ppm; ¹³C n.m.r.: δ , 55.19, 63,46, 68.59, 69.71, 70.05, 72.08, 72.68, 73.32, 73.66, 74.01, 75.07, 75.58, 79.51, 80.97, 97.84, 101.19, 127.62, 127.88, 128.07 (x 2), 128.14 (x 2), 128.25 (x 4), 128.29 (x 2), 128.38 (x 2), 128.46 (x 2), 128.54 (x 4), 128.64 (x 2), 128.67 (x 2), 128.99, 129.57, 129.79, 129.89 (x 2), 129.99 (x 4), 130.03 (x 2), 133.07, 133.39, 133.46, 133.54, 138.04, 138.11, 138.66, 165.34, 165.43, 166.02, 166.28 ppm; HR-FAB MS[M+Na]⁺ calcd for C₆₂H₅₈O₁₅Na⁺ 1065.3673, found 1065.3673.

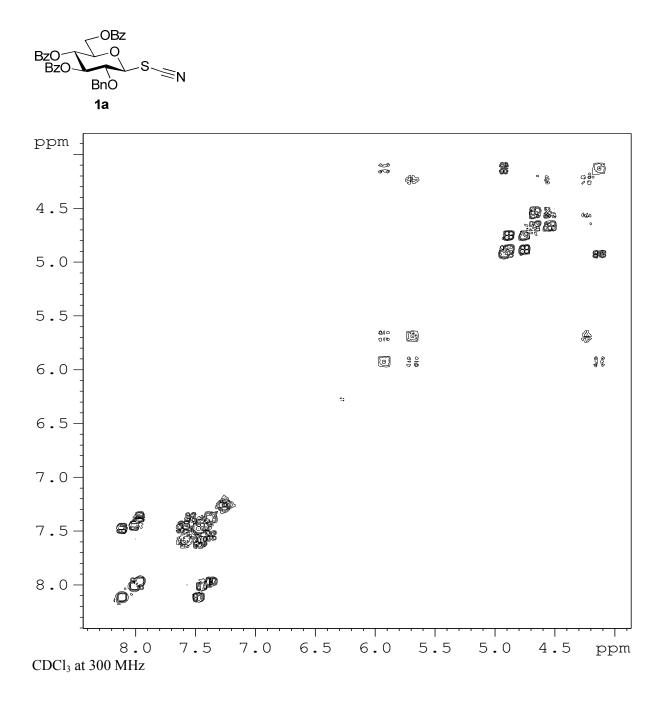
Methyl 2-*O*-(2,3,4,6-tetra-*O*-benzoyl-β-D-glucopyranosyl)-3,4,6-tri-*O*-benzyl-α-Dglucopyranoside (11).¹¹

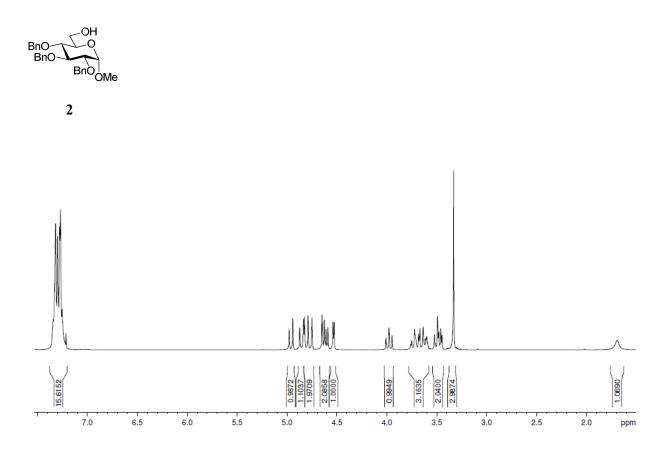


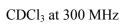
The title compound was synthesized by method D from glycosyl donor 4c (30 mg, 0.04 mmol) and glycosyl acceptor 7 (15 mg, 0.032 mmol) in 75% (25 mg) yield. Analytical data for 11 was in agreement with that reported previously.¹¹

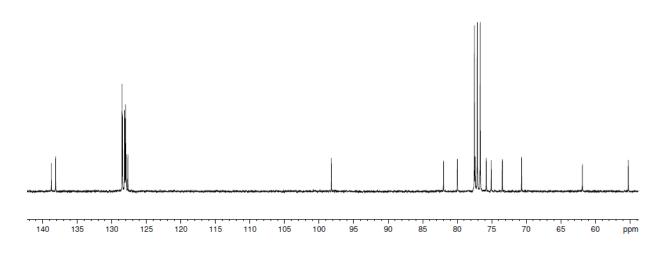
NMR spectra



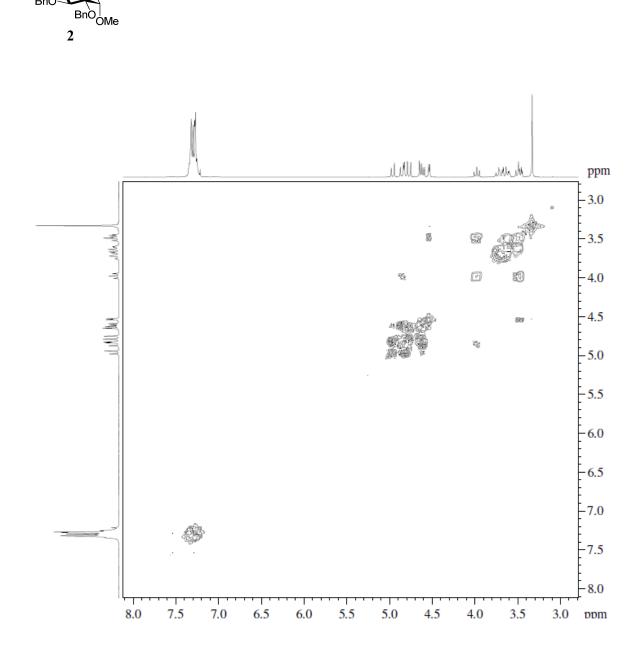






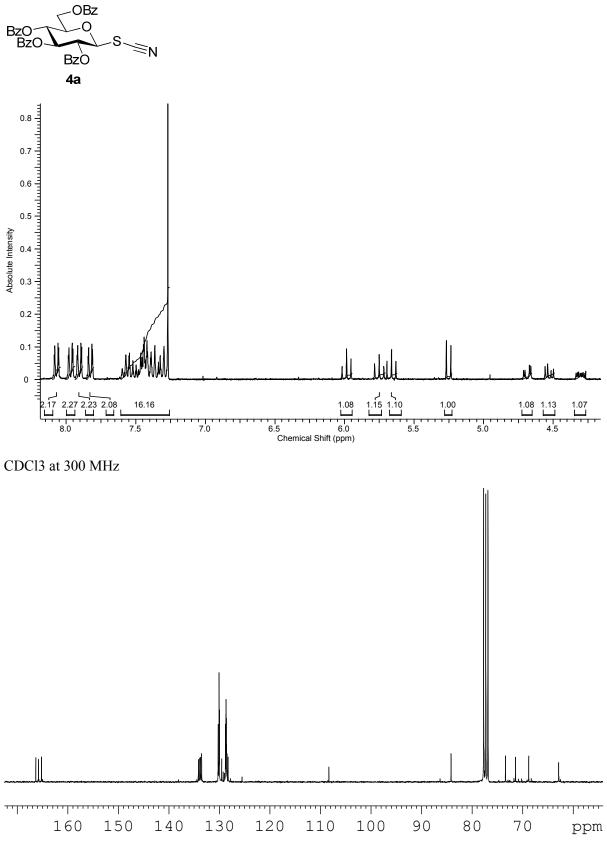


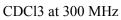
 $CDCl_3$ at 75 MHz

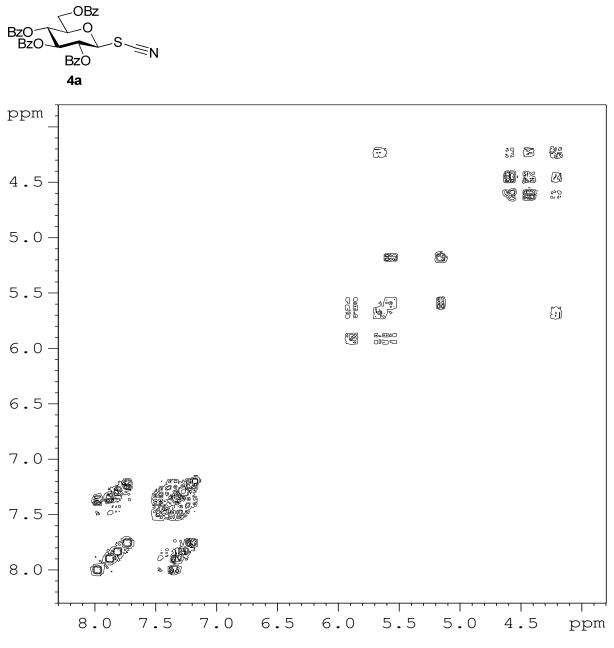


CDCl₃ at 300 MHz

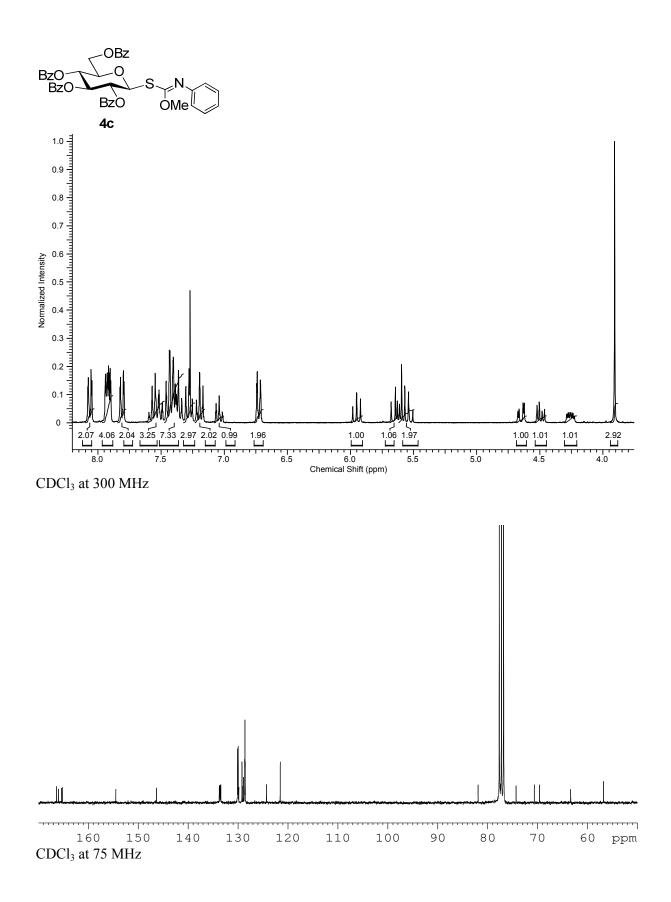
-OH __Q

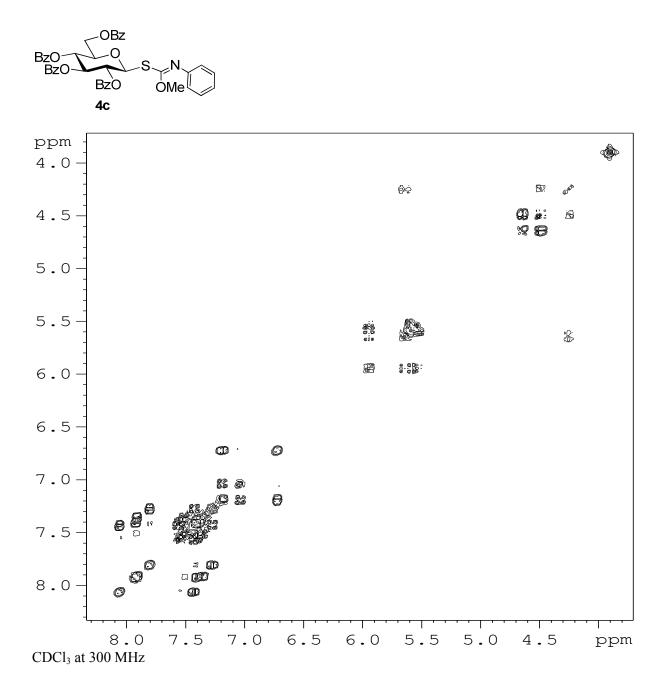


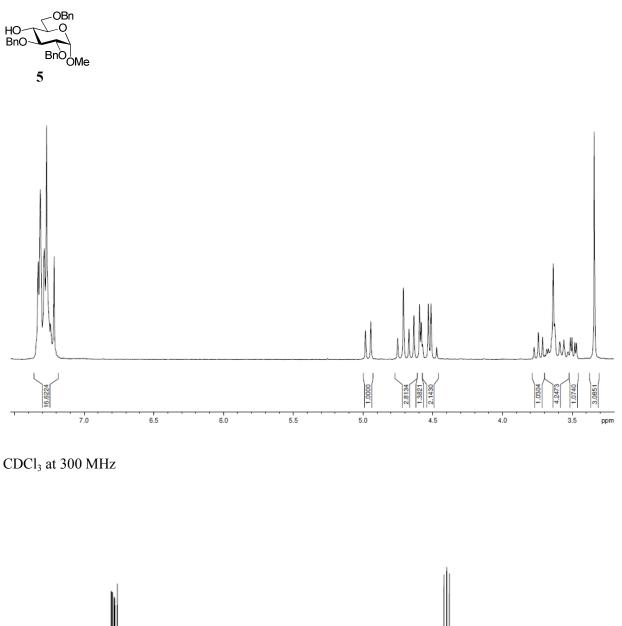


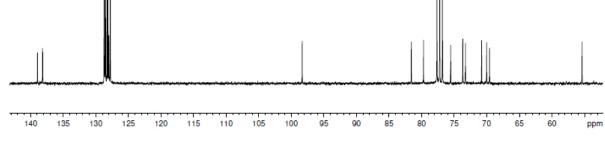


CDCl₃ at 300 MHz

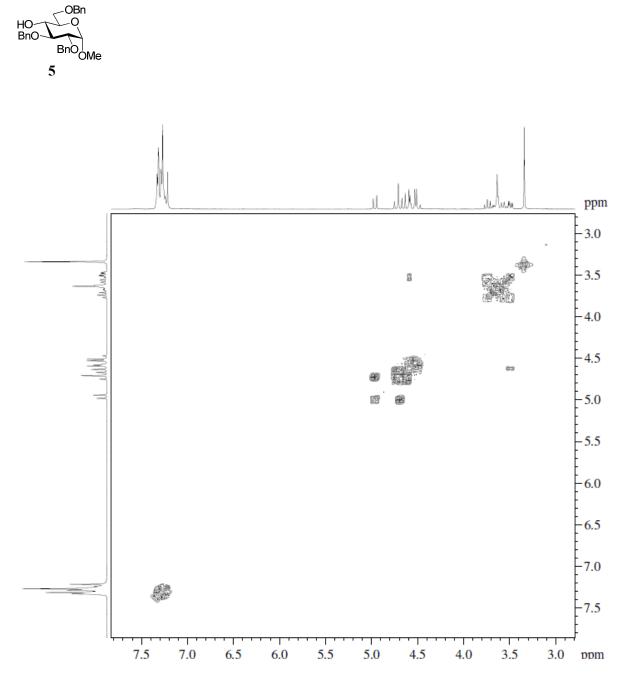




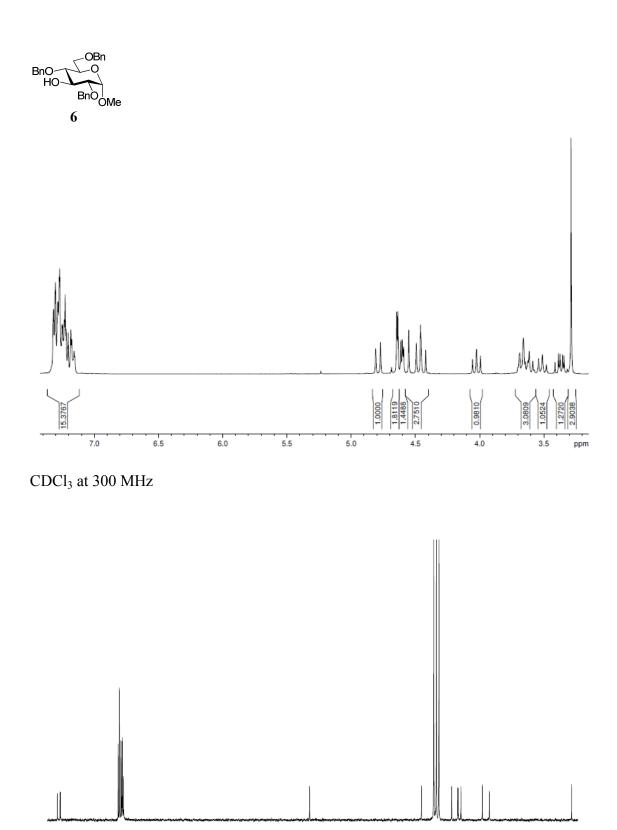




CDCl₃ at 75 MHz

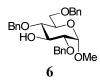


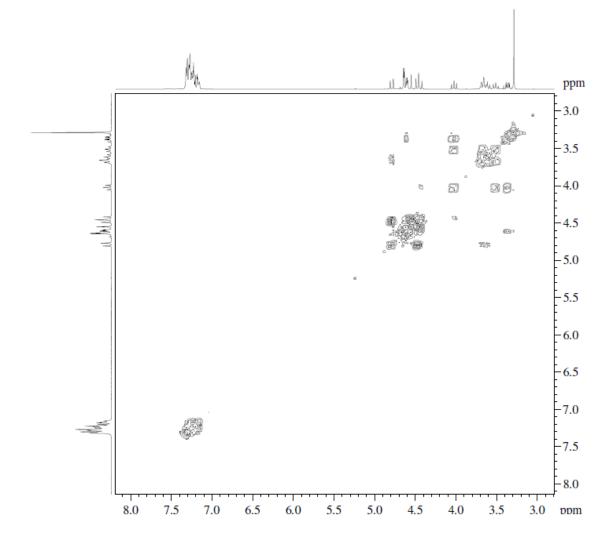
CDCl₃ at 300 MHz



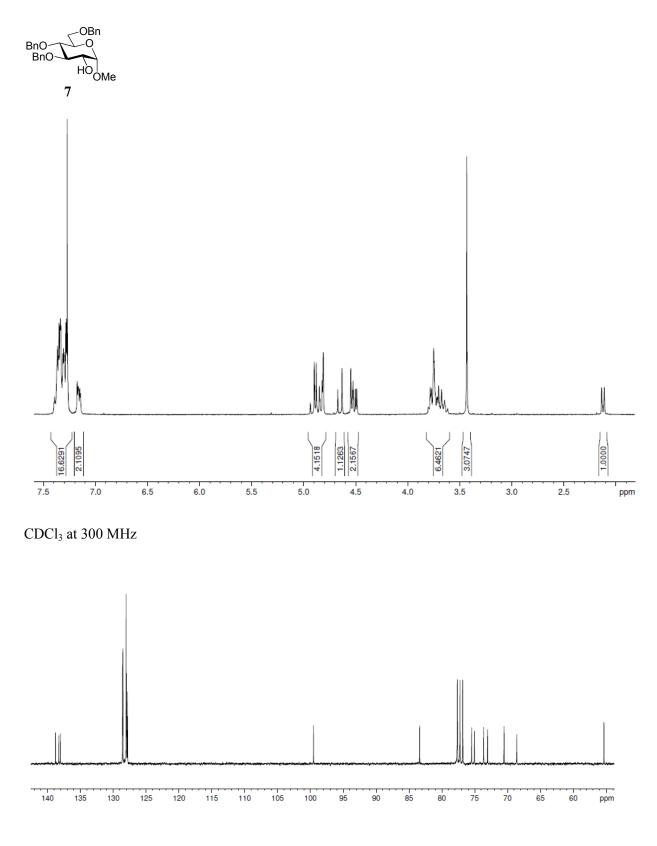
т ppm

 $CDCl_3$ at 75 MHz

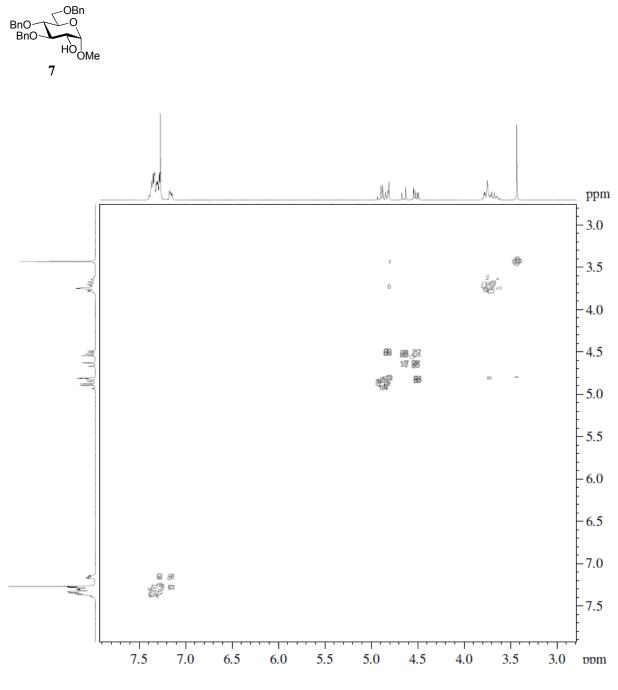




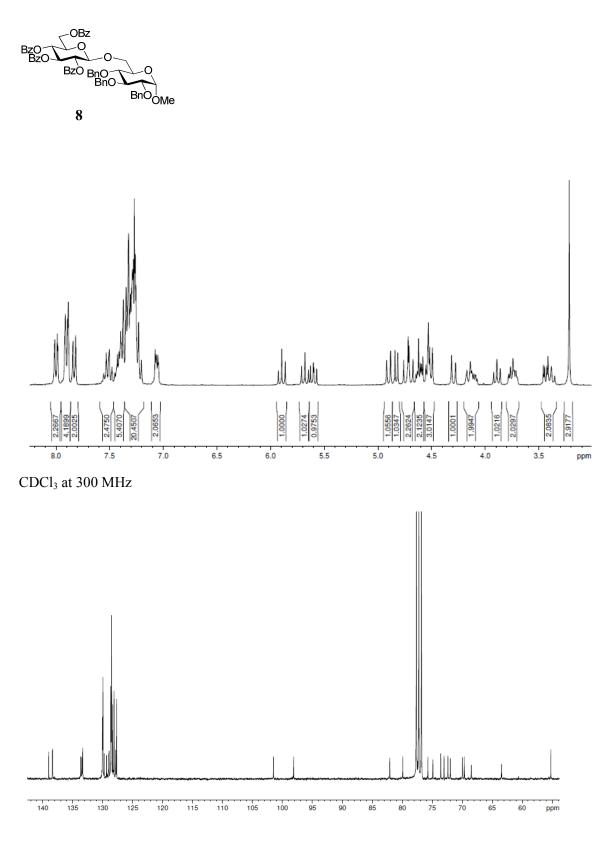
CDCl3 at 300 MHz



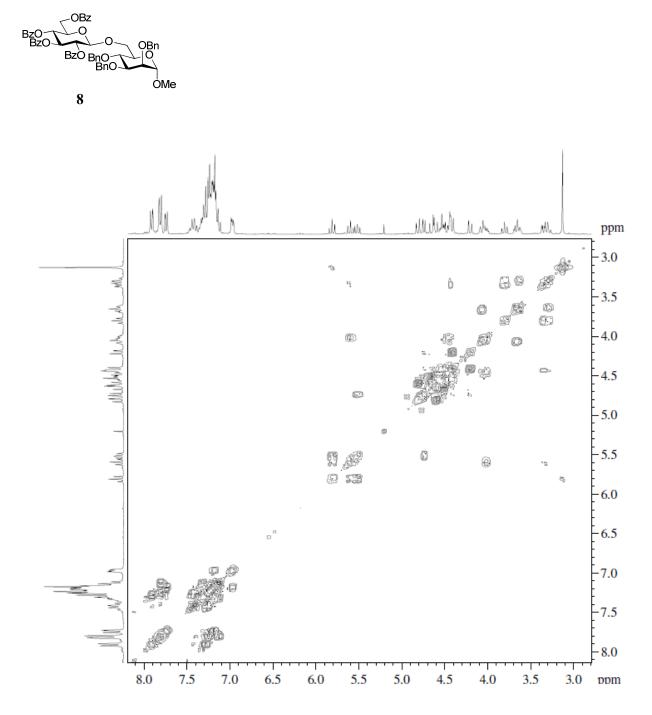
CDCl3 at 75 MHz



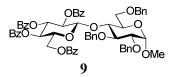
CDCl3 at 300 MHz

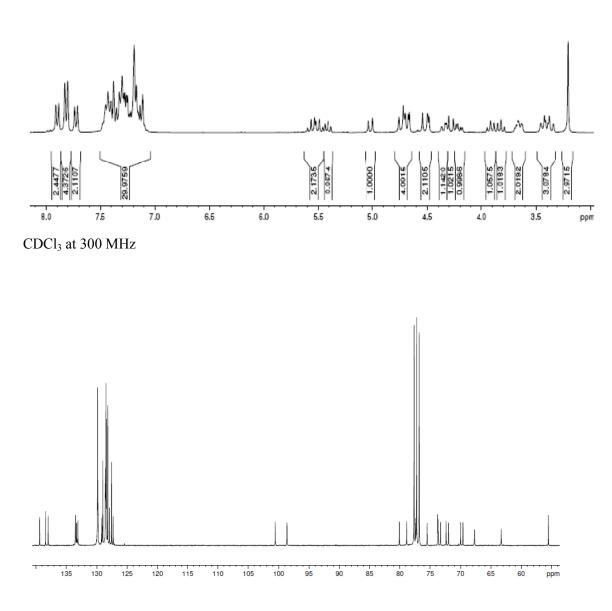


CDCl₃ at 75 MHz

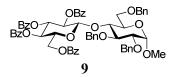


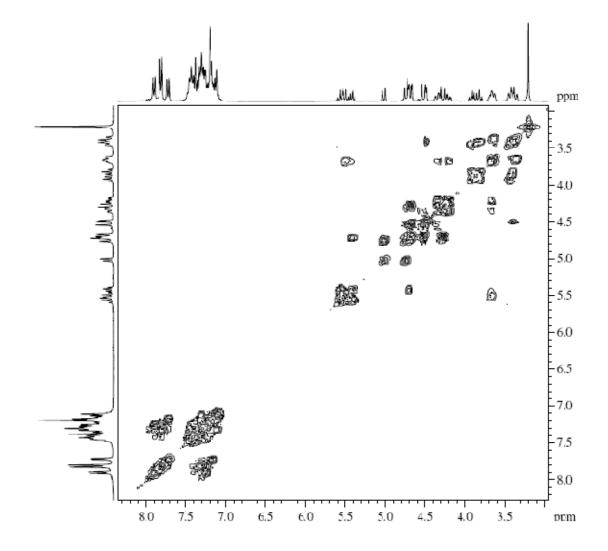
CDCl₃ at 300 MHz



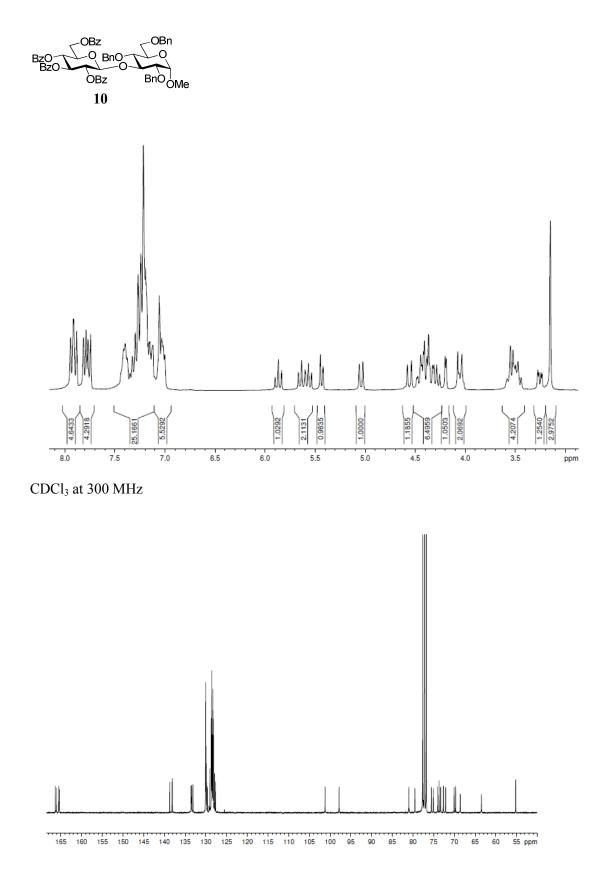


 $CDCl_3$ at 75 MHz

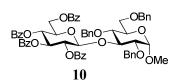


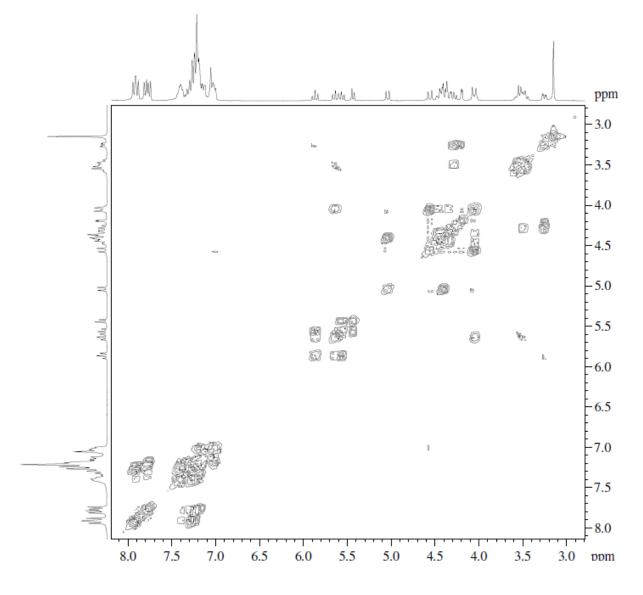


CDCl₃ at 300 MHz

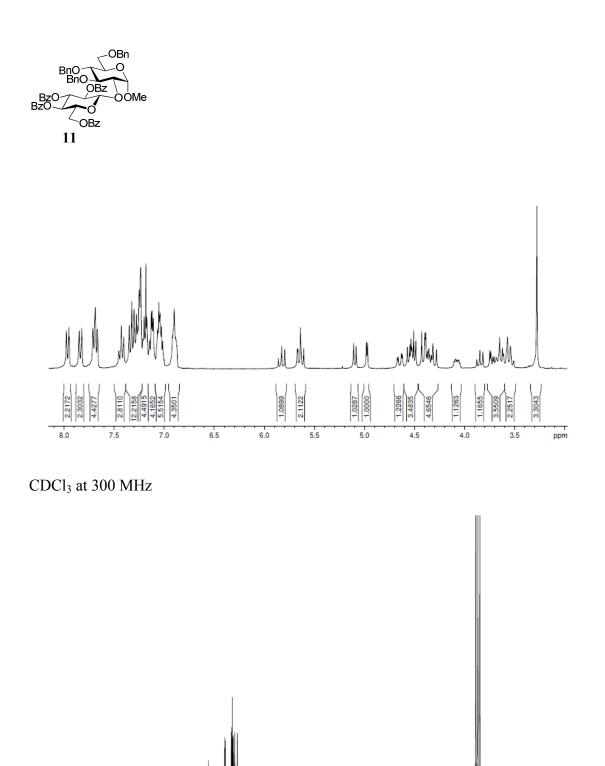


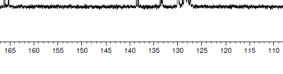
 $CDCl_3$ at 75 MHz





CDCl₃ at 300 MHz



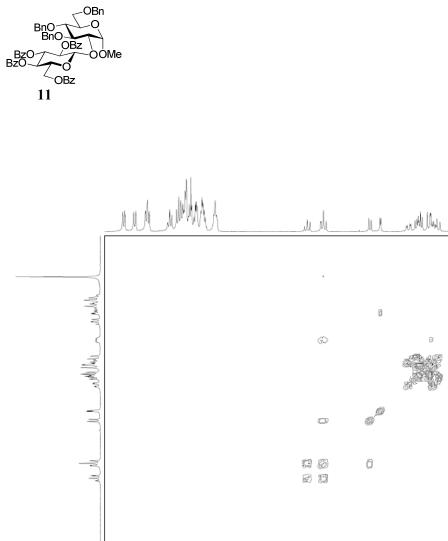


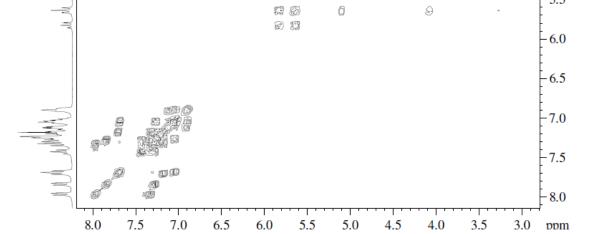
CDCl₃ at 75 MHz

105 100 95 90 85

ppm

80 75 70 65 60





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ppm

- 3.0

-3.5

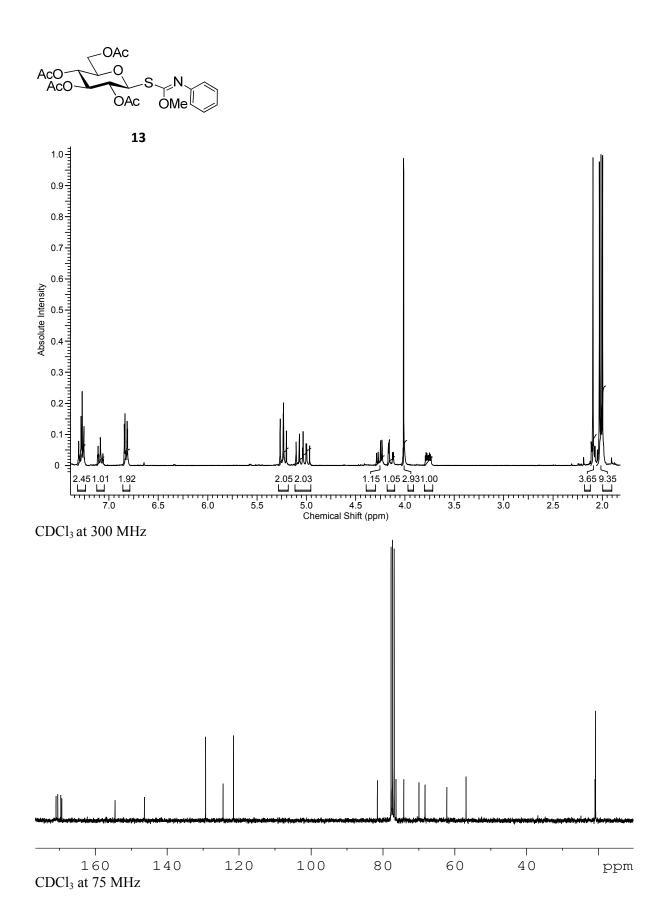
-4.0

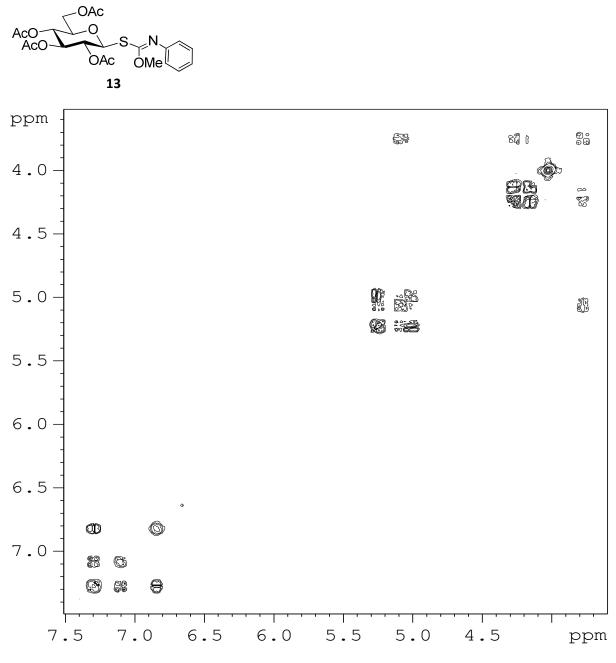
-4.5

5.0

- 5.5

CDCl₃ at 300 MHz





CDCl₃ at 300 MHz

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