

SUPPLEMENTARY ONLINE DATA

Phosphorylation of bacterial-type phosphoenolpyruvate carboxylase at Ser⁴²⁵ provides a further tier of enzyme control in developing castor oil seeds

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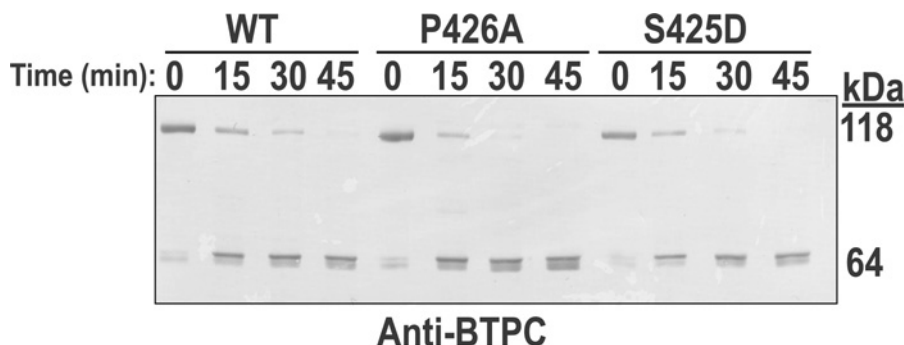


Figure S1 Sensitivity of recombinant COS BTPC mutants towards endogenous COS proteases

Purified recombinant Class-2 PEPCs (15 μg) containing wild-type PTPC (AtPPC3) and wild-type and mutant forms of COS BTPC (RcPPC4) were incubated at 30 °C with 50 μl of clarified stage VII COS endosperm extract (containing approx. 1 mg of protein) to observe the rate and extent of BTPC proteolysis. Aliquots were removed at various time points, boiled in SDS sample buffer, and subjected to SDS/PAGE followed by immunoblotting with anti-BTPC antibody. Each lane contained approx. 70 ng of BTPC.

Table S1 Primers used for site-directed mutagenesis

The base pair alterations leading to the missense mutation are highlighted in **bold**. Base pair alterations that introduced a new restriction site are underlined.

Target	Sequence	New site
<i>AtPpc3</i> R644A	Forward: 5'-GTGGTGGT <u>ACCG</u> TCGGAGCAGGAGGTGGTCTACTCATC-3' Reverse: 5'-GATGAGTAGGACCCCTCCT <u>GCT</u> CCGAC <u>GGTAC</u> CCACCAC-3'	KpnI
<i>RcPpc4</i> S425D	Forward: 5'-GCTAATTCTAGTGGAGATCCGCGGGCATCTTTC-3' Reverse: 5'-GAAAGATGCCCGCGGATCTCCACTAGAAATTAGC-3'	None
<i>RcPpc4</i> S425A	Forward: 5'-GCTAATTCTTCTGGAGCTCCTCGAGCATC-3' Reverse: 5'-GATGCTCGAGGAGCTCCAGAAGAATTAGC-3'	None
<i>RcPpc4</i> P426A	Forward: 5'-CTAATTCTAGTGGATCTGCGGGGCATCTTTTCAG-3' Reverse: 5'-CTGAAAGATGCCCGCGCAGATCCACTAGAAATTAG-3'	None
<i>RcPpc4</i> R760A	Forward: 5'-CGTGGAGGATCCATTGGTGGTGGTGGCCCCACATA-3' Reverse: 5'-TATGTGGGGCCACCACCAGACCAATGGATCCTCCACG-3'	BamHI

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Table S2 Purification of recombinant Class-2 PEPC from combined extracts originating from 5 g of AtPPC3_R644A- and 10 g of RcPPC4-expressing *E. coli*

Step	Activity (units)	Protein (mg)	Specific activity (units/mg of protein)	Purification (fold)	Yield (%)
Combined extracts	50	3000	0.016	1	100
Ni ²⁺ -affinity FPLC	43	162	0.27	16	86
Superdex-200 FPLC	29	6.5	4.5	281	58
Superose-6 FPLC	12	1.8	6.8	425	24

Table S3 Purification of recombinant Class-2 PEPC from combined extracts originating from 5 g of AtPPC3- and 11 g of RcPPC4_R670A-expressing *E. coli*

Step	Activity (units)	Protein (mg)	Specific activity (units/mg of protein)	Purification (fold)	Yield (%)
Combined extracts	71	2196	0.033	1	100
Ni ²⁺ -affinity FPLC	70	26.5	2.6	81	98
Superdex-200 FPLC	34	8.9	3.9	117	49
Superose-6 FPLC	7.6	1.6	4.8	145	11

Table S4 Purification of recombinant Class-2 PEPC from combined extracts originating from 5 g of AtPPC3_R644A- and 15 g of RcPPC4_S425D-expressing *E. coli*

Step	Activity (units)	Protein (mg)	Specific activity (units/mg of protein)	Purification (fold)	Yield (%)
Combined extracts	172	2198	0.079	1	100
Ni ²⁺ -affinity FPLC	130	80	1.6	20	75
Superdex-200 FPLC	52	9.6	5.5	70	31
Superose-6 FPLC	40	4.2	9.5	120	25

Table S5 Purification of recombinant Class-2 PEPC from combined extracts originating from 5 g of AtPPC3_R644A- and 15 g of RcPPC4_S425A-expressing *E. coli*

Step	Activity (units)	Protein (mg)	Specific activity (units/mg of protein)	Purification (fold)	Yield (%)
Combined extracts	37	4800	0.008	1	100
Ni ²⁺ -affinity FPLC	4.3	21.2	0.2	27	12
Superose-6 FPLC*	3.7	0.8	4.6	575	10

*Owing to low total activity, a Superose-6 16/50 prep grade column was used in place of the two gel filtration columns that were normally used.

Table S6 Purification of recombinant Class-2 PEPC from combined extracts originating from 5 g of AtPPC3_R644A- and 15 g of RcPPC4_P426A-expressing *E. coli*

Step	Activity (units)	Protein (mg)	Specific activity (units/mg of protein)	Purification (fold)	Yield (%)
Combined extracts	114	2850	0.04	1	100
Ni ²⁺ -affinity FPLC	84	88	0.95	24	74
Superdex-200 FPLC	38	8.1	4.7	118	34
Superose-6 FPLC	11	1.4	8.4	210	10