

Supporting Information

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SI Text

Multispin Analysis of NMR Distance Measurements.

¹³C{³¹P}REDOR analysis of citrate distances from apatite. ¹³C{³¹P} dipolar coupling measurements by ¹³C{³¹P}REDOR NMR can provide insights into the distances of various citrate carbons from the apatite surface. Fig. 3A and B shows ¹³C{³¹P} REDOR data for all five spectrally resolved carbon sites in citrate bound to apatite in bone. While many systems studied by REDOR can be approximated in terms of two or few interacting spins, a ¹³C spin near an apatite surface experiences the dipolar fields of many ³¹P nuclear magnets. The initial decay depends on the second moment of the ¹³C-³¹P couplings

$$M_2 = \sum_n \omega_{C-P,n}^2 = \sum_n C_n(\alpha, \beta, \gamma) 1/r_{C-P,n}^6,$$

where $r_{C-P,n}$ is the distance between the carbon and the n th ³¹P spin, and $C_n(\alpha, \beta, \gamma)$ is a purely orientation-dependent prefactor. At sufficiently large distances of the ¹³C from the apatite surface ($z_{C-P} > 1$ nm), the second moment depends only on the height z_{C-P} above the top phosphate layer, independent of the lateral (x-y) position of the ¹³C. However, when the height z_{C-P} is not much larger than the average P-P distance of 0.45 nm in apatite, the x-y position of the ¹³C spin needs to be taken into account. Fig. S1A shows a contour plot of the distance-only heteronuclear second moment

$$M_{2r} = \sum_n 1/r_{C-P,n}^6$$

as a function of ¹³C position at a fixed height of $z_{C-P} = 0.35$ nm above a (10 $\bar{1}$ 0)₁ hydroxyapatite surface. The plot shows pronounced maxima above ³¹P (of slightly different height for two types of phosphate at different depths from the apatite surface) and wide flat regions around the Ca ions. Negatively charged citrate is expected to be repelled by the negatively charged phosphate and rather bound to calcium. Therefore, the regions of high M_{2r} near ³¹P can mostly be disregarded. In addition, the regions of high M_{2r} account for only a small fraction of the total area, see the histogram of M_{2r} in Fig. S1B. Calcium ions are located 0.05 nm below the top phosphate layer, so $z_{C-Ca} = z_{C-P} + 0.05$ nm.

Fig. S1C shows simulated REDOR curves and data from Fig. 3B for the 15 positions B1–B5, C1–C5, and D1–D5 marked in Fig. S1A and z_{C-P} values between 0.3 and 0.5 nm. The height of the ¹³C nucleus above calcium is larger by 0.05 nm. The data of Fig. 3 show that the central and one terminal COO group are closest to the surface, while the other terminal carboxylate is at a 0.1-nm greater height but still close to apatite. In the simulations of the REDOR curves, the second moment with REDOR angular dependencies of all the ³¹P-¹³C couplings was calculated for >3,000 crystallite orientations, via orientational averaging of the rotor-axis relative to the crystallite normal, and rotation of the B_0 field around the rotor-axis (with the fixed magic angle between them), as outlined in ref. 1. The total dephasing curve is a superposition, for various rotor-axis and B_0 -field orientations, of Gaussians $\exp(-M_2(Nt_r)^2/2)$, with the second moment M_2 evaluated based on the internuclear distances and the REDOR angular dependencies of the individual C-P couplings at the given rotor-axis and B_0 orientation. The simulation program was validated using a spin-pair model geometry.

Calculation of the citrate density on bone mineral. The area density of citrate on apatite in bone and bound to bone mineral can be calculated from the known structural parameters of the materials. The 3-nm thick nanocrystals in bone, alternating with 3-nm thick collagen layers, have a surface-to-sample-volume ratio of

$$S/V_{\text{bone}} = 2 A/[(3 + 3) \text{ nm A}] = 0.33/\text{nm} = 0.33 \times 10^9/\text{m}$$

and the nanocomposite has an average density of

$$\begin{aligned} \rho_{\text{ave}} &= m_{\text{bone}}/V_{\text{bone}} = (\rho_A V_A + \rho_B V_B)/V_{\text{bone}} \\ &= \rho_A V_A/V_{\text{bone}} + \rho_B V_B/V_{\text{bone}} = (1.15 + 3.2)\text{g/cm}^3 \times 0.5 \\ &= 2.2 \text{ g/cm}^3 = 2.2 \times 10^6 \text{ g/m}^3 \end{aligned}$$

calculated from 50:50 volume fractions and densities of collagen (1.15 g/cm³) and apatite (3.2 g/cm³). From these quantities, the specific surface area (per mass) of bone can be calculated as

$$\begin{aligned} S/m_{\text{tot}} &= (S/V_{\text{tot}})/(m_{\text{tot}}/V_{\text{tot}}) = (S/V_{\text{tot}})/\rho_{\text{ave}} \\ &= 0.33 \times 10^9/\text{m}/(2.2 \times 10^6 \text{ g/m}^3) = 150 \text{ m}^2/\text{g} \\ &= 150 \times 10^{18} \text{ nm}^2/(\text{g bone}). \end{aligned}$$

Also, according to wet chemistry and our spectroscopic analysis, 1 g of bone contains 1.2 wt% = 12 mg citrate, which corresponds to $0.012 \text{ g}/189 \text{ g/mol} = 6.3 \times 10^{-5} \text{ mol}$ of citrate per g of bone, or $n_{\text{citr}}/m_{\text{tot}} = 6.3 \times 10^{-5} \times 6 \times 10^{23} = 3.8 \times 10^{19}$ citrate molecules/(g bone). The area density is thus

$$\begin{aligned} (n_{\text{citr}}/m_{\text{tot}})/(S/m_{\text{tot}}) &= 3.8 \times 10^{19}/(150 \times 10^{18} \text{ nm}^2) \\ &= 0.25/\text{nm}^2 = 1/(2 \text{ nm})^2 \end{aligned}$$

The density of ¹³C-labeled citrate on the NuOss™ bone mineral, with a measured specific surface area of

$$S/m_{\text{tot}} = 60 \text{ m}^2/\text{g}$$

can be calculated from the 0.8 mg of citrate, determined by NMR calibrated on neat citrate, per 0.19 g of bone mineral in the same sample. The total bone-mineral surface area in that sample is

$$\begin{aligned} S &= (S/m_{\text{tot}})m_{\text{tot}} = 60 \text{ m}^2/\text{g} \times 0.19 \text{ g} = 11.4 \text{ m}^2 \\ &= 11.4 \times 10^{18} \text{ nm}^2. \end{aligned}$$

The 0.8 mg of citrate correspond to $0.0008 \text{ g}/189 \text{ g/mol} = 4.2 \times 10^{-6} \text{ mol}$ and thus $n_{\text{citr}} = 4.2 \times 10^{-6} \times 6 \times 10^{23} = 2.54 \times 10^{18}$ citrate molecules. The area density of citrate on bone mineral is thus

$$n_{\text{citr}}/S = 2.54 \times 10^{18}/11.4 \times 10^{18} \text{ nm}^2 = 0.22/\text{nm}^2 = 1/(2 \text{ nm})^2$$

in good agreement with the value for bone obtained above.

CODEX ¹³C NMR simulations. The CODEX NMR data of citrate with ¹³C-labeled terminal COO⁻ groups shown in Fig. 4 were fitted by simulating the spin exchange between 600 ¹³C spins in half as many citrate molecules, distributed on a flat rectangular surface of 10 nm width and ca. 100 nm length. The spin exchange

is characterized by an exchange matrix Π , with off-diagonal elements as

$$\Pi_{nm} = 0.5\pi[2\pi 7.5 \text{ kHz } 10^{-3} \text{ nm}^3 / r_{nm}^3]^2 \langle (P_2(\cos \theta))^2 \rangle F(0)$$

reflecting the coupling between ^{13}C spins n and m . The value of $F(0) = 0.025\text{ms}$ is calibrated by matching the initial decay due to the 0.5-nm intramolecular spin-pair coupling, and is close to literature values for ^{13}COO groups between 0.025 and 0.044 ms (2). The diagonal element Π_{nn} is the negative of the sum of the n th column of off-diagonal elements.

The pointed brackets of $\langle (P_2(\cos \theta))^2 \rangle$ indicate the time averaging, due to magic-angle spinning, of the orientation dependence of the dipolar interaction, which depends on the instantaneous angle θ between the C^n - C^m internuclear vector and the external B_0 field. This average depends on the angle β between the internuclear vector and the rotor-axis. Rather than using only the β -averaged value (0.2) of $\langle (P_2(\cos \theta))^2 \rangle$, we calculated $\langle (P_2(\cos \theta))^2 \rangle$ as a function of $\cos\beta$ numerically and took the variation of $\langle (P_2(\cos \theta))^2 \rangle$ into account by selecting a random value of $\cos\beta$ between -1 and $+1$ for each internuclear vector r_{nm} and using the corresponding value of $\langle (P_2(\cos \theta))^2 \rangle$ in the calculated value of Π_{nm} according to the equation above. This approach provides the same variation of the squared coupling frequencies as full powder averaging and reproduces the observed nonexponential decays (2) better than simulations with a fixed value of $\langle (P_2(\cos \theta))^2 \rangle$.

The normalized CODEX signal $S_{\text{CODEX}}(t)$, which is the fraction of the magnetization remaining on the spin on which it started out, averaged over all spins, is calculated from the standard solution for the N -component vector \mathbf{M} of the z-magnetization values of the N exchanging spins

$$\mathbf{M}(t) = \exp(\Pi t)\mathbf{M}(0)$$

$$S_{\text{CODEX}}(t) = \sum_{n=1}^N (\exp(\Pi t)\mathbf{e}_n)_n = \text{tr}(\exp(\Pi t)).$$

In the intermediate step, \mathbf{e}_n is the unit vector with elements $(\mathbf{e}_n)_m = \delta_{nm}$ (i.e., 1 for $n = m$ and 0 otherwise), representing an initial state $\mathbf{M}(0) = \mathbf{e}_n$ with magnetization only on spin n . For a sufficiently long dephasing and refocusing time Nt_r , the detected CODEX signal from that spin is proportional to the magnetization $M_n(t) = (\exp(\Pi t)\mathbf{e}_n)_n$ remaining on that same spin n . The sum over n provides the summation of the contributions from all N ^{13}C spins; the trace operation (sum of diagonal elements) on the right-hand side of the equation achieves this in a particularly convenient way.

The simulated CODEX curves depend on the density and positional ordering of the citrate molecules. The ^{13}C distributions corresponding to the seven fit curves in Fig. 4 are displayed in Fig. S2. Partial positional ordering is achieved by increasing the excluded area around the center of each molecule. Minimal (“random”) ordering is generated by an excluded-area diameter of $d_{\text{excl}} = 0.8$ nm that corresponds to the diameter of a citrate molecule. For an area density of $1/d_{\text{cc}}^2$ moderate ordering was obtained with $d_{\text{excl}} = 0.6 d_{\text{cc}}$, and partial ordering with $d_{\text{excl}} = 0.75 d_{\text{cc}}$. A higher than average ($>1/(2 \text{ nm})^2$) citrate density is possible for the citrate-covered apatite surfaces, if other surfaces have little or no citrate bound. For instance, the average density of $1/(2 \text{ nm})^2$ can be obtained by combining 50% of surfaces at $1/(1.4 \text{ nm})^2$ (which yield signal) and 50% of surfaces without any citrate (which do not contribute to the measured signal).

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2. Luo W, Hong M (2006) Determination of the oligomeric number and intermolecular distances of membrane protein assemblies by anisotropic H-1-driven spin diffusion NMR spectroscopy. *J. Am. Chem. Soc* 128:7242–7251.

