

Supplementary methods

Cell culture

All cell lines were expanded in RPMI 1640 medium (Invitrogen) containing 10% fetal bovine serum (FBS, Gibco), glutamine (Gibco), and penicillin/streptomycin. Before experiments, CCRCC cells were cultured in this medium until they reached confluence, and then for an additional 1-2 days in the same medium without added serum (changed daily and with or without added inhibitors). pCMVR and pBABE infected populations were selected for 14 days and 4 days with G418 (1 mg/ml) and puromycin (2 µg/ml) respectively, this resulted in significant cell death and selection of stable populations which were then expanded. The 9x HRE VEGF reporter has been described elsewhere (supplementary reference 1).

Small Interfering RNA Transfection, qPCR and ChIP

siRNA reverse transfections were performed in p6-well plates, each well containing 5×10^6 suspended cells and 5 µl LipofectAMINE 2000 (Invitrogen). Oligo concentration was 100 nM unless otherwise indicated. After transfection, cells were allowed to grow for 1-2 days with 10% FBS and then without serum for 1-2 more days before analysis.

ChIP was performed as follows. DNA-protein complexes were cross-linked by adding 0.75% formaldehyde, followed by blocking with glycine. Cells were washed twice with cold PBS and lysed on ice using FA buffer (50 mM HEPES-KOH pH7.5, 140 mM NaCl, 1 mM EDTA pH8., 0, 1% Triton X-100, 0.1% sodium deoxycholate, 0.1% SDS, and protease inhibitors). Chromatin samples were sonicated on ice, diluted in RIPA buffer (50 mM Tris-HCl pH8, 150 mM NaCl, 2 mM EDTA pH8, 1% NP-40, 0.5% sodium deoxycholate, 0.1% SDS and protease inhibitors), centrifuged, and supernatants precleared with protein A sepharose beads pretreated with salmon sperm (Sigma). Antibodies were incubated overnight with the samples on a rotator at 4°C. Fresh protein A sepharose beads with salmon sperm were added afterwards, incubated for 2 hours, and washed at room temperature three times with wash buffer (0.1% SDS, 1% Triton X-100, 2 mM

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EDTA, 20 mM Tris-HCl pH 8, 150 mM NaCl) and once with final wash buffer (0.1% SDS, 1% Triton X-100, 2 mM EDTA, 20 mM Tris-HCl pH 8, 500 mM NaCl). DNA was eluted with elution buffer (1% SDS; 100mM NaHCO₃), reverse cross-linked and purified by chloroform/phenol extraction.

siRNA target sequences and qPCR/ChIP primers

siRNA:

For the firefly luciferase gene see (<http://www.rockefeller.edu/labheads/tuschl/>)

HIF-1 α oligo A 5` - CTGATGACCAGCAACTTGA

HIF-1 α oligo B 5` - CAATCAAGAAGTTGCATTA

HIF-1 α oligo C 5` - CGTTGTGAGTGGTATTATT

HIF-2 α oligo A 5` - CAGCATCTTTGATAGCAGT

HIF-2 α oligo B 5` - GCGACAGCTGGAGTATGAA

HIF-2 α oligo C 5` - GCAAATGTACCCAATGATA

NEDD9 oligo A 5` - CACCCAAGAACAAGAGGTA

NEDD9 oligo B 5` - GGAAAGGGATGGTGTTTAT

qPCR:

VEGF-F 5` -TGCCAAGTGGTCCCAG

VEGF-R 5` -GTGAGGTTTGATCCGC

AurkA-F 5` -TTCAGGACCTGTTAAGGCTAC

AurkA-R 5` -TCTGCTTCTGATTCTGAACC

NEDD9-isoform 1-F 5` -CCTCCTTCTCATACCACTCA

NEDD9-isoform 1-R 5` -GCGGGATGTCATACACC

NEDD9-isoform 2-F 5` -CCAGAGAGATGGGCAAGTGT

NEDD9-isoform 2-R 5` -CAAGCCTCCAAACTCAGGAC

p130CAS-F 5` -AACCCCACTGACAAGACCAG

p130CAS-L 5` -TCATAGTCCTCCATCCAGCC

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Claudin1-F 5'-GCTTTAAATCGCGGCGCCAG

Claudin1-R 5'-CTCTGGGTCGGGGTTGGGGTC

ChIP:

EPO-F 5'-CAGGCGTCCTGCCCCCTGCT

EPO-R 5'-CAGCCCGCGAGTACTCACCGTG

NEDD9-1-F 5'-GGAAGGGCTTTGAATCATTGTCCTCCTT

NEDD9-1-R 5'-CCTTGCTTCTCCTCATTGTCCTTAGAGCA

NEDD9-2-F 5'-CTCAGGGACAGCTATTGTCTGGATTAC

NEDD9-2-R 5'-CAGTCTCTCTCCAGCGAAGCACAGAAC

NEDD9-3-F 5'-CCAGGCTCATCAAACAGTGGGCAT

NEDD9-3-R 5'-GTCACACATATTGAGCGACCTTCAGTGG

NEDD9-4-F 5'-CCACTGAAGGTCGCTCAATATGTGTGA

NEDD9-4-R 5'-TGTCAGTCTGCAAGGAGCTGGAAGAC

Supplementary Figure Legends

Supplementary Figure 1-

Western blot using the indicated antibodies and lysates from *VHL*-defective RCC10 cells transfected with siRNA oligos (oligo pairs B and C) for HIF-1 α , HIF-2 α and the firefly luciferase gene (control).

Supplementary Figure 2-

(A-C) The number of nuclei stained with DAPI (3 experiments in duplicate and 8 fields per coverslip were analyzed) and number of cells stained with BrDU (3 experiments in duplicate and 8 fields per coverslip) were counted to assess changes in proliferation in the indicated cell lines and treatments. (D) Percentage of BrdU positive cells among all migrated cells (stained with DAPI); the results correspond to 3 experiments in duplicate (4 fields per coverslip).

Supplementary references

1. Aragonés J, Jones DR, Martín S, San Juan MA, Alfranca A, Vidal F, Vara A, Merida I, Landazuri MO: Evidence for the involvement of diacylglycerol kinase in the activation of hypoxia-inducible transcription factor 1 by low oxygen tension. *J Biol Chem* 276: 10548-55, 2001

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NEDD9-1-R 5'-CCTTGCTTCTCCTCATTGTCCTTAGAGCA

NEDD9-2-F 5'-CTCAGGGACAGCTATTGTCTGGATTAC

NEDD9-2-R 5'-CAGTCTCTCTCCAGCGAAGCACAGAAC

NEDD9-3-F 5'-CCAGGCTCATCAAACAGTGGGCAT

NEDD9-3-R 5'-GTCACACATATTGAGCGACCTTCAGTGG

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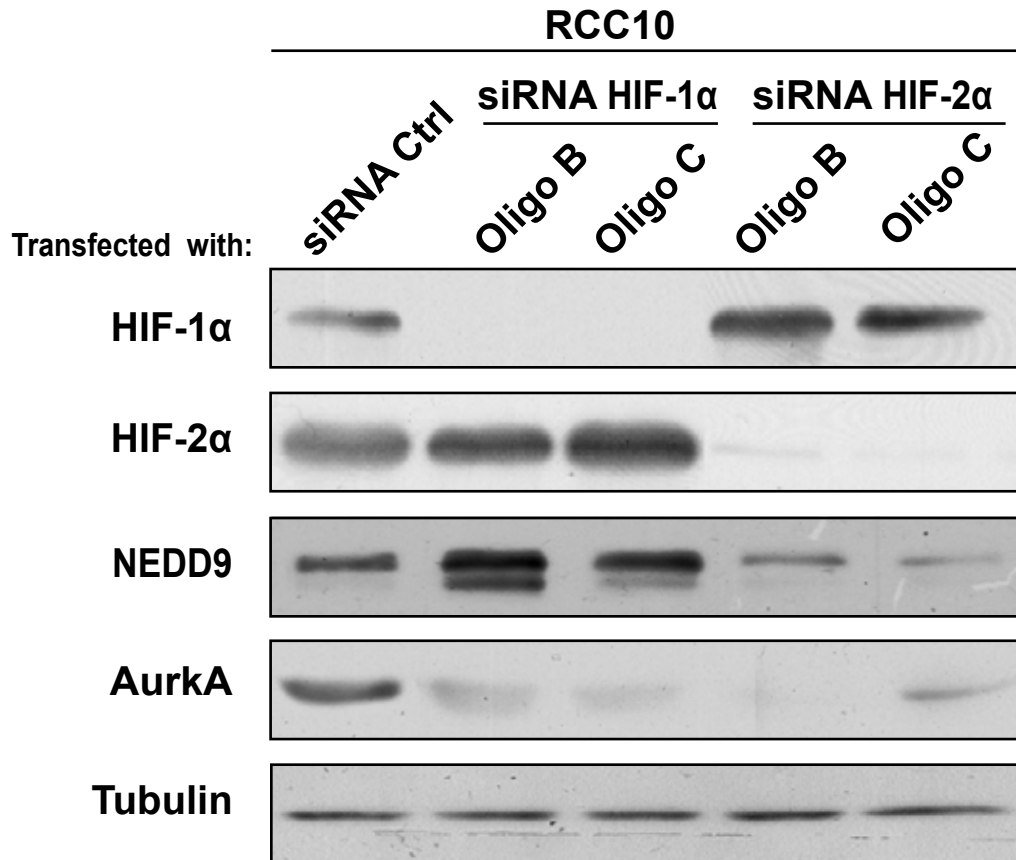
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Suppl. Fig. 1



Suppl. Fig. 2

