## **Support information**

## On-Plate Desalting and SALDI-MS Analysis of Peptides on Hydrophobic Silicate Nanofilms on a Gold Substrate

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Surface	SPR angle	Angle shift	Standard Deviation
Blank Au	62.21±0.01	-	-
8Layer-flat	62.40±0.06	0.19	32%
4Layer-rough	63.00±0.12	0.79	15%
8Layer-rough	64.12±0.26	1.91	14%



**Figure S1.** AFM images for calcinated substrates. (a) Au-covered steel substrate; (b) 8 layers of silicate fabricated with water washing between deposition of each polyelectrolytes; (c) 4 layers of silicate fabricated without water washing; (d) 8 layers of silicate fabricated without water washing. Bright streaks were observed across the images, corresponding to ridges and wrinkles on the SST substrate.



**Figure S2.** Effect of OTS modification on LDI performance of calcinated substrate. (a) Effect of OTS concentration, reaction time was 30 s; (b) effect of reaction time, OTS concentration was 10 mM. The analyte was 20 pmol of [Sar1, Thr8]-angiotensin II.



**Figure S3.** Dependency of signal-to-noise ratio for peptide protonated ions on laser intensity in SALDI-MS detection with C18-modified (square) and unmodified (circle) glassy substrates. Substrates were covered by 5 layers of silicate. The analyte was 20 pmol of [Sar1, Thr8]-angiotensin II. Error bars are omitted for simplicity.



**Figure S4.** Comparison of SALDI performance with different substrates. STT: stainless steel tape; Au-STT: gold-coated stainless steel tape; C18-Au-STT: gold-coated steel substrate treated with 1-octadecanethiol; OTS-glass: OTS modified glass substrate; OTS-calcinated film: OTS-modified calcinated substrate (8-layer smooth surface); CHCA-MALDI: MALDI with CHCA matrix. The analyte was mixture of [Sar1, Thr8]-angiotensin II and neurotensin, 20 pmol each.



**Figure S5.** Contact angle measurement on different substrates. (a) Bare gold substrate; (b) substrate coated with 8-layer S-film (with washing); (c) substrate coated with 8-layer R-film (without washing); (d) substrate coated with 8-layer S-film after OTS modification; (e) substrate coated with 8-layer R-film after OTS modification. CA: contact angle.



**Figure S6.** MALDI and SALDI mass spectra of peptide mixtures in the presence of detergents. (a) 0.2% SDS, CHCA matrix; (b) 0.2% SDS, OPD/SALDI on an OTS-modified calcinated surface; (c) 0.1% CHAPs, CHCA matrix; (d) 0.1% CHAPs, OPD/SALDI on an OTS-modified calcinated surface. The analyte was the mixture of [Sar1, Thr8]-angiotensin II (M1) and neurotensin (M2), 2 pmol each. Squares ( $\Box$ ) represent peaks from CHCA related ions; Circles ( $\circ$ ) represent peaks from detergents. NL: normalized level.



**Figure S7**. SALDI-MS analysis of peptides treated by different desalting methods: (a) off-probe desalting with ZipTip-C18 pipette tip; (b) on-plate desalting with OTS-modified 8-layer R-film. The analyte was the mixture of [Sar1, Thr8]-angiotensin II (M1) and neurotensin (M2) in 200 mM NaAc, 100 fmol each.

Table S2. Detected peptides in 40fmol cytochrome c digest in the presence of 50 mM NH <sub>4</sub> HCO <sub>3</sub>							
Peaks	Calculated m/z	Sequence	Position	$pI^{a}$	<b>GRAVY</b> <sup>a</sup>	MALDI-MS	SALDI-MS
CT1	779.448	MIFAGIK	81-87	8.50	1.600	*	*
CT 2	790.405	TDANKNK	50-56	8.26	-2.457		*
CT 3	955.500	TGPNLHGLF	29-37	6.40	0.067	*	*
CT 4	1065.521	GEREDLIAY	90-98	4.14	-0.733		*
CT 5	1193.616	KGEREDLIAY	89-98	4.68	-1.050	*	*
CT 6	1220.653	HKTGPNLHGLF	27-37	8.76	-0.591	*	*
CT 7	1321.711	KKGEREDLIAY	88-98	6.18	-1.309	*	*
CT 8	1456.670	TGQAPGFSYTDANK	41-54	5.50	-0.993	*	
CT 9	1584.765	KTGQAPGFSYTDANK	40-54	8.50	-1.187	*	
CT 10	1633.819	IFVQKCAQCHTVEK	10-23	8.06	0.021	*	*
CT 11	1880.858	VQKCAQCHTVEKGGKHK	12-28	9.31	-1.106	*	
CT 12	2060.223	AGIKKKGEREDLIAYLKK	84-101	9.70	-0.911		*
CT 13	2138.047	GITWGEETLMEYLENPKK	57-74	4.49	-0.961	*	
		Matched Peptides				10	9
		Sequence Coverage (%)				77.1	50.5
<sup>a</sup> pI and	Grand average	of hydropathicity index (GRAV	Y) were	calculat	ed by using	g the ProtPara	m tool (http:
//ca.expa	asy.org/tools/protpa	aram.html). Asterisks (*) represents	the identifie	d peptic	les in MS det	ection.	

Table S3. Identified β-casein peptides in MS detection								
Peaks	Calculated m/z	Sequence	Position	$pI^a$	<b>GRAVY</b> <sup>a</sup>	MALDI-MS	SALDI-MS	
BT1	830.452	AVPYPQR	192-198	8.79	-0.929	*		
BT2	1138.640	VKEAMAPKHK	113-122	9.70	-1.030	*		
BT3	1383.799	LLYQEPVLGPVR	206-217	6.00	0.283	*	*	
BT4	2061.828	FQpSEEQQQTEDELQDK <sup>b</sup>	48-63	3.77	-2.331	*		
BT5	2107.231	LLYQEPVLGPVRGPFPIIV	206-224	6.00	0.832	*	*	
BT6	2556.093	FQpSEEQQQTEDELQDKIHPF <sup>b</sup>	48-67	4.08	-1.740	*		
Matched Peptides					6	2		
		Sequence Coverage (%)				25.0	8.5	
<sup>a</sup> see Table S2. <sup>b</sup> phosphorylation at serine position. Asterisks (*) represents the identified peptides in MS detection.								

Table S4. Identified α-casein peptides in MS detection								
Peaks	Calculated m/z	Sequence	Position	$pI^a$	<b>GRAVY</b> <sup>a</sup>	MALDI-MS	SALDI-MS	
AT1	824.430	YPELFR	161-166	6.00	-0.717	*	*	
AT2	971.498	FYPELFR	160-166	6.00	-0.214	*	*	
AT3	1267.704	YLGYLEQLLR	106-115	6.00	0.070	*	*	
AT4	1337.681	HIQKEDVPSER	95-105	5.45	-1.755	*		
AT5	1759.945	HQGLPQEVLNENLLR	23-37	5.40	-0.753	*	*	
AT6	1951.952	YKVPQLEIVPNpSAEER <sup>b</sup>	119-134	4.79	-0.794	*	*	
		Matched Peptides				6	5	
Sequence Coverage (%) 27.6						22.4		
<sup>a</sup> see Table S2. <sup>b</sup> Phosphorylation at serine position. Asterisks (*) represents the identified peptides in MS detection.								

	Table S5. Detected peptides of 40 pmol BSA digests in the presence of 800 mM urea								
	Calculated m/z	Sequence	Position	Modification	$pI^a$	<b>GRAVY</b> <sup>a</sup>	MALD-MS	SALDI-MS	
1	789.472	LVTDLTK	257-263		5.84	0.429		*	
2	853.424	CASIQKF	223-229	CYS_CAM $(233)^{b}$	8.22	0.486		*	
3	1017.584	GFQNALIVR	425-433		9.75	0.578	*	*	
4	1075.521	YANKYNGVF	180-188		8.50	-0.567	*	*	
5	1170.525	SQYLQQCPF	52-60	CYS_CAM (58)	5.24	-0.567	*	*	
6	1180.547	GFQNALIVRY	425-434		8.75	0.390	*	*	
7	1226.703	ISLLLLFSSAY	7-17		5.52	1.873		*	
8	1351.685	LSHKDDSPDLPK	127-138		5.30	-1.558		*	
9	1439.812	RHPEYAVSVLLR	360-371		8.75	-0.133		*	
10	1479.795	LGEYGFQNALIVR	421-433		6.00	0.292		*	
11	1639.938	KVPQVSTPTLVEVSR	437-451		8.75	-0.067		*	
12	1673.77	QEPERNECFLSHK	118-130	CYS_CAM (125)	5.50	-1.723		*	
13	1880.921	RPCFSALTPDETYVPK	508-523	CYS_CAM (510)	6.06	-0.537		*	
	Matched Peptides							13	
		Sequence C	overage (%	6)			4.6	16.8	

position is shown in parentheses. Asterisks (\*) represents the identified peptides in MS detection.