SUPPLEMENTAL FIGURE LEGENDS

- Fig. S1. **Relative mRNA expression of mouse PC7 in different tissues.** Quantitative polymerase chain reactions were performed on RNA isolated from mouse tissue using specific oligonucleotides for mouse PC7 and normalized to 10⁶ S16 mRNA levels, as described in the Experimental Procedures.
- Fig. S2. PC7 traffics through an unconventional pathway in COS-1 cells. Immunostaining of PC7 (*red labeling*) and LDLR (*green labeling*) under non-permeabilizing conditions in COS-1 cells expressing either r-PC7 or human LDLR as control, treated or not with BFA (5 μg/ml) for 6h. Nuclei were stained with Hoescht 33258 (*blue labeling*). Bars = 10 μm.
- Fig. S3. **PC7 does not induce ER stress.** Xbp-1 mRNA splicing from HEK293 cells expressing, or not, r-PC7 or treated for 4h with tunicamycin (5 μ g/ml), was assessed by RT-PCR. A 263 bp amplicon (*s*) was generated from spliced Xbp-1 in cells treated with tunicamycin, whereas only an unspliced (*u*) Xbp-1 amplicon (289 bp) was detected in HEK293 cells transfected with r-PC7, demonstrating that PC7 does not induce ER stress.



Figure S1



Figure S2



Figure S3

Table S1. Oligonucleotides used for site-directed mutagenesis of PC7 and for the swapping of CT and TMCT of PC7 and Furin.

Mutants	Sense	Antisense
h-PC7 (C699A, C704A)	CAAGTTGCTAGGAGTGGACCC GCCCACTGGCCCC	GTGGGCGGGTCCACTCCTAGC AACTTGATTGG
h-PC7 _{CT-Furin}	GGAAGTATATTTGCGCTCTGGC TTTAGTTTTCG	CCAGAGCGCAAATATACTTCCA GCATGTAGTAAAC
h-PC7 _{TMCT-Furin}	CTCAAGACCGTGGTGGCCGG CC	GGCCACCACGGTCTTGAGGGT GTTGGGGG
h-Furin _{CT-PC7}	GCAGCTGAGCCAGAGGAATGT GG	CCTCTGGCTCAGCTGCAGGAC CAGG
h-Furin _{TMCT-PC7}	GCCTGAGCTGGTGCTGGTAGG CTG	GCACCAGCTCAGGCAGGTGTG AGGG

Table	S2: Oligonucl	eotides used	l for qu	uantitative i	real time-PCR.
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Mutants	Sense	Antisense	
Mouse PC7	GTTATCAGGGATGTAGGAGA	AAGGGTCTTGAGTGTGTTAG	
Mouse S16	AGGAGCGATTTGCTGGTGTGG	GCTACCAGGGCCTTTGAGATG	