

Supplementary Information

GGTase-I deficiency hyperactivates macrophages and induces erosive arthritis in mice

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SUPPLEMENTARY METHODS

Computerized Tomography

Peripheral quantitative computerized tomography (PQCT) scans were performed *ex vivo* on the tibia with a PQCT XCT RESEARCH M (version 4.5B; Norland), operating at a resolution of 70 μm as described (1). For quantification of trabecular bone mineral density (BMD), the scan was performed in the metaphyseal part of the proximal tibia; for cortical bone parameters, the scan was performed in the mid-diaphyseal region. Micro-CT analyses were performed on the distal femur with a Skyscan 1072 scanner (SkyScan) and imaged with an X-ray tube (100 kV, 98 μA). The scanning angular rotation was 180°, the angular increment was 0.90°, and the isotropic volume-pixel size was 6.51 μm . Images were generated by reconstructing datasets with a modified Feldkamp algorithm and segmenting them into binary images by adaptive local thresholding.

T lymphocyte–dependent and –independent immune responses in vivo

T lymphocyte–independent inflammation was induced by injecting 30 μl of olive oil in a hind paw which primarily recruits neutrophils. The thickness of the paw was measured before and 24 h after the injection (2). T lymphocyte–dependent inflammation was evaluated with the delayed-type hypersensitivity reaction induced by 4-ethoxymethylene-2-phenyloxazolone as described (2).

Immunocytochemistry

Differentiated BM macrophages were plated in 8-well chamber slides (10^4 cells/well). The cells were incubated with 10 μM GGTI-298 (Sigma, G5169) and 5 μM FTI (Lonafarnib, Schering-Plough) for 16 h. Cells were fixed in methanol and incubated with antibodies recognizing RAC1 (ARC03) as described in Methods and prelamin A as described (3).

SUPPLEMENTARY REFERENCES

1. Windahl, SH, Vidal, O, Andersson, G, Gustafsson, JA, and Ohlsson, C. Increased cortical bone mineral content but exchanged trabecular bone mineral density in female ER beta(-/-) mice. *J. Clin. Invest.* 1999;104:895–901.
2. Verdrengh, M, Jonsson, IM, Holmdahl, R, and Tarkowski, A. Genistein as an anti-inflammatory agent. *Inflamm. Res.* 2003;52:341–346.
3. Liu, M, et al. Targeting the protein prenyltransferases efficiently reduces tumor development in mice with K-RAS-induced lung cancer. *Proc. Natl. Acad. Sci. U S A* 2010;107:6471–6476.

SUPPLEMENTARY FIGURE LEGENDS

Supplementary Figure 1 (A) Micro-CT scans of the distal femur showing the articular surface of the knee joint of a *Pggt1b^{fl/+}LC* and a *Pggt1b^{fl/fl}LC* mouse. Arrows indicate site of erosion. (B,C,D) PQCT scanning was used to quantify trabecular bone mineral density (BMD) (B), cortical BMD (C), and cortical thickness (D) in the tibia of 12-week-old *Pggt1b^{fl/+}LC* and *Pggt1b^{fl/fl}LC* mice ($n = 10/\text{genotype}$). (E) Hematoxylin and eosin–stained sections of tissues from 12-week-old *Pggt1b^{fl/+}LC* and *Pggt1b^{fl/fl}LC* mice. Similar histology was seen in four additional mice/genotype. Scale bar, 400 μm . (F) Immunohistochemical staining of lung and bone marrow sections of 12-week-old mice to document the specificity of the non-prenylated (np)-RAP1A antibody. Note the absence of staining in tissues from a control *Pggt1b^{fl/+}LC* mouse. Scale bar, 35 μm . (G) Neutrophil infiltration measured as an increase in paw thickness of mice injected subcutaneously with olive oil ($n = 5/\text{genotype}$). (H) T lymphocyte–dependent inflammation measured as an increase in ear thickness after an oxa-mediated delayed-type hypersensitivity reaction ($n = 5/\text{genotype}$). Data are expressed as mean \pm SEM.

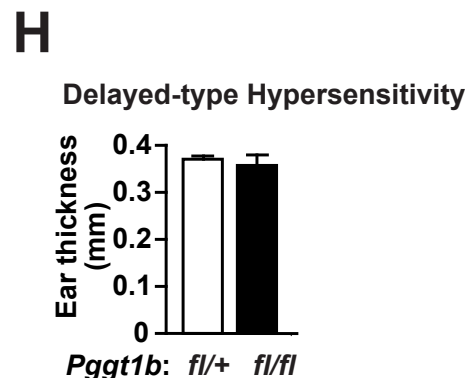
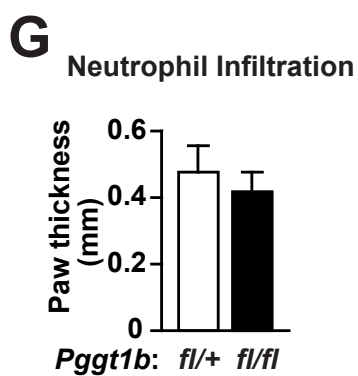
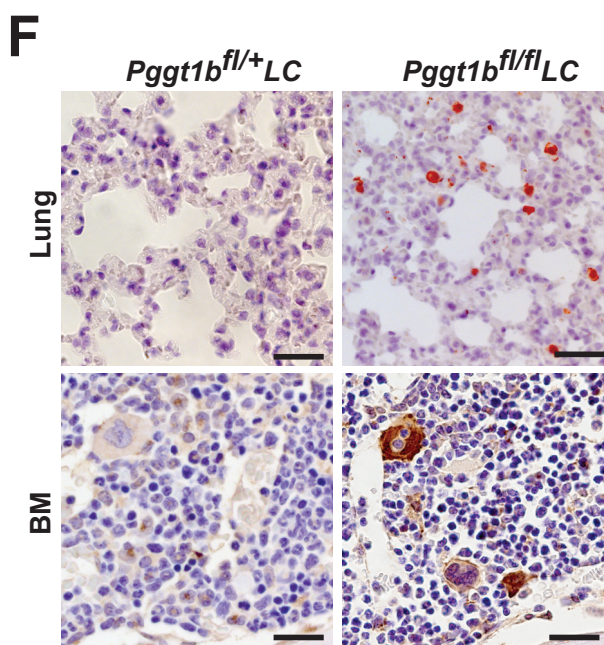
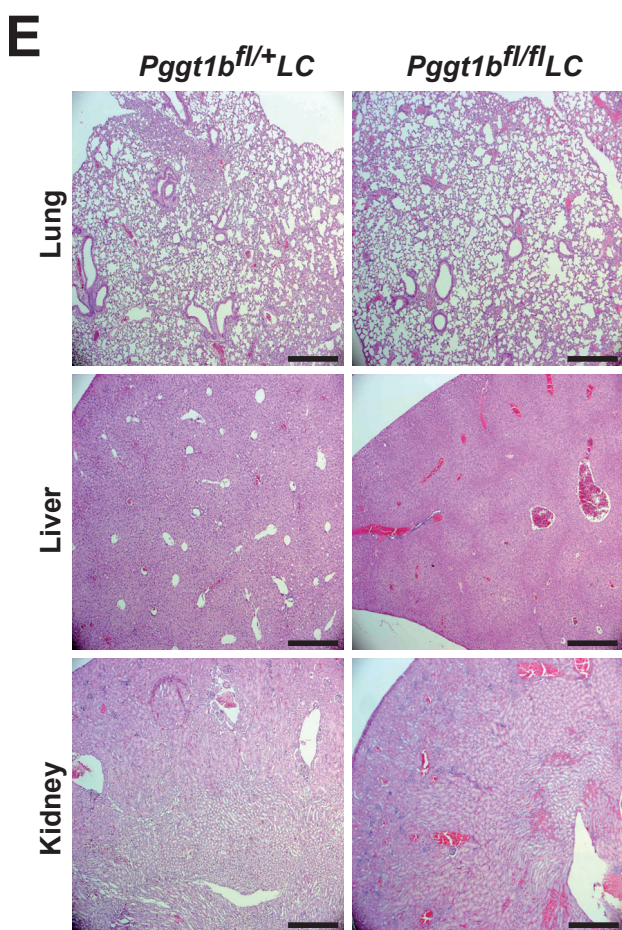
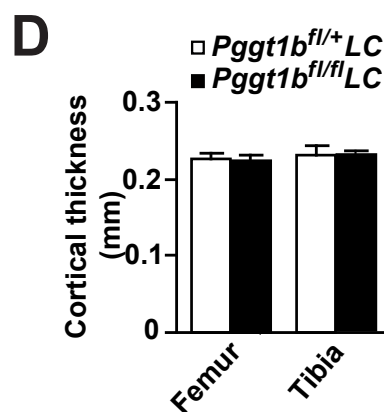
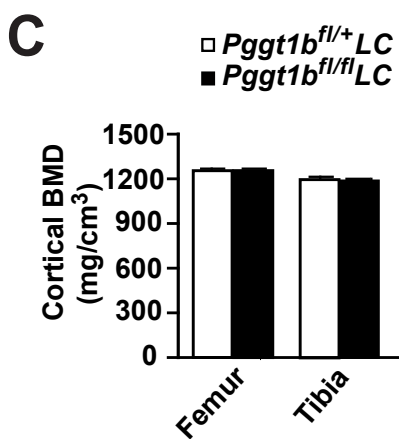
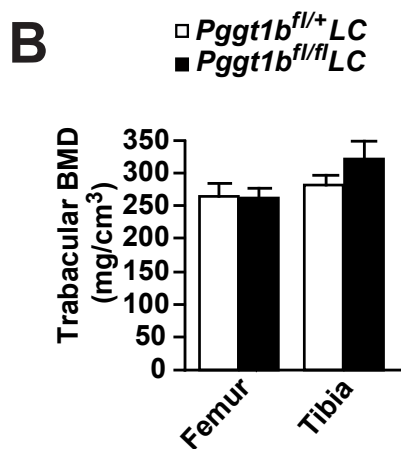
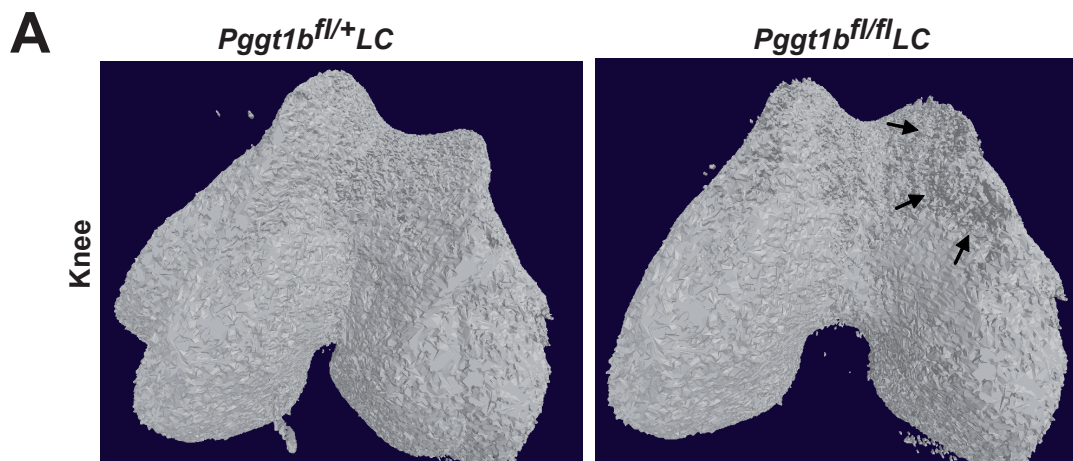
Supplementary Figure 2 (A) Hematoxylin and eosin–stained sections of joints from wild-type (8 weeks old at transplant) and *Pggt1b^{fl/fl}LC* (12 weeks old at transplant) mice that were lethally irradiated and transplanted with *Pggt1b^{fl/fl}LC* and wild-type

BM, respectively. The mice were killed 14 weeks after transplantation. E, erosion; S, synovium. Scale bar, 200 μm (B) Synovitis and bone erosion evaluated in knee, ankle, metatarsal, elbow, wrist, and metacarpal joints of wild-type (8 weeks old at transplant) and *Pggt1b^{fl/fl}*LC (12 weeks old at transplant) mice transplanted with wild-type and *Pggt1b^{fl/fl}*LC BM, respectively ($n = 4/\text{genotype}$). The mice were killed 14 weeks after transplantation. Data are expressed as mean \pm SEM.

Supplementary Figure 3 (A) Western blots illustrating the reduced electrophoretic mobility of affinity purified RAC1 in extracts of *Pggt1b^{fl/fl}*LC compared to *Pggt1b^{fl/+}*LC BM macrophages in multiple experiments. Protein extracts were resolved on 10% or 12% Tris-HCl Protean gels with the 20 kDa molecular weight marker run to the lower end of the gel. (B) Western blot illustrating the reduced migration of affinity purified RAC1 in extracts of *Pggt1b^{fl/fl}*LC BM macrophages and *Pggt1b^{fl/+}*LC BM macrophages incubated with 10 μM GGTI for 24 h. (C) Western blots of lysates from *Rac1^{fl/fl}* mouse embryonic fibroblasts incubated with adenoviruses expressing Cre (to produce *Rac1* null cells) or β -gal (to produce control parental cells) to document the specificity of the RAC1 antibodies from Millipore (used in Figure 3D, 3F and 3G) and Pierce (used in Figure 3A and Supplementary Figure 3A). (D) Confocal micrographs documenting the specificity of the RAC1 antibody used for immunocytochemistry (ARC03, Cytoskeleton) in Figure 3E and Supplementary Figure 3D. Note reduced immunofluorescence of RAC1 in cells incubated for 3 d with a *RAC1*-shRNA lentivirus compared to control *RAC2*-shRNA lentivirus. Scale bar, 100 μm . (E) Confocal micrographs showing immunofluorescence staining of RAC1 and prelamin A in *Pggt1b^{fl/fl}*LC and *Pggt1b^{fl/+}*LC BM macrophages incubated with 5 μM FTI and 10 μM GGTI for 24 h. Evidence that the GGTI was inhibiting GGTase-I is illustrated by the reduced cell adhesive area of the *Pggt1b^{fl/+}*LC cells. Evidence that the FTI was inhibiting FTase is illustrated by the prelamin A staining (prelamin A is normally farnesylated by FTase and rapidly processed to mature lamin A; when FTase is inhibited, nonfarnesylated prelamin A accumulates and is detected by the antibody). Scale bars, 10 μm .

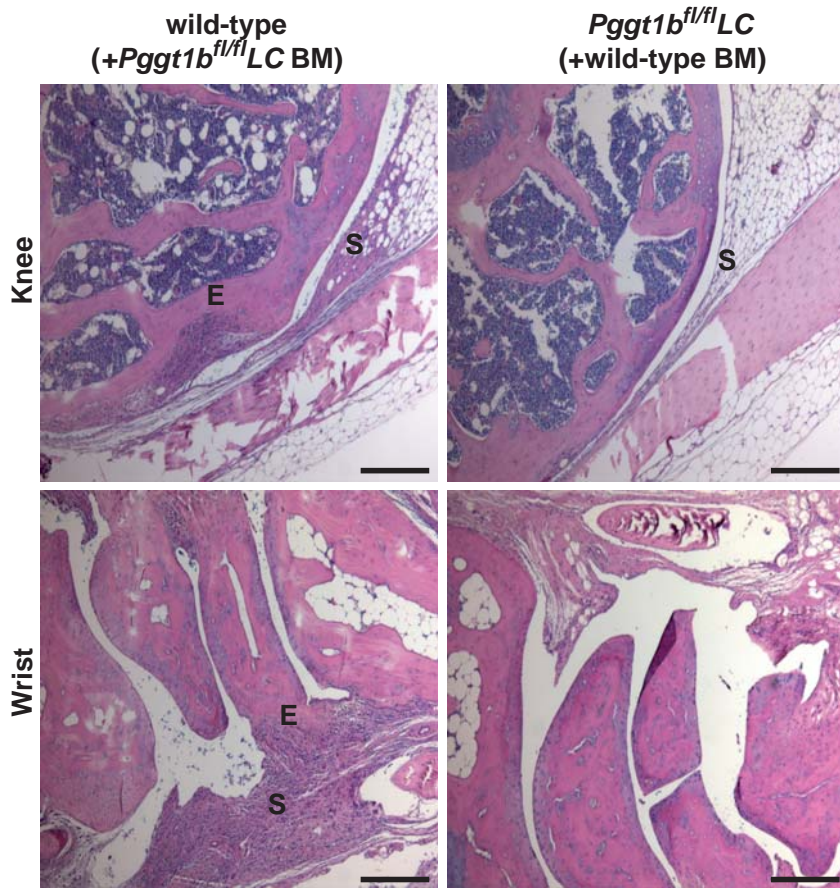
Supplementary Figure 4 (A) RT-QPCR analyses confirming the increased expression of the indicated genes in LPS-stimulated *Pggt1b^{fl/fl}*LC compared to *Pggt1b^{fl/+}*LC BM macrophages ($n = 3/\text{genotype}$). The primers were different from those used in Figures 4E and F. (B) RT-QPCR analyses showing relative expression of the genes in A using cDNA from LPS-stimulated *Pggt1b^{fl/fl}*LC and *Pggt1b^{fl/+}*LC intraperitoneal (IP) macrophages. Data are expressed as the mean ($n = 3/\text{genotype}$).

Supplementary Figure 1

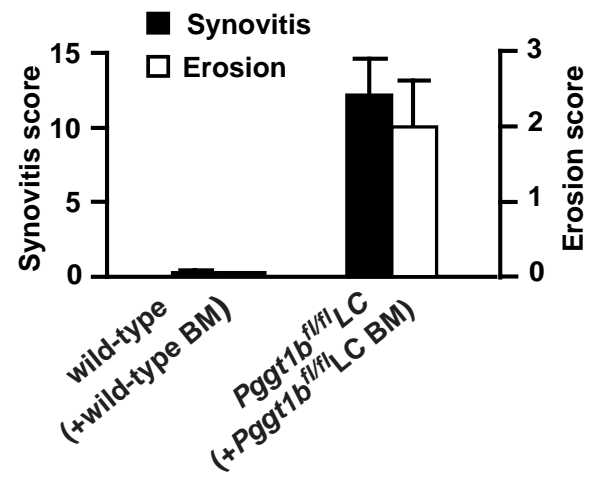


Supplementary Figure 2

A

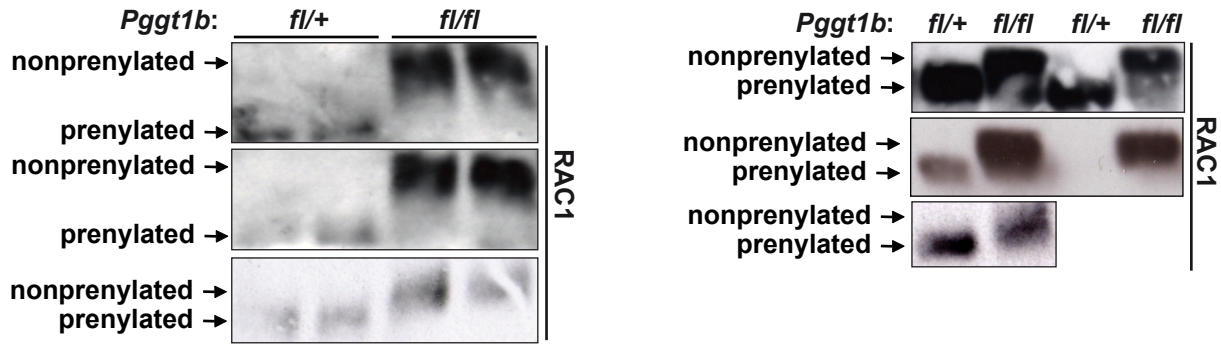


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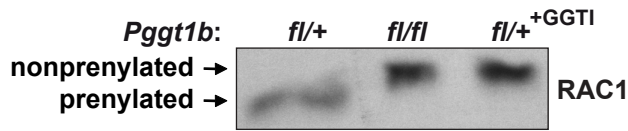


Supplementary Figure 3

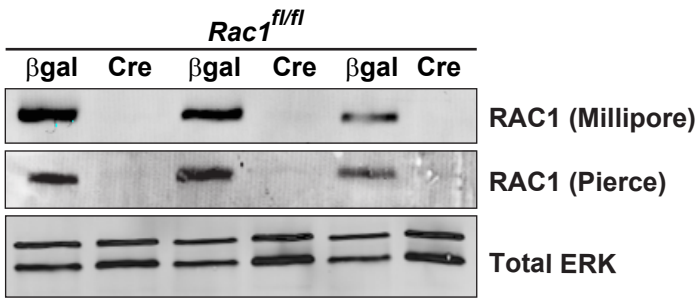
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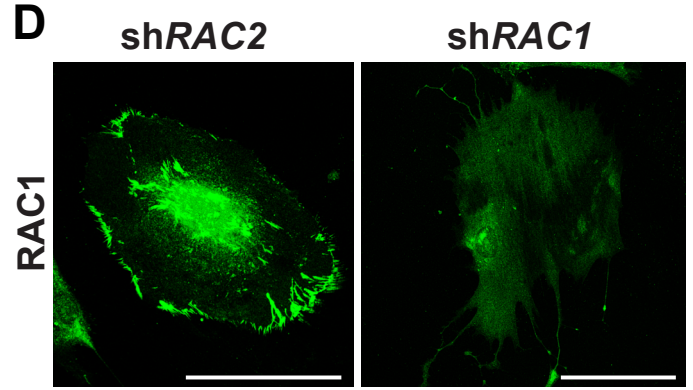
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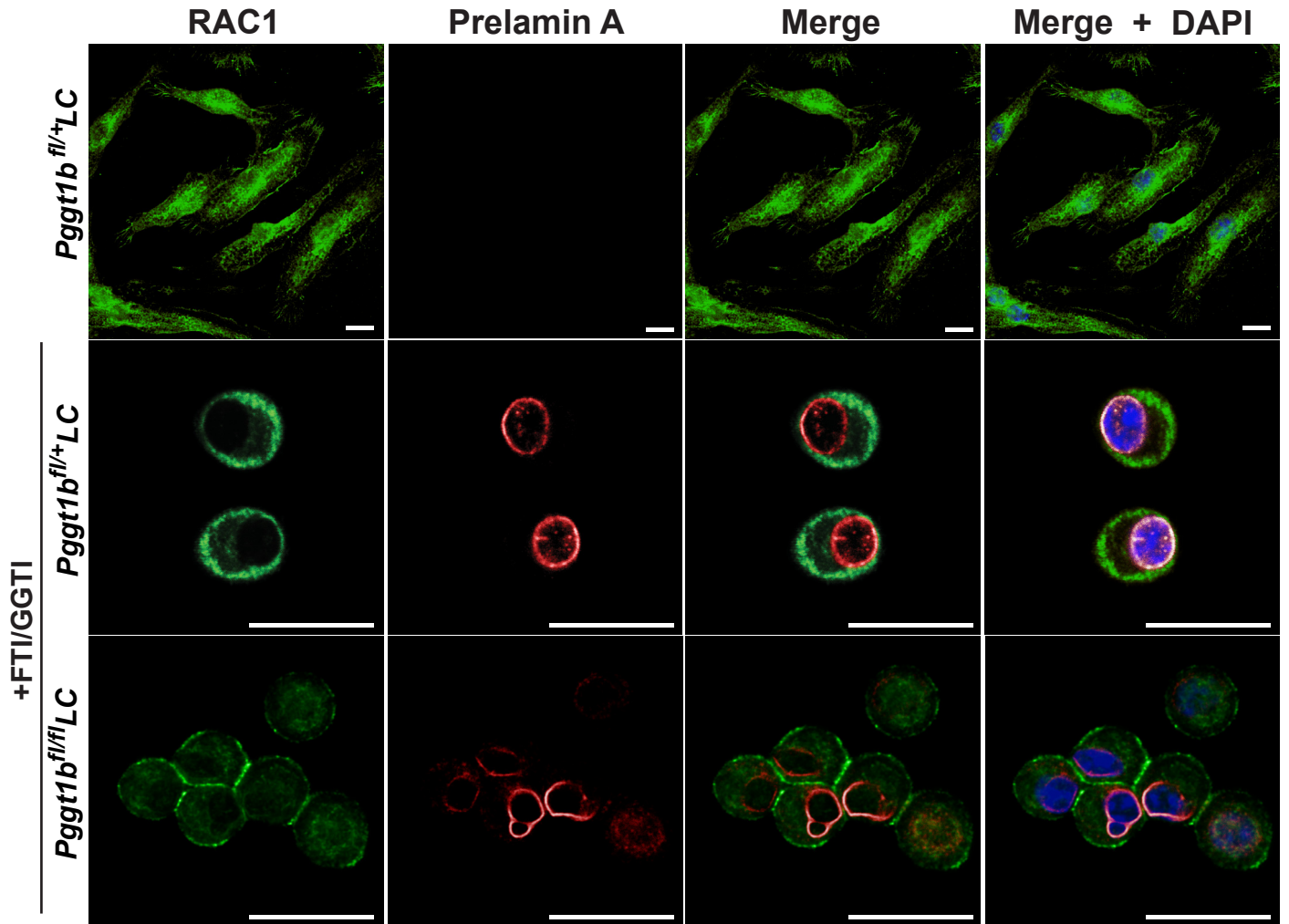
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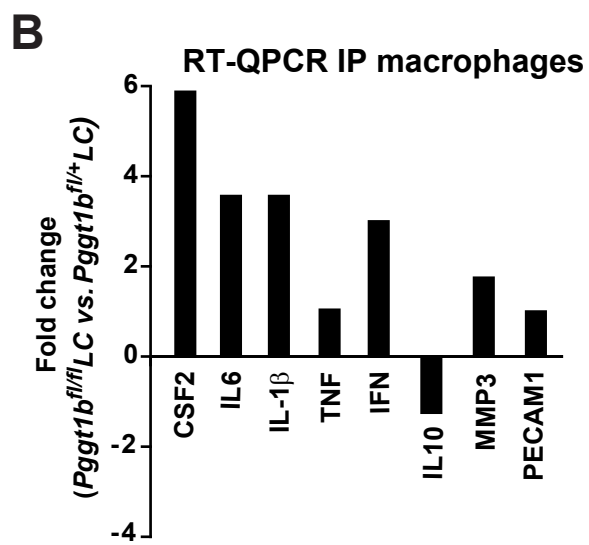
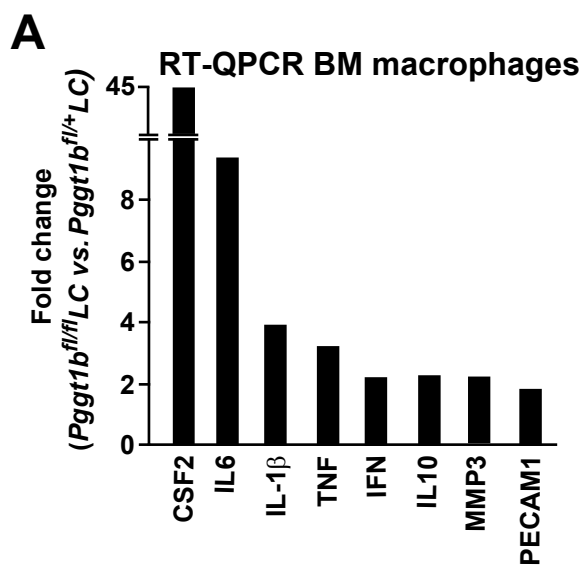
D



E



Supplementary Figure 4



Supplementary Table 1

Mouse NF κ B Signaling Pathway PCR Array (*Pggt1b^{fl/fl}*LC vs *Pggt1b^{fl/+}*LC)

Gene	Fold change	p value	Gene	Fold change	p value
Akt1	1.28	0.435	Nfkb1	1.73	0.254
Atf1	1.40	0.196	Nfkb2	1.17	0.686
Atf2	2.15	0.112	Nfkbia	1.31	0.556
Bcl10	1.59	0.292	Pcaf	1.34	0.227
Bcl3	2.27	0.060	Eif2ak2	0.95	0.816
C3	1.31	0.521	Raf1	1.63	0.035
Card10	1.95	0.121	Rel	2.06	0.031
Nod1	1.40	0.486	Rela	1.22	0.481
Casp1	1.11	0.764	Relb	1.20	0.614
Casp8	1.37	0.473	Ripk1	1.17	0.702
Ccl2	2.74	0.072	Ripk2	1.48	0.408
Cflar	2.42	0.113	Slc20a1	1.24	0.576
Chuk	1.64	0.154	Smad3	0.64	0.365
Crebbp	1.53	0.150	Stat1	0.84	0.638
Csf2	44.75	0.00001	Tbk1	1.46	0.270
Csf3	15.82	0.00006	Tgfr1	0.71	0.312
Edg2	1.66	0.176	Tgfr2	0.50	0.007
Egr1	2.05	0.144	Tlr1	1.29	0.304
Elk1	0.98	0.957	Tlr2	0.89	0.619
F2r	2.91	0.012	Tlr3	0.49	0.010
Fadd	1.14	0.815	Tlr4	1.70	0.154
Fasl	1.64	0.246	Tlr6	1.34	0.442
Fos	0.71	0.525	Tlr7	1.04	0.935
Gja1	4.26	0.012	Tlr8	0.56	0.153
Htr2b	1.07	0.892	Tlr9	0.50	0.136
Icam1	1.27	0.509	Tnf	4.00	0.010
Irfng	2.87	0.058	Tnfaip3	2.98	0.043
Ikbkb	1.61	0.190	Tnfrsf10b	1.04	0.876
Ikbke	1.01	0.971	Tnfrsf1a	1.07	0.615
Ikbkg	1.82	0.232	Tnfrsf1b	1.21	0.459
Il10	2.39	0.123	Cd40	1.65	0.179
Il1a	5.31	0.002	Cd27	1.98	0.174
Il1b	5.73	0.001	Tnfsf10	0.41	0.001
Il1r1	1.09	0.867	Tnfsf14	0.97	0.906
Il6	8.06	0.001	Tollip	1.19	0.610
Irak1	1.06	0.902	Tradd	1.10	0.710
Irak2	1.70	0.134	Traf2	0.80	0.491
Irf1	1.18	0.524	Traf3	1.17	0.650
Jun	1.13	0.629	Zap70	2.03	0.194
Lta	2.83	0.007	Gusb	0.96	0.841
Ltbr	1.09	0.793	Hprt1	0.69	0.585
Map3k1	0.93	0.882	Hsp90ab1	1.54	0.046
Mapk3	1.01	0.991	Gapdh	1.09	0.669
Myd88	1.06	0.901	Actb	0.90	0.585
Nlrp12	2.04	0.109			

Supplementary Table 2

Mouse Extracellular Matrix and Adhesion Molecules PCR Array (*Pggt1b^{fl/fl}*LC vs *Pggt1b^{fl/+}*LC)

Gene	Fold change	p value	Gene	Fold change	p value
Adamts1	4.45	0.025	Lama2	1.67	0.360
Adamts2	4.04	0.184	Lama3	1.12	0.862
Adamts5	2.27	0.194	Lamb2	1.43	0.396
Adamts8	1.94	0.275	Lamb3	1.52	0.415
Ctnna1	2.30	0.170	Lamc1	1.32	0.578
Ctnna2	1.85	0.266	Mmp10	3.72	0.129
Ctnnb1	1.59	0.446	Mmp11	1.24	0.709
Cd44	3.40	0.062	Mmp12	3.56	0.038
Cdh1	1.32	0.698	Mmp13	3.55	0.060
Cdh2	4.84	0.106	Mmp14	1.26	0.660
Cdh3	1.81	0.288	Mmp15	1.27	0.722
Cdh4	2.60	0.293	Mmp1a	4.23	0.056
Cntn1	1.96	0.225	Mmp2	1.28	0.499
Col1a1	1.66	0.699	Mmp3	7.69	0.009
Col2a1	2.60	0.207	Mmp7	2.26	0.203
Col3a1	5.88	0.101	Mmp8	1.62	0.180
Col4a1	2.12	0.175	Mmp9	1.61	0.303
Col4a2	1.84	0.179	Ncam1	2.82	0.067
Col4a3	1.52	0.544	Ncam2	2.01	0.227
Col5a1	3.17	0.240	Pecam1	2.00	0.046
Col6a1	3.41	0.196	Postn	2.61	0.183
Vcan	2.98	0.080	Sele	2.30	0.239
Ctgf	2.98	0.463	Sell	4.72	0.005
Ecm1	2.40	0.062	Selp	3.90	0.062
Emilin1	1.02	0.964	Sgce	9.74	0.126
Entpd1	1.00	0.994	Sparc	3.08	0.388
Fbln1	1.67	0.549	Spock1	1.81	0.289
Fn1	81.06	0.002	Spp1	1.23	0.611
Hapln1	1.81	0.289	Syt1	1.85	0.264
Hc	1.84	0.063	Tgfbi	0.49	0.082
Icam1	1.98	0.201	Thbs1	2.65	0.070
Itga2	3.31	0.148	Thbs2	5.82	0.067
Itga3	2.72	0.180	Thbs3	2.97	0.017
Itga4	1.63	0.273	Timp1	1.81	0.287
Itga5	1.87	0.160	Timp2	0.69	0.419
Itgae	1.63	0.478	Timp3	2.53	0.356
Itgal	1.85	0.323	Tnc	0.58	0.605
Itgam	1.81	0.253	Vcam1	1.86	0.280
Itgav	1.58	0.443	Vtn	1.53	0.579
Itgax	1.26	0.761	Gusb	1.12	0.579
Itgb1	1.64	0.294	Hprt1	0.58	0.530
Itgb2	2.31	0.048	Hsp90ab1	1.45	0.175
Itgb3	1.75	0.382	Gapdh	1.28	0.175
Itgb4	1.31	0.674	Actb	0.82	0.451
Lama1	1.83	0.288			