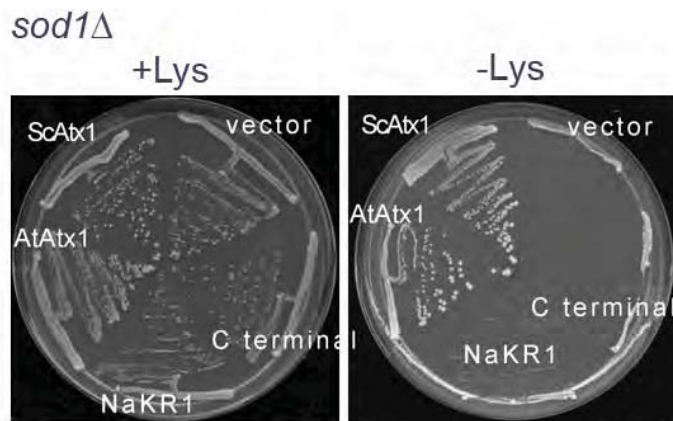


**Supplemental Figure 1. Identification of a 7 bp deletion in At5g02600.** *nkr1-1* and Col-0 WT genomic DNA was hybridized to Arabidopsis thaliana ATTILE 1.0R arrays. The vertical axis is the difference between the mean hybridization of *nkr1-1* and Col-0. Horizontal axis is the base pair position on chromosome 5. The data points represent hybridization to individual 25-mer features on the *A. thaliana* ATTILE 1.0R chip. Microarray data have been submitted to the GEO database with accession: GSE24385.



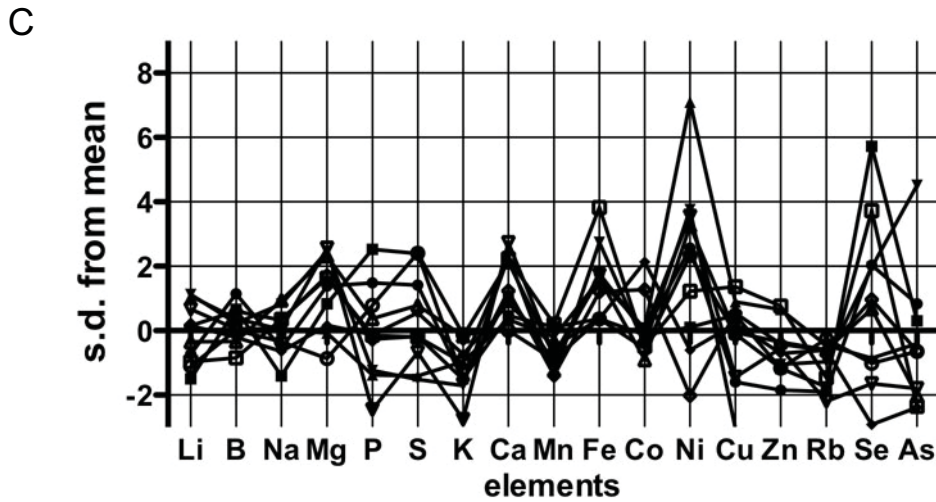
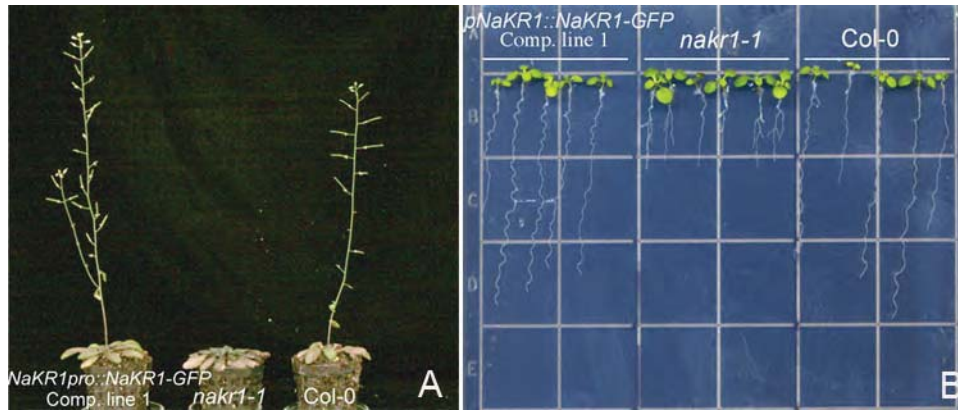
**Supplemental Figure 2. *AtNaKR1* and the C-terminal HMA domain failed to rescue the lysine auxotrophy phenotype of the yeast *sod1Δ* mutant.**

Lysine auxotrophy is an indication for reactive oxygen toxicity. Yeast *sod1Δ* cells transformed with p426GPD vector (vector) alone or *Saccharomyces cerevisiae* *ATX1* (*ScATX1*), *Arabidopsis ATX1* (*AtATX1*), *AtNaKR1* and the C-terminal region (containing the HMA domain) were assayed for growth on SC-Ura and SC-Ura lacking lysine medium.

MKTEEGKLVIWINGDKGYNGLAEVGKKFEKDTGIKVTV  
EHPDKLEEKFQVAATGDGPDIIFWAHDRFGGYAQSG  
LAEITPDKAFQDKLYPFTWDAVRYNGKLIAYPIAVEAL  
SLIYNKDLLPNPKTWEEIPALDKELKAKGKSSALMFNL  
QEPYFTWPLIAADGGYAFKYENGKYDIKDVGVDNAGAK  
AGLTFLVDLIKNKHMNATDYSIAEAAFNKGETAMTIN  
GPWAWSNIDTSKVNYGVTVLPTFKGQPSKPFVGVLSSAG  
INAASPNKELAKEFLENYLLTDEGLEAVNKDKPLGAVA  
LKSYEEELAKDPRIAAATMENAQGEIMPNIPQMSAFWY  
AVRTAVINAASGRQTVDEALKDAQTNSSNNNNNNNN  
NLGIEGRISEFLNQTSLYKKAEGPSRPEGDITMLCASQ  
ASTTTLCSTMDQTSQPSSSSSATIRLGGRAIDRHNPII  
RDGRRLTPPPSPNLNPSSSSSSTYHTPLMTRLGLESS  
QKRLAKRKSKGDSDVGKSPVSCFSSDTPQGSSRYLLS  
NPVFFDGFVDSDPIPIPIDEPEITKADDLNNFHEDRLI  
INASKYLSTSASFLEKKQPDFFEGFLDYEPVLSPDNPF  
SEPTKASPTASLSSLEDKDVSSPDFKFSPPPPPPSP  
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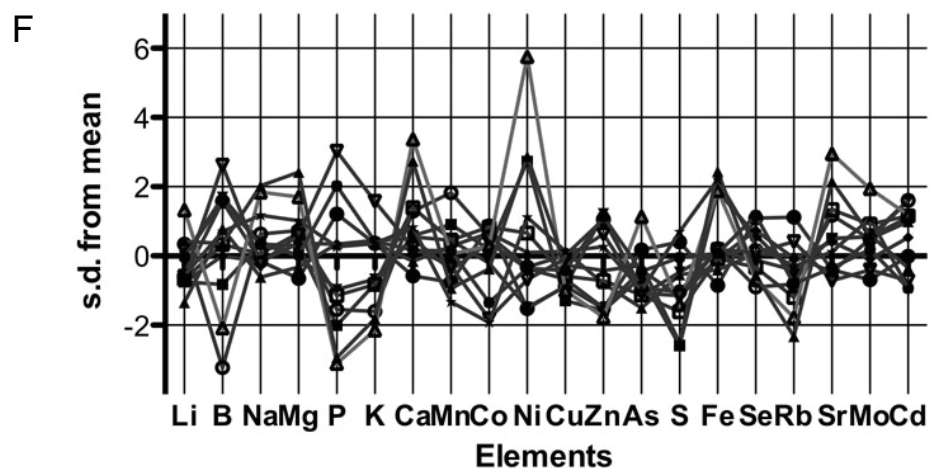
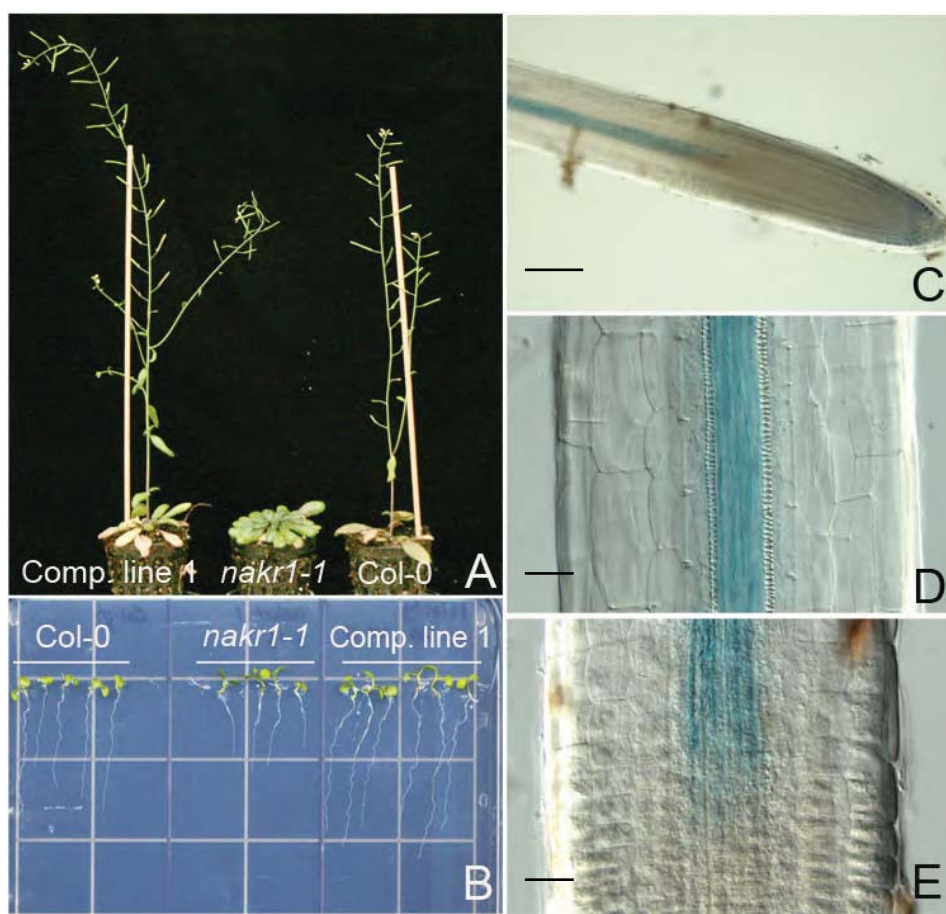
**Supplemental Figure 3. The identity of the purified MBP-NaKR1 fusion protein was confirmed by mass spectrometry analysis.**

Expression of *MBP-NaKR1* in *Escherichia coli* strain BL21-AI was induced by adding IPTG to the liquid culture. Cell lysate was incubated with amylose resin and MBP- tagged proteins eluted with 30 mM maltose and digested with trypsin. The underlined sequences were the unique peptides detected by LC-MS/MS analysis.



**Supplemental Figure 4. *nakr1-1* developmental and  $\text{Na}^+/\text{K}^+/\text{Rb}^+$  accumulation defects were complemented with *NaKR1pro::NaKR1-GFP*.**

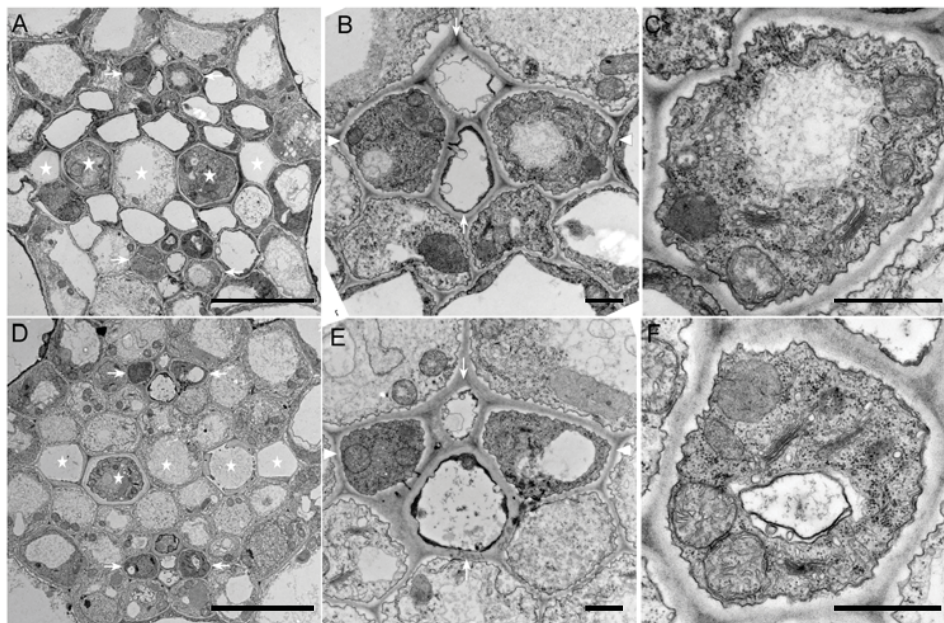
**(A)** From left to right, *nakr1-1* expressing *NaKR1pro::NaKR1-GFP*, *nakr1-1* and Col-0. Plants were grown in the same tray under long day growth conditions. **(B)** Seedlings grown vertically on ATS media. From left to right: *nakr1-1* transformed with *NaKR1pro::NaKR1-GFP*, *nakr1-1* and Col-0. **(C)** Differences in leaf elemental content between individual *NaKR1-GFP* complementation lines and the average of Col-0 plants were plotted as z-values (standard deviations). Wild type and complemented lines were grown in the same tray.



**Supplemental Figure 5. *nakr1-1* developmental and  $\text{Na}^+/\text{K}^+/\text{Rb}^+$  accumulation defects were complemented with *NaKR1pro::NaKR1-GUS*.**

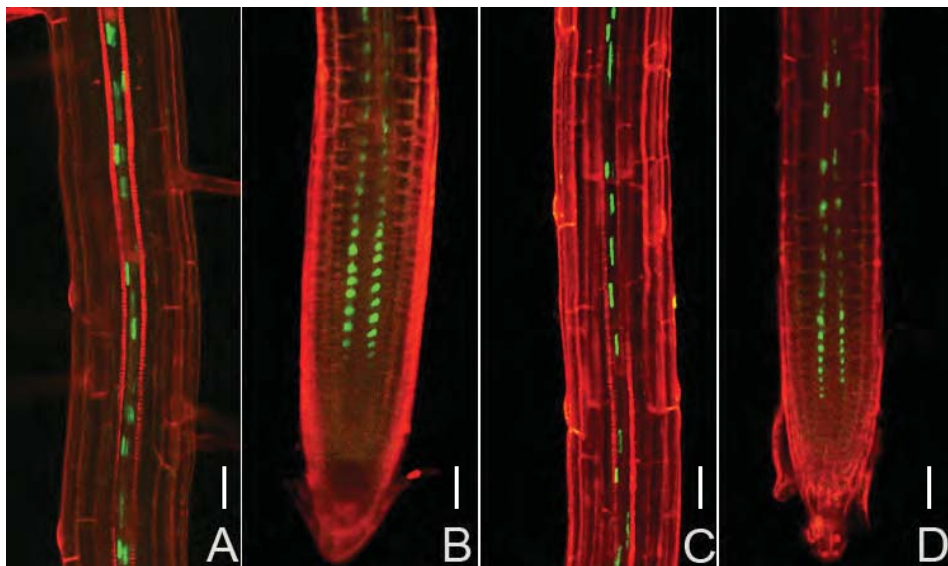
(A) From the left to the right, *nakr1-1* plants expressing *NaKR1pro::NaKR1-GUS*, *nakr1-1* and Col-0 plants grown in the same tray under long day growth conditions. (B) Seedlings grown vertically on ATS plate. From the left to the right: Col-0, *nakr1-1* mutants and *nakr1-1* plants complemented with *NaKR1pro::NaKR1-GUS*. (C-E) GUS staining of root tip (C), mature root (D) and the proximal region of root meristem (E). The scale bar is 100  $\mu\text{m}$  in (C) and 10  $\mu\text{m}$  in (D) and (E). (F) Differences in leaf elemental content between complementation lines and the mean of wild-type Col-0 plants were plotted as z-values (standard deviations).





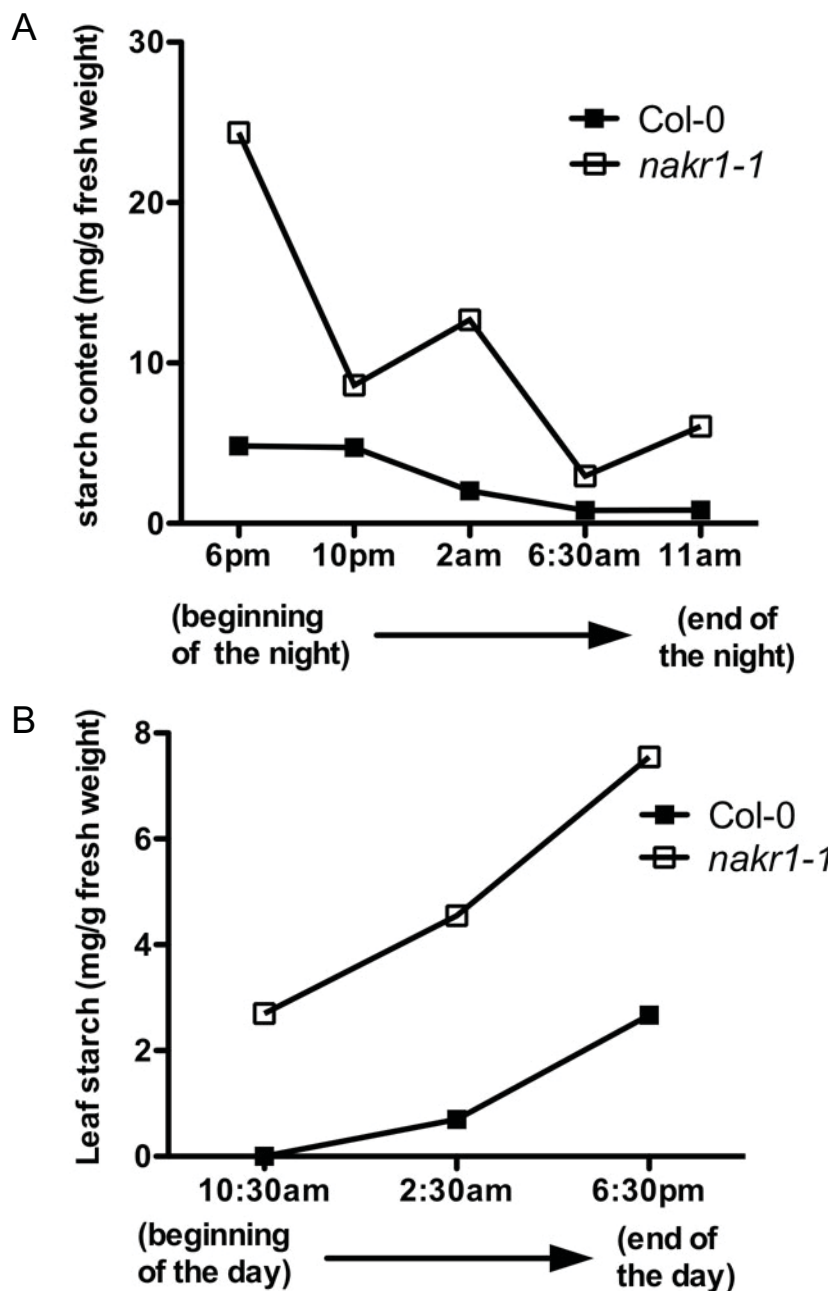
**Supplemental Figure 6. Cellular structures of SE/CC complexes were not affected by the *nakr1-1* mutation as shown by transmission electron microscopy.**

(A) and (D) vascular region of Col-0 (A) and *nakr1-1* (D) roots. Xylem cells and phloem companion cells were labeled with asterisks and arrows, respectively. Scale bars are 10  $\mu\text{m}$ . (B) and (E) SE/CC complex of WT (B) and *nakr1-1* mutant (E), with the arrowheads pointing to the companion cells and the arrows pointing the protophloem (top) and metaphloem cells (bottom) respectively. Scale bars are 1  $\mu\text{m}$ . (C) and (F), the cellular structure of companion cells of WT (C) and *nakr1-1* mutant (F). Scale bars are 1  $\mu\text{m}$ .



**Supplemental Figure 7. The differentiation of protophloem/ metaphloem cells and companion cells was not affected by the *nakr1-1* mutation.**

The expression of *APLpro::GFP-APL* in Col-0 and *nakr1-1* was analyzed by confocal microscopy. (A) and (C) Expression of *APLpro::GFP-APL* in mature roots of Col-0 (A) and *nakr1-1* mutant (C). The GFP fluorescence specifically labeled the companion cells. (B) and (D) Expression of *APLpro::GFP-APL* in root meristem of Col-0 (B) and *nakr1-1* mutant (D), with the GFP fluorescence specifically localized in developing metaphloem and protophloem cells. Scale bars are 40 μm.

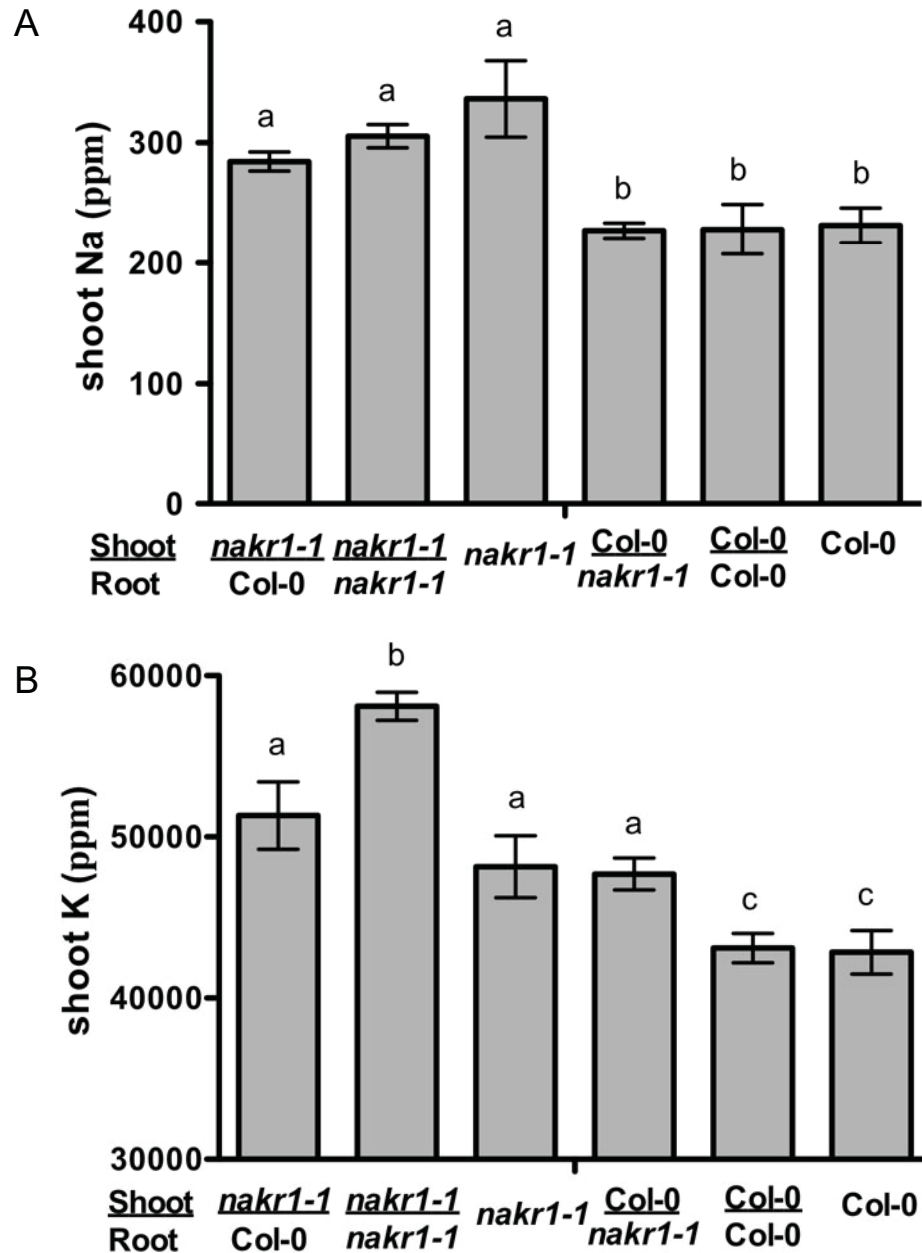


**Supplemental Figure 8. Quantification of starch in Col-0 and *nakr1-1* rosette leaves.**

**(A)** Leaf starch of *nakr1-1* and Col-0 at different time points during the night. The values at each time point are means from two independent experiments.

**(B)** Starch quantification during the day after 3 days of continuous darkness. A higher starch accumulation rate was detected in *nakr1-1* than in Col-0 leaves.





**Supplemental Figure 9. Reciprocal grafting of Col-0 and *nakr1-1* was used to test the contribution of NaKR1 in root and shoot on leaf Na<sup>+</sup> and K<sup>+</sup> content of plants grown on soil.**

**(A)** Leaf Na<sup>+</sup> content analyses of different grafting types (indicated in the figure) and control plants grown in LG<sub>3</sub> soil. **(B)** Leaf K<sup>+</sup> content of the same plants in **(A)**. Na<sup>+</sup> and K<sup>+</sup> were assayed by ICP-MS. Data are presented as mean ± SE, n= 9-12 plants for the reciprocal grafting types and n=5 for self-grafted plants and control plants.