

Supplemental Data

B-myb Positively Regulates Serine-Threonine Kinase Receptor-associated Protein (STRAP) Activity through Direct Interaction

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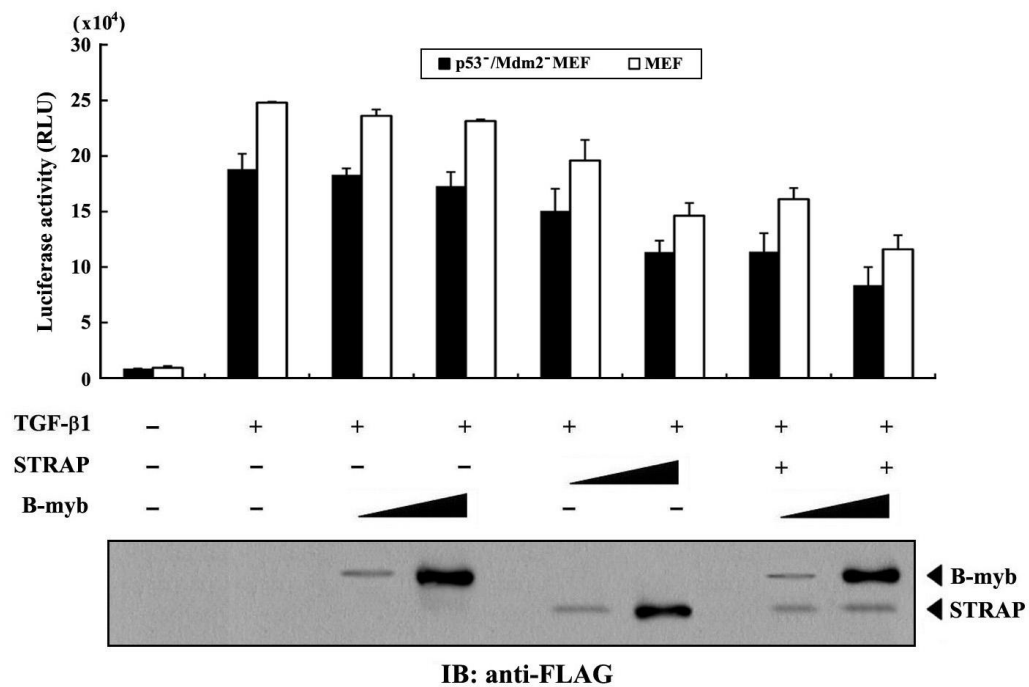
Materials and Methods

Generation of Inducible STRAP(KD) and Smad7(KD) Cell Lines—An inducible STRAP shRNA HEK293 cell line [STRAP(KD)] has been described previously (5). An inducible Smad7 shRNA HepG2 cell line [Smad7(KD)] was generated using the following oligonucleotides: forward, 5'-TCGAGGGCTCAATTCGGACAACAAGTTCAAGAGACTTGTTGTCCGAATTGAGCCTTTTTTA-3'; and reverse, 5'-AGCTTAAAAAAGGCTCAATTCGGACAACAAGTCTCTTGAACCTTGTTGTCCGAATTGAGCCC-3', as described previously (5). The Smad7 sequence was underlined.

RT-PCR—HepG2 or MCF7 cells were transiently transfected with control scrambled siRNA or B-myb-specific siRNA #1 as indicated, and then treated with TGF- β 1 (100 pM, 20 h) or 5FU (0.38 mM, 30 h). Total RNA was isolated from transfected cells using easy-BLUETM reagent according to the manufacturer's instructions (iNtRON Inc., Seongnam, Korea). 1-2 μ g total RNA was reverse transcribed with RevertAidTM reverse transcriptase (Fermentas Inc., Glen Burnie, Maryland). The reaction was conducted at 42°C for 1 h. 2-5 μ l cDNA mixture diluted 5-fold in DEPC-treated water was subjected to PCR. The reaction parameters were 5 min at 94°C, 1 cycle; 30 s at 94°C, 1 min at 50°C, 1.5 min at 72°C, 20-30 cycles; 5 min at 72°C, 1 cycle. The primers used for PCR were as follows: for p21, forward 5'-GTCCGTCAGAACCCATGC-3', reverse 5'-TTAGGGCTTCCTCTTGGAGA-3'; for PAI-1, forward 5'-CAACTTGCTTGGGAAAGGAG-3', reverse 5'-GTGGAGAGGCTCTTGGTCTG-3'; for Smad7, forward 5'-CCAACTGCAGACTGTCCAGA-3', reverse 5'-TGCTGCATAAACTCGTGGTC-3'; for CDK4, forward 5'-GGCCCTCAAGAGTGTGAGAG-3', reverse 5'-CCAACACTCCACATGTCCAC-3'; for Cyclin D1, forward 5'-CGTGGCCTCTAAGATGAAGG-3', reverse 5'-TCCTCCTCTTCCCTCCTC-3'; for p53, forward 5'-TCCCAAGCAATGGATGATTT-3', reverse 5'-ACACGCAAATTTCTTCCAC-3'; for Mdm2, forward 5'-AGATTCCAGCTTCGGAACAA-3', reverse 5'-GTGGCGTTTTCTTTGTCGTT-3'; for Bax, forward 5'-GCCGTGGACACAGACTCC-3', reverse 5'-AACCACCCTGGTCTTGGAT-3'; for β -actin, forward 5'-GGCATCTCACCCTGAAGTA-3', reverse 5'-CCATCTCTTGCTCGAAGTCC-3'; for B-myb, forward 5'-CCCTTCAAACCTTCCAGCC-3', reverse 5'-ATGAGAATGGGCTCGTGACA-3'.

FIG. S1. Effect of B-myb on the mRNA expression of TGF- β and p53 targets. HepG2 (A) or MCF7 (B) cells were transiently transfected with control scrambled siRNA (*Sc*) or B-myb-specific siRNA #1 (*B-myb*) as indicated, and then treated with TGF- β 1 (100 pM, 20 h) or 5FU (0.38 mM, 30 h) (*left panels*). HEK293 cells harboring stably integrated pcDNA4TM/TO/myc-HisA vector containing wild-type B-myb [*inducible B-myb(OE)*] or pSingle-tTS-shRNA vector containing B-myb-specific shRNA [*inducible B-myb(KD)*] treated with TGF- β 1 (100 pM, 20 h) or 5FU (0.38 mM, 30 h) were cultured in the presence or absence of 1 μ g/ml doxycycline (*Dox*) for 72 h (*right panels*). The mRNA expression of TGF- β (A) or p53 (B) targets was analyzed by RT-PCR as described in “Materials and Methods”. The amount of β -actin mRNA was used as a loading control.

A



B

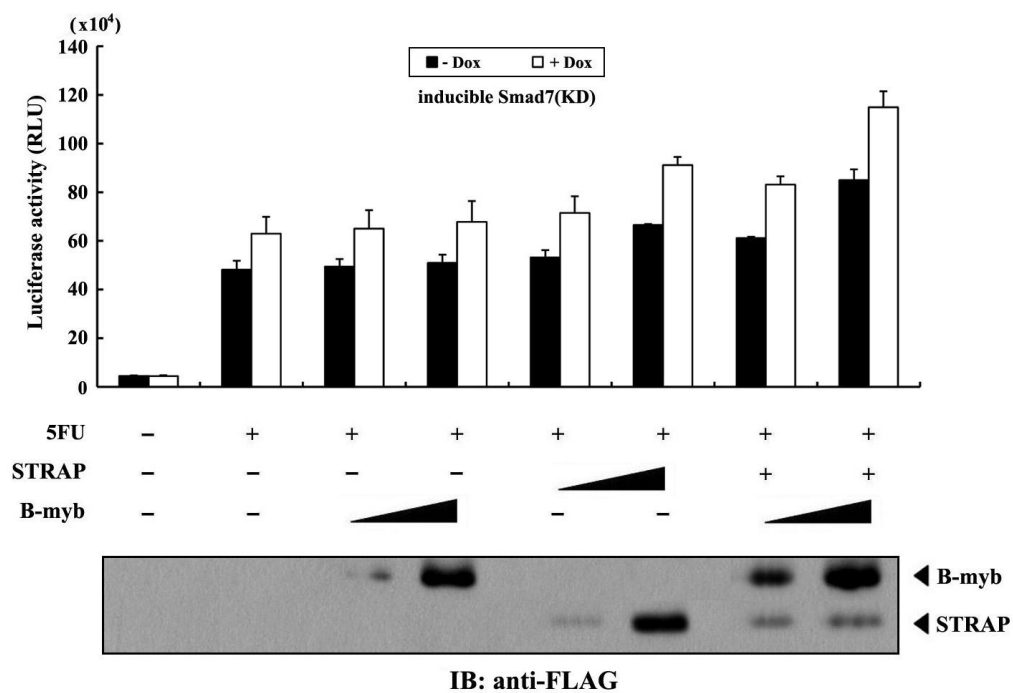


FIG. S2. B-myb-STRAP complex functions as an intermediate of cross-talk between the TGF- β and p53 signaling pathways. *A*, effect of B-myb on STRAP-mediated inhibition of TGF- β -induced transcription in p53-null system. Wild-type MEF or p53/Mdm2 double null MEF cells (6) were transiently transfected with 0.3 μ g p3TP-Lux reporter, 0.1 μ g expression plasmid for β -galactosidase as an internal control, and increasing amounts (0.3 and 0.6 μ g) of FLAG-tagged B-myb or STRAP, and subsequently stimulated by TGF- β 1 (100 pM, 20 h) and 5FU (0.38 mM, 30 h). *B*, effect of B-myb on STRAP-mediated stimulation of p53 transactivation in Smad7-knockdown system. HepG2 cells harboring stably integrated pSingle-tTS-shRNA vector containing Smad7-specific shRNA [*inducible Smad7(KD)*] were transfected with increasing amounts (0.3 and 0.6 μ g) of FLAG-tagged B-myb or STRAP as indicated, together with 0.2 μ g p53-Luc reporter plasmid and 0.1 μ g β -galactosidase internal control, in the presence of TGF- β 1 (100 pM, 20 h) and 5FU (0.38 mM, 30 h), and cultured in the presence or absence of 1 μ g/ml doxycycline (*Dox*) for 72 h. Luciferase activity was measured 48 h after transfection and normalized to β -galactosidase activity. The expression level of FLAG-tagged B-myb and STRAP was analyzed by anti-FLAG antibody immunoblot.

FIG. S3. STRAP is required for B-myb-mediated regulation of TGF- β and p53 signaling. *A*, effect of STRAP on the expression of TGF- β target genes. Parental HEK293 or HEK293 cells harboring stably integrated pSingle-tTS-shRNA vector containing STRAP-specific shRNA [*inducible STRAP(KD)*] cells were transfected with FLAG-B-myb as indicated, and then treated with 100 pM TGF- β 1 for 20 h and cultured in the presence or absence of 1 μ g/ml doxycycline (*Dox*) for 72 h. Cell lysates were subjected to immunoblot analysis using anti-PAI-1, anti-p21, anti-Smad7, anti-CDK4, anti-cyclin D1, anti-FLAG, anti-STRAP, and anti- β -actin antibodies. *B*, effect of STRAP on the expression of p53 target genes. Parental HEK293 or inducible STRAP(KD) cells were transfected with or without FLAG-B-myb, and then treated with 0.38 mM 5FU for 30 h and cultured in the presence or absence of 1 μ g/ml doxycycline (*Dox*) for 72 h. Cell lysates were subjected to immunoblot analysis using anti-p53, anti-p21, anti-Mdm2, anti-Bax, anti-FLAG, anti-STRAP, and anti- β -actin antibodies. The relative level of the expression of TGF- β and p53 target genes was quantified by densitometric analysis. The fold increase relative to untreated HEK293 cells in the absence of B-myb was calculated.

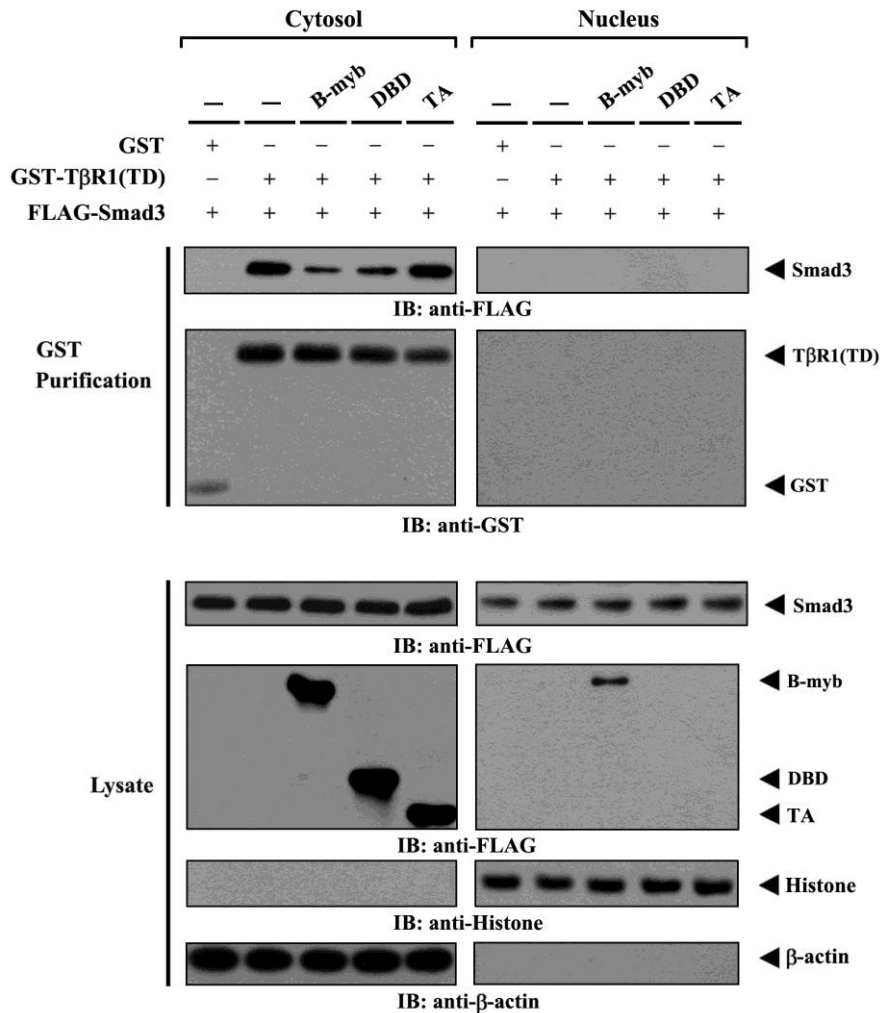


FIG. S4. B-myb decreases the activated type I TGF- β receptor-Smad3 complex formation in the cytoplasm. Cytoplasmic and nuclear fractions from the extracts of HEK293 cells transfected with the indicated combinations of plasmid vectors expressing GST-T β R1(TD), FLAG-Smad3, and FLAG-tagged wild-type B-myb or its deletion mutants DBD and TA were separated from each other, and the each fraction was subjected to precipitation with glutathione-Sepharose beads (*GST purification*). Complex formation between T β R1(TD) and Smad3 was analyzed by anti-FLAG antibody immunoblot (*top panels*). *Cytosol* and *Nucleus* indicate the cytoplasmic and nuclear fractions of cell extracts, respectively.