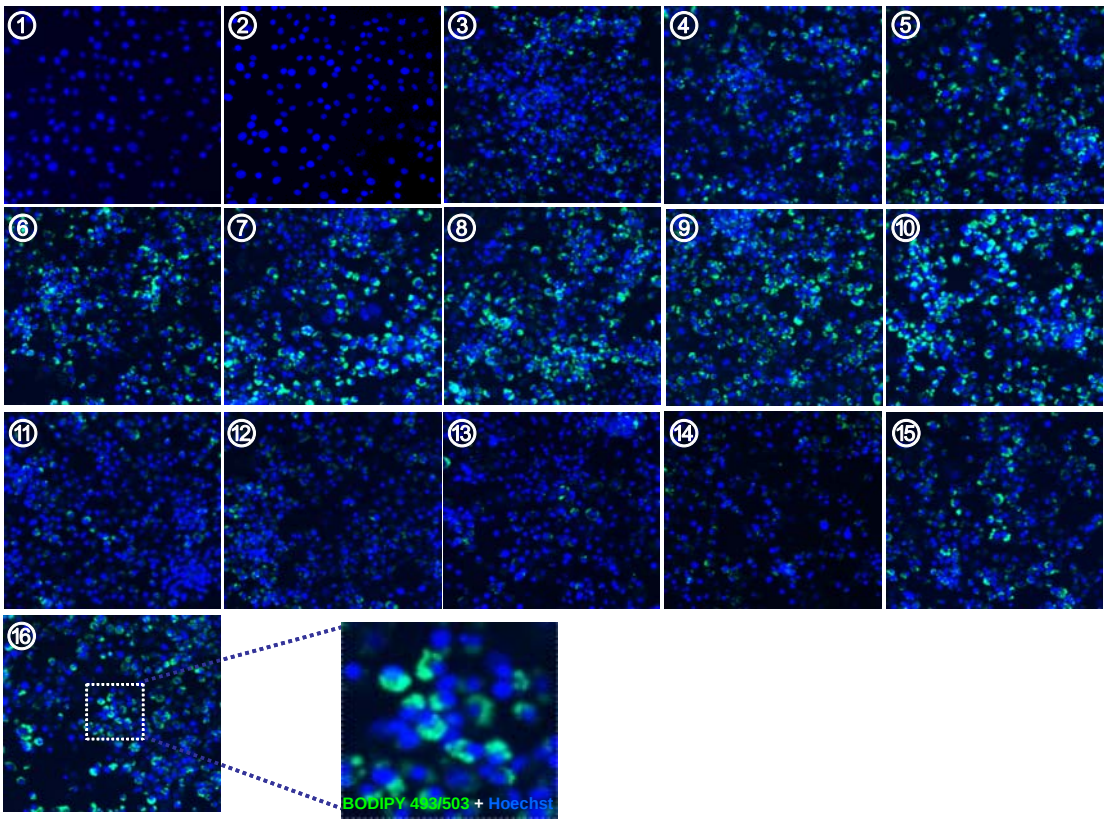
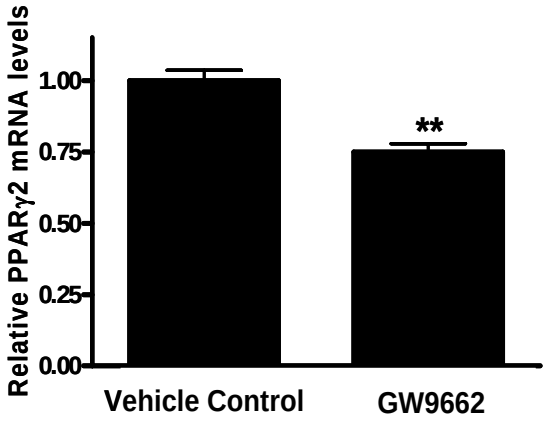
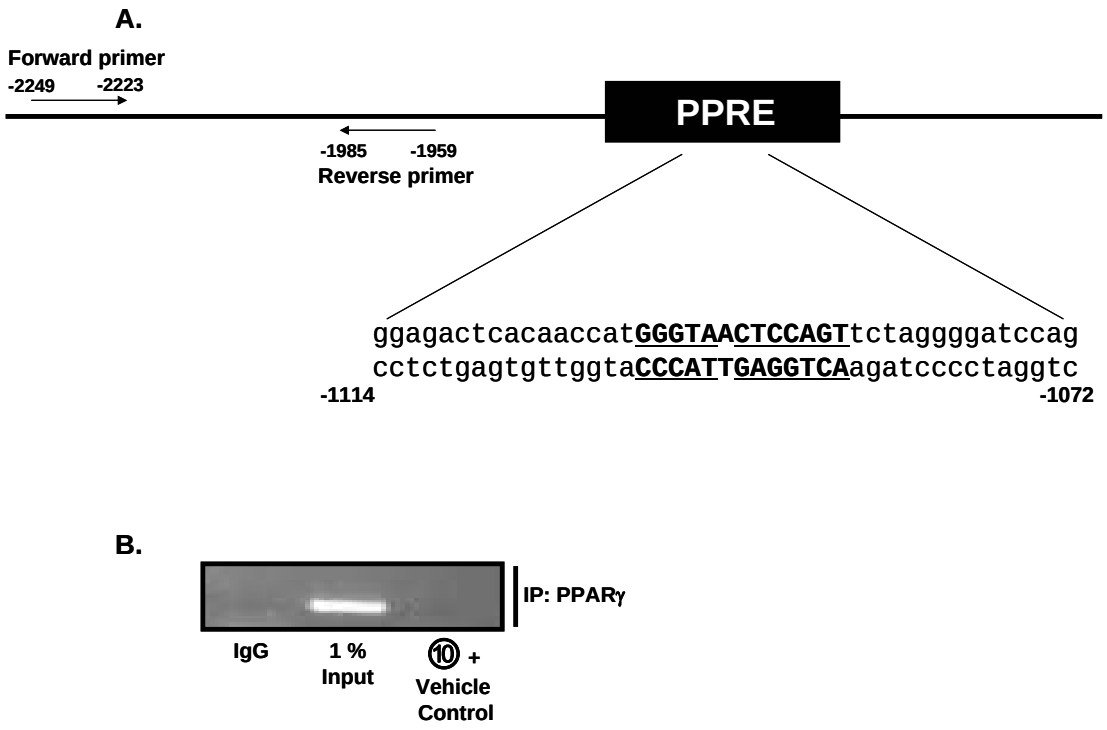


Supplementary Figure 1. Visualization of lipid in differentiated 3T3-L1 cells at day 7. 3T3-L1 preadipocytes were treated as described in Figure 1A for 7 days and lipid and nucleus were visualized with BODIPY 493/503 and Hoechst, respectively. Fluorescent signals were captured by a high throughput Cellomics^{TM}Arrayscan V^{TI} automated fluorescence imager.



Supplementary Figure 2. Effects of GW 9662 on PPARγ2 mRNA expression. 3T3-L1 adipocytes were treated for 7 days with conditional culture medium ⑩, described in Figure 1A. Following differentiation, adipocytes were treated with GW 9662 (20 μM) for 20 hr and real-time RT-PCR performed to quantify PPARγ2 mRNA levels; shown as the fold difference *vs* control normalized to 18S rRNA expression levels. All data represent three independent experiments, each carried out in duplicate and significance was tested using Student's *t* test, where ** represents $p<0.05$ *vs* Vehicle Control.



Supplementary Figure 3. PPARγ-ChIP using a negative PCR control. A. Identification of putative PPRE sequences in the promoters of the mouse GIPR gene and the location of primers used for the negative PCR control. B. PPARγ-ChIP. 3T3-L1 preadipocytes were treated for 7 days with conditional culture medium ⑩, described in Figure 1A. PPARγ were immunoprecipitated from intact chromatin isolated from 3T3-L1 cells (an aliquot of lane 11 in Figure 6B, ⑩ + Vehicle Control) using anti-PPARγ antibody. Precipitated DNA fragments were analysed by PCR using primers > 500 bps away from the PPRE site in the GIPR promoter. An isotype-matched IgG was used as negative control and 1% Input (PCR product of one-hundredth of the total isolated DNA used in the ChIP assay) as positive control, respectively.