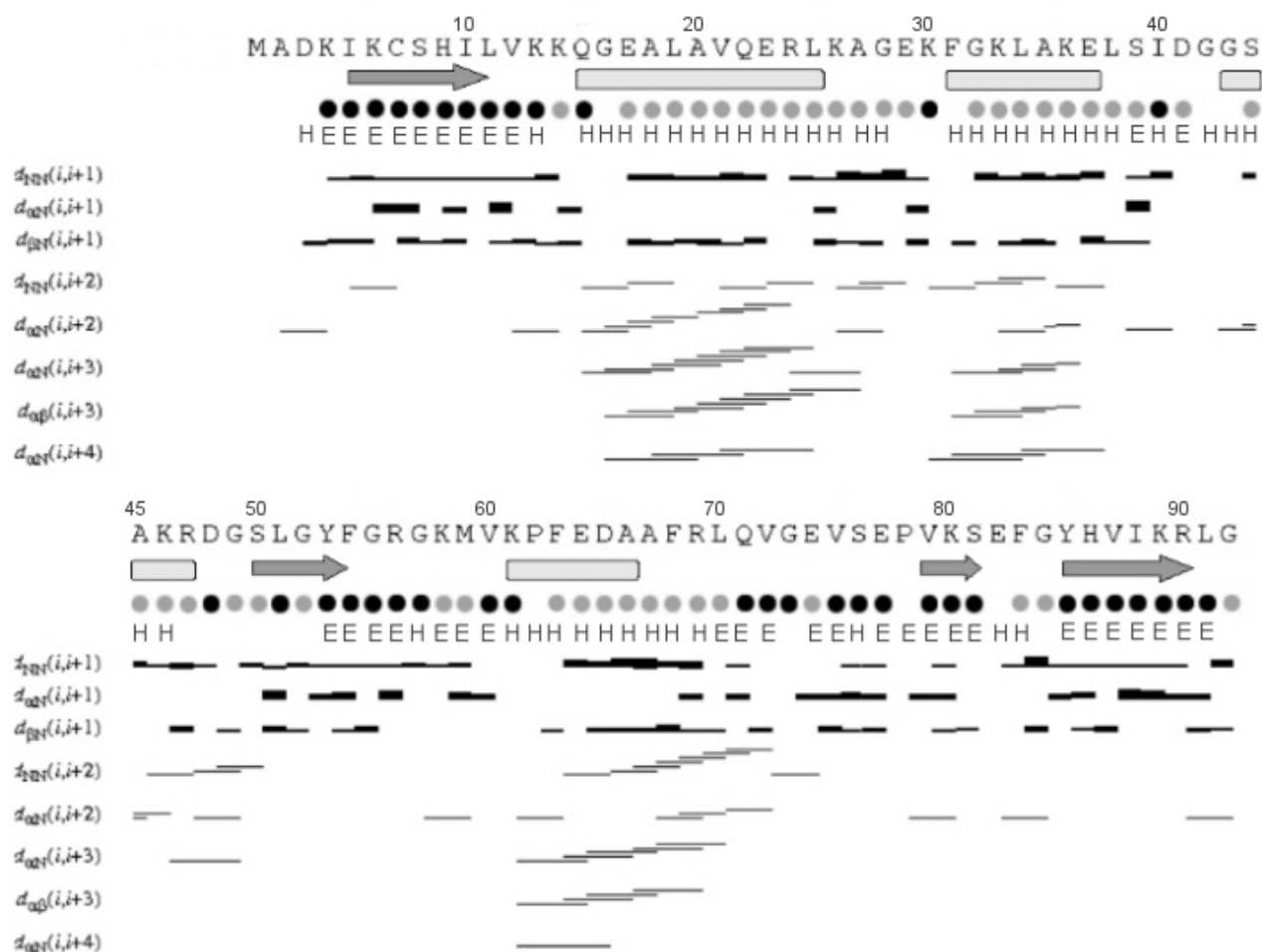


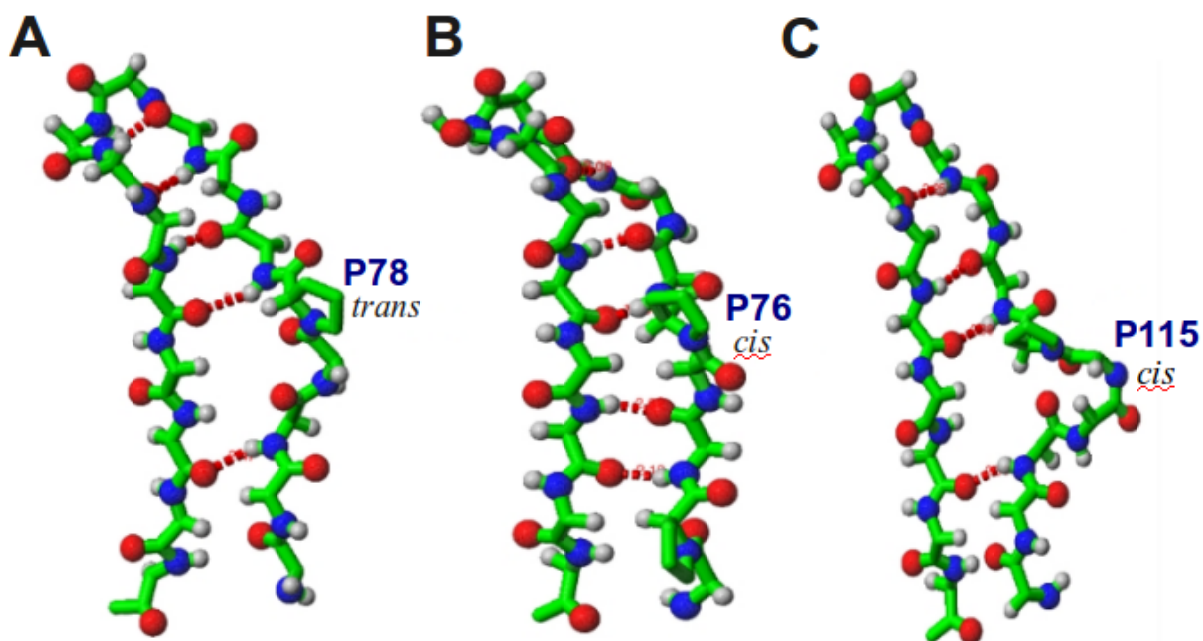
## Supplementary Materials

### Structure and dynamics of the first archaeal parvulin reveal a new functionally important loop in parvulin-type prolyl isomerases

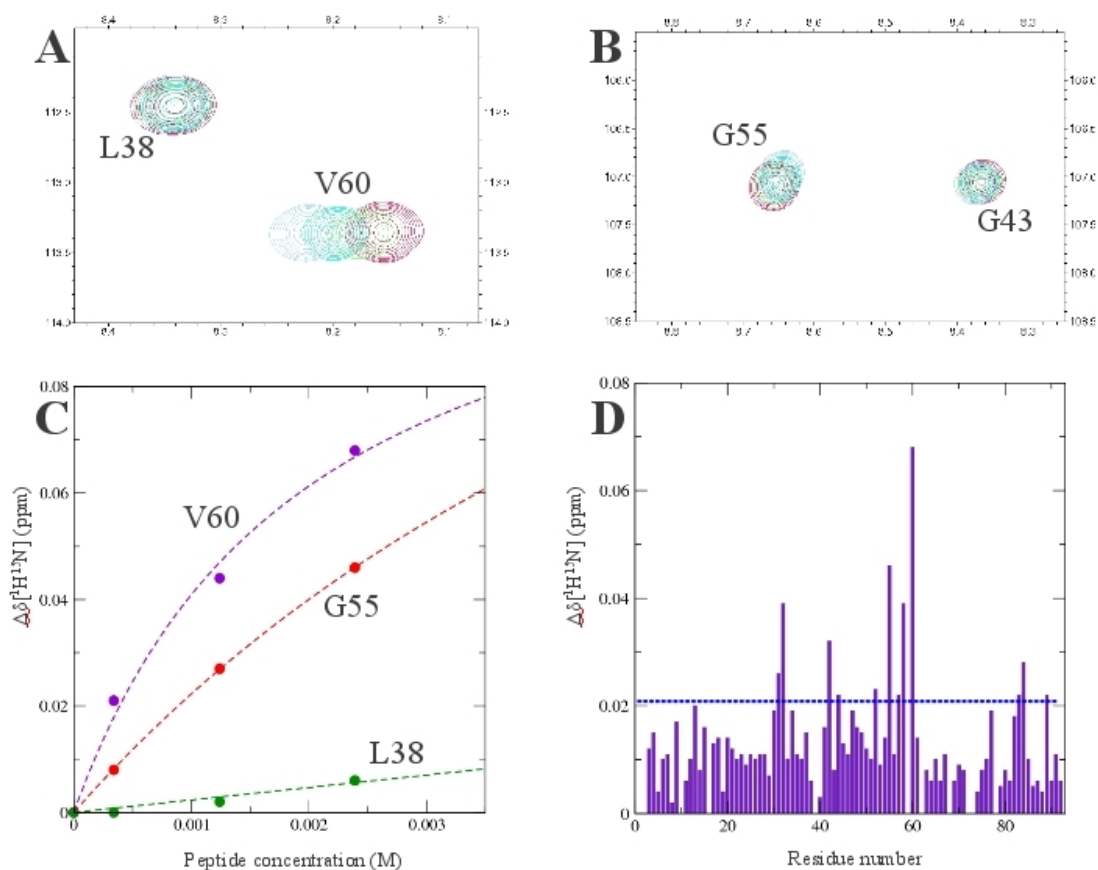
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Andrzej Ejchart, Peter Bayer, and Igor Zhukov



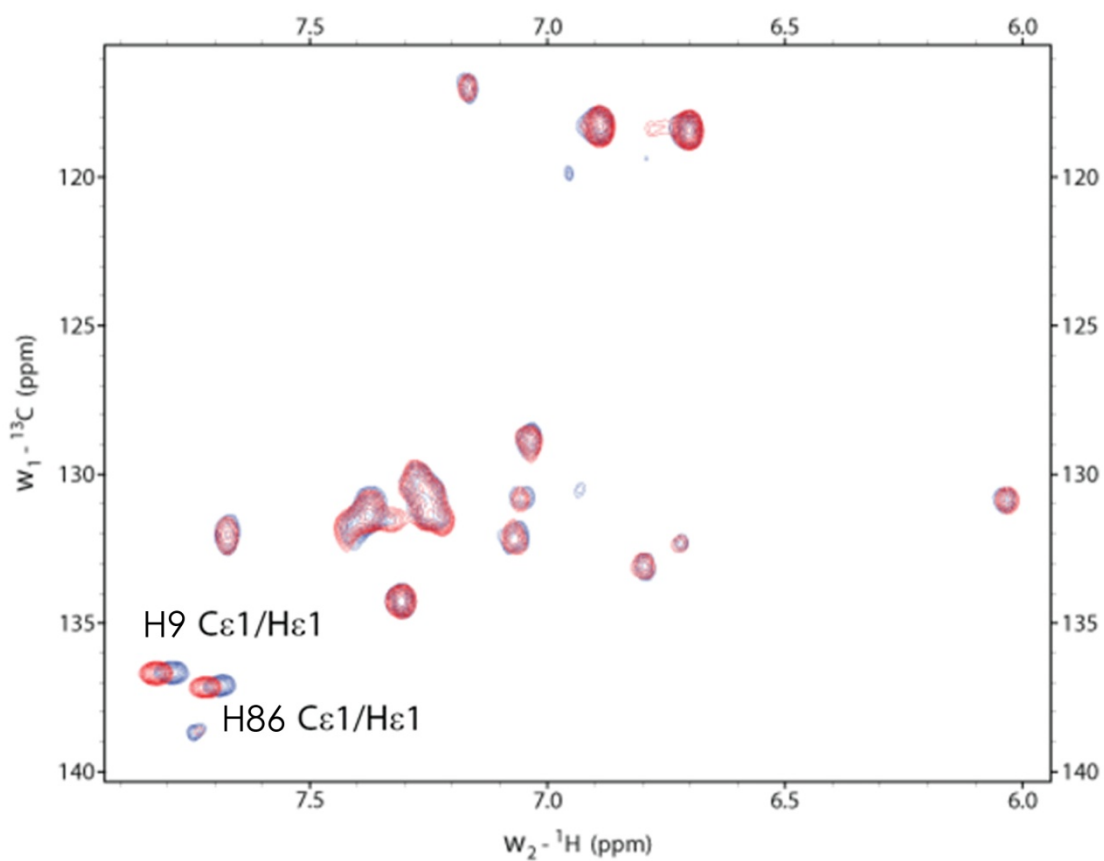
**Figure S10:** Short and medium range NOE connectivities observed in  $^{15}\text{N}$ -edited 3D NOESY-HSQC spectrum. Positions of secondary structure elements in PinA are denoted as black arrows ( $\beta$ -sheets) and gray bars ( $\alpha$ -helices). Values of  $^2J_{N(i)C\alpha(i-1)}$  couplings are shown as black (bigger than -7.2Hz) and gray (less than -7.2Hz) circles (Ding & Gronenborn, 2004) taking the sign of coupling into account (Kozminski et al., 2005). Please note that scalar  $^2J_{N(i)C\alpha(i-1)}$  couplings correlate with  $\psi$  backbone torsion angles of their preceding residues. Predictions originating from the TALOS+ software for 40 residues are indicated as 'H' (helix) or 'E' (extended), respectively.



**Figure S11:** Conformation of a conserved Xaa-Pro motif in different parvulins. Conformation and hydrogen bonding network between the third and fourth beta-strands are shown for (A) *C. symbiosum* PinA (PDB 2RQS), (B) *E. coli* Par10 (PDB 1JNS) and (C) *H. sapiens* Par14 (PDB 1EQ3). The conserved Xaa-proline at the beginning of the third beta-strand shows *trans* conformation in *C. symbiosum* PinA. However, the Gly76-Pro77 bond in *E. coli* Par10 as well as the Pro114-Pro115 bond in human Par14 exhibit *cis* conformation, which was additionally confirmed by chemical shift analysis (Shen & Bax, 2010).



**Figure S12:** Binding of the peptide HQSPWHH to PinA from *C. symbiosum*. **(A, B)** Part of  $^1\text{H}$ - $^{15}\text{N}$  HSQC spectra demonstrating chemical shifts after the addition of peptide. Cross peaks for **(A)** Val60 and **(B)** Gly55 are shown. Protein/peptide ratios were 1:0, 1:1, 1:4 and 1:8 shown in red, orange, yellow and green, respectively. **(C)** Best fit of chemical shift perturbations for Leu38, Gly55 and Val60. **(D)** Chemical shift perturbations in PinA upon binding to the peptide HQSPWHH at 283 K. The threshold of 0.022 ppm is indicated by a blue line.



**Figure S13:** Titration of the PinA protein with the peptides selected by phage display was also monitored with  ${}^1\text{H}$ - ${}^{13}\text{C}$  correlation spectra. Only for the best binding peptide HQSPWHH chemical shift changes for the Cε1/Hε1 correlation of the two conserved histidines, His9 and His86, could be observed.

**Supplementary table 1:** Peptide sequences found after screening a phage displayed 7-mer peptide library using PinA as bait

7-mer motifs	Remarks	
HKRPRNN (5x)*	Besides the HQSPWHH peptide, HKRPRNN was also tested for HSQC titrations. This peptide however only gave minor chemical shift perturbations and was not investigated further.	
HQSPWHH (3x)*		
APSPMIW		
WDPSQMR		
SLHSRPN		
TIEQHPP		
VYLTGPS		* These peptides have been identified multiple times.
LDRANVF		
NQLTTLN		** According to the manufacturer of the 7-mer library (NEB), this is a target-unrelated peptide that might obscure whole panning procedures. It occurred only once during our screening, proving the selection to be effective.
SHTIRML		
YVHQQRH		
TMCYCT		
GLCCSRL		
HAIYPRH**		