

## SUPPLEMENTAL DATA

### **STRUCTURAL BASIS OF MOLECULAR RECOGNITION BY LEISHMANIA SMALL HYDROPHILIC ENDOPLASMIC RETICULUM-ASSOCIATED PROTEIN, SHERP, AT MEMBRANE SURFACES**

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FIGURE S1 2D  $^1\text{H}$ - $^{15}\text{N}$  HSQC NMR spectrum of *L. major* SHERP at 4 °C in the absence of SDS or phospholipids. The poor amide  $^1\text{H}$  dispersion is indicative of a lack of tertiary structure.

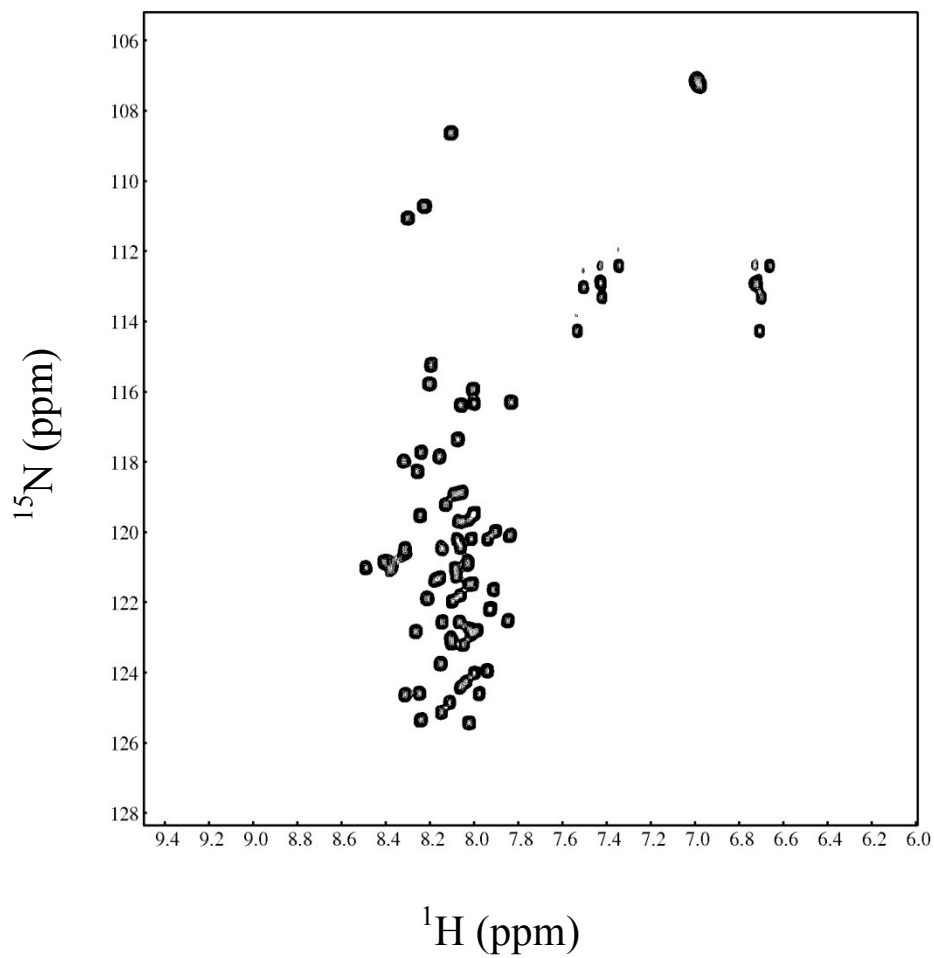
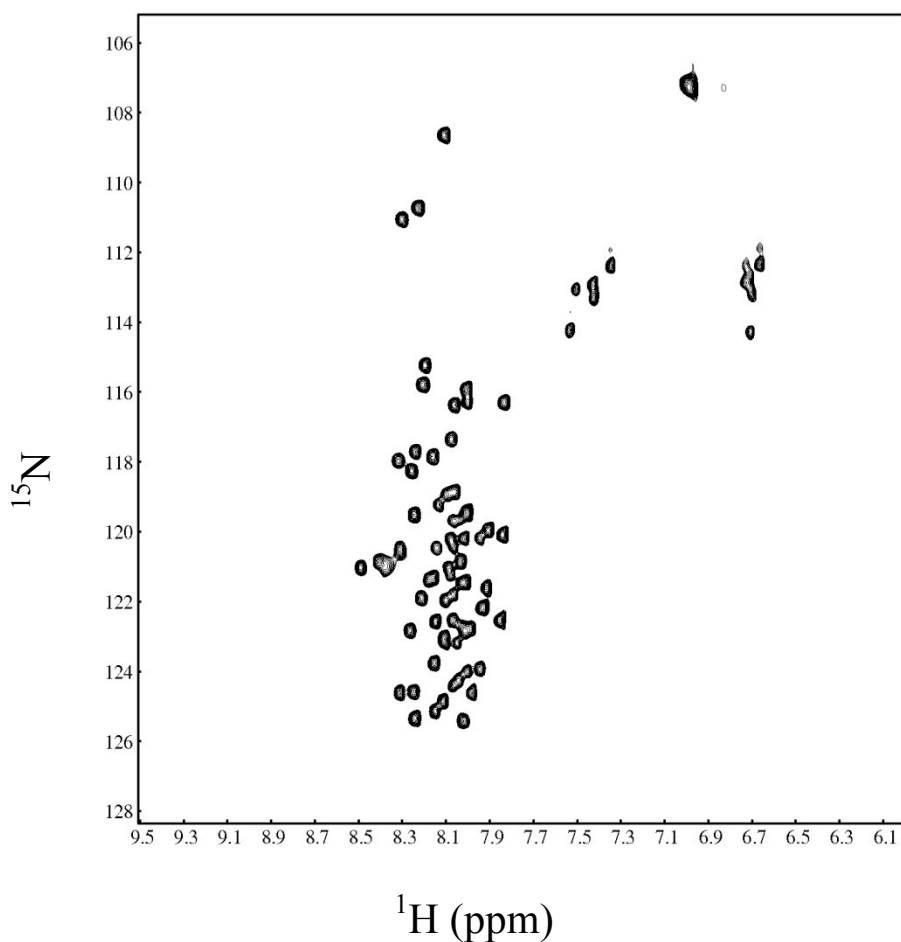


FIGURE S2  $^1\text{H}$ - $^{15}\text{N}$  heteronuclear NOE spectrum of *L. major* SHERP at 4 °C in the absence of SDS or phospholipids. The spectrum shown is a summation of the reference and NOE spectrum corresponding to (0.8 times the reference – NOE spectrum). Residual signals thus correspond to those with a significant  $^1\text{H}$ - $^{15}\text{N}$  NOE contribution arising from extensive internal motion on the ps timescale, which for the free SHERP is observed in essentially all residues (for comparison with Figure S2).



**FIGURE S3 SDS-PAGE avidin-column fractions analyzed to identify SHERP-SBED complexes containing a biotinylated label.** Lanes labeled correspond to the flow-through (FT), wash (W) and elution (E) fractions. Bands labeled with numbers correspond to sample numbers shown in Table S1 below.

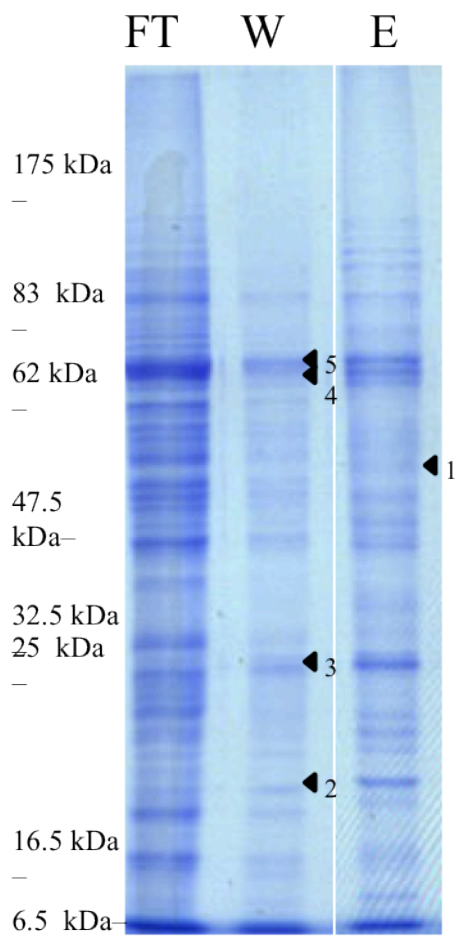


FIGURE S4 Amino acid sequence alignment between the *L. major* and *T. thermophilus* V-ATPase subunit B. A ClustalW2 alignment is shown. Identical residues (blue), conserved substitutions (green) and semi-conserved substitutions (yellow) are colored according to ClustalW2 conventions (see <http://www.ebi.ac.uk/Tools/clustalw2/index.html> and Larkin M.A., et al. (2007) *Bioinformatics* **23**, 2947-2948).

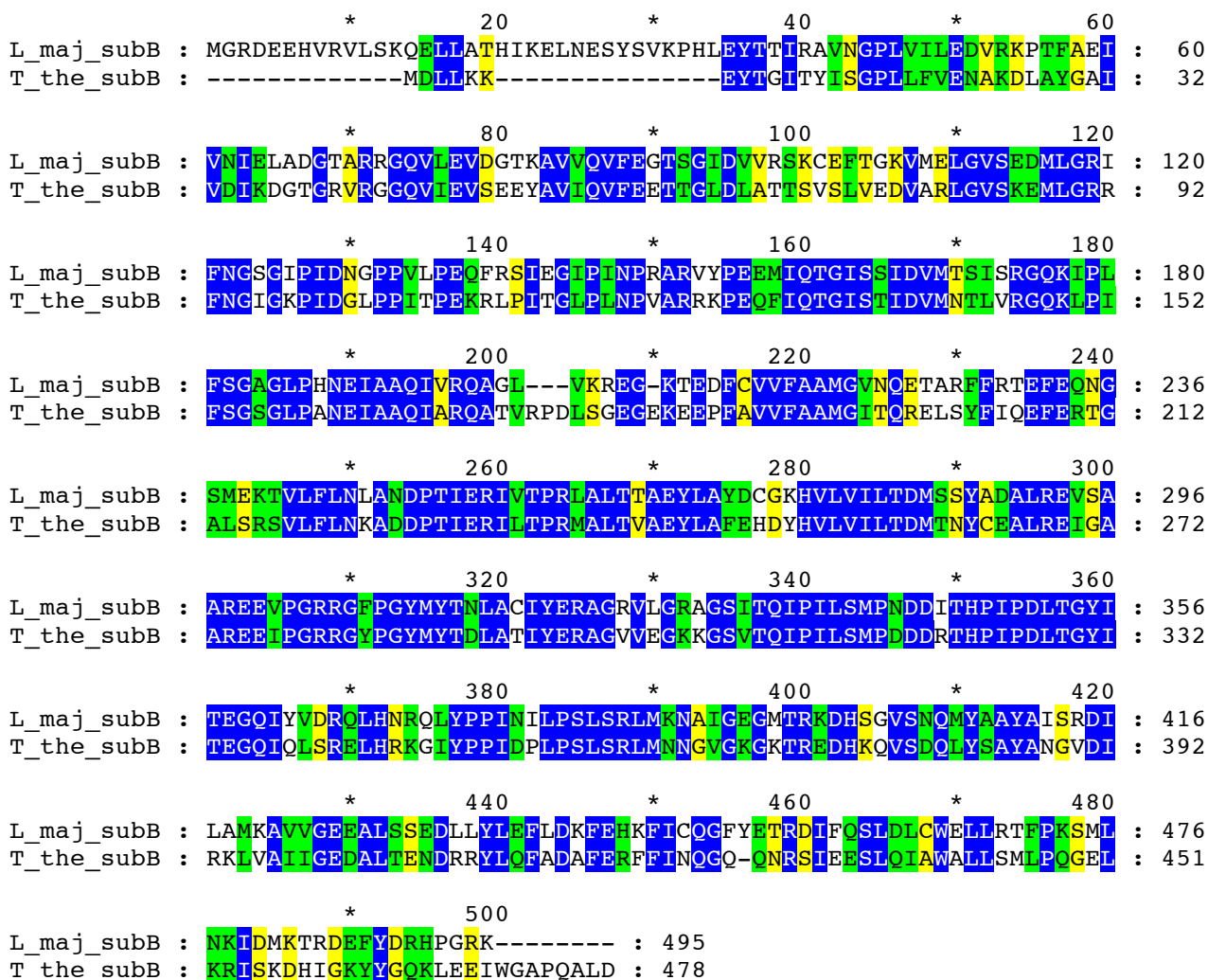
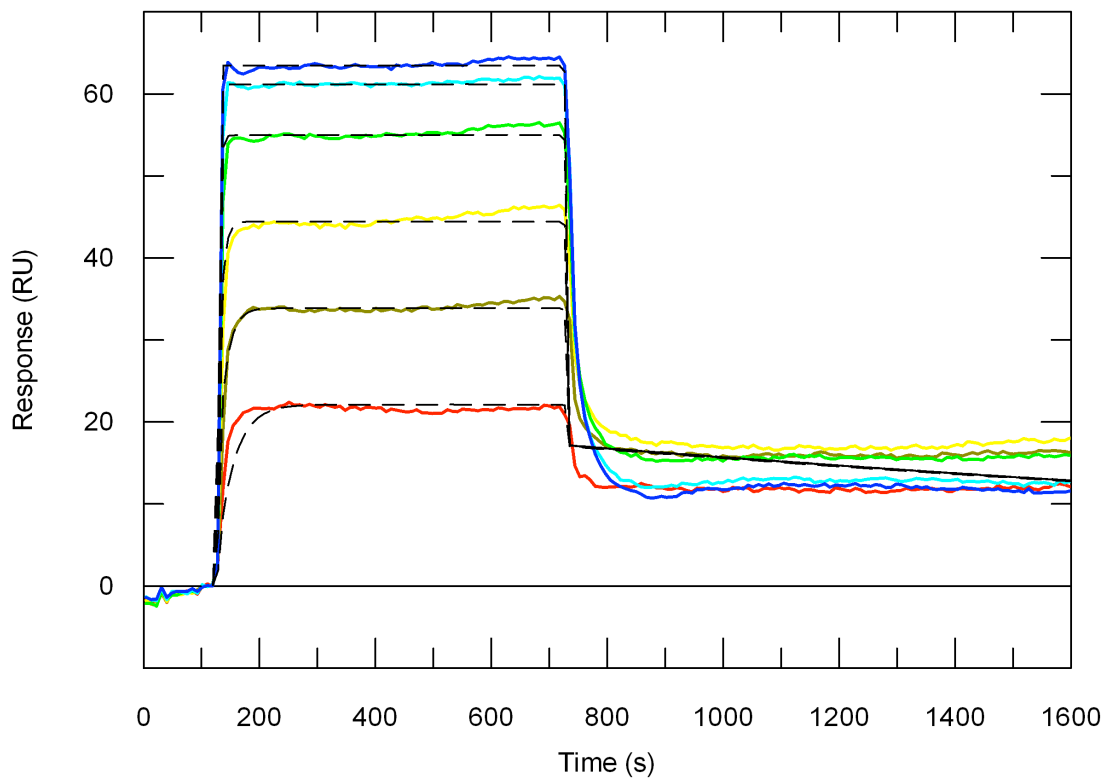


FIGURE S5 **Recombinant *L. major* SHERP binding to *T. thermophilus* V-ATPase demonstrated by surface plasmon resonance.** *T. thermophilus* V-ATPase at 0.2 mg/ml in 10 mM sodium acetate buffer, pH 5.0 was immobilized to a CM5 chip by amine coupling. Injection of *L. major* SHERP at a flow rate of 5 ul/min is shown for concentrations ranging from 0.5  $\mu$ M (red) to 15  $\mu$ M (dark blue) are shown along with the associated fits (dashed lines) to the raw data based upon a 1:1 Langmuir binding model. Binding was measured as response units over time. See main article for more details on the experimental data and analysis.



**TABLE S1**

**Summary of mass spectrometric analyses of *Leishmania major* proteins isolated from cross-linking studies with recombinant *L. major* SHERP**

<b>Sample No.</b>	<b>Molecular Mass (kDa)</b>	<b>MOWSE Score<sup>a</sup></b>	<b>GeneDB Identifier</b>	<b>Description</b>
<b>1</b>	<b>55</b>	<b>301</b>	LmjF28.2430	vacuolar ATP synthase subunit B, putative
<b>2</b>	<b>25</b>	<b>121</b>	LmjF23.0040	peroxidoxin tryparedoxin peroxidase
<b>3</b>	<b>27</b>	<b>381</b>	LmjF36.5010	40S ribosomal protein SA, putative
<b>4</b>	<b>68</b>	<b>111</b>	LmjF29.1760	paraflagellar rod protein 1D, putative
<b>5</b>	<b>71</b>	<b>574</b>	LmjF28.2780	heat-shock protein hsp70, putative

<sup>a</sup> Pappin, D. J., Hojrup, P., and Bleasby, A. J. (1993) *Curr. Bio.* **3**, 327–332