

Supplementary material

Figure S1: Expression of GFP-lamin B1 in HEK 293 cells is not masking epitopes of a monoclonal Nup88 antibody against residues 314-425 (first row). In the presence of GFP-lamin A epitope masking is not occurring for epitopes recognized by the monoclonal antibody mAb414 (second row), while expression of GFP-lamin A is masking monoclonal antibodies directed against residues 314-425 of Nup88 to different extents (third and fourth row).

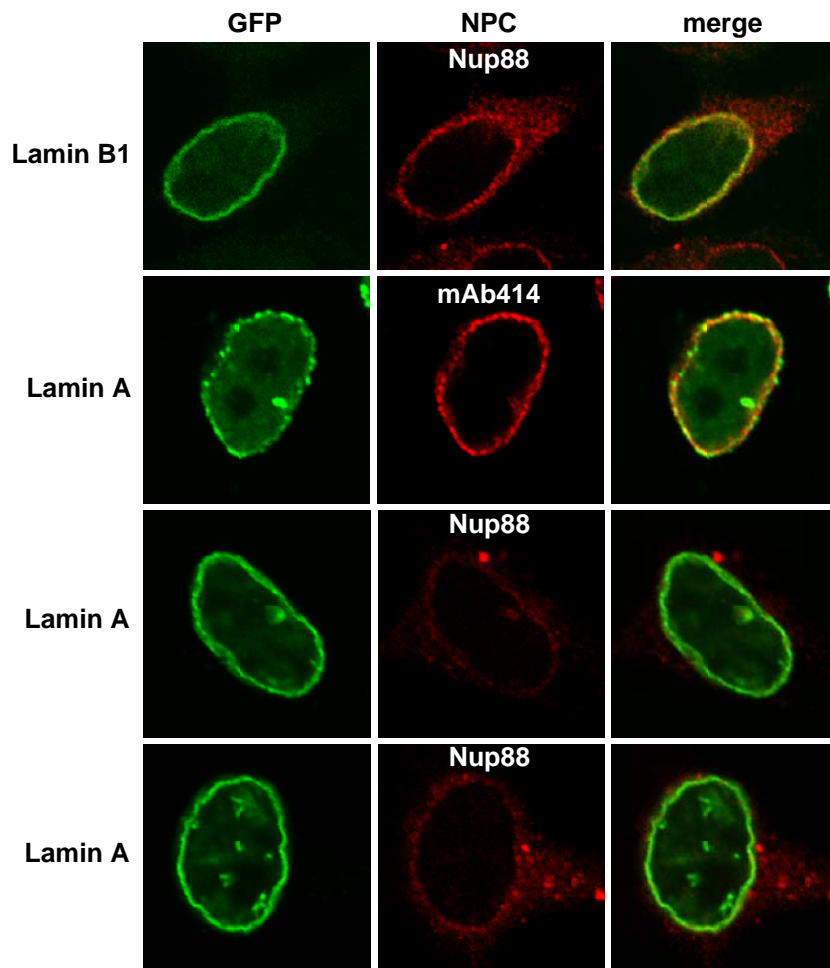
Figures S2: Bacterially expressed GST-Nup88 and GST were bound to glutathione sepharose beads and incubated with *in vitro* synthesized ³⁵S-labeled full-length lamin A, lamin A tail domain (residues 243 to 664), and the Ig-fold of lamin A (residues 463 to 545), as well as the two lamin A Ig-fold mutants R453W and R482W, respectively. Unbound and bound fractions were analyzed by SDS-PAGE and autoradiography. Whereas the lamin A tail and lamin A Ig-fold were able to bind GST-Nup88 but not GST, the lamin A Ig(R453W) and lamin A Ig(R482W) did not bind to GST-Nup88 or GST.

Figure S3: (A) Nup88-lamin A complex is not associated with transport. Bacterially expressed GST-Nup88 and GST were bound to glutathione sepharose beads and incubated with *in vitro* synthesized ³⁵S-labeled full-length lamin A. The reticulocyte lysate used for *in vitro* expression and ³⁵S-labeling of lamin A and the bound fractions to GST-Nup88 and GST were then analyzed by SDS-PAGE and immunoblotting with antibodies against CRM1, importin α and importin β . Neither CRM1, importin α nor importin β were found to be associated with the Nup88-lamin A complex. (B) HeLa cells were transfected with cyclophilin or lamin A siRNA respectively, or mock

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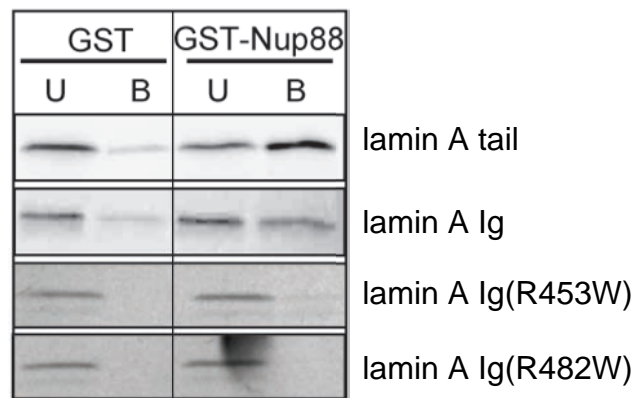
transfected. Protein levels were analyzed by immunoblot 48 hours after transfection with antibodies against lamin A, Nup88 or β -tubulin as loading control. (C) Protein levels of Nup88 were quantified using MultiGauge (FujiFilm) and normalized with β -tubulin protein levels. Nup88 protein levels of mock-transfected cells are set to 100%.

Figure S1



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Figure S2



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Figure S3

