

Supporting Information

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SI Text

S1: Oligonucleotide Sequences Used for Cloning. PstI_1-2225_SmaI: Pc-BioI (CTGCAGTCGGCCACTCCTCACTACC)

Pc-BioCtermII (CCCGGGAGCTACTGGCGACGAATCGC); SmaI_2226-3712_PstI: Pc-BioCtermIII (CCCGGGTGAGCTTCGACCCAAACAATG)

Pc-BioIV (CTGCAGTGGCGCCTTATCATC).

Linker sequence 1 (GAGAATTTGTATTTTCAGGGTATG-GCCTCCTCGTGCAGATCCTGGATTCCCAGAAGA-TGGAGTGGCGCTCGAACGCGCGGCTCCGACTACA-AGGACGACGATGACAAGGCTTGA)

Linker sequence 2 (TCAAGCCTTGTATCGTCGTCCTTG-TAGTCGGAGCCGCGGCGTTTCGAGCGCCACTCCATCT-TCTGGGAATCCAGGATCTGGCGCAGCGAGGAGGCCA-TACCTGAAAATACAAATTCTC)

NotI-Flag-BirA (GCGGCCGCATGGCTGACTACAAGGA-CGACGATGACAAGATGAAGGATAACACCGTGC)

S2: Mass Spectrometric Analysis of Pc-Bio Pull-downs. 2 μ l of the dissolved peptide mixture was analyzed by LC-MS/MS. The setup of the μ RPLC-MS system was as described previously (1). The hybrid LTQ-FT-ICR mass spectrometer was interfaced to a nanoelectrospray ion source (both Thermo Electron) coupled online to a Tempo 1D-plus nanoLC (Applied Biosystems/MDS Sciex). Peptides were separated on an RP-LC column (75 μ m \times 15 cm) packed in-house with C18 resin (Magic C18 AQ 3 μ m; Michrom BioResources) using a linear gradient from 98% solvent A and 2% solvent B (98% acetonitrile, 2% water, 0.15% formic acid) to 30% solvent B over 40 minutes at a flow rate of 0.3 μ l/min. Each survey scan acquired in the ICR-cell at 100,000 FWHM was followed by MS/MS scans of the three most intense precursor ions in the linear ion trap with enabled dynamic exclusion for 20 seconds. Charge state screening was employed to select for ions with at least two charges and to reject ions with undetermined charge state. The normalized collision energy was set to 32%, and one microscan was acquired for each spectrum. Before database searching, the acquired raw files (with Xcalibur 2.0 SR1) were converted to the mzXML data format employing the ReadAW 3.5.1 software (2). MS/MS-spectra were exported as dta files without further processing using the mzXML2other program (2). The MS/MS-spectra were searched using the X!Tandem search tool (3) (Version: X! TANDEM 2 (2007.07.01.2)) against the Flybase protein database (release version 5.7, April 2008) as well as known contaminants such as porcine trypsin and human keratins (Non-Redundant Protein Database, National

Cancer Institute Advanced Biomedical Computing Center, 2004, <ftp://ftp.ncifcrf.gov/pub/nonredun/>; total number of protein sequences searched: 21129). The search was performed with full-tryptic cleavage specificity, mass tolerance of 25 ppm, methionine oxidation as variable modification, and cysteine carbamidomethylation as fixed modification. The database search results were further processed and analyzed using the Trans-Proteomics Pipeline 3.5 (TPP) (4). First, the identified peptides were validated using the PeptideProphet software (5) and then assigned for protein identification using the ProteinProphet program (6). Both software tools employed allow filtering and evaluation of large proteomic datasets with assessment of predictable sensitivity and false positive identification error rates. In this study, only peptides with a PeptideProphet score of ≥ 0.55 were considered. Moreover, only proteins identified with a minimum of four peptides were accepted as correct calls for further analysis and all of the 22 identified proteins had a ProteinProphet score of ≥ 0.9 . This represents a false error rate of less than 5% on the peptide and less than 1% on the protein level as determined by Peptide- and ProteinProphet, respectively. Protein coverage and number of unique and total peptides found for specifically enriched proteins are displayed in Table S3 together with the peptide sequences identified.

S3: Analytical Gel Filtration Experiments. Analytical gel filtration experiments were performed on a Superose 6 HR column (23.56 ml, 1 cm \times 30 cm, GE Healthcare) equilibrated with three column volumes of DB-3. After that, 200 μ l of nuclear extract (2.5 mg protein) was loaded onto the column, followed by two column volumes of DB-3 for elution. Fractionation was carried out using a linear flow rate of 0.5 mL/min and the obtained samples were acetone precipitated and dissolved in sample buffer for SDS/PAGE and Western blotting with specific antibodies. The column was calibrated with a selection of molecular weight standards for gel filtration (GE Healthcare).

S4: Eye Pigment Measurements. Freshly hatched flies were aged for three days at 29°C until stored at -20°C. For quantitative measurements, heads were isolated (30 heads for each sample) and ground in 1.5 mL tubes with a micropestle in 80 μ l EPE buffer (30% Ethanol-HCL pH 2). Extraction was allowed to proceed at room temperature for 1 h and followed by sedimentation at 13,000 rpm. Supernatants were collected and the OD at 480 nm was measured with a nanodrop spectrophotometer.

- Schmidt A, et al. (2008) An integrated, directed mass spectrometric approach for in-depth characterization of complex peptide mixtures. *Mol Cell Proteomics* 7:2138–2150.
- Pedrioli PG, et al. (2004) A common open representation of mass spectrometry data and its application to proteomics research. *Nat Biotechnol* 22:1459–1466.
- Craig R, Beavis RC (2004) TANDEM: Matching proteins with tandem mass spectra. *Bioinformatics* 20:1466–1467.
- Keller A, Eng J, Zhang N, Li XJ, Aebersold R (2005) A uniform proteomics MS/MS analysis platform utilizing open XML file formats. *Mol Syst Biol* 1:2005 0017.

- Keller A, Nesvizhskii AI, Kolker E, Aebersold R (2002) Empirical statistical model to estimate the accuracy of peptide identifications made by MS/MS and database search. *Anal Chem* 74:5383–5392.
- Nesvizhskii AI, Keller A, Kolker E, Aebersold R (2003) A statistical model for identifying proteins by tandem mass spectrometry. *Anal Chem* 75:4646–4658.
- Sato T, Denell RE (1985) Homoeosis in *Drosophila*: Anterior and posterior transformations of Polycomb lethal embryos. *Dev Biol* 110:53–64.
- Messmer S, Franke A, Paro R (1992) Analysis of the functional role of the Polycomb chromo domain in *Drosophila melanogaster*. *Gene Dev* 6:1241–1254.

Rescue of embryonic lethality of Pc transheterozygous mutants by Pc-Bio

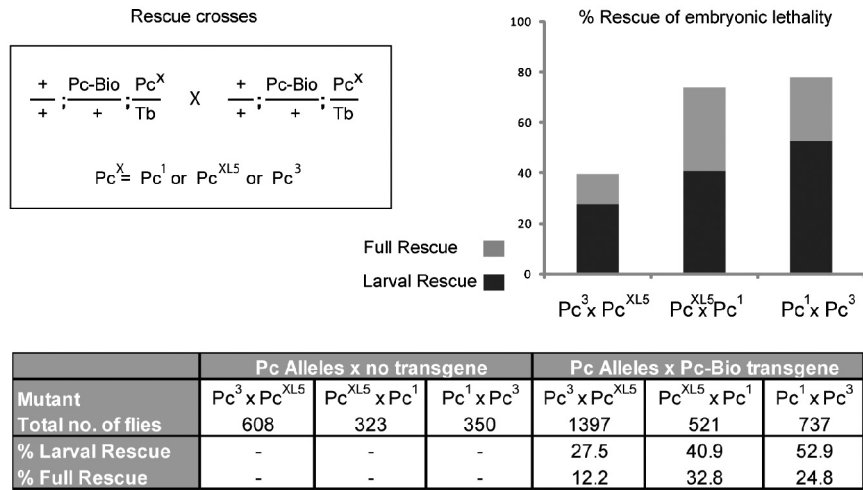


Fig. S1. Rescue of embryonic lethality of transheterozygous Pc mutants by Pc-Bio. Animals heterozygous for the Pc-Bio transgene were combined with Pc¹, Pc³ or Pc^{XL5} alleles (7, 8). Transheterozygous mutants missing an intact copy of endogenous Pc protein were identified by selection of wild-type larvae lacking the tubby (Tb) balancer chromosome. Larval rescue (black bars) was scored by the number of larvae that were able to pupariate and full rescue (gray bars) was attested for hatching flies.

Pc-Bio is specifically released by TEV cleavage

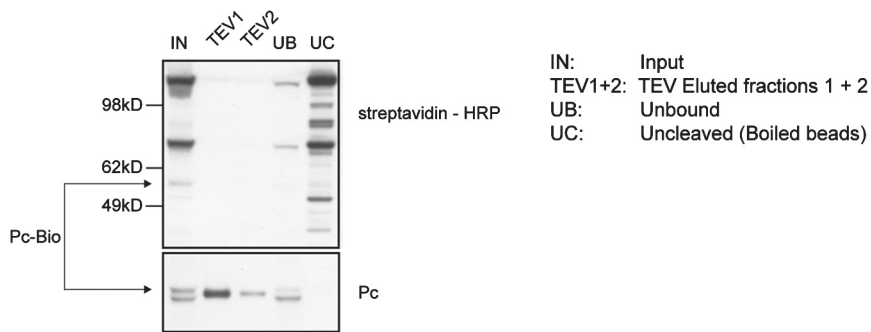


Fig. S2. Pc-Bio is specifically released by TEV cleavage. Elution of biotinylated Pc-Bio from streptavidin beads was performed in 100 μ l using 15 U of AcTEV protease (Invitrogen). Endogenously biotinylated proteins were not eluted and could not be detected in the supernatant (TEV1 + 2 lanes) but became visible after boiling of the beads in sample buffer (lane uncleaved UC).

Estimation of a minimal peptide cut off (total peptides/protein) for randomly identified proteins

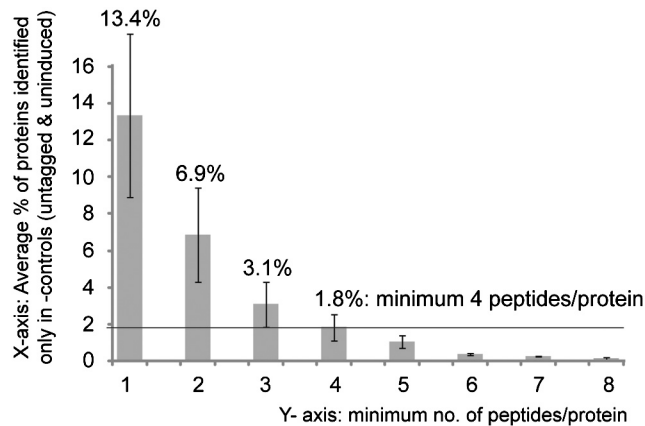


Fig. S3. Calculation of a minimal peptide cut off for excluding randomly identified proteins. Peptides specifically found in negative controls (untagged and uninduced) were assumed to be detected at random and their contribution to the sample assessed. If only proteins identified by four peptides minimum are considered, the amount of randomly detected proteins \leq 1.8%.

The *vtd4* allele suppresses the sex comb phenotype
 Suppression by *Rad21ex3* can be rescued by expressing full length Rad21-myc

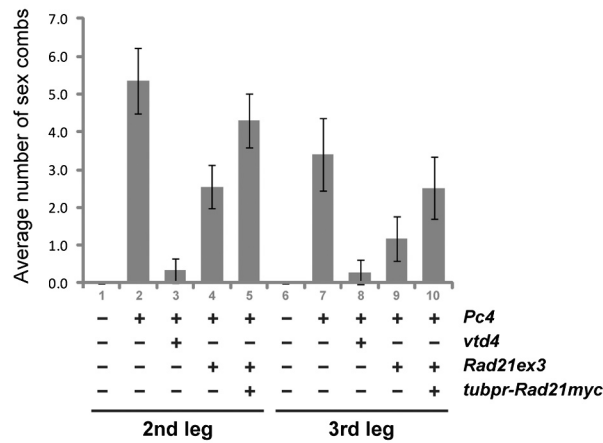


Fig. S4. Rad21 alleles suppress extra sex comb phenotype. The Polycomb *Pc4* mutant shows extra sex combs on second and third legs (lanes 2 and 7). This phenotype is suppressed in a *vtd4* (*verthandi*) mutant background (*Pc4/vtd4*; lanes 3 and 8), as has been published previously (1). *vtd4* is a known allele of the *Rad21* locus that can be rescued by transgenic Rad21 (2). Suppression also occurs in the *Rad21ex3* background (lanes 4 and 9) (3) and the extra sex combs phenotype can be rescued by expressing a full-length form of myc-tagged Rad21 (lanes 5 and 10).

- Schulze S, et al. (2001) Essential genes in proximal 3L heterochromatin of *Drosophila melanogaster*. *Mol Gen Genet* 264:782–789.
- Hallson G, et al. (2008) The *Drosophila* cohesin subunit Rad21 is a trithorax group (trxG) protein. *Proc Natl Acad Sci USA* 105:12405–12410.
- Pauli A, et al. (2008) Cell-type-specific TEV protease cleavage reveals cohesin functions in *Drosophila* neurons. *Dev Cell* 14:239–251.

Pc remains bound to polytene chromosomes in the absence of cohesin

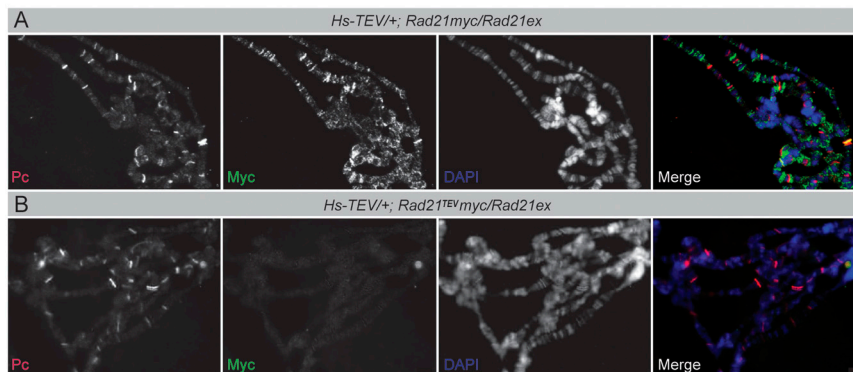


Fig. S5. Pc remains bound to polytene chromosomes in the absence of cohesin. Animals expressing TEV cleavable and myc-tagged Rad21^{TEV}myc protein in a Rad21 null background (1) were stained with myc and Pc antibodies. (A) Coimmunostainings reveal mutual exclusive localization of the two proteins except for one prominent band. (B) Rad21 is removed from chromosomes after heat shock induction of TEV protease. Polycomb localization is not altered after loss of Rad21 binding.

- Pauli A, et al. (2008) Cell-type-specific TEV protease cleavage reveals cohesin functions in *Drosophila* neurons. *Dev Cell* 14:239–251.

Control experiments for the pairing sensitive silencing reporter

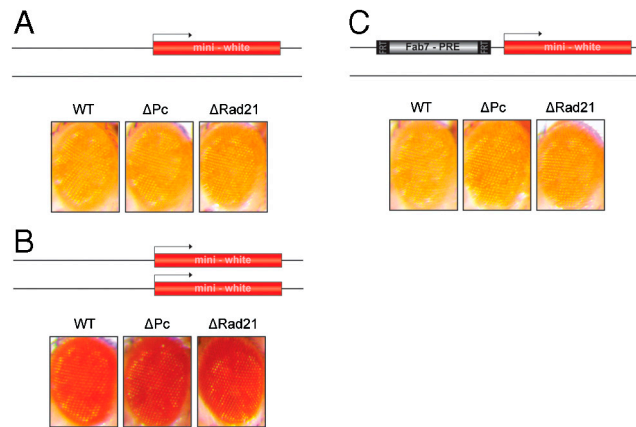


Fig. 56. Control experiments for pairing sensitive silencing reporter. The reporter shows PRE-dependent pairing sensitive silencing (PSS). Heterozygous animals with or without a PRE or homozygotes lacking the PRE do not show repression of the miniwhite reporter and do not respond to Pc and Rad21 mutations.

Table S1. Library of UAS-BirA ligase strains

Ligase Name	L4.2	L4.1	L4.7	L4.3	L4.5	L4.11	L4.14	L6.2
Chromosome	X	II	II	III	III	III	III	X
Promoter	pUASp	pUASp	pUASp	pUASp	pUASp	pUASp	pUASp	pUASp
Marker	white	white	white	white	white	white	white	white
Epitope Tag	Flag	Flag	Flag	Flag	Flag	Flag	Flag	-
Biotinylation competent	n.t.	n.t.	n.t.	n.t.	n.t.	n.t.	n.t.	Yes
Ligase Name	L6.1	L11	L8.1	L3.8	L3.24	L3.6	L3.10	L3.16
Chromosome	II	II	III	X	X	II	II	II
Promoter	pUASp	pUASp	pUASp	pUASp	pUASp	pUASp	pUASp	pUASp
Marker	white	white	white	eGFP	eGFP	eGFP	eGFP	eGFP
Epitope Tag	-	-	-	-	-	-	-	-
Biotinylation competent	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes
Ligase Name	L3.3	L3.12	L3.15	L5.5	L5.2	L5.3	L5.6	L5.8b
Chromosome	III	III	III	X	II	III	III	III
Promoter	pUASp	pUASp	pUASp	pUASp	pUASp	pUASp	pUASp	pUASp
Marker	eGFP	eGFP	eGFP	yellow	yellow	yellow	yellow	yellow
Epitope Tag	-	-	-	Flag	Flag	Flag	Flag	Flag
Biotinylation competent	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes

The table displays different ligase strains, which were generated in the course of this study. Transgene markers used were either white, eGFP, or yellow. Some ligases further feature a Flag-Tag. We tested the expression of the ligase transgenes by glass or *daughterless-Gal4* induction in *Drosophila* heads and subsequent Western blot of head protein extracts.

Table S2. List of fly strains used in this study

Name	Genotype	Source and Notes
Mutants		
Pc[1]	<i>w[1118]; ; Pc[1]/TM6B, Tb</i>	Paro Lab stock collection
Pc[3]	<i>w[1118]; ; Pc[3]/TM6B, Tb</i>	Paro Lab stock collection
Pc[XL5]	<i>w[1118]; ; Pc[XL5]/TM6B, Tb</i>	Paro Lab stock collection
Pc[4]	<i>w[1118]; ; Pc(4) p(p) e(s)/TM6B, Hu Sb e Tb ca</i>	1
rad21 excision alleles	<i>+</i> ; <i>Rad21[ex3 or ex 8 or ex15 or ex16];</i>	2
rdx[1]	<i>cn[1]; ry[506] P{PZ}mei – P19 [03477] rdx[1]/ TM3, ry[RK] Sb [1] Ser[1]</i>	Bloomington stock no: 11592
rdx[2]	<i>w[1118]; ; Rdx[2]/TM3</i>	Kornberg Lab (UCSF)
rdx[5], rdx[6]	<i>w[1118]; ; Rdx[5or6]/TM6B, Tb</i>	3
vtd[4]	<i>Eip74EF^{v4}vtd^A/TM3, st^{2A}Sb¹</i>	Bloomington stock no: 5050
ebi [WKS-24]	<i>w[1118]; ebi[WKS-24]/CyO, P{GMR – p21.Ex}2; +</i>	Bloomington stock no: 8398
ebi [CCS-6]	<i>w[1118]; ebi[CCS-6]; +</i>	Bloomington stock no: 8396
ebi [E90], ebi [E4]	<i>w[1118]; Ebi[E90 or E4]/CyO; +</i>	4
In vivo biotinylation system		
Pc-Bio (C-term, TEV- Bio- Flag)	<i>w[1118]; P{Pc – Bio(23), GFP};</i>	This work
BirA ligase	<i>w[1118]; ; P{BirAL3.15,GFP}</i>	This work (see Table S1)
da-Gal4	<i>w[1118]; ;P{da – Gal4.w}</i>	Paro Lab stock collection, white transgene marker
TEV cleavable Rad21 system		
TEV degradable & Myc-tagged Rad21	<i>w-; ; Rad21ex, tubPr.-Rad21(3xTEV)myc 10</i>	2
Myc-tagged Rad21	<i>w-; ; Rad21ex, tubPr.-Rad21myc 10</i>	2
Pc-GFP, ΔRad21, hs TEV	<i>w-; Pc-GFP,hsTEV/CyO; Rad21ex/TM6Bubi-GFP</i>	Pc-GFP (5) Rad21ex (2)
PSS reporter		
pFas (Cre)	<i>w- ; pFas;</i>	6
pFas (Flp; Cre)	<i>w- ; pFas (Flp) ;</i>	6

1 Kennison JA, Tamkun JW (1988) Dosage-dependent modifiers of polycomb and antennapedia mutations in *Drosophila*. *Proc Natl Acad Sci USA* 85:8136-8140.

2 Pauli A, et al. (2008) Cell-type-specific TEV protease cleavage reveals cohesin functions in *Drosophila* neurons. *Dev Cell* 14:239-251.

3 Kent D, Bush EW, Hooper JE (2006) Roadkill attenuates Hedgehog responses through degradation of *Cubitus interruptus*. *Development* 133:2001-2010.

4 Tsuda L, Nagaraj R, Zipursky SL, Banerjee U (2002) An EGFR/Ebi/Sno pathway promotes delta expression by inactivating Su(H)/SMRTER repression during inductive notch signaling. *Cell* 110:625-637.

5 Dietzel S, Niemann H, Bruckner B, Maurange C, Paro R (1999) The nuclear distribution of Polycomb during *Drosophila melanogaster* development shown with a GFP fusion protein. *Chromosoma* 108:83-94.

6 Schmitt S, Prestel M, Paro R (2005) Intergenic transcription through a polycomb group response element counteracts silencing. *Genes Dev* 19:697-708.

Table S3. Proteins and peptides identified by mass spectrometry

Psc	
LPTLAPFVGFNPLQNPAAGK	QAGNLPM[147]SAPPNK
TTTTTTPPTTTTTTAAAAAEATTNADK	AITPPSPSVQQSASPK
ENQEQLLAVEVASSK	CEMVINNAKPN
LPTLAPFVGFNPLQNPAAGK	PNPFANIPNDVNR
QNSVTIIDMSDPER	SEEDPTAAVAASSTATTTSDLATTSR
SDTTLQAIVYK	SSPCTPVSSPSEPNIK
SVVSNANSSGNSSK	DVAATPPTETLK
VYESPQLVKPAPR	SPSPLTVPPLTIR
SEEDPTAAVAASSTATTTSDLATTSR	YESPQLVKPAPR
DRPEEALATPEQR	SENTLATTANAALAAATTTTTATPALATGK
LPVTNGNSSGTASPK	IMSPSGVSTLSR
M[147]GPPALPATTPSQGNK	LVNGGQSQAQK
QNSVTIIDM[147]SDPER	VGFNPLQNPAAGK
VGNEVFNDYLQK	IEKPLM[147]PPPAKPPM[147]LAPR
VTSGAFSEDPK	GNSSNNYLNLFNSSK
SEEDPTAAVAASSTATTTSDLATTSRPR	SPVNNYIEIVK
ATSEDASSNGGASADEEK	PQLVKPAPR
LPDQPQDQVQAAGK	FSIDIM[147]YK
PGLYER	YLQCPAMCR
CEM[147]VINNAKPNIK	PELVDTLRPR
LTAATAPQTK	TDSEPELVDTLR
MGPPALPATTPSQGNK	SEPELVDTLRPR
NPT[181]PPPSLPAVGGK	S[167]PSPLTVPPLTIR
SPQPLVKPAPR	CEM[147]VINNAKPNIK
VYESPQLVKPAPR	LALSQSQK
PPLPTVDFK	FVYDKFEIDAQR
ESPQLVKPAPR	QLLFAACSIK
VYESPQLVK	GGNGGSLGGLFPSPTK
YLQCPAM[147]CR	ASRPNPFANIPNDVNR
LSPPLPTVDFK	SQLSTLAK
TDSEPELVDTLRPR	E[111]IVKPLKPEK
TDSEPELVDTLRPR	VKVES[167]PER
VTPLKPVLTPTQVDK	PPPPSSPR
NPTPPPSLPAVGGK	RVLPLK
LVPGLYER	SFALKPIK
RYQPILPK	YDKFEIDAQR
GNLDDSLM[147]KPPSCM[147]PPK	C[143]EM[147]VINNAKPNIK
FEIDAQR	VEPVSLPEDQK
AIT[181]PPS[167]PSVQQSASPK	KLALSQSQK
EIVKPLKPEK	
Ph-p	
YADKDVSDPEPK	GPTATLVPIGSPK
VVGHLTTVQQQQATNLQVVNAAGNK	HLVNAM[147]GM[147]K
LSGIASAPGSDM[147]VACEQCGK	QEFPTHHTSGSGTELK
TEIGQVAGQNK	AAVQPPSSTTPNSVSGK
M[147]VVM[147]STTGTPITLQNGQTLHAATAAGVDK	GPTATLVPIGS[167]PK
SSTPATVSASVEASSSTGEALSNGDASDR	STASSGGGSIPATPTK
VVGHLTTVQQQQATNLQVVNAAGNK	IIQEANEPPVTR
SSEVNGTDRPPISSWSVDDVSNFIR	LSGIASAPGSDMVACEQCGK
AQQQQAVAAQQQAVAAQQQQQR	QQQLQLFQK
QQQQQQVGTNNQTTQQQLAVATAQLQQQQQLTAAALQR	LSTASSGGGSIPATPTK
NQPDGTQM[147]FIQQQPATQLTQQNQIIQCNTQTPTK	TQLDALAPK
M[147]VVMSTTGTPITLQNGQTLHAATAAGVDK	QEFPTHHTSGSGTELK
PASSVSTQTAQNQSLLK	ATMQEDIK
QIIQCNTQTPTK	LDEAM[147]AEEK
NGIGGVGSGETNGLGTGGIVGVDAM[147]ALVDR	SVALPTLAPLSVVTSGAAPK
QSNAAVQPPSSTTPNSVSGK	VIDGFIIQEANEPPVTR
Q[111]EFPHTTSGSGTELK	LTTVQQQQATNLQVVNAAGNK
ATM[147]QEDIK	ADKDVSDPEPK
QSNAAVQPPSSTTPNSVSGK	LDEAMAEK
YADKDVSDPEPK	VIDGFIIQEANEPPVTR
QLAAATGGVGGDWTQGR	VLTHVIDGFIIQEANEPPVTR
PTHHTSGSGTELK	TVQQQQATNLQVVNAAGNK
MVVMSTTGTPITLQNGQTLHAATAAGVDK	AAGLQPFGNQIILR
QSNAAVQPPSSTTPNSVSGKEEPK	EVPPPGEAK
IGQVAGQNK	Q[111]QQQLQLFQK
HTSLTLEK	QSNAAVQPPSSTTPN
PPSSTTPNSVSGK	YCSPGCSR
HLVNAM[147]GMK	LS[167]ESFPILGASTEVPMSLP

Psc

DKDVSDEPPK	HLVNAMGM[147]K
PVRPALATLK	
Sce/dRing	
DPGHSGTSAASAITASNAAPSSSANSNGASTSATR	KPQEVITDSTEIAVSPR
FNQTQSQQALVNSINEGIK	LTLDLGADLPEACR
GHSGTSAASAITASNAAPSSSANSNGASTSATR	M[147]QVDDASNPPSVR
NASNQM[147]HVHDTASNDSNSNTNSIDR	SEESSEDSQMDCR
SLHSELM[147]CPICLDM[147]LK	SEESSEDSQM[147]DCR
DPGHSGTSAASAITASNAAPSSSANSNGASTSATR	KPQEVITDSTEIAVSPR
PGHSGTSAASAITASNAAPSSSANSNGASTSATR	AM[147]SVLTSER
GGGGGGGGGNGNGAANVAAPPAGAPTAVGR	STPSPVPSNSSSSK
NASNQM[147]HVHDTASNDSNSNTNSIDR	ANATVDHLSK
GGGGGGGGGNGNGAANVAAPPAGAPTAVGR	FNQTQSQQALVNSINEGIK
S[167]TPSPVPSNSSSSKPK	SLHSELM[147]CPICLDMLK
S[167]TPS[167]PVPSNSSSSKPK	PVPSNSSSSKPK
Q[111]SQNRPQR	CSDCIVTALR
LQSQNRPQR	VNKPM[147]EM[147]YYSWK
AAPSSSANSNGASTSATR	TANATVDHLSK
FCSDCIVTALR	PSPVPSNSSSSKPK
MQVDDASNPPSVR	STPSPVPSNSSSSKPK
PPAPGAPTAVGR	VAAPPAGAPTAVGR
TTANATVDHLSK	ADPNFDLLISK
TWELSLYELQR	SPVPSNSSSSKPK
PNFDLLISK	TTANATVDHLSK
TPSPVPSNSSSSKPK	YEAIQEK
DPGHSGTSAASAITASN	KPQEVITDSTEIAVS[167]PR
FNQTQSQQALVN	EYEAIQEK
LQSQNRPQR	PSNSSSSKPK
M[147]TSLDPAPNK	NATVDHLSK
PSPVPSNSSSSKPK	DPNFDLLISK
Ph-d	
VVGHLLTVQQQQATNLQVVNAAGNK	SSASSGGGAGFPATPTK
VVGHLLTVQQQQATNLQVVNAAGNK	QEFPTHHTSGSGTELK
M[147]VVM[147]STTGTPITLQNGQTLHAATAAGVDK	LSSASSGGGAGFPATPTK
NGIGVGSGETNGLGTGGIVGVDAM[147]ALVDR	SASSGGGAGFPATPTK
SSTPATVSASVEASSSTGEALNGDASDR	LSGIASAPGSDMVACEQCGK
MVVMSTTGTPITLQNGQTLHAATAAGVDK	QQQLQLFQK
NQPDGTQGM[147]FIQQPATQLTQQQNIIQCNTQTPTK	GPTATLVPIIDS[167]PK
M[147]VVMSTTGTPITLQNGQTLHAATAAGVDK	LDEAM[147]AEEK
Q[111]QQQLQLFQK	YCSPGCSR
GPTATLVPIIDSPK	AAVQPPSSTIPNSVSGK
LSGIASAPGSDM[147]VACEQCGK	Q[111]JEFPTHHTSGSGTELK
TEIGQVAGQNK	ATM[147]QEDIK
LDEAMAEK	YADKDVSEPPK
TQLDALAPK	ENHLVNAM[147]GM[147]K
QEFPTHHTSGSGTELK	YADKDVSEPPK
ATMQEDIK	AAGLQPFGSNQILR
PPGDVKD	QSNAAVQPPSSTIPNSVSGK
ENHLVNAMGM[147]K	
Su(z)2	
INQDIEPEHSVR	ILLYDNEQTK
NIGLKPIEQPLQSSASNPDSK	NQDIEPEHSVR
LSQAQAM[147]ASSYAAK	SVTFAEDLESEIDSGSPR
SHSLASGELDLQK	LHAEISSQTDGK
KSYVDAEDFELK	PGLYQR
LDSTSTSEALNR	PPALINLR
SYVDAEDFELK	TNPAKPLSSNNNR
YQSTPSSIASAANK	EAEPESPVSNFK
INQDIEPEHSVR	NQPAAASTAASISK
DLTLPTSPPLPPLSLFK	AAALAEAPPVLSNAAK
PEQEQLLPR	SPPLSVALSGQR
TTAVALRPEPK	LVPGLYQR
SMSLDESHPAK	LSM[147]PLSAGPR
SM[147]SLDESHPAK	QDIEPEHSVR
AVYCPECK	PAKPLSSNNNR
Q[111]FHDLITCR	AYTPSTTPTAPTAVAGGKPK
SEPEQEQLLPR	LM[147]GPPAALPK
SVTFAEDLESEIDSGSPR	EINELNLK
YSDYAVSK	PNPSALAFR
LHAEISSQTD	NDENNLRS
GLTM[147]PPLSPPATSSAR	
Pc	

Psc

TSPDGPTIKPQPTQQVTPSQQQPFQDQQQAEK	IDHSSSSNSSFTHN
EPDPEPESEEDYFTTENDVDTHQATTSSATHDK	LIDIYEQTNK
EKEPDPEPESEEDYFTTENDVDTHQATTSSATHDK	RLIDIYEQTNK
EKEPDPEPESEEDYFTTENDVDTHQATTSSATHDK	YNTWEPEVNILDR
TSPDGPTIKPQPTQQVTPSQQQPFQDQQQAEK	PQPTQQVTPSQQQPFQDQQQAEK
E[111]KEPDPEPESEEDYFTTENDVDTHQATTSSATHDK	KGVVEYR
EKEPDPEPESEEDY[243]FTTENDVDTHQATTSSATHDK	PCNNLAINQK
TSPDGPT[181]IKPQPTQQVTPSQQQPFQDQQQAEK	FWLPAK
IDHSSSSNSSFTH	IASEAATQLK
DNATDDPVDLVYAAEK	PFQDQQQAEK
IDHSSSSNSSFTH	IDHSSSSNSSFTHN
QQVTPSQQQPFQDQQQAEK	SFVPEPDSNSSSESDQLIGTK
PVDLVYAAEK	EKEPDPEPESEEDYFTTENDVDTH
IDHSSSSNSSF	QPLTPLSPR
NNLAINQK	PLTPLSPR
ASEAATQLK	IDIYEQTNK
TSPDGPTIKPQPTQQVTPSQQQPF	SESPLTHH
SEAATQLK	QPLTPLS[167]PR
Smc1	
TALFEIISGSGLLK	YSM[147]VDLESSK
FEDEIENESQR	SEQDLDGETNR
NFLVFQGAVENIAM[147]K	FDALALDGTIFYQK
VQSTNEEFENAR	FM[147]EAIIVDTEK
ELVM[147]QQR	QSELATVESQIK
SGLISGGSHDLAR	SELDVNR
SVQDEEDALEGLK	HEKMEADR
SSQAALREEQNR	VIGGSS[167]EYR
AVIGGSSEYR	
Smc3/Cap	
LDLTIVDLNDEVQDGNK	INQM[147]ATAADSYR
LPGEVTFM[147]PLNR	QELM[147]STLSSQDQR
STGLDCVTLGDGDQVSSK	TEYTSQIAEFEK
AYYGPVIENFSCDK	LDNLLINNLFR
TIEDRL	TELVSAEK
EVENSDAFTGIGIR	EIDQLNDDIR
SVLM[147]TEQQQLLR	GSLTGGYFNTSR
VLDSFVER	LQTLSEEEKEELKEYQK
IASLGAVPLVDPSTYR	VSHIDCVTR
dSfmbt	
GELYSVLNLTN	GNIDPSVPIQK
ISDLIAQLK	TPTDNNTQSVK
M[147]NFTFDEYYSDGK	IEPVNRPGLVLK
DNNFDDNGSELEPK	LGLNLECVDKDR
LVCVATVAR	INDSLQSR
PP1-87B	
IFCCHGGLSPDLTSM[147]EQIR	ICGDIHGQYYDLLR
YSENFLLR	AHQVVEDGYEFFAK
HEFDLICR	IYGFYDECK
TFTDCFNCLPVAIAIVDEK	
Ebi	
DLLASGSGDSTAR	PGTGVAGSAGGNK
IEPGTGVAGSAGGNK	APSGAVTIR
LASTLGQHK	HLASGSFDK
Enok	
TDVSSLVEQPVK	VVSSVNVVSTAR
VQLLEEIER	TDQQLVQVK
STPVASSTPEK	
Grh	
TAVHGSQNSPTTSLVDTSTNGSTR	VVGPNGEQQIISR
IEFDENQIIR	NEAGEHILTR
ISTTSINNIYR	
Eaf6	
NEGSVVSGM[147]VSGSTNSGGNK	KAETSEQLANLER
DLLGTPTSNTK	AETSEQLANLER
SSITSM[147]AICNPER	
Pho	
KGDNVINYNIHENDK	VLTNLSNNDINTEESGVVDK
GNLSQENNISER	NIGYGENQETSK
Fs(1)h	
NNAAGNAAGGAAGAAAGAGSVGGVGGAGAAGGGNASK	ELELEPVAK
GGRGAKGS[167]GAGGVGASNNAAGNAAGGA	GSSSAGAGGGVGGANASAGGAGAR
CG1845	

Psc

SLNNSSPVGINR	LLLAAPASEGIVQK
GCILNADAPPLEENAPWAR	EPVDTSEVPDYTDIVK
Rdx	
VAITDVDHEVLK	DFLLDEANGLLPEDK
SSTFSAGANDK	M[147]ADDLLAAADK
Rad21	
YLLADCNEAFVK	DSLELSQLTSGNSR
AHVFETNIEK	
26–29 kd proteinase	
HGPLSVAIDASPK	LYGAVTPVK
DEIPDQYDWR	

This table displays the peptide coverage as well as the number of unique and total peptides found for the 23 proteins, which were specifically enriched in the sample and could be identified by a minimum of four peptides. Below the table, the individual peptides are listed.