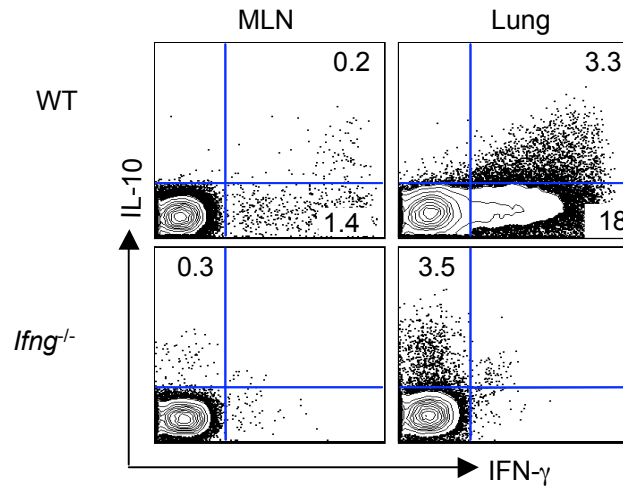


# **CD4<sup>+</sup> T cell help and innate-derived IL-27 induce Blimp-1 dependent IL-10 production by anti-viral CTL**

Jie Sun, Haley Dodd, Emily K. Moser, Rahul Sharma and Thomas J. Braciale

# Supplementary Figure 1

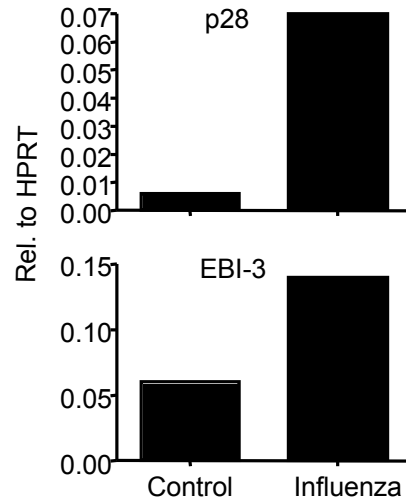


**Supplementary Figure 1. IFN- $\gamma$  is dispensable for the induction of IL-10 producing CTL during respiratory virus infection.**

WT or *Ifng*<sup>-/-</sup> mice were infected with influenza. At d7 p.i., the production of IL-10 and IFN- $\gamma$  by CTL was measured by ICS following flu-infected BMDC restimulation.

Numbers are the percentages of cells in gated population.

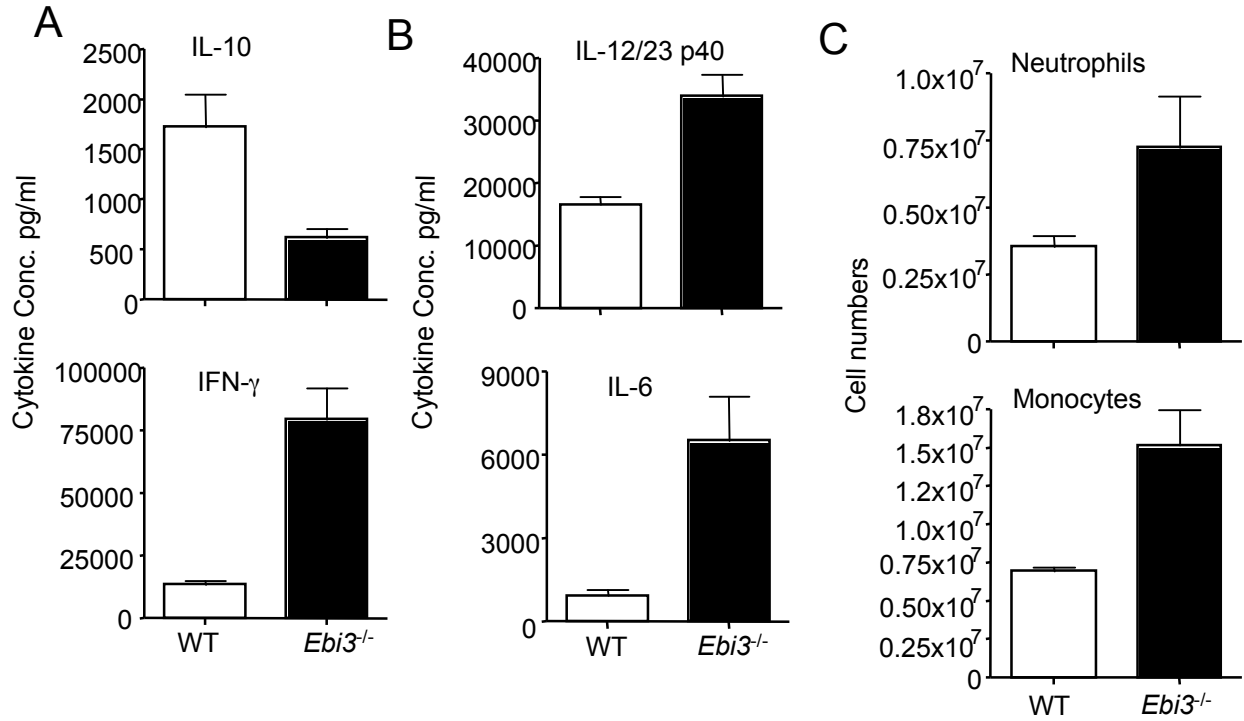
## Supplementary Figure 2



### **Supplementary Figure 2. Influenza infection induces IL-27 expression in the respiratory tract.**

WT mice were infected with influenza. At d5 p.i., the expression of IL-27 subunits p28 and EBI-3 by lung cells were determined through quantitative RT-PCR. Lung cells from uninfected mice (Control). Lung cells from d5 infected mice (Influenza).

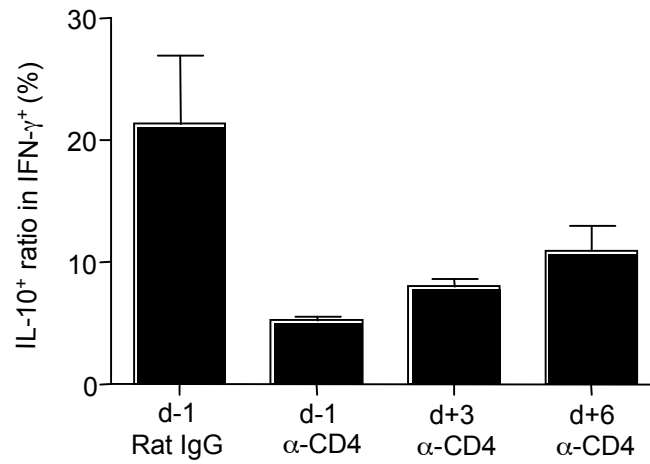
## Supplementary Figure 3



### Supplementary Figure 3. *Ebi3*<sup>-/-</sup> mice exhibit diminished IL-10 production and enhanced pulmonary inflammation during influenza infection.

WT or *Ebi3*<sup>-/-</sup> mice were infected with influenza. At d7 p.i., the levels of IL-10 and IFN- $\gamma$  (a) or IL-12/23 p40 and IL-6 (b) in BALF were determined through ELISA. (d) At d9 p.i., the numbers of lung monocytes/macrophages and neutrophils were measured through flow cytometry.

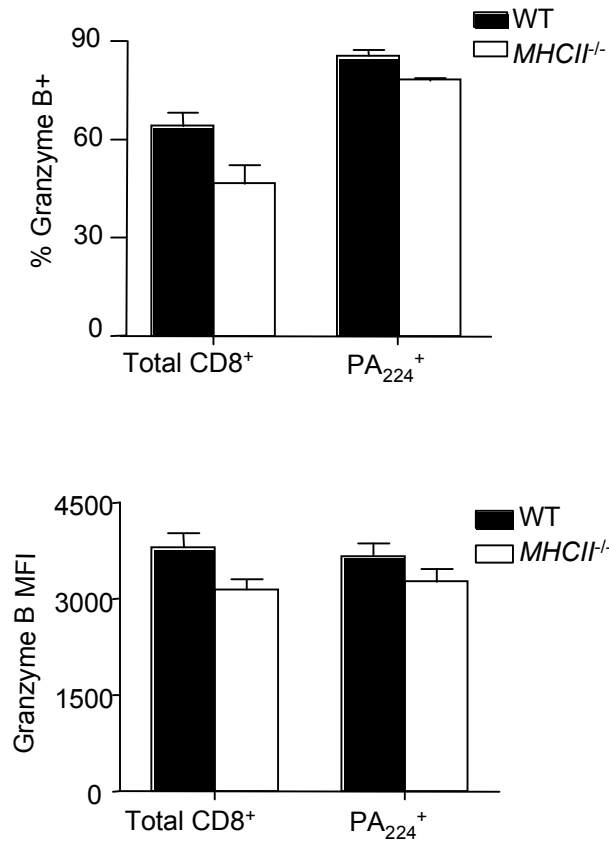
## Supplementary Figure 4



**Supplementary Figure 4. Impact of CD4<sup>+</sup> T cell depletion at different days on the induction of IL-10 producing CTL.**

WT mice were injected with Rat IgG control mAb or  $\alpha$ -CD4 depleting mAb at various days p.i.. At d8 p.i., the production of IL-10 and IFN- $\gamma$  by CTL was measured by ICS following flu-infected BMDC restimulation. The normalized percentages of IL-10<sup>+</sup> in influenza-specific CD8<sup>+</sup> T cells (reflected as IFN- $\gamma$ <sup>+</sup>) are depicted.

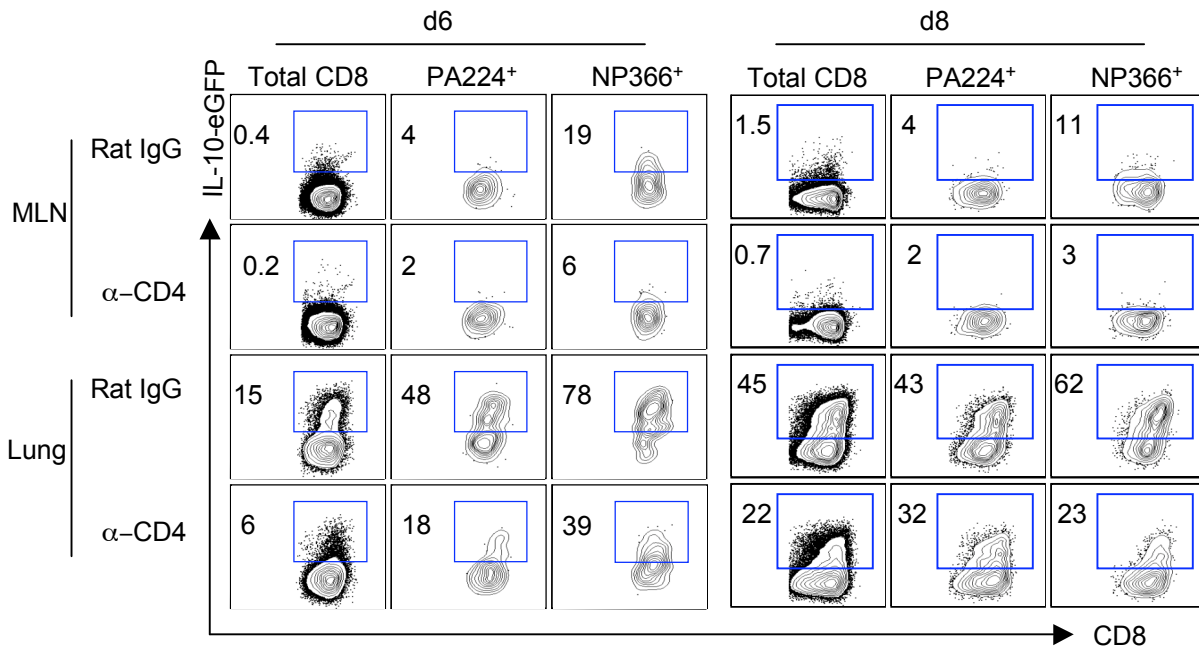
## Supplementary Figure 5



### Supplementary Figure 5. CD4<sup>+</sup> T cell help has a minimal impact on Granzyme B expression by CTL.

WT or *MHCII*<sup>-/-</sup> mice were infected with influenza. At d7 p.i., the expression of Granzyme B (*Gzmb*) by total lung CD8<sup>+</sup> T cells or PA<sub>224</sub> tetramer positive cells was determined through ICS. The percentages of *Gzmb*<sup>+</sup> cells and the MFI of *Gzmb* in *Gzmb*<sup>+</sup> cells are depicted.

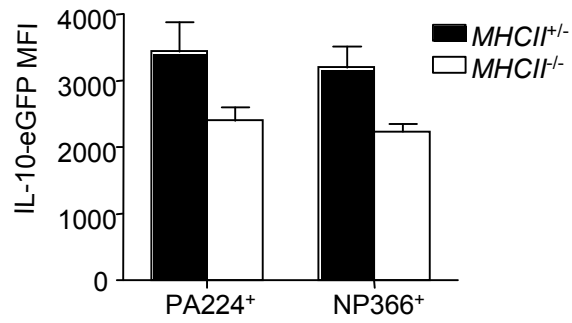
## Supplementary Figure 6



### Supplementary Figure 6. CD4<sup>+</sup> T cell depletion impairs the *in vivo* development of IL-10-expressing CTL

Vert-X mice were injected with either Rat IgG or  $\alpha$ -CD4 Ab and infected with influenza. At d6 and 8 p.i., the percentages of IL-10-eGFP<sup>+</sup> in total CD8<sup>+</sup> T cells or influenza specific PA<sub>224</sub> or NP<sub>366</sub> tetramer positive cells were measured through flow cytometry. Numbers are the percentages of cells in gated population.

## Supplementary Figure 7

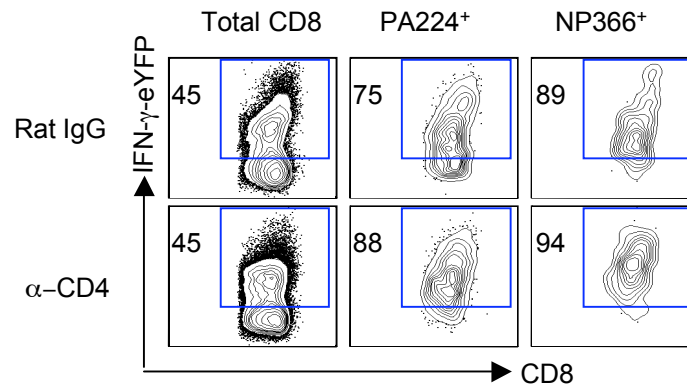


**Supplementary Figure 7. The absence of CD4<sup>+</sup> T cell impairs IL-10 expression at the level of individual cells**

*MHCII<sup>+/-</sup>*-Vert-X or *MHCII<sup>-/-</sup>*-Vert-X mice were infected with influenza. At d7 p.i., the MFI of IL-10-eGFP in IL-10-eGFP<sup>+</sup> influenza specific CTL is depicted.



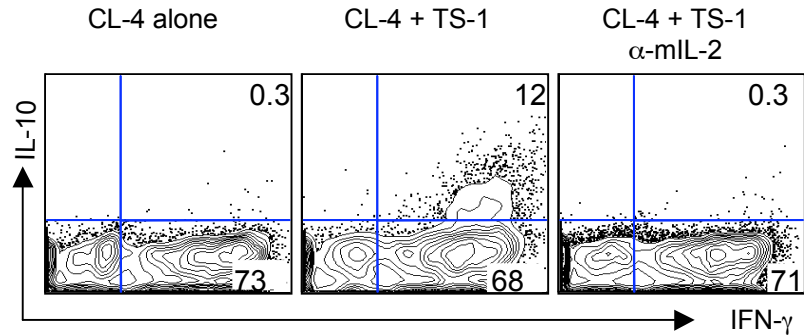
## Supplementary Figure 8



### Supplementary Figure 8. CD4<sup>+</sup> T cell depletion minimally affects the induction of IFN- $\gamma$ producing cells *in vivo*

IFN- $\gamma$  reporter (Yeti) mice were injected with either Rat IgG or  $\alpha$ -CD4 Ab and infected with influenza. At d7 p.i., the expression of IFN- $\gamma$ -eYFP in total CD8<sup>+</sup> cells or influenza specific PA<sub>224</sub> or NP<sub>366</sub> tetramer positive cells was measured by flow cytometric analysis. Numbers are the percentages of cells in gated population.

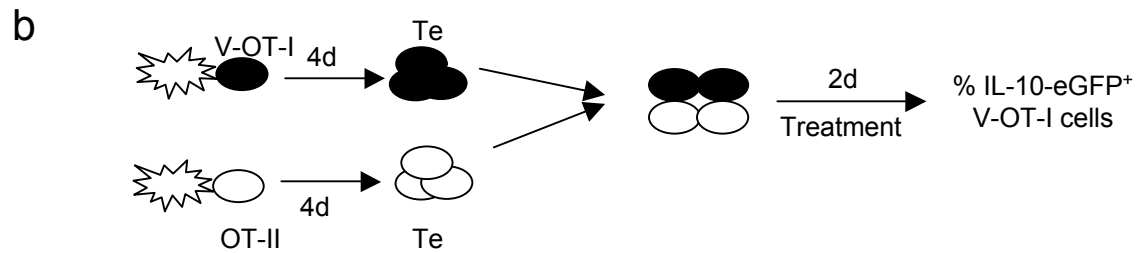
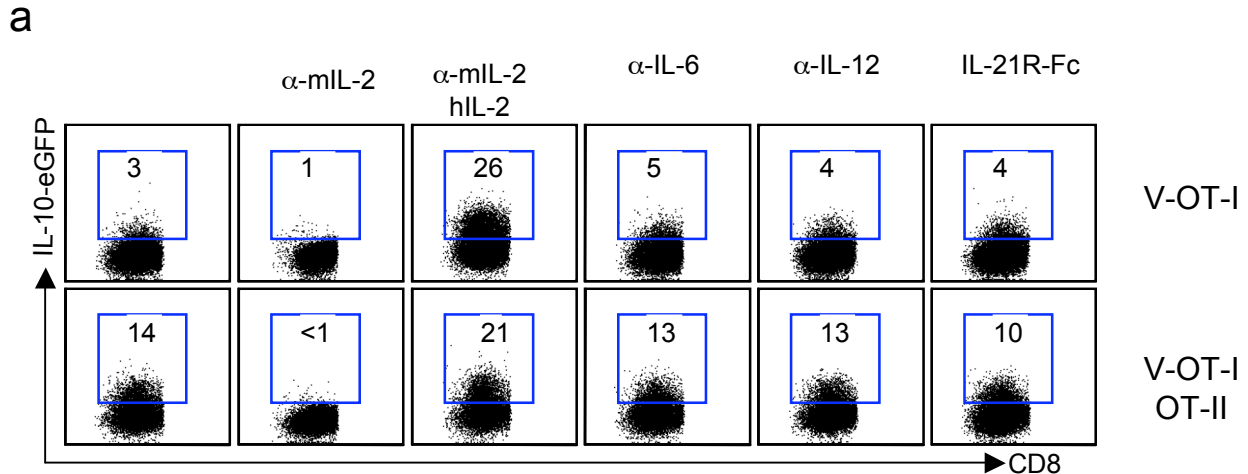
## Supplementary Figure 9



### Supplementary Figure 9. In vitro generated IL-10-producing cells are type 1 effectors

CD8<sup>+</sup> CL-4 TCR transgenic T cells were stimulated with influenza-infected DC in the presence or absence of CD4<sup>+</sup> TS-1 TCR transgenic T cells for 4d. The cultured cells were treated with various conditions as indicated. Then the production of IL-10 and IFN- $\gamma$  by CL-4 cells were measured through ICS following restimulation with HA<sub>533-541</sub> peptide. Numbers are the percentages of cells in gated population.

# Supplementary Figure 10

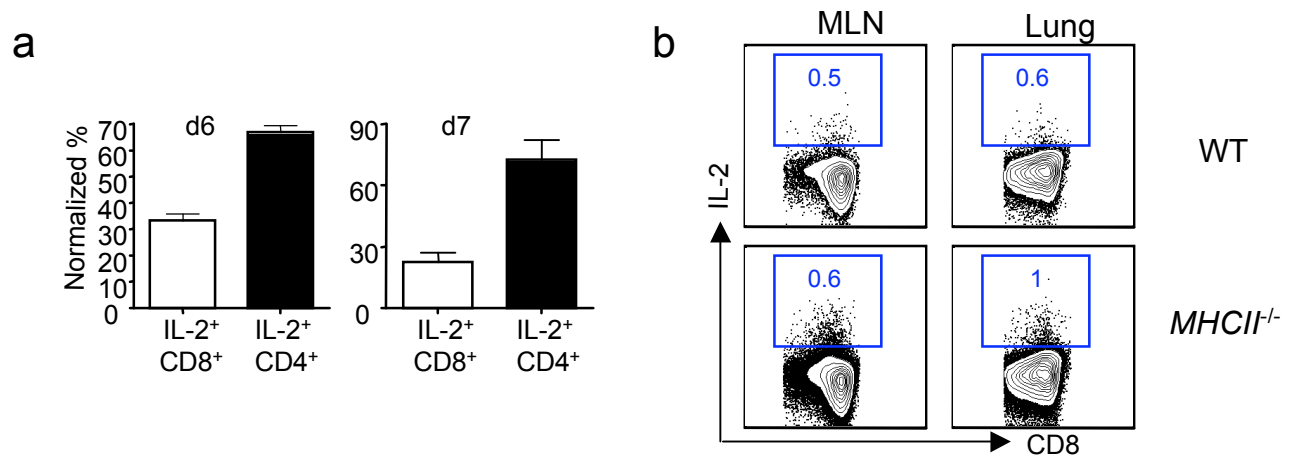


**Supplementary Figure 10. CD4<sup>+</sup> T cell help for the induction of IL-10 producing CTL by is dependent on IL-2 and not IL-6, IL-12 or IL-21.**

(a) CD8<sup>+</sup> Vert-X-OT-I (V-OT-I) double transgenic T cells were stimulated with influenza infected DC in the absence or presence of OT-II for 4d. The cultured cells were treated with various conditions as indicated. The expression of IL-10-eGFP by V-OT-I cells was measured by flow cytometry. Numbers are the percentages of cells in gated population.

(b) Schematic depiction of the culture condition to assess the role of activated CD4<sup>+</sup> effector T cells (Te) providing the “help” to activated CD8<sup>+</sup> Te.

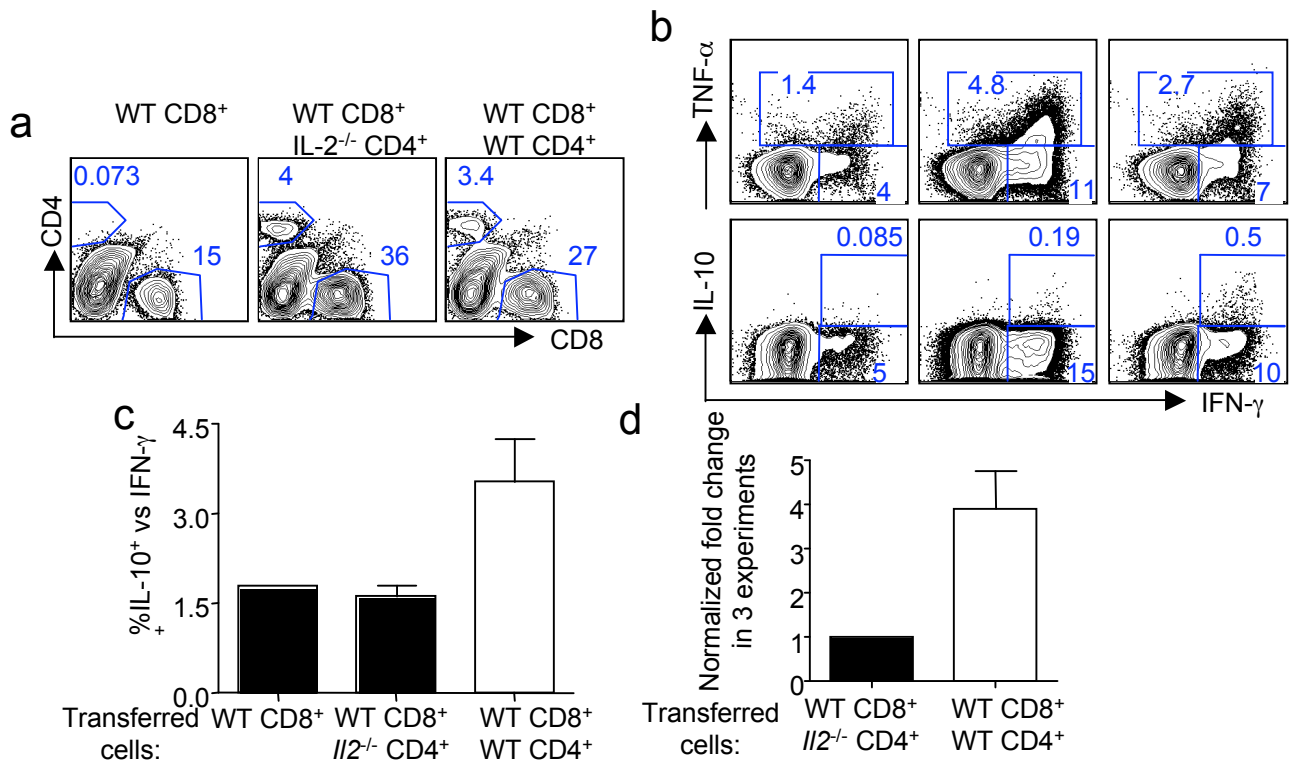
## Supplementary Figure 11



**Supplementary Figure 11. CD4<sup>+</sup> T cells (and not CD8<sup>+</sup> T cells) are the predominant source of IL-2 necessary to support optimal IL-10 expression by CTL.**

(a) WT mice were infected with influenza. At indicated days p.i., the production of IL-2 by gated Thy1<sup>+</sup> cells were measured by ICS following influenza infected BMDC restimulation. The normalized percentages of CD4<sup>+</sup> IL-2<sup>+</sup> cells or CD8<sup>+</sup> IL-2<sup>+</sup> cells in total IL-2<sup>+</sup> cells are depicted. (b) WT or *MHCII*<sup>-/-</sup> mice were infected with influenza. At d7 p.i., the production of IL-2 by gated Thy1<sup>+</sup> CD8<sup>+</sup> cells were measured by ICS following restimulation with influenza infected BMDC.

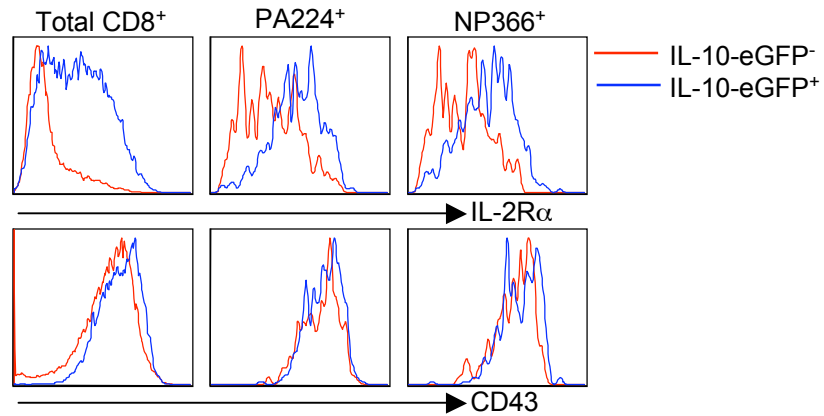
## Supplementary Figure 12



### Supplementary Figure 12. IL-2 derived from CD4<sup>+</sup> T cells is required for the optimal induction of IL-10 producing CTLs *in vivo*.

WT CD8<sup>+</sup> T cells were co-transferred either with WT or *Il2*<sup>-/-</sup> CD4<sup>+</sup> T cells into *Rag2*<sup>-/-</sup> mice. The recipient mice were subsequently infected with influenza. (a) At d9 p.i., the percentages of CD4<sup>+</sup> T cells or CD8<sup>+</sup> T cells in the lung were determined by flow cytometry. (b) The production of TNF, IL-10 and IFN-γ by CTL was measured by ICS following restimulation with influenza-infected BMDC. (c) The normalized percentages of IL-10<sup>+</sup> cells among influenza-specific (IFN-γ<sup>+</sup>) CD8<sup>+</sup> T cells from infected lungs of transferred mice are depicted. (d) The normalized fold changes of the percentages of IL-10<sup>+</sup> cells in antigen specific IFN-γ<sup>+</sup> secreting CTL detected in infected recipients of transferred WT or *Il2*<sup>-/-</sup> CD4<sup>+</sup> T cells from three individual experiments are depicted. Normalization method for panel d: % of IL-10<sup>+</sup> cells in antigen specific CTL from *Rag2*<sup>-/-</sup> recipient mice receiving *Il2*<sup>-/-</sup> CD4<sup>+</sup> T cells are designated as 1. The corresponding numbers in *Rag2*<sup>-/-</sup> mice receiving WT CD4<sup>+</sup> T cells are calculated as % of IL-10<sup>+</sup> cells in antigen specific CTL in these mice divided by the % of IL-10<sup>+</sup> cells in antigen specific CTL from *Rag2*<sup>-/-</sup> mice receiving *Il2*<sup>-/-</sup> CD4<sup>+</sup> T cells.

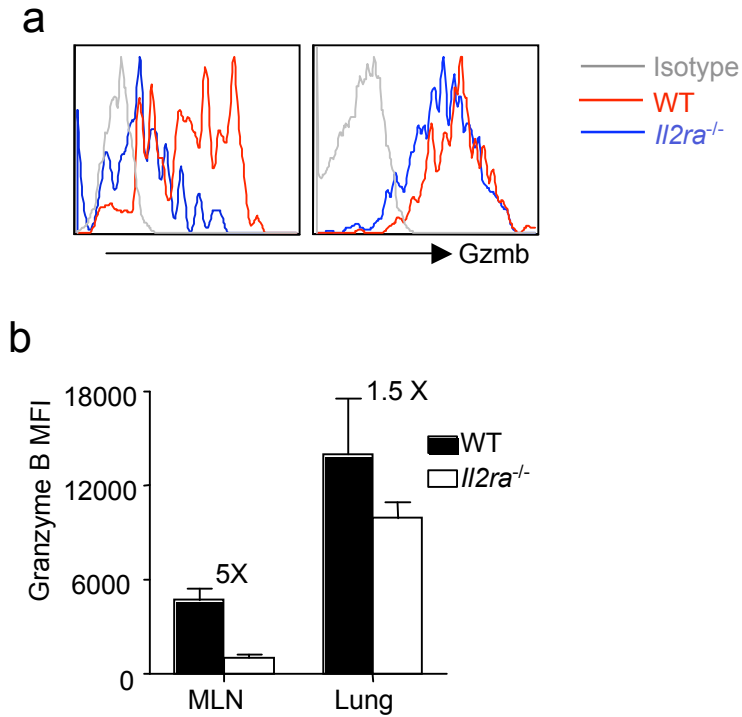
## Supplementary Figure 13



### Supplementary Figure 13. IL-10-expressing cells express IL-2R $\alpha$

Vert-X mice were infected with influenza. At d7 p.i., the expression of IL-2R $\alpha$  and CD43 by lung total CD8<sup>+</sup> cells or influenza specific PA<sub>224</sub><sup>+</sup> or NP<sub>366</sub><sup>+</sup> cells was measured by flow cytometry.

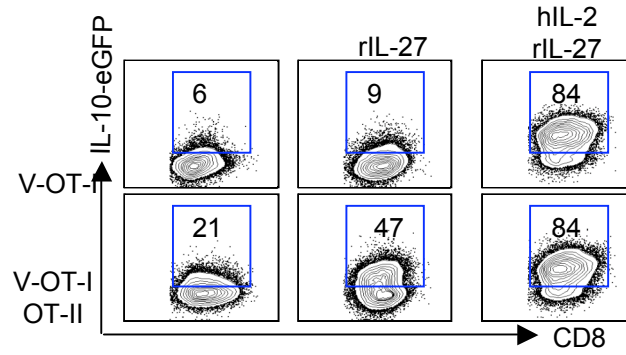
## Supplementary Figure 14



### Supplementary Figure 14. IL-2 signaling is minimally required for Granzyme B expression by CTL in the lung

WT:*Il2ra*<sup>-/-</sup> mixed bone marrow chimera mice were infected with influenza. At d7 p.i., the expression of Granzyme B by influenza specific CD8<sup>+</sup> PA<sub>224</sub><sup>+</sup> cells in MLN or lung was measured ICS.

## Supplementary Figure 15

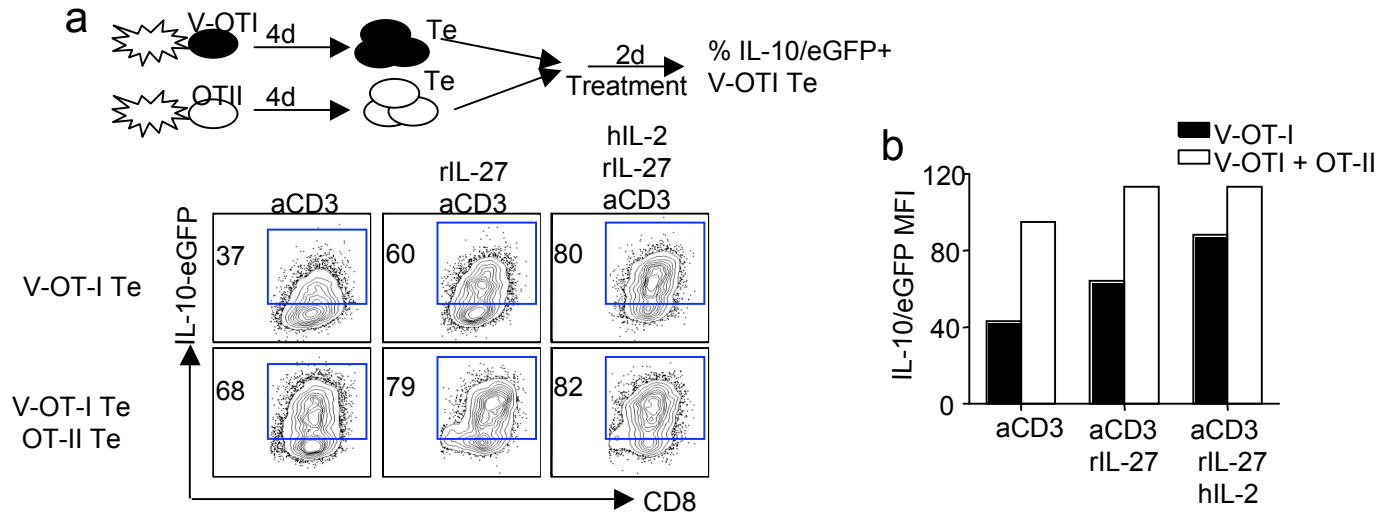


### **Supplementary Figure 15. IL-2 and IL-27 co-operatively induce IL-10 expressing CD8<sup>+</sup> CTL.**

CD8<sup>+</sup> Vert-X-OT-I (V-OT-I) double transgenic T cells were stimulated with influenza-OVA infected DC in the absence or presence of OT-II for 4d. The expression of IL-10-eGFP by V-OT-I cells was measured by flow cytometry.



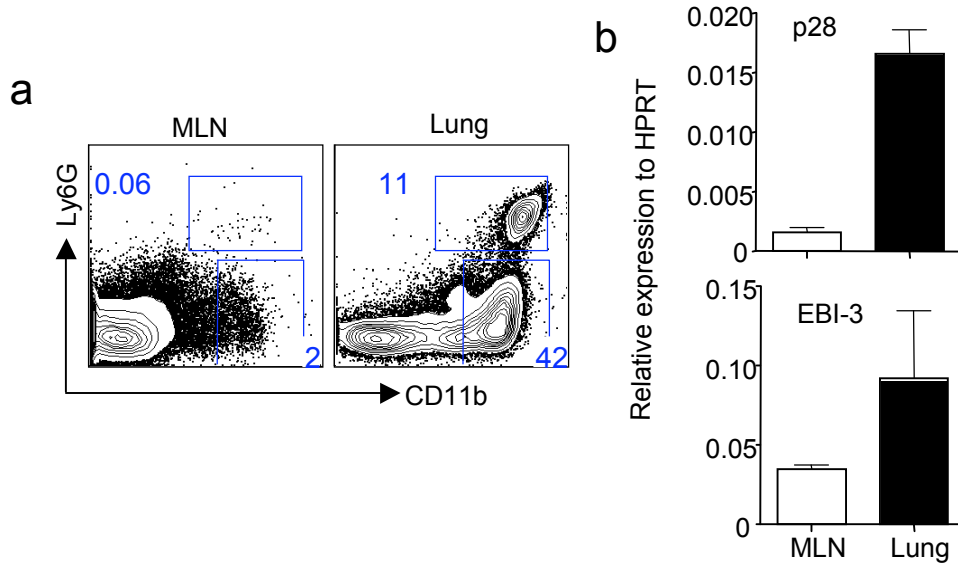
## Supplementary Figure 16



### Supplementary Figure 16. IL-2 and IL-27 co-operatively induce IL-10 expression by CD8<sup>+</sup> CTL at the effector stage.

V-OT-I cells or OT-II cells were activated separately by influenza-OVA infected DC for 4d. The effector V-OT-I were left alone or mixed together with effector OT-II cells. The cells were then stimulated with plate-bound aCD3 and treated with various conditions as indicated. (a) The expression of IL-10-eGFP by V-OT-I was measured through flow cytometry. (b). The MFI of IL-10-eGFP by V-OT-I cells is depicted.

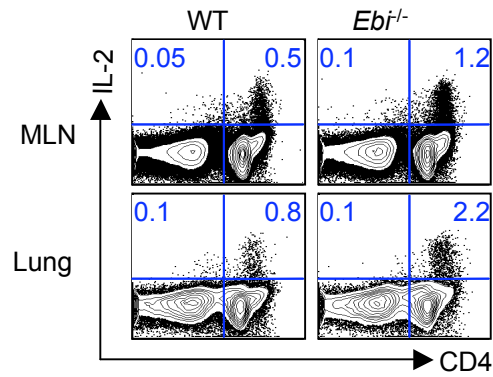
## Supplementary Figure 17



### **Supplementary Figure 17. Neutrophils, monocytes/macrophages and IL-27 message are highly enriched in the influenza infected lungs.**

WT mice were infected with influenza. (a) At d5 p.i., the percentages of neutrophils ( $CD11b^+ Ly6G^+$ ) and monocytes/macrophages ( $CD11b^+ Ly6G^-$ ) were evaluated by flow cytometry. (b) At d5 p.i., the expression of IL-27 subunits p28 and EBI-3 by lung cells were determined through quantitative RT-PCR.

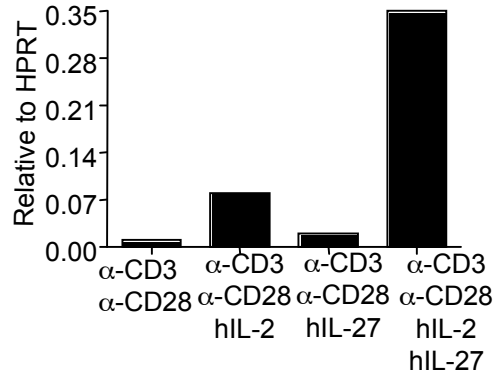
## Supplementary Figure 18



### Supplementary Figure 18. EBI-3 deficiency does not impair IL-2 production by T cells

WT or *Ebi3*<sup>-/-</sup> mice were infected with influenza. At d6 p.i., the production of IL-2 in gated Thy1<sup>+</sup> cells was measured by ICS following restimulation with influenza-infected BMDC.

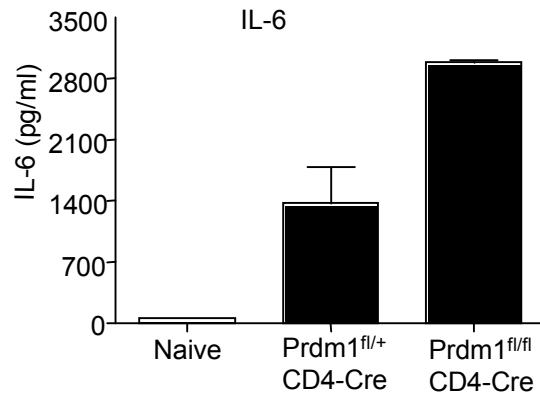
## Supplementary Figure 19



### **Supplementary Figure 19. IL-2 and IL-27 synergistically induce IL-10 expression by human CTL**

Purified human CD8<sup>+</sup> T cells were stimulated with  $\alpha$ -CD3 plus  $\alpha$ -CD28 with indicated conditions for 3d. The expression of IL-10 by CD8<sup>+</sup> T cells was subsequently determined by quantitative RT-PCR.

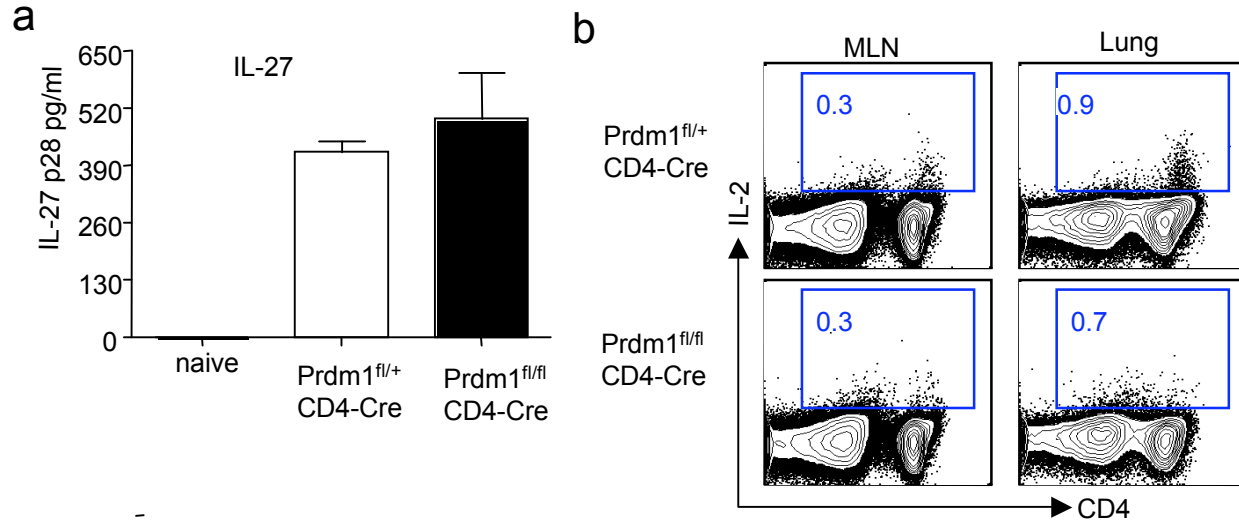
## Supplementary Figure 20



**Supplementary Figure 20. Conditional deletion of Blimp-1 in T cells results in increased IL-6 release in the airway.**

CD4-Cre *Prdm1*<sup>fl/+</sup> or CD4-Cre *Prdm1*<sup>fl/fl</sup> mice were infected with influenza. At d9 p.i., the levels of IL-6 in BALF were determined through ELISA.

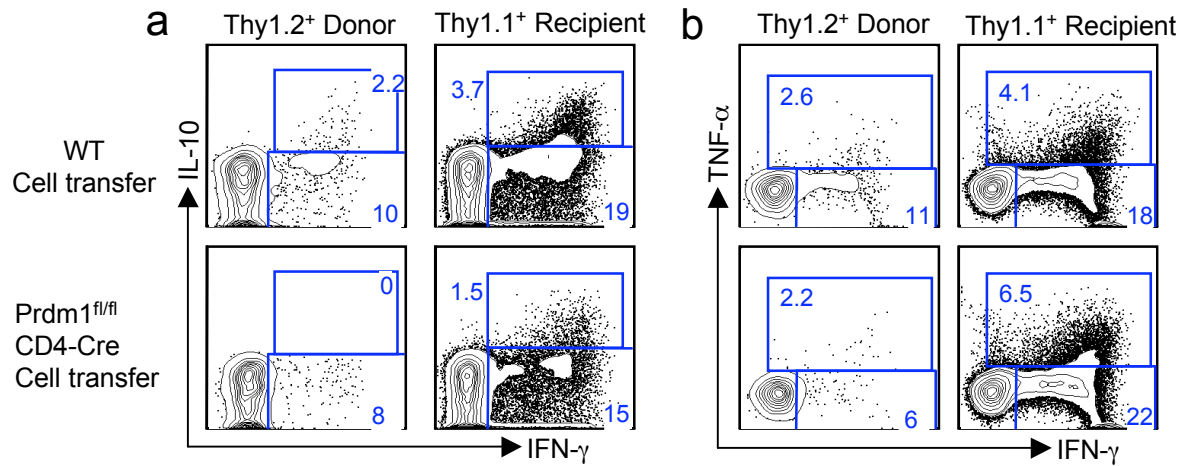
## Supplementary Figure 21



**Supplementary Figure 21. Conditional deletion of Blimp-1 in T cells results in comparable IL-27 release in the airway and moderately diminished IL-2 production by T cells following influenza infection.**

CD4-Cre *Prdm1*<sup>fl/+</sup> or CD4-Cre *Prdm1*<sup>fl/fl</sup> mice were infected with influenza. (a) At d7 p.i., the levels of IL-27 p28 in BALF were determined through ELISA. (b) The production of IL-2 by T cells (Thy1<sup>+</sup>) was measured by ICS following restimulation with influenza-infected BMDC.

## Supplementary Figure 22



### Supplementary Figure 22. The requirement of Blimp-1 in IL-10 production by CTL is CD8<sup>+</sup> T cell intrinsic.

WT or CD4-Cre *Prdm1*<sup>fl/fl</sup> T cells were transferred into Thy1 mis-matched WT mice and infected with influenza. At d7 p.i., the production of IL-10 and IFN- $\gamma$  (a) or TNF and IFN- $\gamma$  (b) by CTL was measured by ICS following restimulation with influenza-infected BMDC.