

Supporting Information

Intercalative Conformations of the 14*R* (+)- and 14*S* (-)-*trans-anti*-DB[*a,l*]P-*N*⁶-dA Adducts: Molecular Modeling and MD Simulations

*Yuqin Cai*¹, *Shuang Ding*¹, *Nicholas E. Geacintov*^{2,*} and *Suse Broyde*^{1,*}

Department of ¹Biology and ²Chemistry, New York University, New York, N.Y. 10003,
U.S.A.

***Corresponding authors:** Nicholas E. Geacintov, nicholas.geacintov@nyu.edu and
Suse Broyde, broyde@nyu.edu

Running Title: NER of DB[*a,l*]P-Adenine Adducts

Keywords: Nucleotide excision repair, conformational heterogeneity, dibenzo[*a,l*]pyrene, MD simulations

MD computation protocols. Our computational protocols follow recommendations of the AMBER development team (<http://amber.scripps.edu/tutorials/basic/tutorial1/section5.htm>). First, the starting models were subjected to 50 steps of conjugate gradient energy minimization, using a distance dependent dielectric function with dielectric constant 4.0. Then each system was neutralized with 20 Na⁺ counterions and solvated with explicit water using the LEAP Module of the AMBER 9 simulation package (1). A periodic rectangular box of TIP3P water (2) with 10.0 Å buffer was created around the DNA for each sequence. The Particle-Mesh Ewald (3, 4) method with 9.0 Å cutoff for the non-bonded interactions was used in subsequent energy minimizations and MD simulations.

Subsequently, 500 steps of steepest descent minimization followed by 500 cycles of conjugate gradient minimization were conducted for the waters and counterions with 500 kcal·mol⁻¹·Å⁻² restraint on the DNA. Then, 500 steps of conjugate gradient minimization were carried out for the whole system without restraints. A 2.0 fs time step and the SHAKE algorithm (5) were applied in the MD simulations. All other parameters were default values in the AMBER 9 simulation package. The system was heated from 0 K to 300 K over 20 ps with the DNA fixed with a weak restraint of 10.0 kcal·mol⁻¹·Å⁻² at constant volume, using the Berendsen coupling algorithm (6) with a 1.0 ps coupling parameter. A 60 ps of constant pressure dynamics at 300 K followed to further equilibrate the system. Finally, a 30.0 ns production was conducted at a temperature of 300 K and constant pressure of 1 Atm. Temperature and pressure coupling constants were both 1.0 ps.

Best representative structures. The best representative structure in the MD trajectory for each adduct was obtained by using the cluster analysis option in MOIL-View (7). The best representative structure is a real frame from the ensemble; as determined by a cluster analysis, it is the most populated conformation or the conformation that is the closest to all other snapshots in an ensemble.

Hydrogen bond quality index analyses. We employed our hydrogen bond quality index (HBI) (8), to quantitatively assess the quality of Watson-Crick hydrogen bonding, in terms of the deviation from ideal Watson-Crick hydrogen bond distances and angles:

$$I_H = \sum_{D-H \cdots A} [(d_{DA} - d_{DA}^0)^2 + (1 + \cos \gamma)^2]$$

where d_{DA} is the instantaneous donor-acceptor distance, d_{DA}^0 is an ideal donor-acceptor distance (9) [O6 (G) to N4 (C) is 2.91 Å, N1 (G) to N3 (C) is 2.95 Å, and N2 (G) to O2 (C) is 2.86 Å] and γ is the instantaneous donor-hydrogen…acceptor (D-H…A) hydrogen bond angle with an ideal value of 180 °. The summation is over all the Watson-Crick hydrogen bonds in a base pair over the trajectory. The lower the value of I_H , the better the quality of the Watson-Crick hydrogen bonding. I_H is zero when the Watson-Crick hydrogen bonding is ideal during the dynamics. However, in real, even unmodified DNA, sequence-dependent deviations from ideal Watson-Crick hydrogen bonding are normal (10). Therefore, comparing the I_H value of a modified step to its analogue step in the unmodified control duplex provides an estimate of the perturbation of its Watson-Crick hydrogen bond upon modification.

Table S1. Ensemble average values with standard deviations (given in parentheses) for structural properties at the lesion site.

	<i>I4R</i> (+)	<i>I4S</i> (-)	Unmodified
Buckle (°)	-26.4 (7.9)	28.2 (7.5)	4.4 (11.1)
Propeller (°)	21.4 (8.9)	-24.5 (7.6)	-6.8 (9.1)
α' ^a (°)	-13.7 (8.4)	15.6 (7.8)	N/A
β' (°)	103.6 (7.7)	-100.9 (7.2)	N/A
γ' (°)	-62.7 (4.9)	61.5 (5.3)	N/A
δ' (°)	24.2 (6.2)	-23.3 (6.4)	N/A
Twist (°)	17.1 (8.5)	-6.6 (10.7)	32.0 (5.4)
Roll (°)	2.9 (8.9)	23.6 (7.1)	-4.2 (6.9)
Rise (Å)	7.6 (0.4)	7.1 (0.4)	3.3 (0.5) 3.4 (0.4)
Tilt (°)	-10.5 (5.7)	-16.0 (4.4)	0.4 (5.7) 1.8 (5.1)
Opening (Å)	12.5 (8.0)	11.3 (6.2)	2.2 (6.2)
χ (°)	-113.0 (18.9)	-58.0 (15.7)	-95.8 (29.5)
Total vdW interaction energy (kcal/mol) with the intercalation pocket	-28.1 (1.7)	-25.0 (2.0)	N/A

^a For α' , we give the values for *R* and *S* as -13.7 (8.4) ° and 15.6 (7.8) °, respectively, to emphasize their inversion. These equal the respective values of 166.3 (8.4) ° and 195.6 (7.8) °, given in Figure S3, Supporting Information.

Table S2. Ensemble average values of duplex helicoidal parameters with standard deviations (in parenthesis) for all cases. The steps highlighted in yellow are the ones where the lesion causes maximum distortion.

	14R (+)	14S (-)	Unmodified
Twist (degrees)			
Step 3	27.9 (7.2)	28.0 (5.1)	30.3 (4.4)
Step 4	16.1 (13.1)	20.5 (7.3)	36.9 (5.3)
Step 5	34.2 (8.4)	46.3 (8.0)	23.9 (10.7)
Step 6	17.1 (8.5)	-6.6 (10.7)	32.0 (5.4)
Step 7	28.9 (7.7)	25.1 (5.8)	34.3 (6.2)
Step 8	34.9 (6.4)	34.8 (6.5)	32.8 (6.9)
Step 9	28.2 (8.8)	29.9 (8.2)	29.1 (4.8)
Roll (degrees)			
Step 3	-0.8 (7.0)	-1.3 (5.5)	-2.9 (6.9)
Step 4	1.0 (6.2)	3.0 (5.4)	-1.6 (7.6)
Step 5	8.1 (9.0)	-6.2 (6.3)	10.5 (7.2)
Step 6	2.9 (8.9)	23.6 (7.1)	-4.2 (6.9)
Step 7	-4.7 (8.4)	-3.9 (7.9)	2.1 (6.7)
Step 8	4.7 (9.9)	6.2 (11.1)	3.4 (8.3)
Step 9	-3.3 (7.6)	-0.5 (6.8)	-10.6 (7.6)
Buckle (degrees)			
Step 3	-5.7 (10.8)	-4.6 (9.7)	-6.8 (10.5)
Step 4	3.7 (9.2)	5.3 (10.1)	-2.9 (10.9)
Step 5	10.9 (10.3)	16.9 (9.8)	-4.6 (11.4)
Step 6	-26.4 (7.9)	28.2 (7.5)	4.4 (11.1)
Step 7	-2.7 (10.6)	-10.5 (9.7)	5.9 (10.3)
Step 8	5.1 (11.1)	3.8 (10.4)	-8.8 (11.8)
Step 9	-1.4 (12.6)	-0.7 (12.4)	-8.6 (11.4)
Propeller (degrees)			

Step 3	-11.4 (8.6)	-10.5 (8.6)	-13.9 (8.8)
Step 4	-10.9 (8.3)	-6.8 (8.2)	-13.8 (8.1)
Step 5	-13.8 (7.5)	-10.3 (8.1)	-6.1 (9.7)
Step 6	21.4 (8.9)	-24.5 (7.6)	-6.8 (9.1)
Step 7	8.2 (10.7)	16.4 (8.6)	-13.4 (9.0)
Step 8	-12.6 (9.5)	-7.5 (9.5)	-21.1 (11.0)
Step 9	-13.6 (9.9)	-12.8 (9.5)	-7.2 (8.4)

Opening (degrees)

Step 3	1.5 (11.3)	3.4 (5.4)	3.9 (5.7)
Step 4	0.2 (7.0)	3.4 (5.6)	1.4 (5.9)
Step 5	-2.0 (4.2)	0.12 (3.7)	1.6 (3.8)
Step 6	12.5 (8.0)	10.4 (6.0)	2.2 (6.2)
Step 7	2.7 (4.6)	1.5 (6.3)	1.4 (4.1)
Step 8	0.9 (6.2)	1.5 (6.3)	9.4 (9.5)
Step 9	4.1 (6.3)	4.9 (5.8)	0.6 (6.6)

Rise (degrees)

Step 3	3.2 (0.5)	3.1 (0.3)	3.2 (0.4)
Step 4	3.3 (0.4)	3.2 (0.3)	3.4 (0.4)
Step 5	7.6 (0.4)	3.5 (0.3)	3.3 (0.5)
Step 6	3.0 (0.4)	7.1 (0.4)	3.4 (0.4)
Step 7	3.3 (0.5)	3.2 (0.3)	3.6 (0.4)
Step 8	3.5 (0.5)	3.4 (0.4)	3.2 (0.5)
Step 9	3.2 (0.5)	3.3 (0.5)	2.9 (0.5)

Tilt (degrees)

Step 3	3.9 (6.3)	1.3 (4.3)	1.3 (5.1)
Step 4	9.5 (5.7)	3.4 (4.6)	5.1 (6.0)
Step 5	-10.5 (5.7)	8.5 (5.0)	0.4 (5.7)
Step 6	2.0 (4.6)	-16.0 (4.4)	1.8 (5.1)

Step 7	7.5 (5.6)	0.0 (5.0)	7.7 (6.4)
Step 8	6.6 (7.1)	3.1 (5.9)	2.8 (7.0)
Step 9	2.7 (8.9)	-0.3 (4.5)	3.7 (6.6)

Table S3 (a). Ensemble average values with standard deviations (in parentheses) for minor groove widths (\AA)

	14R (+)	14S (-)	Unmodified
P6-P21	8.2 (3.5)	6.9 (1.7)	6.4 (1.6)
P7-P20	10.8 (3.5)	9.5 (1.5)	7.2 (1.8)
P8-P19	13 (1.7)	12.6 (2.9)	7.0 (1.5)
P9-18	9.3 (1.9)	12.1 (2.9)	5.5 (1.5)
P10-P17	7.2 (1.8)	9.4 (1.9)	6.3 (1.6)
Total	48.5	50.5	32.4

Table S3 (b). Ensemble average values with standard deviations (in parentheses) for major groove widths (\AA)

	14R (+)	14S (-)	Unmodified
P3-P17	17.2 (1.9)	16.4 (1.6)	14.4 (1.7)
P4-P16	17.2 (1.5)	18.7 (1.4)	13.4 (2.0)
P5-P15	16.8 (1.9)	17.8 (1.9)	14.5 (1.9)
P6-P14	15.9 (1.7)	17.6 (1.8)	14.4 (1.9)
Total	67.1	70.5	56.7

Table S4 (a) Ensemble average values with standard deviations (in parentheses) for van der Waals interaction energies between adjacent base pairs and between DB[*a,l*]P and nearby base pairs ^a

Base pair step	Van der Waals interaction energy (kcal·mol ⁻¹)		
	14R (+)	14S (-)	Unmodified
A3:T20-T4:A19	-15.2 (1.3)	-15.5 (1.0)	-15.5 (0.9)
T4:A19-C5:G18	-14.9 (1.0)	-14.9 (1.0)	-14.9 (1.0)
C5:G18-A6*:T17	-1.4 (0.4)	-12.8 (1.0)	-14.5 (1.3)
A6*:T17-C7:G16	-14.3 (1.4)	-3.2 (0.6)	-15.6 (1.0)
C7:G16-T8:A15	-14.7 (1.1)	-14.4 (1.2)	-14.8 (1.0)
T8:A15-A9:T14	-13.9 (1.1)	-13.9 (1.1)	-14.2 (1.0)
C5:G18-DB[<i>a,l</i>]P	-15.0 (1.0)	-1.2 (0.1)	
DB[<i>a,l</i>]P-A6*T17	-12.0 (1.3)	-11.8 (1.3)	
DB[<i>a,l</i>]P-C7:G16	-1.1 (0.2)	-12.1 (1.6)	
Total	-103	-99.8	-89.5
Difference between modified and unmodified duplexes	-13.5	-10.3	

^a Only the aromatic rings are considered.

^b Note that the ~3 kcal/mol difference between the 14R (+) and 14S (-) adducts is at the interactions highlighted in yellow.

Table S4 (b) Ensemble average values with standard deviations (in parentheses) for van der Waals interaction energies (kcal·mol⁻¹) between the DB[*a,l*]P aromatic ring system and adjacent bases in the intercalation pocket^{a,b}

	14R (+)	14S (-)
DB[<i>a,l</i>]P-C5	-5.5 (1.1)	-0.6 (0.1)
DB[<i>a,l</i>]P-A6*	-3.1 (1.1)	-2.9 (1.1)
DB[<i>a,l</i>]P-C7	-0.3 (0.1)	-2.2 (0.7)
DB[<i>a,l</i>]P-G16	-0.8 (0.2)	-9.8 (1.1)
DB[<i>a,l</i>]P-T17	-9.0 (0.7)	-8.9 (0.7)
DB[<i>a,l</i>]P-G18	-9.4 (1.0)	-0.5 (0.1)
Total	-28.1 (1.7)	-25.0 (2.0)
DB[<i>a,l</i>]P-(A6* and T17)^c	-12.1 (1.3)	-11.8 (1.3)

^a Only the aromatic rings are considered.

^b Note that the ~3 kcal/mol difference between the 14R (+) and 14S (-) adducts is at the interactions highlighted in yellow.

^c Note that the entry DB[*a,l*]P-(A6* and T17) shows the interaction energies between the DB[*a,l*]P aromatic rings and the damaged base pair A6:T17 (Figures 6a and 6b), and are derived from the sum of DB[*a,l*]P-A6* and DB[*a,l*]P-T17.

Table S5 (a) Hydrogen bond occupancies (%)^a

		14R (+)	14S (-)	Unmodified
A3:T20	N6-H61...O4	93.7	93.4	93.6
	N3-H3...N1	99.4	95.3	99.3
T4:A19	N3-H3...N1	98.6	96.6	97.6
	N6-H62...O4	91.9	93.4	94.6
C5:G18	N4-H41...O6	98.1	98	97.3
	N2-H21...O2	99.9	99.4	99.7
	N1-H1...N3	99.8	99.9	99.9
A6:T17	N-HN...O4	95.6	97.7	95.2
	N3-H3...N1	98.4	98.6	99.2
C7:G16	N4-H41...O6	98.1	97.5	95.8
	N2-H21...O2	99.3	99.5	99.4
	N1-H1...N3	98.9	99.6	99.6
T8:A15	N3-H3...N1	97.5	98.1	99.1
	N6-H62...O4	96.4	95.3	79
A9:T14	N6-H61...O4	90.7	90.2	96.6
	N3-H3...N1	99.3	99.3	95.4
<hr/>				
Extra HBs				
	O13-HO13...N7 (A6)	92.6	30.6	
	(C7)N4-H42...O12	68.9	45.0	

^a Criteria for hydrogen bonding: heavy atom-heavy atom distance < 3.3 Å, and donor-hydrogen-acceptor angle > 140 °.

Table S5 (b) Hydrogen bond quality for the Watson-Crick base pairs adjacent to the lesion site and their corresponding unmodified control base pairs.

HBI	14R (+)	14S (-)	Unmodified
C5:G18	318	321	343
A6:T17	375	397	416
C7:G16	357	956	390

Table S6a. Partial charges, atom types and topologies for the 14R (+)- and 14S (-)-*trans-anti*-DB[*a,l*]P-*N*⁶-dA adducts

	Atom name	Atom type	Topology	Partial charge	
				14R (+)	14S (-)
1	P	P	M	1.2553	1.2687
2	O1P	O2	E	-0.795	-0.795
3	O2P	O2	E	-0.795	-0.795
4	O5'	OS	M	-0.477	-0.476
5	C5'	CT	M	-0.115	-0.169
6	H5'1	H1	E	0.1096	0.1079
7	H5'2	H1	E	0.1096	0.1079
8	C4'	CT	M	0.2273	0.3142
9	H4'	H1	E	0.1026	0.0639
10	O4'	OS	S	-0.426	-0.451
11	C1'	CT	B	-0.029	0.2203
12	H1'	H2	E	0.17	0.1191
13	N9	N*	S	-0.029	-0.081
14	C8	CK	B	0.03	0.1243
15	H8	H5	E	0.1868	0.1856
16	N7	NB	S	-0.405	-0.661
17	C5	CB	S	0.0547	0.0524
18	C6	CA	B	0.3264	0.4648
19	N	N2	B	-0.245	-0.535
20	HN	H	E	0.2853	0.3617
21	C14	CT	B	-0.014	-0.025
22	HC14	H1	E	0.2105	0.1766

23	C13	CT	3	0.0083	0.0299
24	HC13	H1	E	0.112	0.1574
25	O13	OH	S	-0.604	-0.625
26	HO13	HO	E	0.4232	0.3875
27	C12	CT	3	0.1817	0.2376
28	HC12	H1	E	0.1019	0.0834
29	O12	OH	S	-0.662	-0.659
30	HO12	HO	E	0.4314	0.4245
31	C11	CT	3	0.1404	0.1874
32	HC11	H1	E	0.0712	0.0222
33	O11	OH	S	-0.695	-0.637
34	HO11	HO	E	0.4424	0.4095
35	C16	CA	S	-0.041	-0.019
36	C10	CA	B	-0.256	-0.11
37	HC10	HA	E	0.1545	0.134
38	C17	CA	B	0.0666	0.0099
39	C18	CA	E	0.0226	0.0406
40	C9	CA	B	-0.159	-0.192
41	HC9	HA	E	0.1338	0.1575
42	C8	CA	B	-0.226	-0.216
43	HC8	HA	E	0.1593	0.1477
44	C20	CA	B	0.0519	0.0808
45	C21	CA	E	0.1033	0.1306
46	C7	CA	B	-0.185	-0.209
47	HC7	HA	E	0.1499	0.1559
48	C6	CA	B	-0.169	-0.207
49	HC6	HA	E	0.1479	0.1425

50	C5	CA	B	-0.163	-0.155
51	HC5	HA	E	0.1521	0.1894
52	C22	CA	S	-0.019	0.0278
53	C23	CA	S	0.0285	0.0635
54	C4	CA	B	-0.179	-0.217
55	HC4	HA	E	0.135	0.1477
56	C3	CA	B	-0.104	-0.146
57	HC3	HA	E	0.1313	0.1399
58	C2	CA	B	-0.153	-0.146
59	HC2	HA	E	0.1528	0.1255
60	C1	CA	B	-0.15	-0.18
61	HC1	HA	E	0.1249	0.1492
62	C24	CA	S	0.0427	0.0454
63	C19	CA	S	-0.046	-0.087
64	C15	CA	E	0.0147	-0.062
65	N1	NC	S	-0.753	-0.702
66	C2	CQ	B	0.6729	0.6403
67	H2	H5	E	0.0278	0.0355
68	N3	NC	S	-0.775	-0.787
69	C4	CB	E	0.4191	0.4804
70	C3'	CT	M	0.1698	0.2263
71	H3'	H1	E	0.069	0.0398
72	C2'	CT	B	-0.05	-0.056
73	H2'1	HC	E	0.0394	0.0257
74	H2'2	HC	E	0.0394	0.0257
75	O3'	OS	M	-0.472	-0.467

Table S6b. Added force field parameters^a

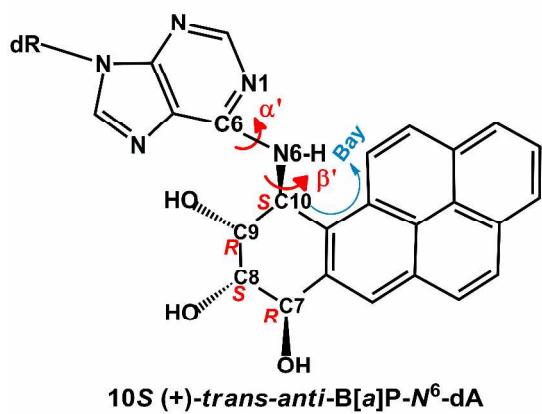
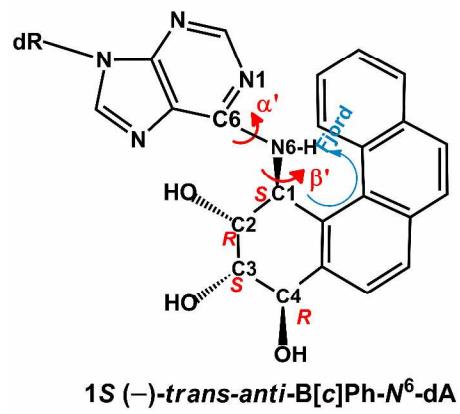
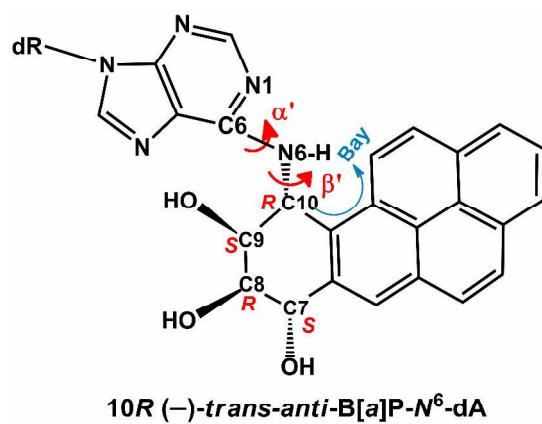
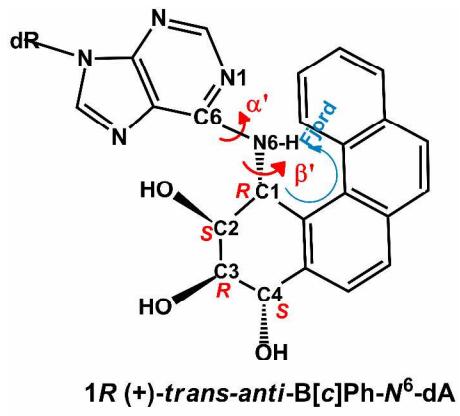
Angle	K _θ (kcal·mol ⁻¹ ·rad ⁻²)	θ _{eq} (deg)
N2-CT-CA	80.0	111.20
H1-CT-CA	50.0	109.50
OH-CT-CA	50.0	109.50

^a Missing angle parameters were added to the force field by analogy to chemically similar atom types already exist in the parm99 parameter set.

Table S7. Box size and number of waters added to the MD simulation starting models

	14R (+)	14S (-)	Unmodified
Box size (Å ³)	51 X 48 X 74	49 X 51 X 69	50 X 51 X 67
Number of waters added	4157	3911	3872

(a)



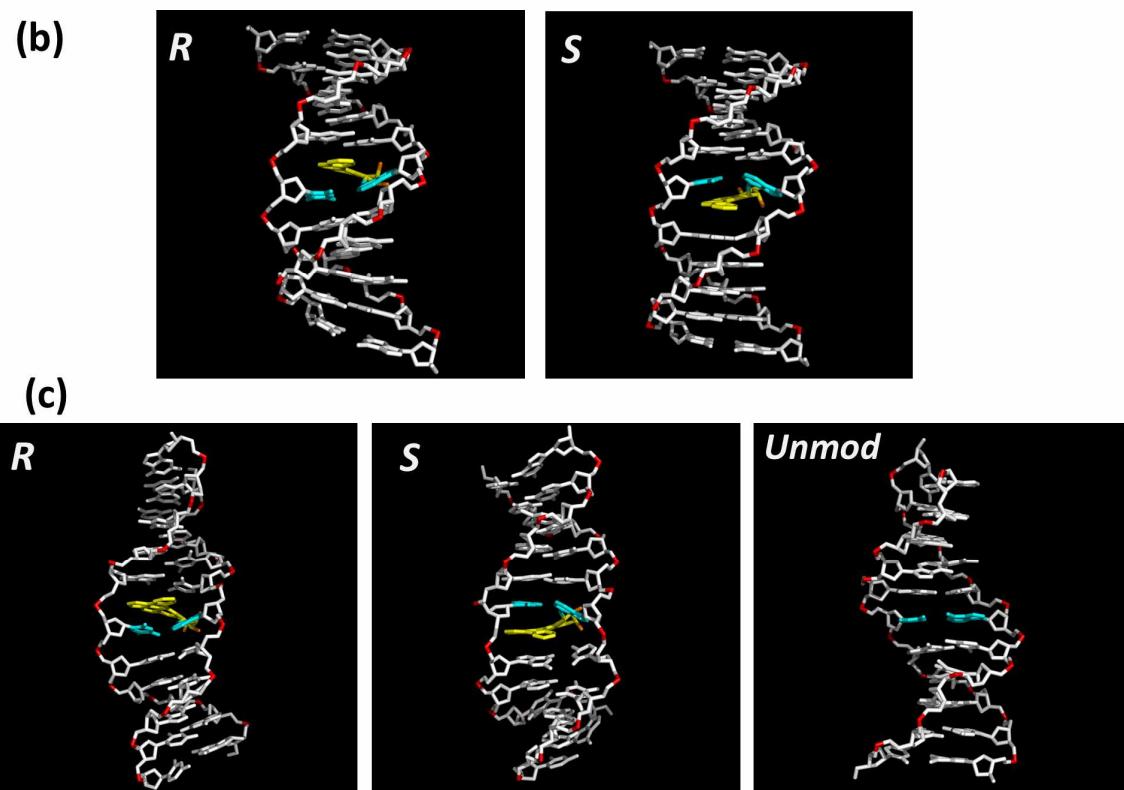


Figure S1. (a) Chemical structures of the 1*R* (+)- and 1*S* (-)-*trans-anti*-B[*c*]Ph-*N*⁶-dA adducts as well as the 10*R* (-)- and 10*S* (+)-*trans-anti*-B[*a*]P-*N*⁶-dA adducts. (b) NMR solution structures of the 11 mer duplexes for the 1*R* (+)- and 1*S* (-)-*trans-anti*-B[*c*]Ph-*N*⁶-dA adducts (*I1*, *I2*) (c) Best representative structures (*7*) of the 11 mer duplexes for the 14*R* (+)- and 14*S* (-)-*trans-anti*-DB[*a,l*]P-*N*⁶-dA adducts, as well as the unmodified control (present work). Views are into the minor grooves.

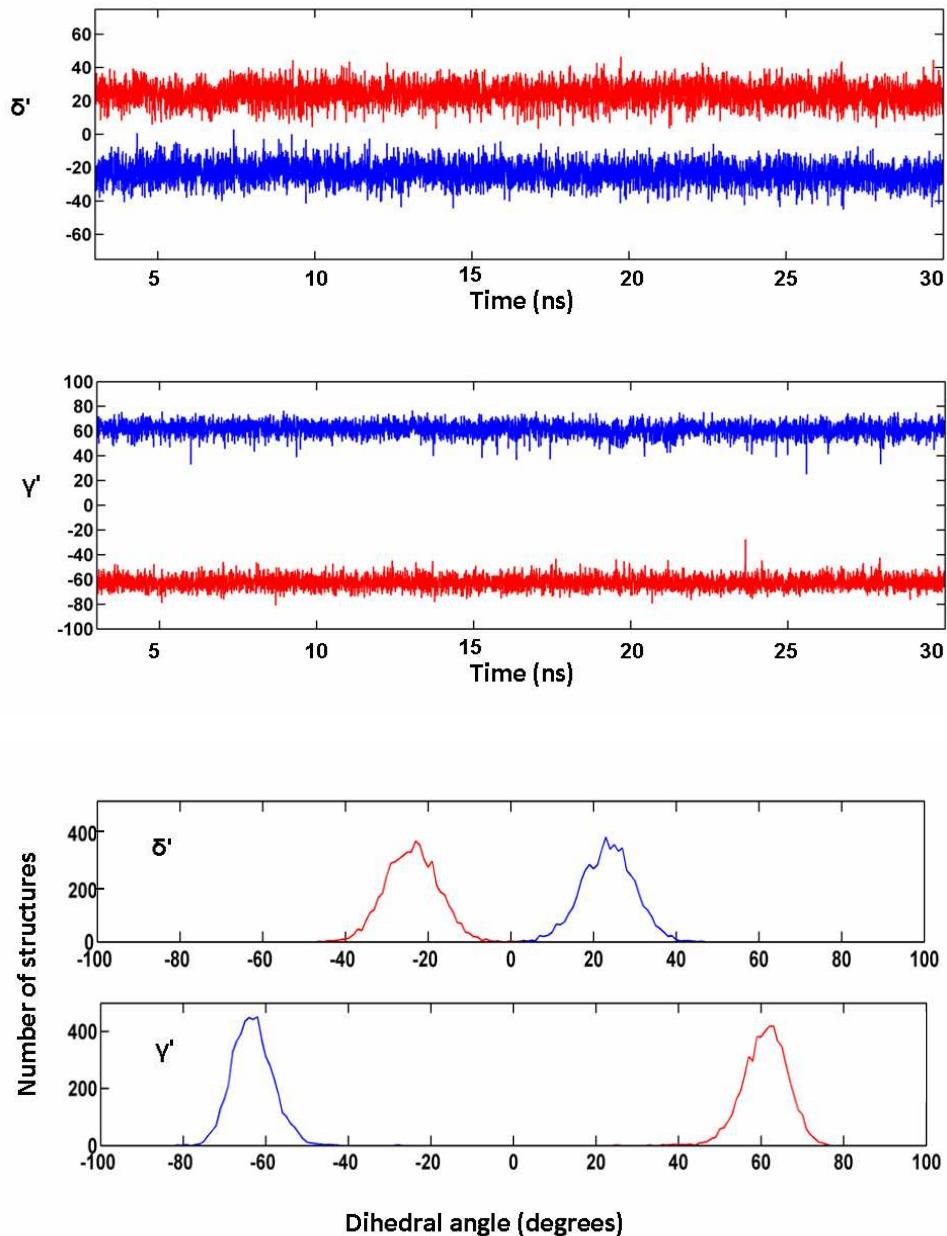


Figure S2. Time-dependence and population distribution of the fjord region twist angle δ' and benzylic ring pucker torsion angle γ' . The 14*R* (+)-adduct is red, the 14*S* (-)-adduct is blue, and the unmodified control duplex is green. Means and standard deviations are given in Table S1.

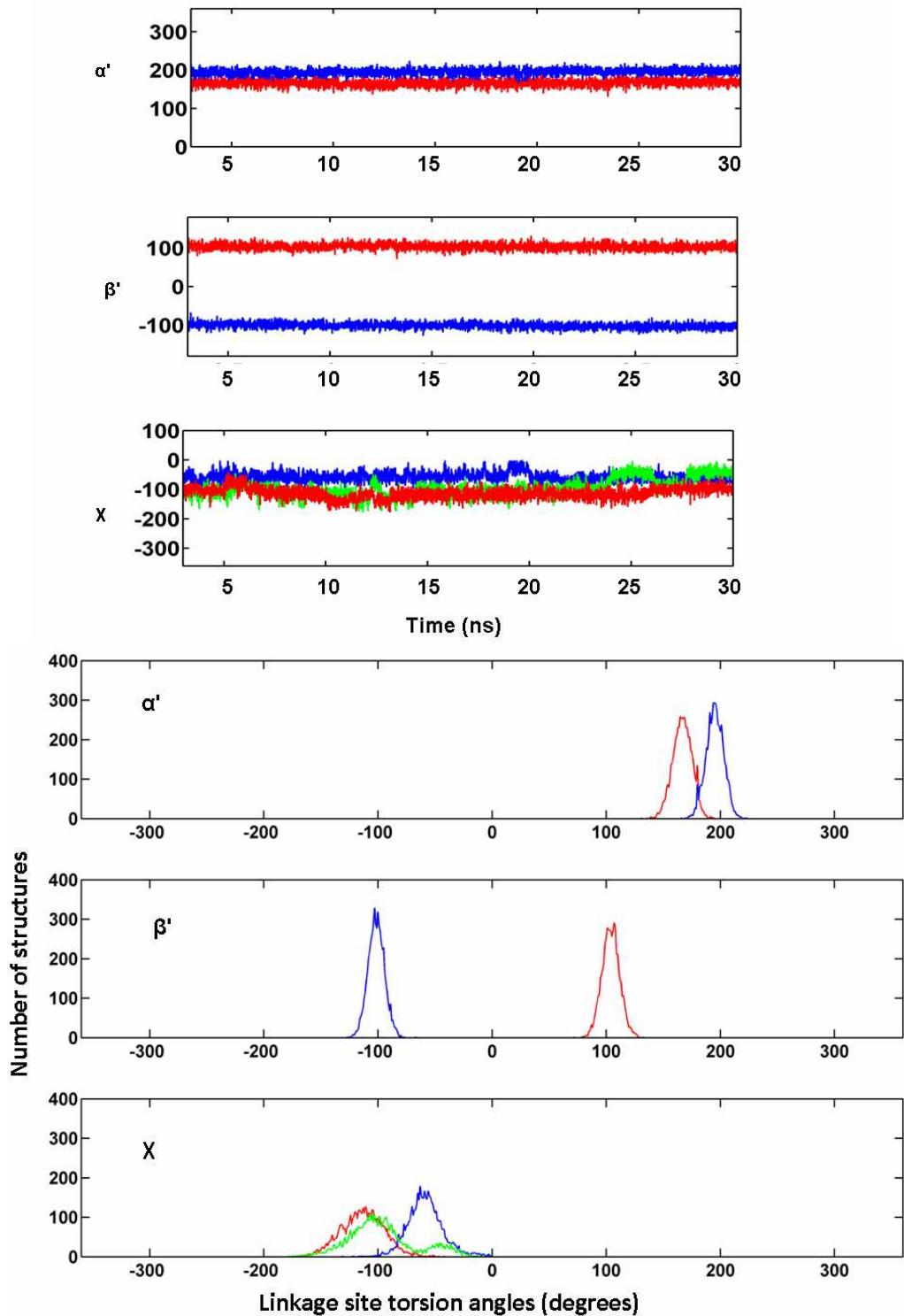
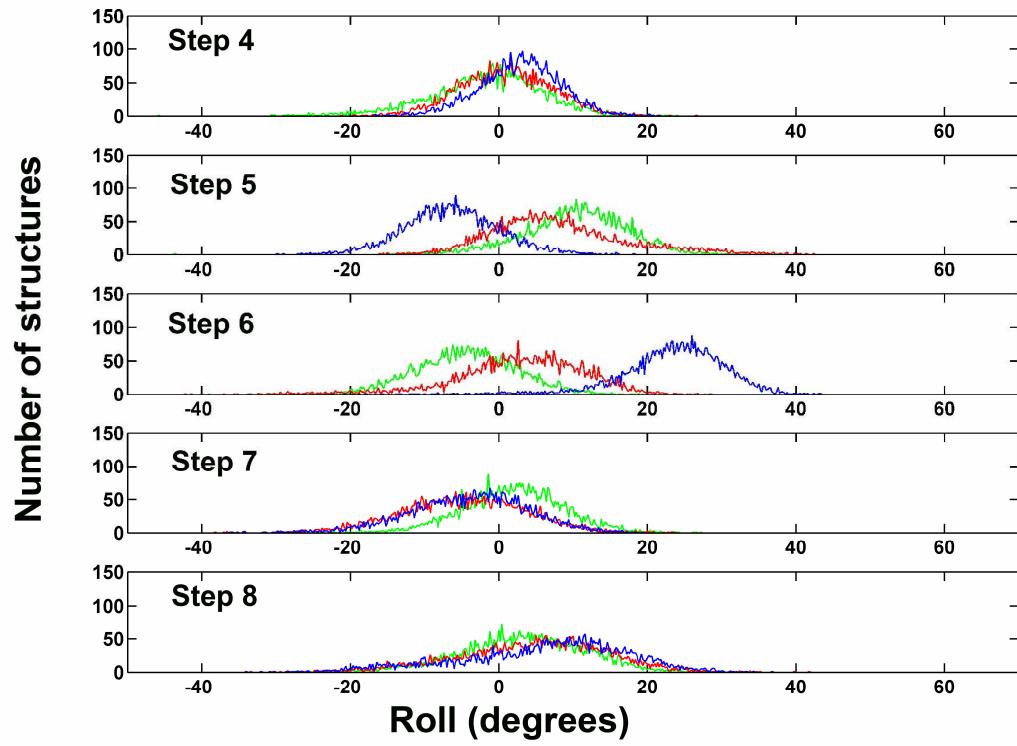
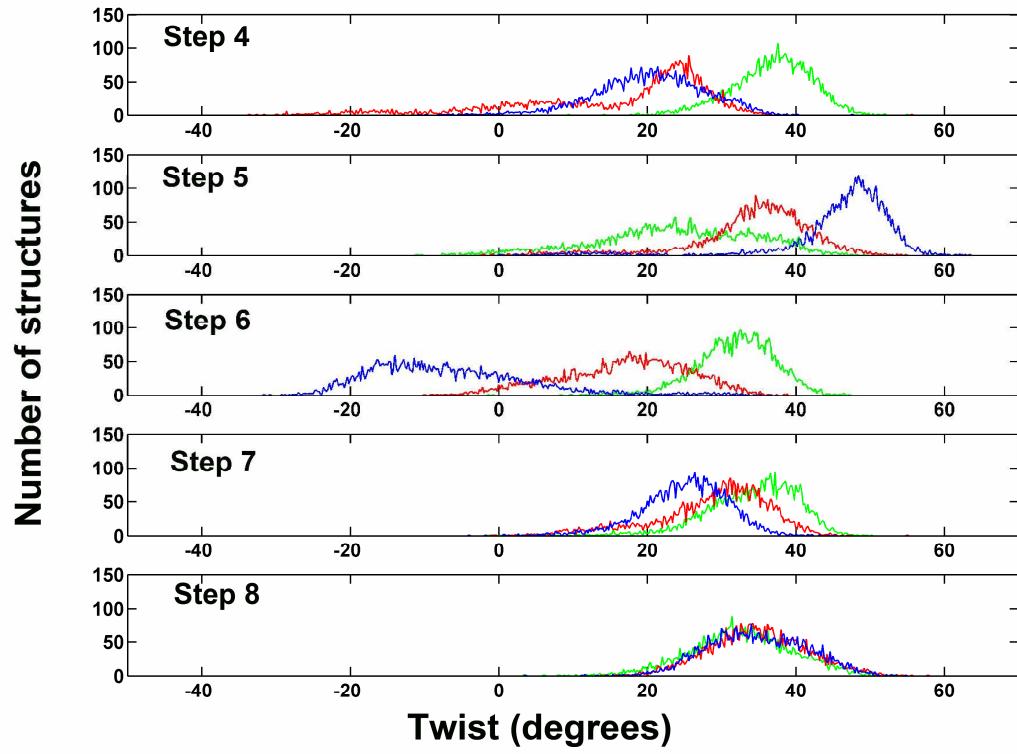
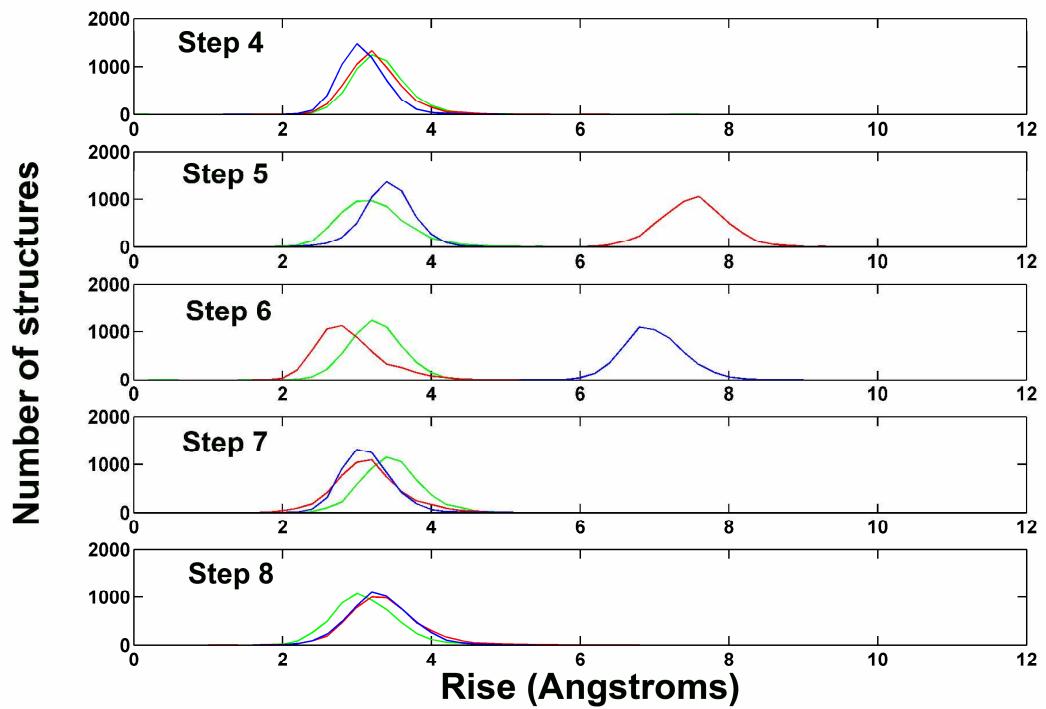
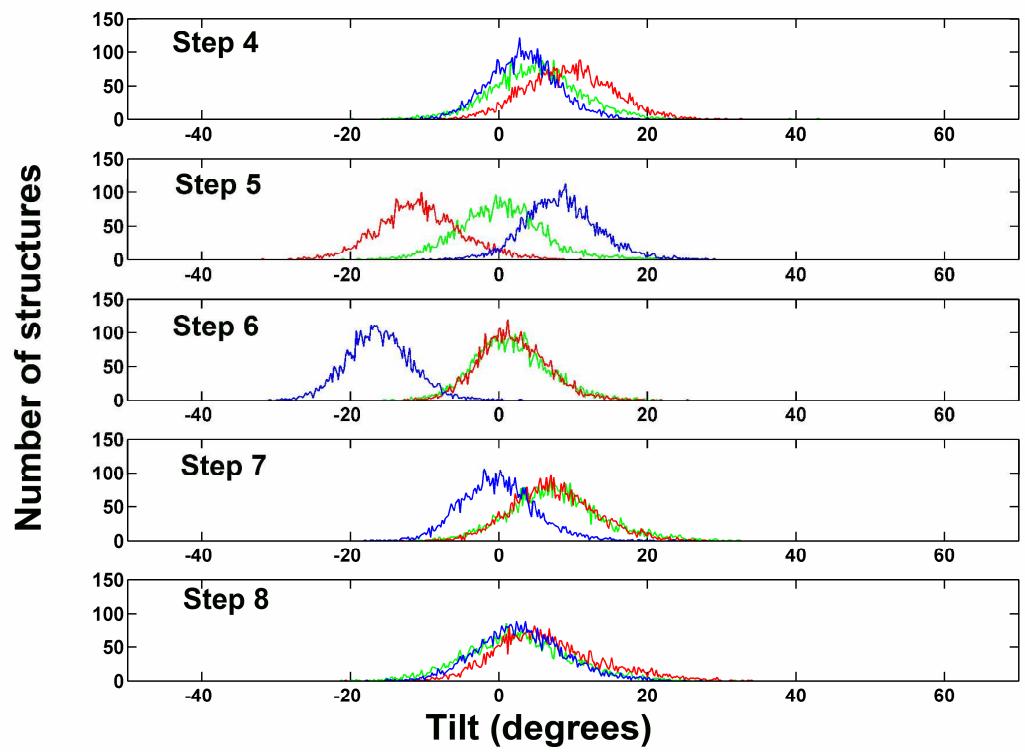
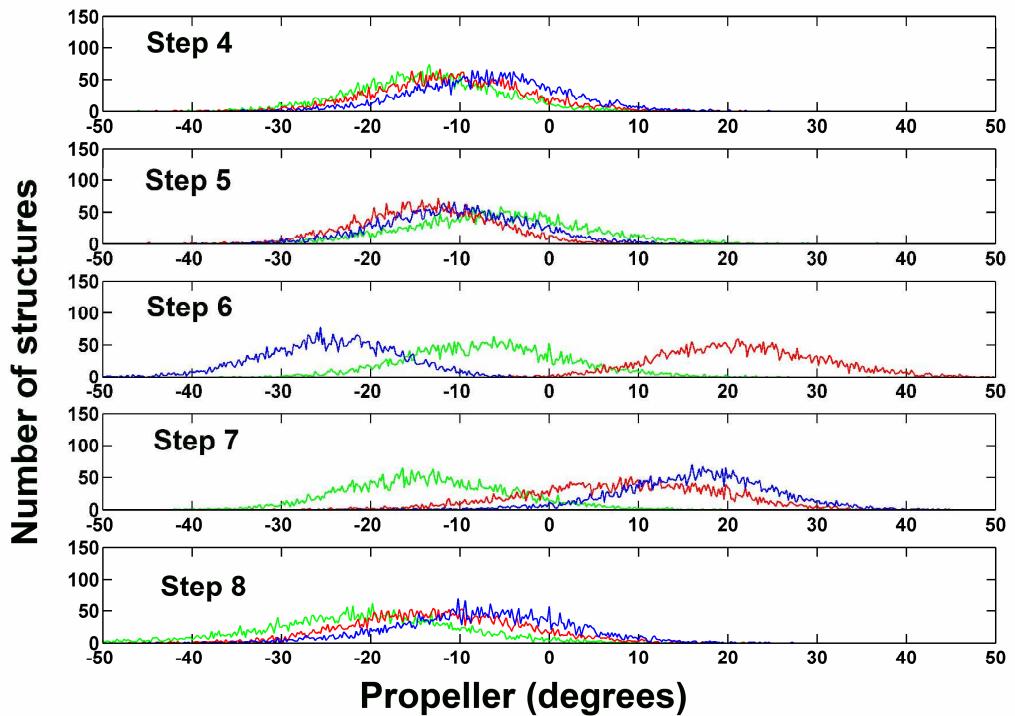
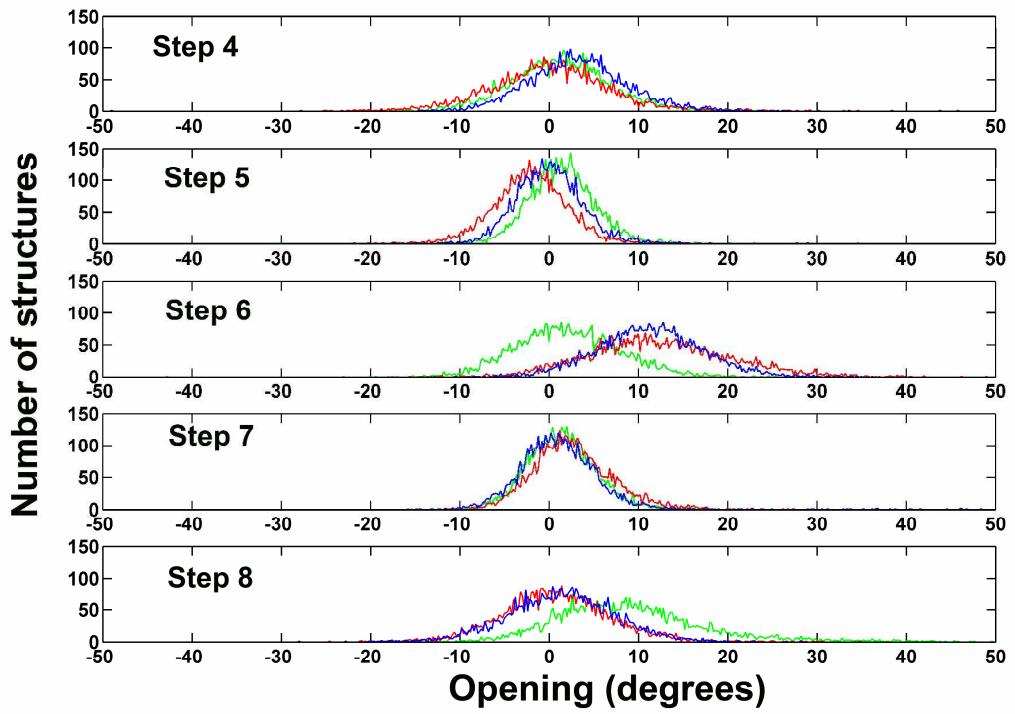


Figure S3. Time-dependence and population distributions of the carcinogen-DNA linkage site and glycosidic torsional angles α' , β' and χ . The 14R (+)-trans-anti-DB[*a,l*]P- N^6 -dA adduct is red, the 14S (-)-trans-anti-DB[*a,l*]P- N^6 -dA adduct is blue, and the unmodified control duplex is green. Means and standard deviations are given in Table S1.







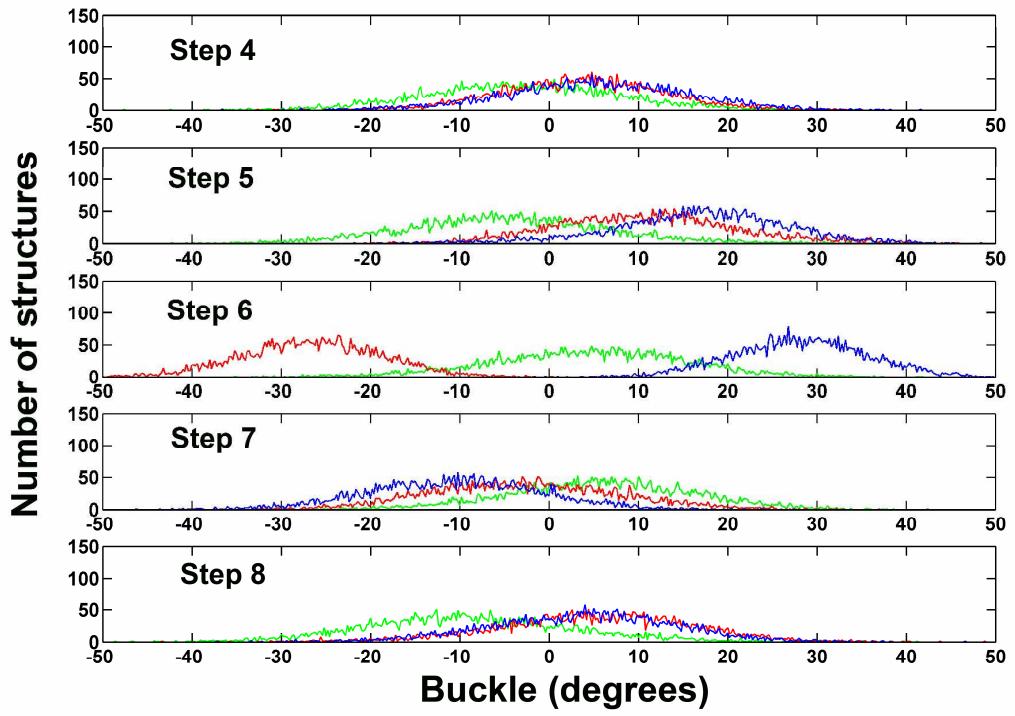
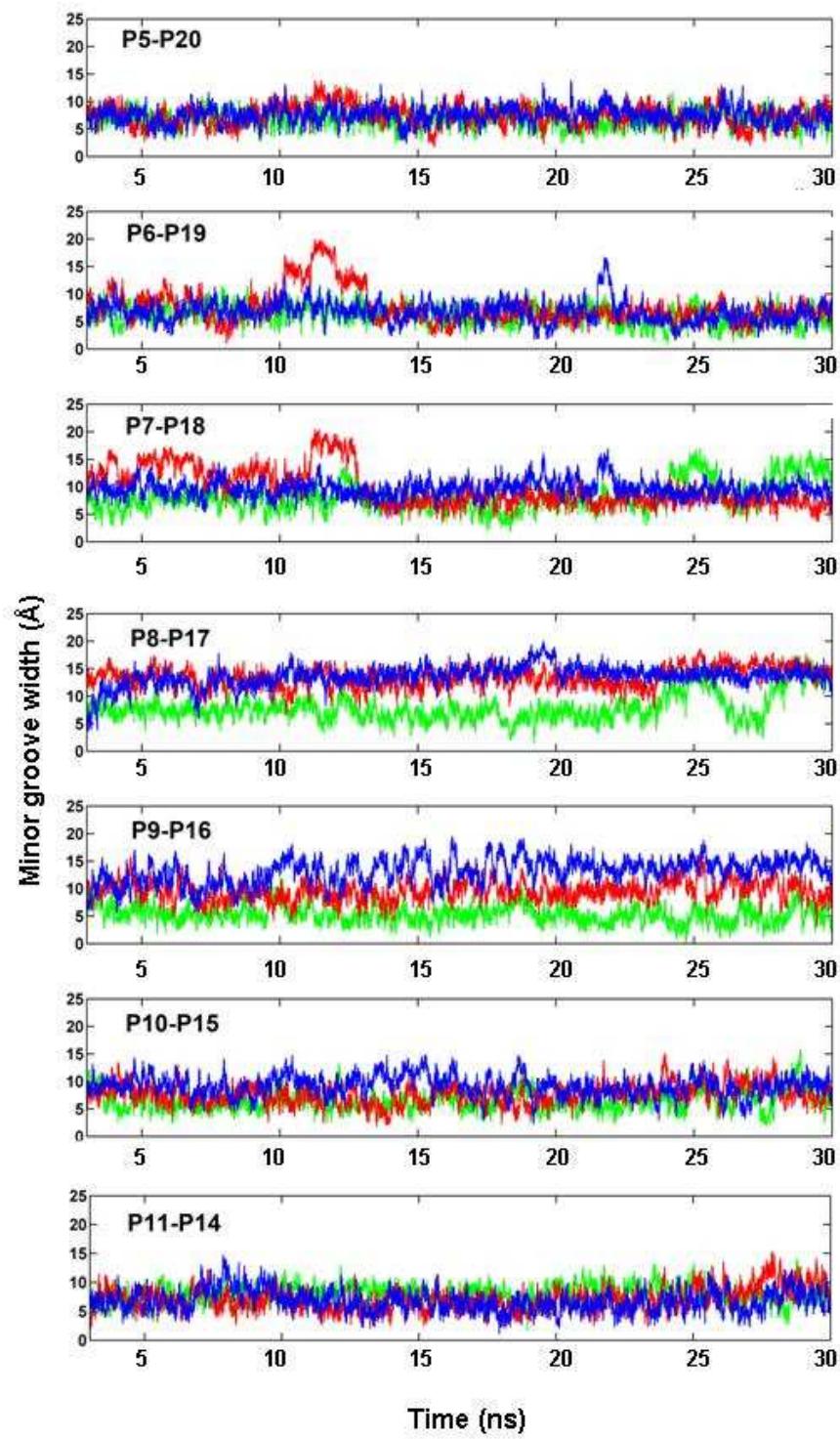


Figure S4. Population distributions of duplex helicoidal parameters at and surrounding the lesion site. The 14R (+)-adduct is red, the 14S (-)-adduct is blue, and the unmodified control duplex is green. Means and standard deviations are given in Table S2.



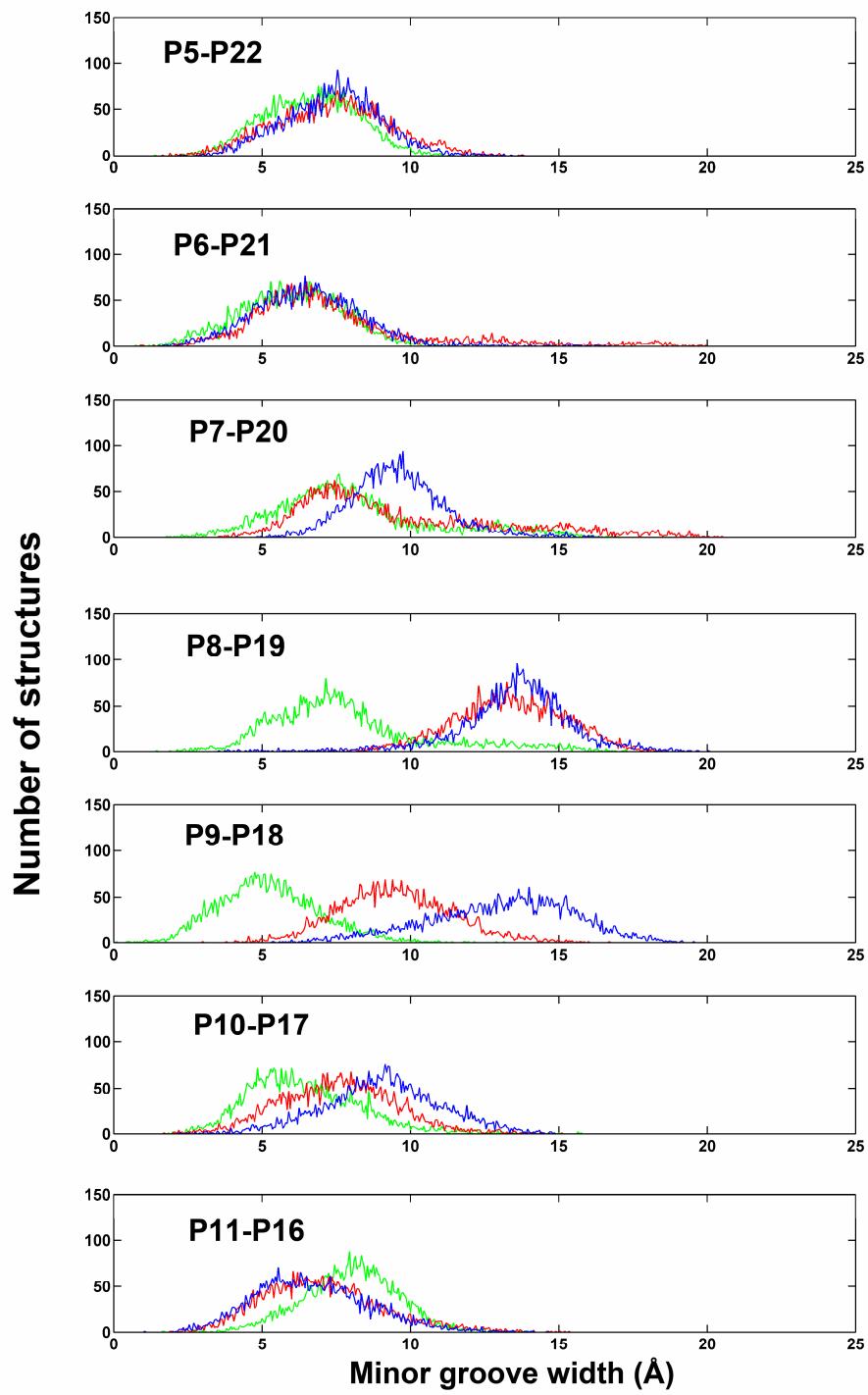


Figure S5. Time-dependence and population distributions of the minor groove widths. The $14R$ (+)-*trans-anti*-DB[*a,l*]P- N^6 -dA adduct is red, the $14S$ (-)-*trans-anti*-DB[*a,l*]P- N^6 -dA adduct is blue, and the unmodified control duplex is green. Means and standard deviations are given in Table S2.

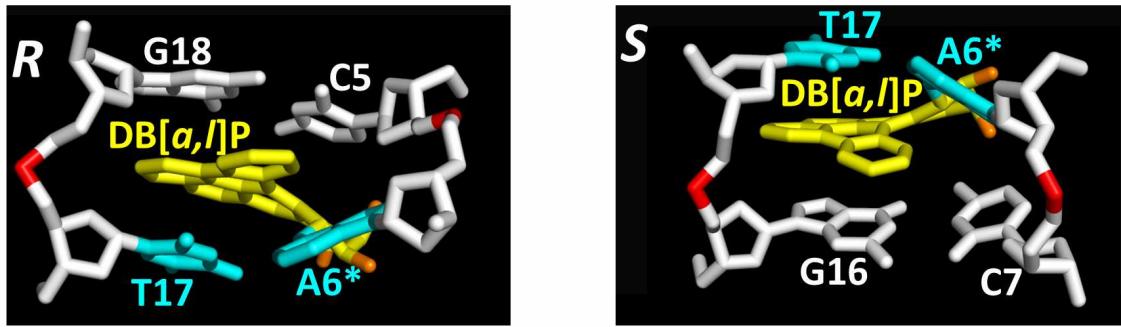


Figure S6. Stick views looking into the minor groove of the intercalation pocket, the $((C5\text{-}A6^*)\bullet(T17\text{-}G18))$ segment for the $14R$ (+)-adduct and the $(A6^*\text{-}C7)\bullet(G16\text{-}T17)$ segment for the $14S$ (-)-adduct.

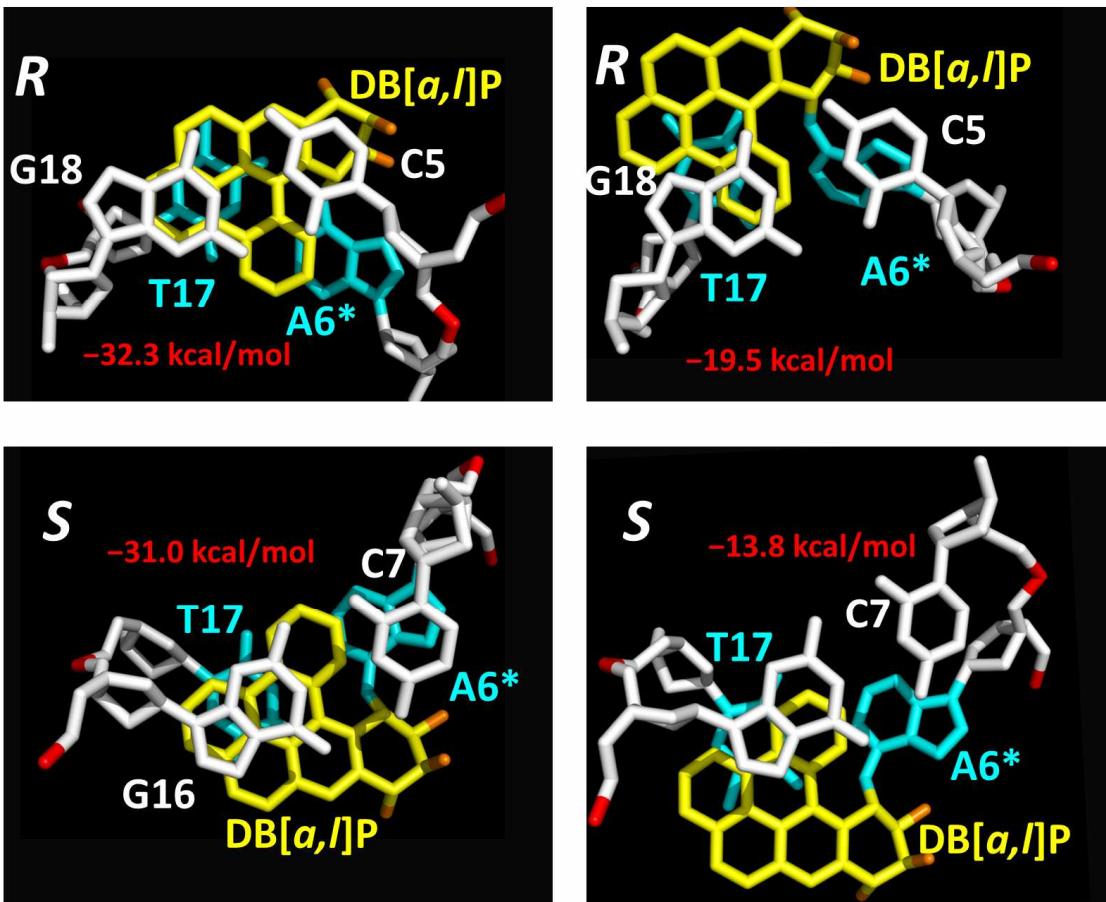


Figure S7 Maximally and minimally stacked structures from the ensembles (R stereoisomer: at 9,521 ps and 6,036 ps, respectively; S stereoisomer: at 5,926 ps and 3,456 ps, respectively). Energies associated with these structures are given. Note that in the S stereoisomer, C7 is never in proximity with the DB[a,I]P ring system, while C5 can overlap with it in the R stereoisomer.

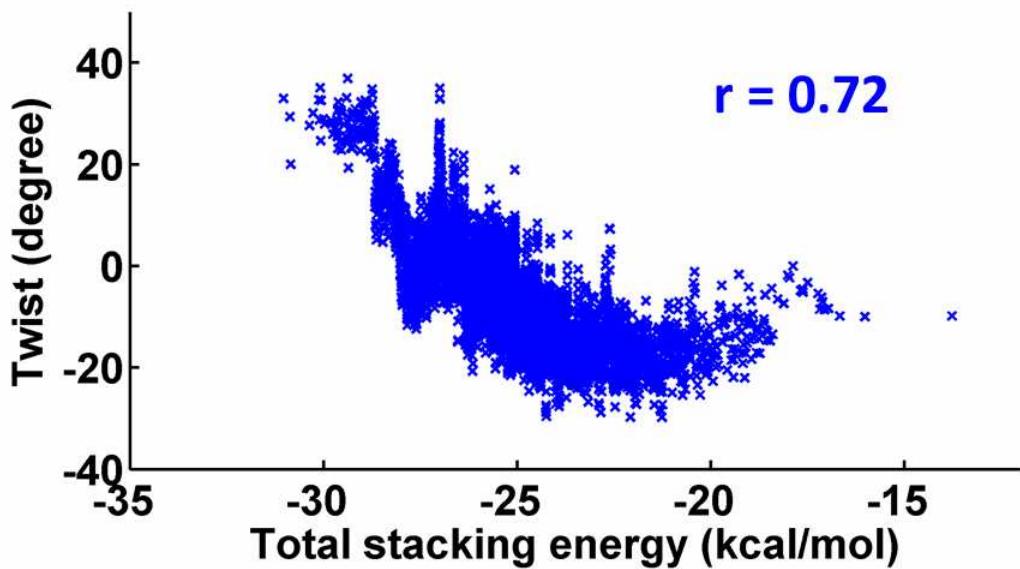


Figure S8 Scatter plot showing the correlation between total van der Waals interaction energy in the intercalation pocket and Twist in the $14S$ ($-$)-*trans-anti*-DB $[a,l]$ P- N^2 -dA adduct. The correlation coefficient is shown.

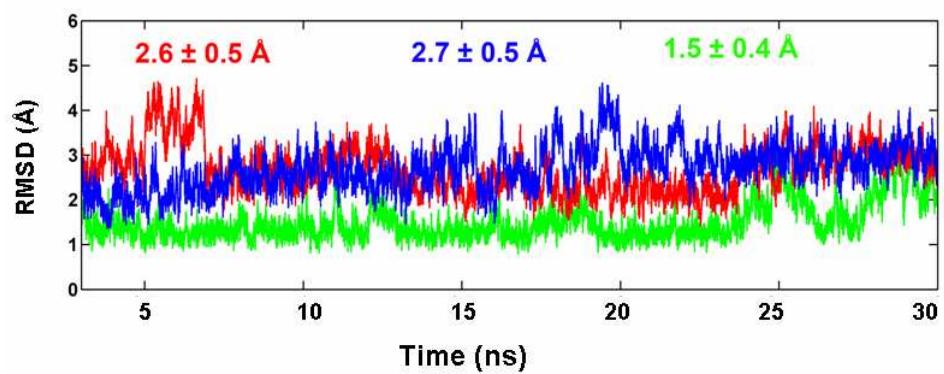


Figure S9. Time-dependence of RMSDs for the 14*R* (+)- (red) and 14*S* (-)-*trans-anti*-DB[*a,l*]P-*N*⁶-dA (blue) adducts as well as the unmodified control duplex (green). Ensemble average values and standard deviations are given. Only the central 9-mer is considered in this analysis.

References

1. Case, D. A., Darden, T. A., Cheatham III, T. E., Simmerling, C. L., Wang, J., Duke, R. E., Luo, R., Merz, K. M., Pearlman, D. A., Crowley, M., Walker, R. C., Zhang, W., Wang, B., Hayik, S., Roitberg, A., Seabra, G., Wong, K. F., Paesani, F., Wu, X., Brozell, S., Tsui, V., Gohlke, H., Yang, L., Tan, C., Mongan, J., Hornak, V., Cui, G., Beroza, P., Mathews, D. H., Schafmeister, C., Ross, W. S., and Kollman, P. A. (2006) AMBER 9, University of California, San Francisco, CA.
2. Jorgensen, W. L., Chandrasekhar, J., Madura, J. D., Imrey, R. W., and Klein, M. L. (1983) *J. Chem. Phys.* 79, 926-935.
3. Darden, T., York, D., and Pedersen, L. (1993) *J. Chem. Phys.* 98, 10089-10092.
4. Essmann, U., Perera, L., Berkowitz, M. L., Darden, T., Lee, H., and Pederson, L. G. (1995) *J. Chem. Phys.* 103, 8577-8593.
5. Ryckaert, J. P., Ciccotti, G., and Berendsen, H. J. C. (1977) *J. Comp. Phys.* 23, 327-341.
6. Berendsen, H. J. C., Postma, J. P. M., van Gunsteren, W. F., DiNola, A., and Haak, J. R. (1984) *J. Chem. Phys.* 81, 3684-3690.
7. Simmerling, C., Elber, R., and Zhang, J. (1995), pp 241-265, Netherlands: Kluwer.
8. Hingerty, B. E., Figueroa, S., Hayden, T. L., and Broyde, S. (1989) *Biopolymers* 28, 1195-1222.
9. Saenger, W. (1983) In *Principles of Nucleic Acid Structure* p226, Springer-Verlag, New York.
10. Calladine, C. R., and Drew, H. R. (1997), Morgan Kaufmann.
11. Cosman, M., Fiala, R., Hingerty, B. E., Laryea, A., Lee, H., Harvey, R. G., Amin, S., Geacintov, N. E., Broyde, S., and Patel, D. (1993) *Biochemistry* 32, 12488-12497.
12. Cosman, M., Laryea, A., Fiala, R., Hingerty, B. E., Amin, S., Geacintov, N. E., Broyde, S., and Patel, D. J. (1995) *Biochemistry* 34, 1295-1307.