

Cappell and Dohlman Supplemental Material

Table S1
Strains used in this study

Strain Name	Genotype	Source
BY4741*	<i>MATa leu2Δ met15Δ his3-1 ura3Δ</i>	(1)
TetO ₇ -STT4	<i>MATa URA3::CMV-tTA kan^R-TetO₇-TATA_{CYCI}-STT4</i>	(2)
TetO ₇ -PIK1	<i>MATa URA3::CMV-tTA kan^R-TetO₇-TATA_{CYCI}-PIK1</i>	(2)
TetO ₇ -WT	<i>MATa URA3::CMV-tTA</i>	(2)
TetO ₇ -PIK1 <i>kss1Δ</i>	<i>MATa URA3::CMV-tTA kan^R-TetO₇-TATA_{CYCI}-PIK1 kss1::LEU2</i>	This study
TetO ₇ -PIK1 <i>ste4Δ</i>	<i>MATa URA3::CMV-tTA kan^R-TetO₇-TATA_{CYCI}-PIK1 ste4::LEU2</i>	This study
TetO ₇ -PIK1 <i>ste5Δ</i>	<i>MATa URA3::CMV-tTA kan^R-TetO₇-TATA_{CYCI}-PIK1 ste5::LEU2</i>	This study
TetO ₇ -PIK1 <i>ste7Δ</i>	<i>MATa URA3::CMV-tTA kan^R-TetO₇-TATA_{CYCI}-PIK1 ste7::LEU2</i>	This study
TetO ₇ -PIK1 <i>ste11Δ</i>	<i>MATa URA3::CMV-tTA kan^R-TetO₇-TATA_{CYCI}-PIK1 ste11::LEU2</i>	This study
TetO ₇ -PIK1 <i>ste20Δ</i>	<i>MATa URA3::CMV-tTA kan^R-TetO₇-TATA_{CYCI}-PIK1 ste20::LEU2</i>	This study
TetO ₇ -PIK1 <i>ste50Δ</i>	<i>MATa URA3::CMV-tTA kan^R-TetO₇-TATA_{CYCI}-PIK1 ste50::LEU2</i>	This study
TetO ₇ -PIK1 <i>cla4Δ</i>	<i>MATa URA3::CMV-tTA kan^R-TetO₇-TATA_{CYCI}-PIK1 cla4::LEU2</i>	This study
TetO ₇ -PIK1 <i>ssk1Δ</i>	<i>MATa URA3::CMV-tTA kan^R-TetO₇-TATA_{CYCI}-PIK1 ssk1::LEU2</i>	This study
TetO ₇ -PIK1 <i>opy2Δ</i>	<i>MATa URA3::CMV-tTA kan^R-TetO₇-TATA_{CYCI}-PIK1 opy2::LEU2</i>	This study
<i>vps34Δ</i>	<i>MATa vps34::kan^R</i>	Invitrogen

*All strains derived from BY4741

Table S2
Plasmids used in this study

Fig	Plasmid Name	Alias	Description	Source
1	pRS315	SC-2-013	CEN LEU2 vector	(3)
5	pRS425	SC-2-006	2 μ M LEU2 vector	(4)
1	pRS423 FUS1-LacZ	SC-2-001	2 μ M HIS3 P _{FUS1} -lacZ	(5)
1	pRS315-STT4	SC-2-075	CEN LEU2 STT4	(6)
1	pRS315-PIK1	SC-2-076	CEN LEU2 PIK1	(6)
5	pRS425-STE11-4	SC-2-025	2 μ M LEU2 STE11-4	(6)
5	pRS315 GAL1-STE4	SC-2-024	CEN LEU2 P _{GAL1} -STE4	(7)
3	pRS313 GAL1-STE5-CTM	SC-2-035	CEN HIS3 P _{GAL1} -STE5-CTM	This Study
	pGS5-CTM	SC-2-027	CEN TRP1 P _{GAL1} -STE5-CTM	(8)
2	pRS316 STE5-GFPx3	SC-1-055	CEN URA3 GFP-STE5x3-T _{CYC1}	(9)
S2	pRS426 GFP-2xPH-PLC δ	SC-3-008	2 μ M URA3 GFP-2xPH-PLC δ	(10)
S2	pRS426 GFP-2xPH-FAPP	SC-3-009	2 μ M URA3 GFP-2xPH-FAPP	(10)
S2	pRS416 GFP-Lact-C2	SC-3-010	CEN URA3 GFP-Lact-C2	(11)

Figure S1

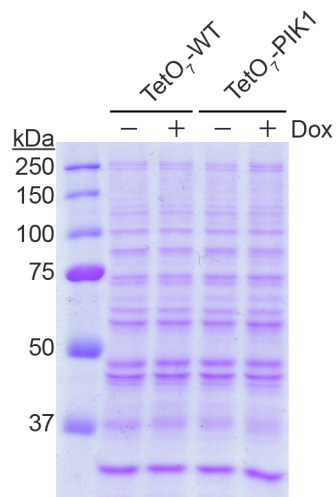


Figure S1. **Knockdown of *PIK1* does not affect global protein expression.** Whole cell lysates stained with Coomassie Brilliant Blue. Cells were treated with 10 μ g/ml doxycycline (Dox) for 15 h.

Figure S2

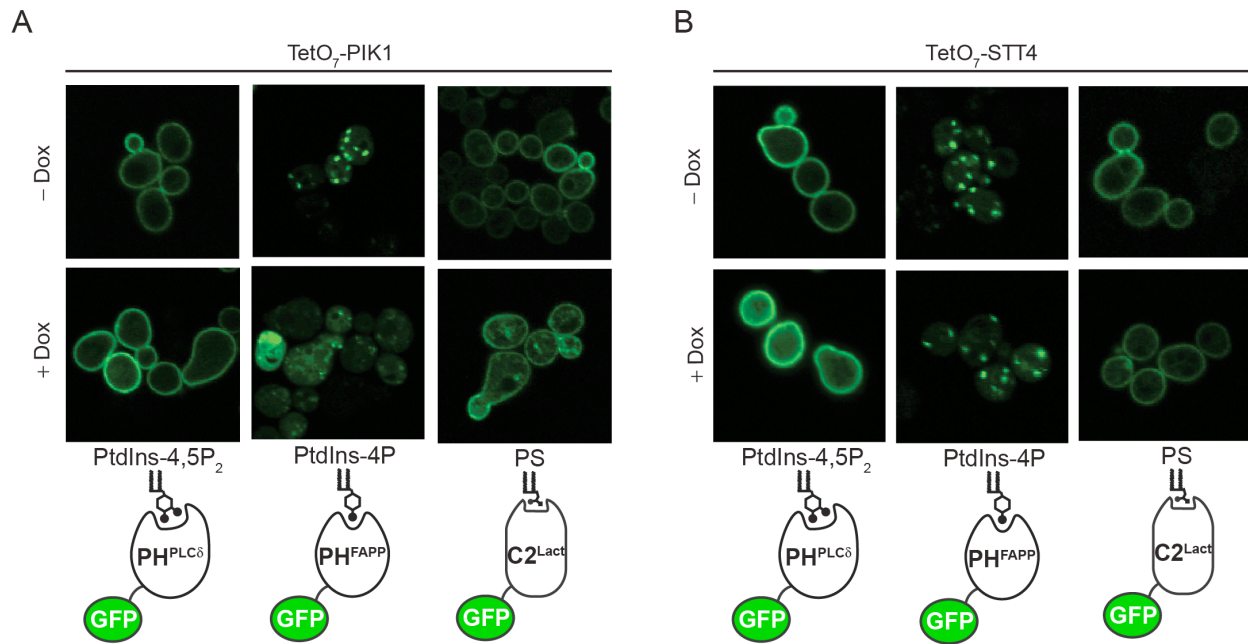


Figure S2. **Knockdown of *PIK1* or *STT4* results in loss of PtdIns 4-P *in vivo*.** GFP fluorescence of cells expressing a single copy plasmid pRS316 containing GFP fusion proteins of PH^{PLCδ}, PH^{FAPP1}, or C2^{Lact} and treated with 10 μg/ml doxycycline (Dox) for 15 h, as indicated. *A*, TetO₇-PIK1. *B*, TetO₇-STT4.

Figure S3

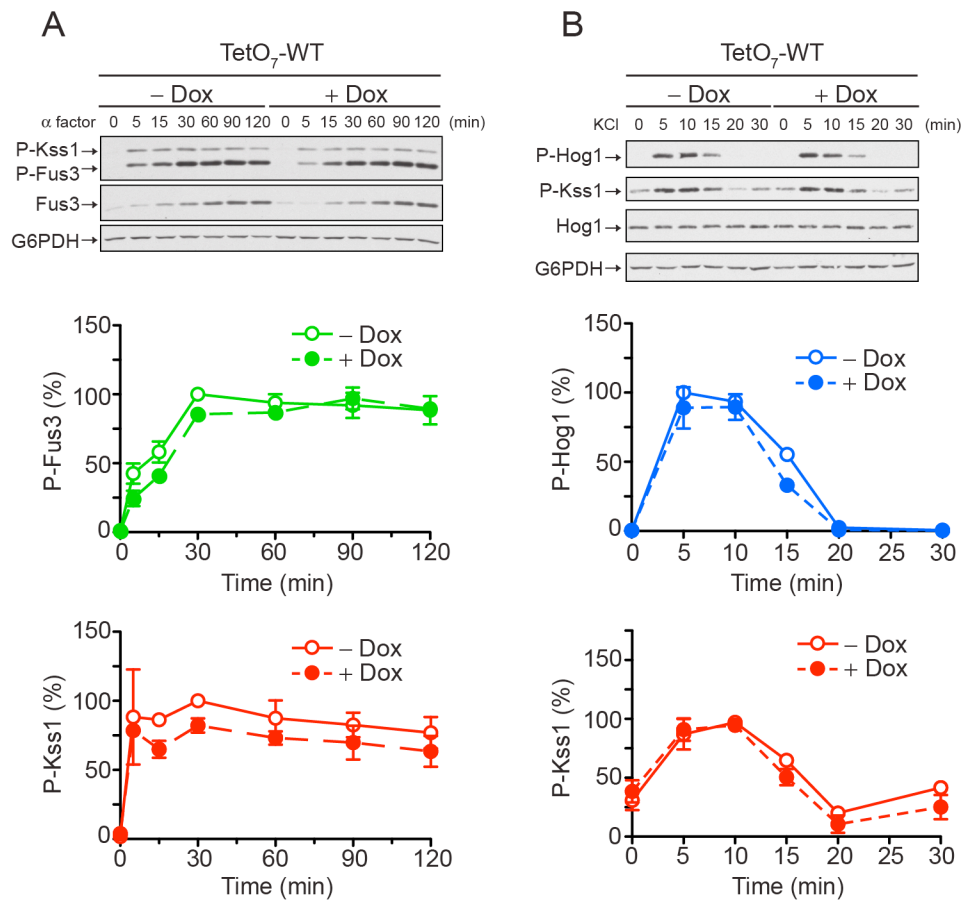


Figure S3. Doxycycline treatment does not affect Fus3, Kss1, or Hog1 activation. *A*, Cells harboring a doxycycline-repressible promoter attached to a non-expressible genetic element (TetO₇-WT) were treated with 10 μ g/ml doxycycline (Dox) for 15 h and 3 μ M α factor pheromone for the times indicated. Cell lysates were resolved by 12.5% SDS-PAGE and immunoblotting with phospho-p44/42 antibodies (P-Fus3, P-Kss1), Fus3 antibodies, Hog1 antibodies, or G6PDH antibodies as a loading control. Note that pheromone stimulation induces *FUS3* but not *KSS1* expression. Bands were quantified by scanning densitometry and analyzed with ImageJ software. Results are the mean \pm S.E. for three individual experiments. *B*, as in *A*, but cells were treated with 0.5 M KCl instead of pheromone.

Figure S4

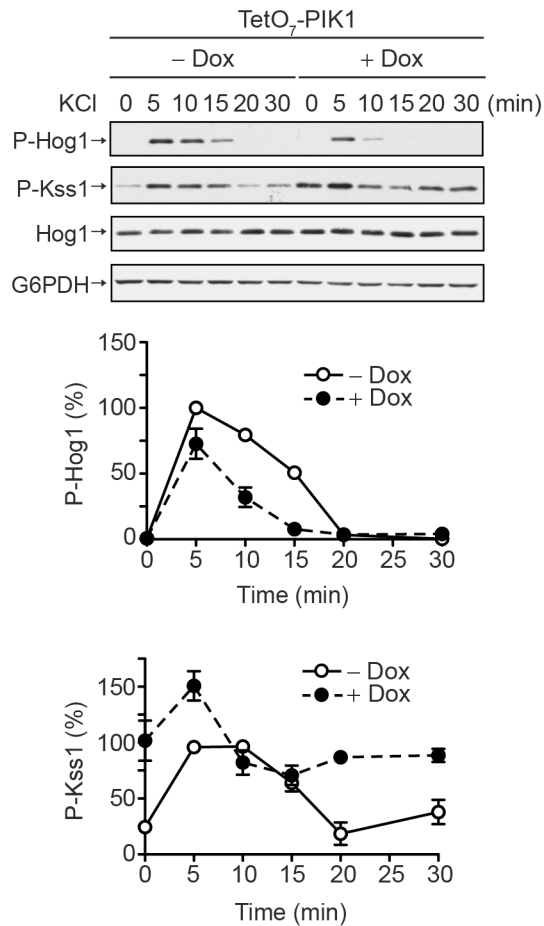


Figure S4. **Knockdown of *Pik1* inhibits *Hog1* activation.** TetO₇-PIK1 cells were treated with doxycycline for 15 h and 0.5 M KCl for the times indicated and analyzed by immunoblotting with phospho-p38 (P-Hog1) antibodies, phospho-p44/42 (P-Kss1) antibodies, Hog1 antibodies, or G6PDH antibodies as a loading control. All bands were quantified by scanning densitometry and analyzed with ImageJ software. Results are the mean \pm S.E. for three individual experiments.

Figure S5

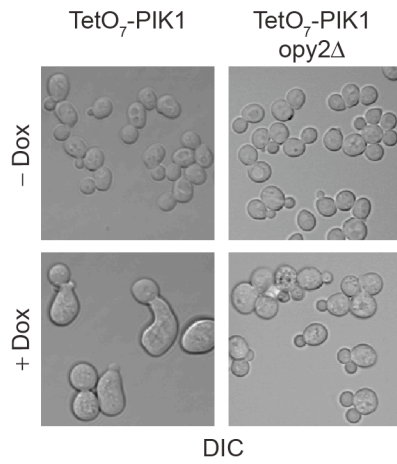


Figure S5. **Deletion of *OPY2* restores normal cell morphology upon *PIK1* knockdown.** DIC image of TetO₇-PIK1 and TetO₇-PIK1 *opy2*Δ cells treated with doxycycline for 15 h where indicated (+ Dox).

SUPPLEMENTAL REFERENCES

1. Brachmann, C. B., Davies, A., Cost, G. J., Caputo, E., Li, J., Hieter, P., and Boeke, J. D. (1998) *Yeast* **14**(2), 115-132
2. Mnaimneh, S., Davierwala, A. P., Haynes, J., Moffat, J., Peng, W. T., Zhang, W., Yang, X., Pootoolal, J., Chua, G., Lopez, A., Trochesset, M., Morse, D., Krogan, N. J., Hiley, S. L., Li, Z., Morris, Q., Grigull, J., Mitsakakis, N., Roberts, C. J., Greenblatt, J. F., Boone, C., Kaiser, C. A., Andrews, B. J., and Hughes, T. R. (2004) *Cell* **118**(1), 31-44
3. Sikorski, R. S., and Hieter, P. (1989) *Genetics* **122**(1), 19-27
4. Christianson, T. W., Sikorski, R. S., Dante, M., Shero, J. H., and Hieter, P. (1992) *Gene* **110**(1), 119-122
5. Hohmann, S. (2002) *Microbiol Mol Biol Rev* **66**(2), 300-372
6. Cappell, S. D., Baker, R., Skowyra, D., and Dohlman, H. G. (2010) *Mol Cell* **38**(5), 746-757
7. Dohlman, H. G., Apaniesk, D., Chen, Y., Song, J., and Nusskern, D. (1995) *Mol Cell Biol* **15**(7), 3635-3643
8. Pryciak, P. M., and Huntress, F. A. (1998) *Genes Dev* **12**(17), 2684-2697
9. Winters, M. J., Lamson, R. E., Nakanishi, H., Neiman, A. M., and Pryciak, P. M. (2005) *Mol Cell* **20**(1), 21-32
10. Stefan, C. J., Audhya, A., and Emr, S. D. (2002) *Mol Biol Cell* **13**(2), 542-557
11. Yeung, T., Gilbert, G. E., Shi, J., Silvius, J., Kapus, A., and Grinstein, S. (2008) *Science* **319**(5860), 210-213