

Figure S1. Regulation of Ste9 in wild type and ste9-shut off diploid cells during meiosis

Meiosis was induced from G1 phase using *pat1-114* mutation in wild type and *Puhp1-HA-ste9* diploid strains. Ste9 and Cdc2 were analyzed by immunoblotting using anti-Ste9 and Cdc2 antibodies. Ste9 is mostly phosphorylated in the vegetatively growing phase (indicated as v). Ste9 was dephosphorylated in G1 caused by nitrogen starvation (0 hr). Following this Ste9 became highly phosphorylated at 2 hrs after meiosis induction and remained phosphorylated until the end of meiosis. In *Puhp1-HA-ste9* strain, Ste9 protein levels are almost undetectable until 6 hrs after meiotic induction.



Figure S2. Expression of Fzr2 in meiosis.

Samples from *pat1*-induced synchronized meiosis were analyzed by immunoblotting using anti-HA, anti-Cdc2 and anti-Fzr2 antibodies. Asterisk indicates a non-specific crossreacting band.



Figure S3. Synchronized meiosis in WT, *slp1-NF410*, *fzr2* Δ *fzr3* Δ and *fzr1* Δ *fzr2* Δ *fzr3* Δ diploid cells.

pat1-114-induced synchronization of meiosis was performed in WT, slp1-NF410, $fzr2\Delta fzr3\Delta$ and $fzr1\Delta fzr2\Delta fzr3\Delta$ mutant diploid strains. Meiotic progression was examined microscopically for the number of nuclei in a cell. Protein levels were analyzed by immunoblotting using specific antibodies at indicated time points.



Figure S4. Fzr1/Mfr1 protein levels in synchronized meiosis in wild type and *slp1-NF410* cells.

Samples from *pat1-114*-induced synchronized meiosis in wild type (WT) and *slp1-NF410* were analyzed by immunoblotting using anti-Fzr1 and anti-Cdc2 antibodies. Fzr1/Mfr1 expression started around 4.5 hrs and continued till the end of meiosis. Fzr1/Mfr1 was more accumulated in *slp1-NF410* mutant cells than WT.



Figure S5. Mes1 levels in $fzr2\Delta fzr3\Delta$ and $fzr1\Delta fzr2\Delta fzr3\Delta$ mutant cells.

pat1-114-induced synchronization of meiosis was performed in WT, slp1-NF410, $fzr2\Delta fzr3\Delta$ and $fzr1\Delta fzr2\Delta fzr3\Delta$ mutant diploid strains expressing HA-tagged Mes1 under the native promoter. HA-Mes1 levels were analyzed by immunoblotting using anti-HA antibody.



Figure S6. In vitro ubiquitylation assay with S. pombe APC/C

(A) *S. pombe* APC/C purified from *lid1*⁺-*TAP* strain was used for in vitro ubiquitylation assay with Ste9 and *S. pombe* E2s. Efficient ubiquitylation of Cut2 and Cdc13 was observed only when APC/C, E2s and Ste9 were all present. (B) Destruction box-dependency of the reconstituted ubiquitylation reaction. In vitro ubiquitylation was performed using ³⁵S-labeled wild type or D-box mutant of N-terminal 70 residues of Cdc13 (N70 or N70dm, respectively) as subtrates. Both Slp1 and Ste9 induced ubiquitylation of N70 but not N70dm.



Figure S7. In vitro translation of S. pombe APC/C co-activators

Slp1, Ste9, Fzr1/Mfr1, Fzr2 and Fzr3 were prepared in coupled transcription/translation in reticulocyte lysates in the presence of ³⁵S-methionine (TNT; Promega). Band intensities quantified are indicated at the bottom. Knowing the number of methionine residues in the co-activators allowed estimation of the relative levels of each. To prepare co-activators for cell-free destruction and ubiquitylation assay in this paper, cold methionine (0.02mM) was used instead of ³⁵S-methionine.



Figure S8. S. pombe APC/C-dependent destruction assay in Xenopus egg extracts.

Endogenous APC/C and Fizzy/Cdc20 were depleted from *Xenopus* CSF extracts using anti-Apc3 and anti-Fizzy antibodies. Purified *S. pombe* APC/C and in vitro translated Slp1 as well as recombinant *S. pombe* E1 (Uba1) and E2s (Ubc4 and Ubc11) were supplemented into the extracts and destruction assays were performed with ³⁵S-labelled Cdc13 or Mes1 as a substrate. Cdc13 and Mes1 were destroyed in a *S. pombe* APC/C and Slp1-dependent manner. Addition of Ubc4 and Ubc11 enhanced the destruction.

Table S1. Strain List

Strains	Genotype	Source
HYY718	h-/h- pat1-114/pat1-114 mes1::kanMX/mes1::kanMX slp1-	This study
	NF410::kanMX/slp1-NF410::kanMX ade6-M210/ade6-M216	
	ura4-d18/ura4-d18	
HYY719	h-/h- pat1-114/pat1-114 mes1::kanMX/mes1::kanMX	This study
	fzr1::hphMX/fzr1::hphMX ade6-M210/ade6-M216 ura4-	
	d18/ura4-d18	
HYY729	h-/h- pat1-114/pat1-114	Our stock
	mes1WT::ura4+::kanMX/mes1WT::ura4+::kanMX ade6-	
	M210/ade6-M216 ura4-d18/ura4-d18	
HYY733	h-/h- pat1-114/pat1-114 mes1::Pint-3HA-ura4+-	Our stock
	kanMX/mes1::Pint-3HA-ura4+-kanMX ade6-M210/ade6-M216	
	ura4-d18/ura4-d18	
HYY734	h-/h- pat1-114/pat1-114 HA-mes1WT::ura4+::kanMX/HA-	Our stock
	mes1WT::ura4+::kanMX ade6-M210/ade6-M216 ura4-	
	d18/ura4-d18	
HYY738	h-/h- pat1-114/pat1-114 HA-K0mes1::ura4+::kanMX/HA-	Our stock
	K0mes1::ura4+::kanMX ade6-M210/ade6-M216 ura4-	
	d18/ura4-d18	
HYY742	h-/h- pat1-114/pat1-114 HA-mes1WT::ura4+::kanMX/HA-	This study
	mes1WT::ura4+::kanMX ade6-M210/ade6-M216 ura4-	
	d18/ura4-d18 slp1-NF410::kanMX/slp1-NF410::kanMX	
HYY743	h-/h- pat1-114/pat1-114 HA-mes1WT::ura4+::kanMX/HA-	This study
	mes1WT::ura4+::kanMX ade6-M210/ade6-M216 ura4-	
	d18/ura4-d18 fzr1::hphMX/fzr1::hphMX	
HYY744	h-/h- pat1-114/pat1-114 HA-mes1WT::ura4+::kanMX/HA-	This study
	mes1WT::ura4+::kanMX ade6-M210/ade6-M216 ura4-	
	d18/ura4-d18 slp1-NF410/slp1-NF410	
	fzr1::hphMX/fzr1::hphMX	
HYY745	h-/h- pat1-114/pat1-114 HA-mes1WT::ura4+::kanMX/HA-	This study
	mes1WT::ura4+::kanMX ade6-M210/ade6-M216 ura4-	
	d18/ura4-d18 fzr2::ura4+/fzr2::ura4+	
HYY793	h-/h- pat1-114/pat1-114 HA-mes1WT::ura4+::kanMX/HA-	This study
	mes1WT::ura4+::kanMX ade6-M210/ade6-M216 ura4-	
	d18/ura4-d18 Puhp1-HA-ste9::hphMX/Puhp1-HA-	
	ste9::hphMX	
HYY790	h-/h- pat1-114/pat1-114 HA-mes1WT::ura4+::kanMX/HA-	This study
	mes1WT::ura4+::kanMX ade6-M210/ade6-M216 ura4-	
	d18/ura4-d18 jzr5::kanMX/jzr3::kanMX	m1 · 1
HYY1026	h-/h- pat1-114/pat1-114 HA-mes1W1::ura4+::natMX/HA-	This study
	$mes_1w_1::ura_4+::natMX a a eo-M_210/a deo-M_210 ura_4-$	
	$a_{18}/ura_{4}-a_{18}$ $Jzr_{2\Delta}$:: $ura_{4}+/Jzr_{2\Delta}$:: $ura_{4}+/Jzr_$	
111/1/1027	JZr5::KUNNIX/JZr5::KUNNIX	
HYY1027	<i>n-/n- pati-114/pati-114 HA-mes1W1::ura4+::natMX/HA-</i>	This study
	$mes_1 w_1$:: ura_4 +:: nat_MX $aaeo-M_2 10/aaeo-M_2 10 ura_4$	
	$a_{10}/ura_{4-a_{10}}$ $J_{2r_{10}}$ $J_{2r_{10}}/ura_{4+a_{10}}$ $J_{2r_{10}}/ura_{4+a_{10}}$	
KCW2050	$JZr 2\Delta :: Ura4 + JZr 5 :: KanMA / JZr 5 :: KanMA $	V. C 1 1
KGY2050	n- IIa1-TAP::KanMX mts3-1 mad2::ura4+ mad3::ura4+	K. Gould