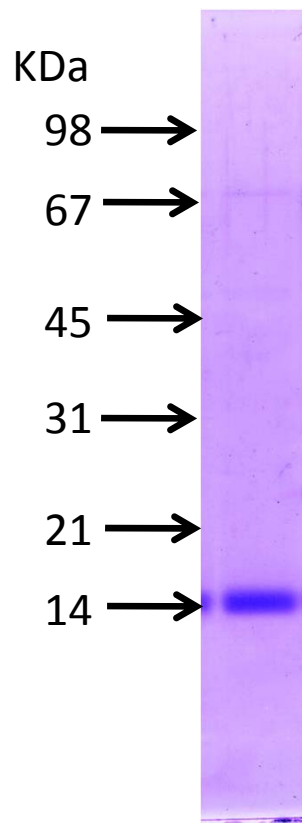
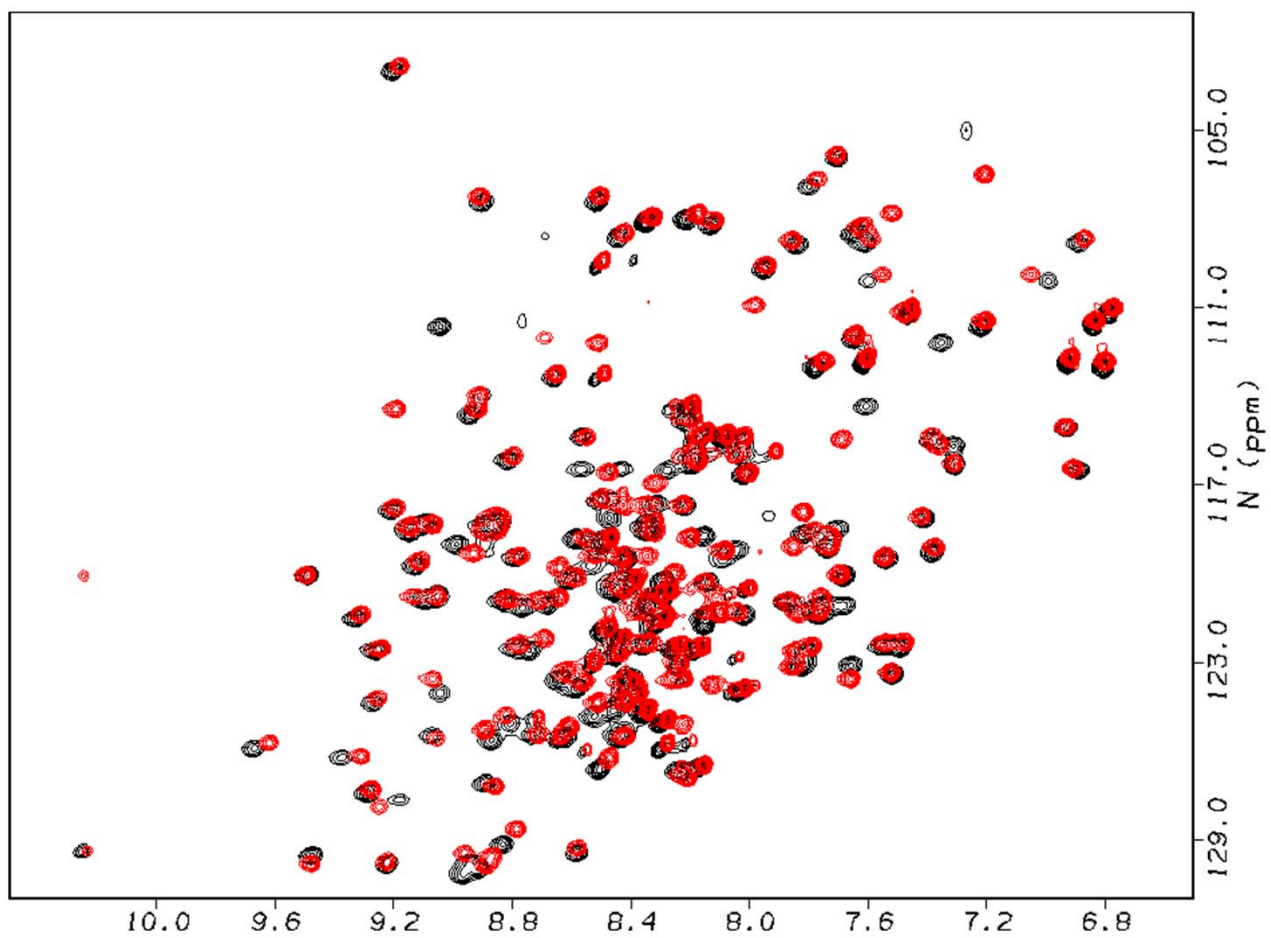


## Supplementary Fig 1



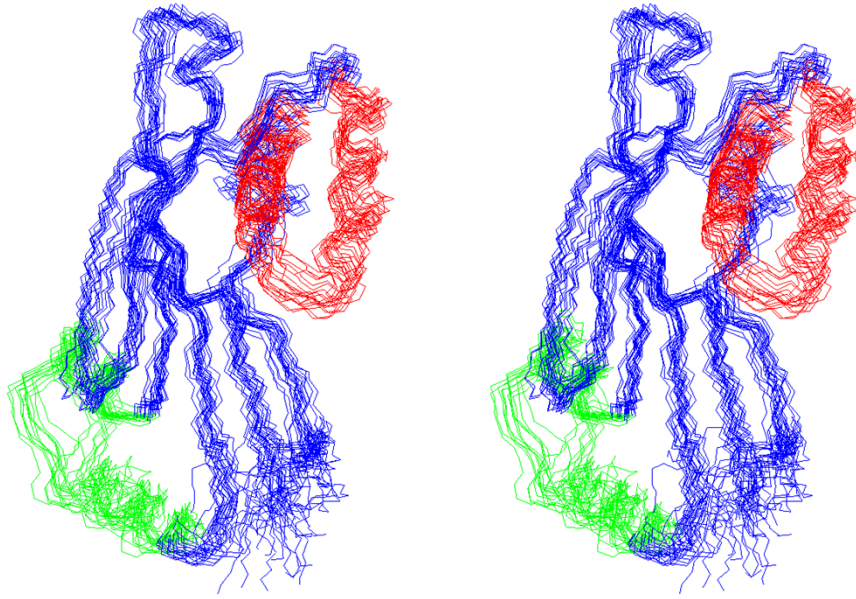
**SDS-PAGE of the purified E1064-WND- $\Delta$  used for NMR structure determination.** Positions of the molecular weight markers and their mass in Kda are shown on the left.

## Supplementary Fig 2



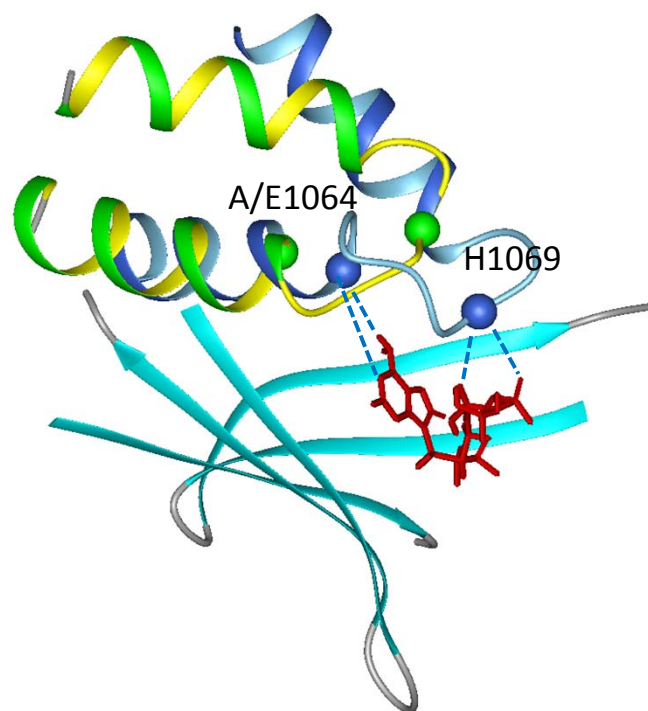
**Overlay of the  $^1\text{H}$ ,  $^{15}\text{N}$ -HSQC spectra of the wild type (*red*) and E1064A (*black*) N-domains**

### Supplementary Fig 3



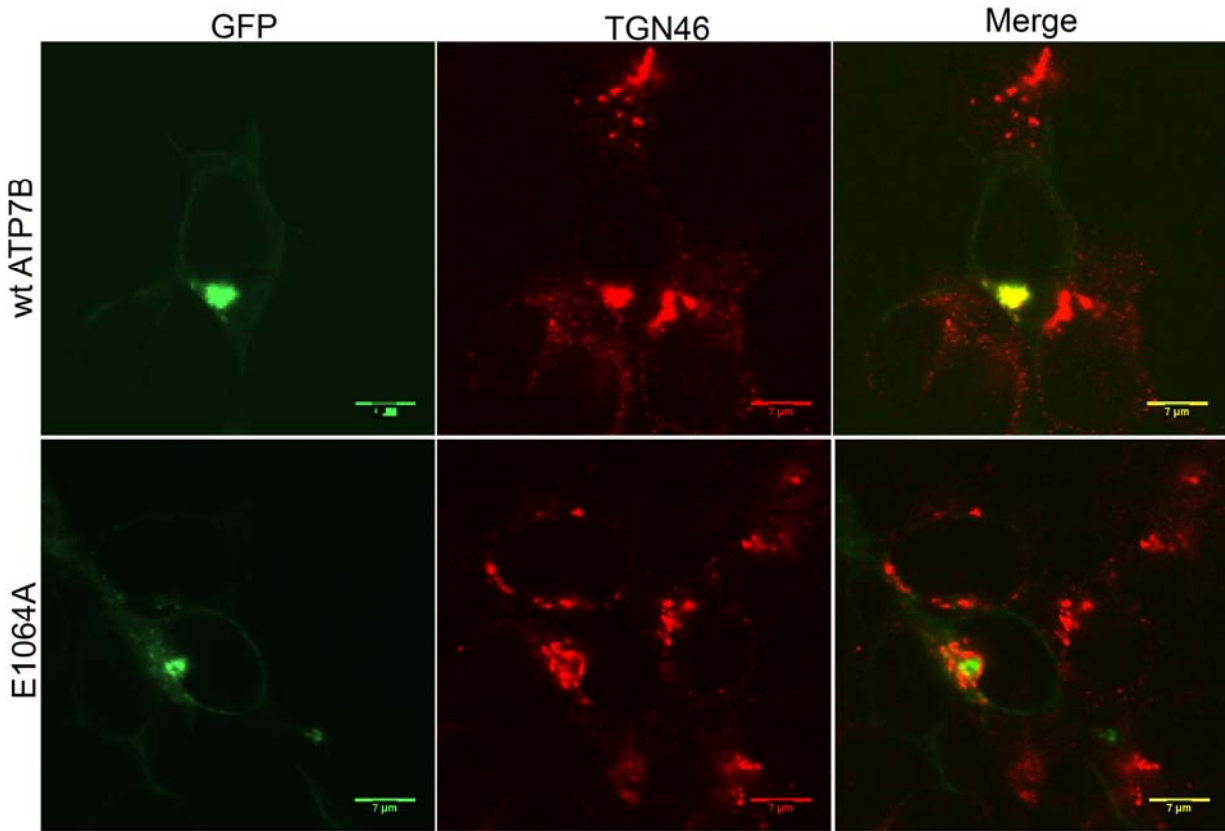
**Stereoview of the ensemble of the 20 lowest energy structures of E1064A-WND- $\Delta$ .** The  $\alpha$ 1- $\alpha$ 2 helical hairpin (residues 1053-1083) is shown in *red*, the  $\alpha$ 3- $\alpha$ 4 helical hairpin (residues 1151-1173) in *green*, and the core  $\beta$ -sheet and the connecting loops in *blue*.

## Supplementary Fig 4



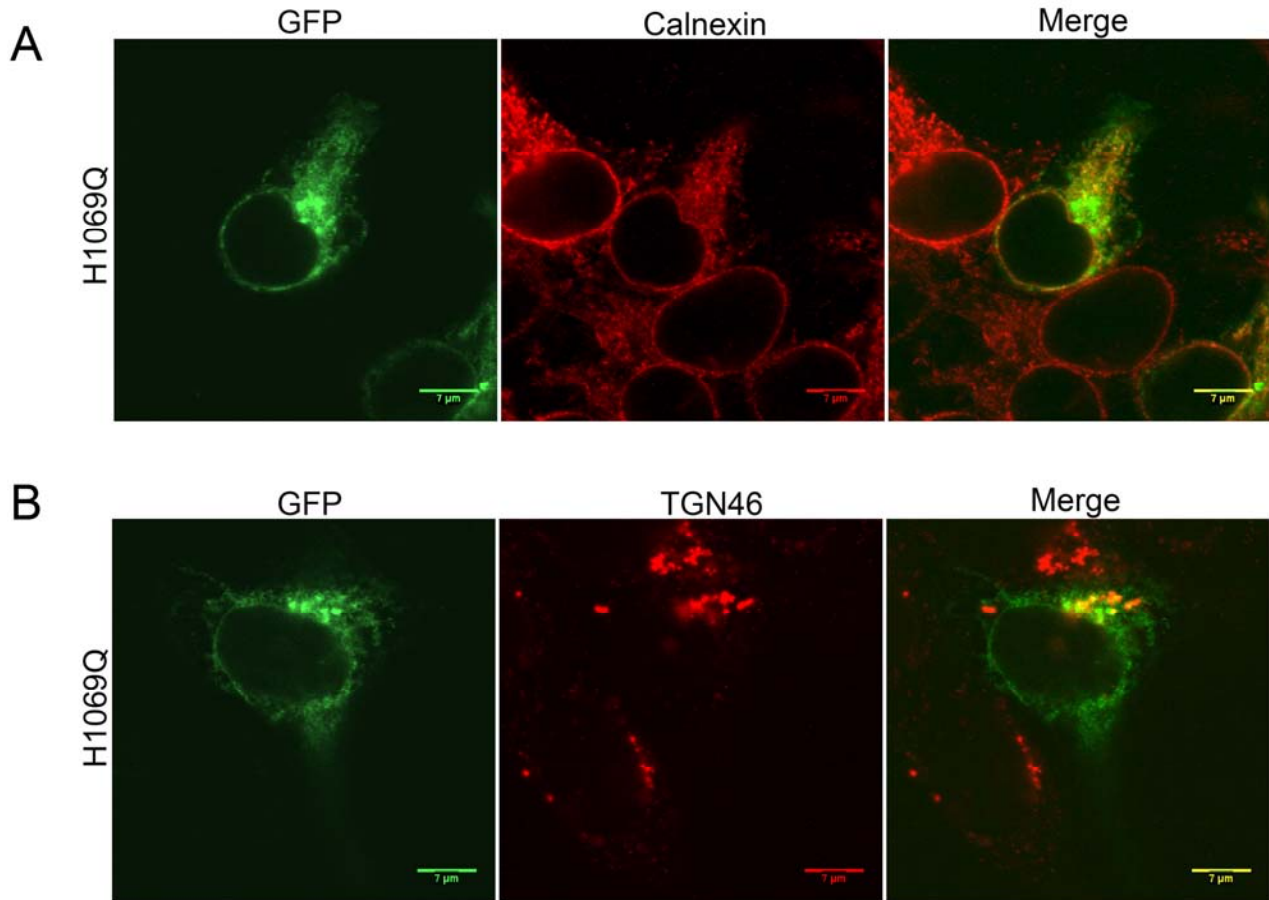
**ATP molecule modeled into the structure of the wild type N-domain** (2ARF, *blue*) based on the structure of the CopA ATP-binding domain with AMPPCP (3A1C). The position of the  $\alpha 1$ - $\alpha 2$  helical hairpin in the E1064-WND $\Delta_{1115-1138}$  when the core  $\beta$ -sheets in both structures are aligned to minimize r.m.s.d. is shown in *green*.

## Supplementary Fig 5



**Subcellular localization of E1064A mutant** HEK293T cells were transfected with the plasmids expressing a GFP- tagged wt ATP7B (upper panel), or the E1064A mutant (lower panel). Following protein expression at 37°C , cells were immunostained with the anti-TGN46 antibody (red) to visualize TGN and compare with the ATP7B pattern (green). In transfected cells both GFP-tagged wt and mutant were co-localized with the TGN marker (yellow). Scale bar is 7 μm

## Supplementary Fig 6



**Co-localization of H1069Q mutant with the organelle markers.** HEK293TRex cells were transfected with the plasmid expressing the GFP tagged H1069Q mutant (green) and following protein expression at 37°C cells were immunostained with the organelle markers (red) to determine the co-localization of H1069Q with the markers (yellow). (A) Immunostaining with the antibody against the ER marker calnexin. (B) Staining with the with anti-TGN46 antibody. Scale bar is 7 μm

Supplementary Table 1

STATISTICS FOR STRUCTURE CALCULATION OF E1064A-WND	
<b>Number of NOE-derived distance restraints</b>	
Total	1379
Intra-residue	272
Sequential ( $ i-j  = 1$ )	533
Medium-range ( $1 <  i-j  < 5$ )	228
Long-range ( $ i-j  > 4$ )	346
<b>Hydrogen bond restraints</b>	68
<b>RMS violations per distance restraint (Å):</b>	0.0194±0.0028
<b>RMS violations per dihedral angle constraint (°):</b>	0.43±0.11
<b>Average number of violations per conformer:</b>	
Distance restraints	5±2
Dihedral angle constraints	1±1
Van der Waals	1±1
<b>Mean NOE violations larger than 0.25 Å</b>	0
<b>Maximum NOE violation (Å)</b>	0.35
<b>Average RMSD to the mean (Å)<sup>a</sup></b>	
backbone	1.35±0.23
all heavy atoms	1.77±0.21
residual CYANA Target Function (Å <sup>2</sup> ) <sup>b</sup>	2.38±0.48
<b>Ramachandran plot quality<sup>c</sup></b>	
% of residues in most favorable regions	82.5
% of residues in allowed regions	17.1
% of residues in generously allowed regions	0.4
% of residues in disallowed regions	0.0
<b>Close contacts</b>	0
<b>RMS deviations from ideal geometry</b>	
Bond lengths (Å)	0.001
Bond angles (°)	0.2
<b>MolProbity Clashscore<sup>d</sup></b>	-2.33

<sup>a</sup> RMSD values calculated for residues 7-140 corresponding to residues 1038-1114,1139-1195 in the full length ATP7B

<sup>b</sup> Structure calculations were performed with CYANA 2.1. A total of 1000 random conformers were subjected to 20000 steps of a simulated annealing process. Twenty lowest energy structures were selected for deposition.

<sup>c</sup> As reported by PROCHECK for residues 7-140

<sup>d</sup> As reported by PSVS for residues 7-140