

# Supplementary Figures

## **Cas3 is a single-stranded DNA nuclease and ATP-dependent helicase in the CRISPR/Cas immune system**

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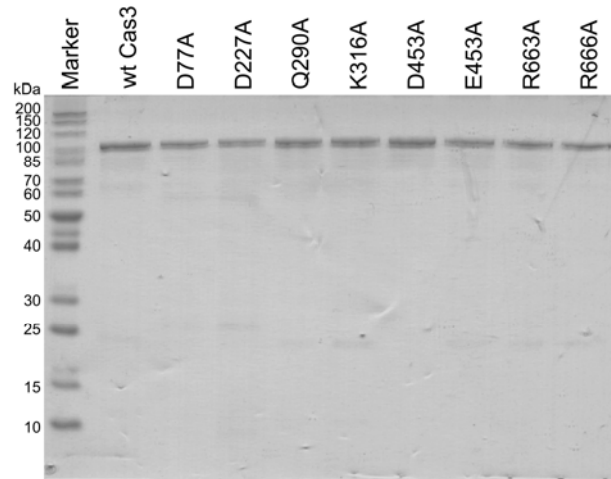
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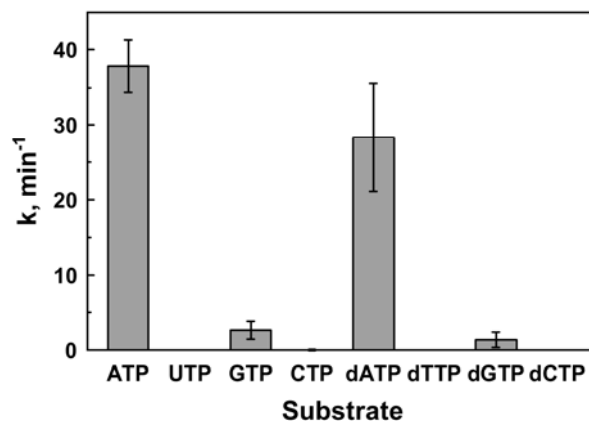
Tel: +370-5-2602108; Fax: +370-5-2602116; E-mail: siksnys@ibt.lt



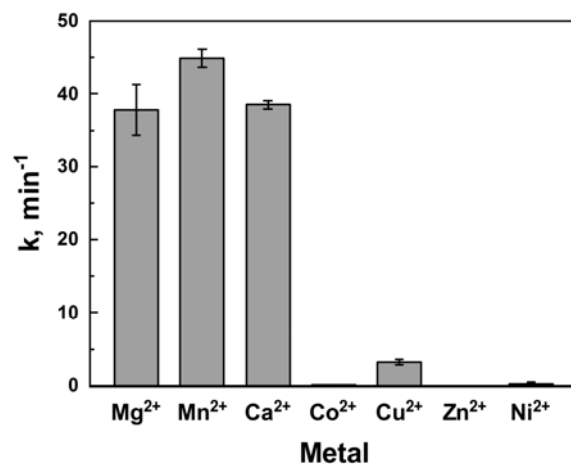
**Supplementary Figure 1. Multiple alignment of the conserved regions in the (A) HD and (B) DExD/H domains of Cas3 protein sequence.** Alignments were assembled using “COBALT” (Papadopoulos & Agarwala, 2007) tool and visualised with “BioEdit 7.0.5” (Hall, 1999). The numbers preceding and following the alignment indicate the number of amino acid residues between the protein termini and the proximal and distal aligned blocks, respectively; those between individual blocks indicate the number of residues separating them. Homologous sequences with different similarity to Cas3 from *Streptococcus thermophilus* DGCC7710 were used for construction of alignment. GeneBank identification numbers of proteins are given in the left-hand column. Stars below the columns (black-shaded) indicate conservative amino acid residues. Grey-shaded columns indicate similar amino acid residues. Positions of point mutations to alanin which were made in this work are indicated above the alignment columns. Roman numbers below the alignment columns in the figure B indicate regions of conservative motifs of DExD/H helicases.



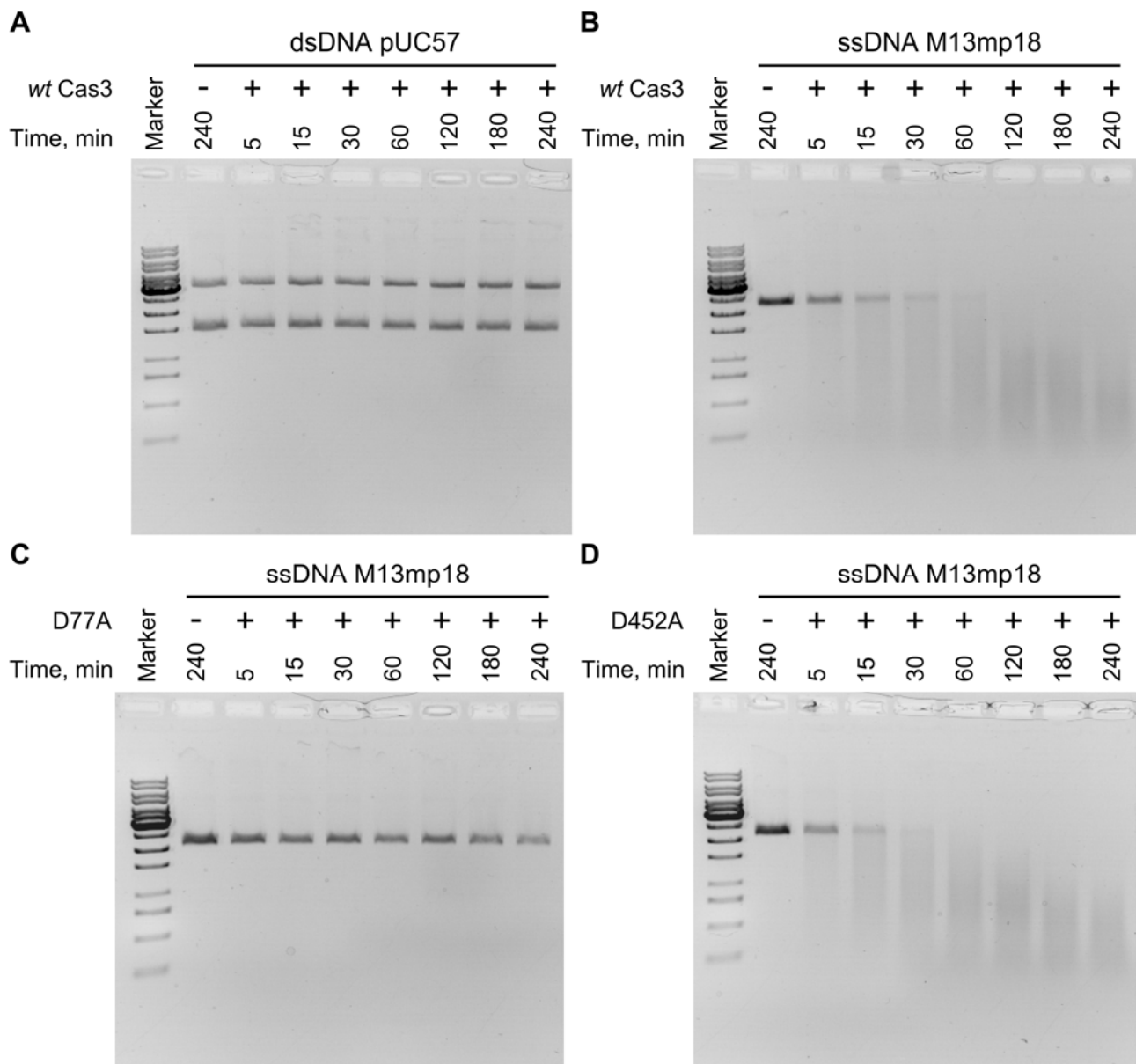
**Supplementary Figure 2.** *SDS-PAGE of the purified Cas3 proteins and its mutants.* Marker – protein molecular marker “PageRuler Unstained Protein Ladder” (Fermentas, Vilnius, Lithuania). The numbers shown on the left indicate markers’ size in kDa. Samples corresponding to 750 ng of each protein were loaded on the gel.



**Supplementary Figure 3. *Cas3* ATPase activity dependence on nucleotides and deoxynucleotides.** ATPase reactions were conducted at 30 °C in a reaction buffer containing 10 mM Tris–HCl (pH 7.5 at 25C), 30 mM KCl, 5% glycerol, 2 mM MgCl<sub>2</sub>, 0.1 mg/ml of BSA, and 3.0 nM of ssDNA (M13mp18), 250 nM of Cas3 and 500 μM of various nucleotides and deoxynucleotides. Malachite green assay was used to measure ATP hydrolysis through the detection of liberated free phosphate from ATP. Reaction rate constant  $k$  (min<sup>-1</sup>) calculated from the linear slopes of respective times courses.



**Supplementary Figure 4. *Cas3* ATPase activity dependence on divalent metal ions.** ATPase reactions were conducted at 30 °C in a reaction buffer containing 10 mM Tris-HCl (pH 7.5 at 25 °C), 30 mM KCl, 5% glycerol, 0.1 mg/ml of BSA, and 3.0 nM of ssDNA (M13mp18), 250 nM of Cas3 and 2 mM of different divalent metal ions. Malachite green assay was used to measure ATP hydrolysis through the detection of liberated free phosphate from ATP. Reaction rate constant  $k$  ( $\text{min}^{-1}$ ) calculated from the linear slopes of respective times courses.



**Supplementary Figure 5.** Time courses of nuclease activity of Cas3 and mutant proteins. Nuclease reactions were conducted at 37 °C in a reaction buffer containing 10 mM Tris-HCl (pH 7.5 at 25 °C), 60 mM KCl, 10% glycerol (v/v), 10 mM MgCl<sub>2</sub>, 0.1 mg/ml of BSA, and 4 nM of dsDNA supercoiled form of pUC57 plasmid (A) or ssDNA of M13mp18 (C-D), and 500 nM Cas3 (A-B) or mutant proteins (D77A – C, D452A – D). Time points of the reactions indicated above the figures. Marker – DNA molecular marker “GeneRuler™ 1 kb DNA Ladder” (Fermentas, Vilnius, Lithuania).

### Supplementary references

Hall TA (1999) BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucl Acids Symp Ser* **41**: 95-98

Papadopoulos JS, Agarwala R (2007) COBALT: constraint-based alignment tool for multiple protein sequences. *Bioinformatics* **23**(9): 1073-1079