

Table S1. List of primers used for qRT-PCR

Human gene

Hepcidin
(HAMP) 5'-CTGACCAGTGGCTCTGTTTTTC-3'
 5'-GAAGTGGGTGTCTCGCCTC-3'

Zebrafish gene

RPL 5'-TCTGGAGGACTGTAAGAGGTATGC-3'
 5'-AGACGCACAATCTTGAGAGCAG-3'

zHepcidin 5'-CCTGGCTGCTGTCGTCAT-3'
 5'-TGGTTCTCCTGCAGTTCTTCAC-3'

Mouse gene

Hepcidin
(mHamp-1) 5'-AAGCAGGGCAGACATTGCGAT-3'
 5'-CAGGATGTGGCTCTAGGCTATGT-3'

18S rRNA 5'-CGGCTACCACTCCAAGGAA-3'
 5'-GCTGGAATTACCGCGGCT-3'

Primers used to quantify transcript levels in human HepG2 cells, zebrafish, and mice by quantitative real-time PCR.

Table S2. Treatment of wild-type mice with LDN-193189 for 30 days does not impact circulating erythrocyte, myeloid, lymphocyte, or thrombocyte lineages

	Mean (n=4)	Mean (n=5)
Total WBC	1.9±0.3	2.3±0.2
% Neutrophil	30.8±5.4	24.0±3.1
% Lymphocyte	63.1±7.6	69.7±4.4
% Monocyte	3.8±0.7	4.2±0.9
RBC	9.0±0.2	8.5±0.1
Hb	11.9±0.1	12.5±0.2
Hct	43.8±0.9	42.3±0.3
MCV	48.5±1.0	50.1±0.7
MCHC	27.3±0.3	29.6±0.3
RDW	18.6±0.0	17.7±0.2
Platelet count	830±73	628±86
% Reticulocyte	2.6±0.6	2.9±0.1

Treatment of wild-type mice with LDN-193189 (3 mg/kg i.p. daily) for 30 days did not impact circulating numbers of erythrocyte, myeloid, lymphocyte, or thrombocyte lineages. Data shown are mean±SEM, with no significant differences detected between vehicle or LDN-193189-injected groups.

Figure S1. Kinetics of IL-6-mediated hepcidin induction in human hepatoma (HepG2) cells, and synergy with BMP signaling

(A) RNA was extracted from HepG2 cells incubated with IL-6 (100 ng/ml) for 0 to 120 min. Hepcidin mRNA levels were measured by qRT-PCR (values shown are mean±SEM, UUn=3, *p<0.05 vs. untreated control). Under some conditions (B)U, treatment of HepG2 cells with BMP6 (5 ng/ml) and IL-6 (25 ng/ml) appeared to induce hepcidin mRNA expression in an additive manner as compared to either treatment alone (n=4 per group, one-way ANOVA p<0.01, *p<0.05 vs. untreated control, #p<0.01 vs cells treated with BMP6 alone), whereas, under other conditions (C), treatment of HepG2 cells with BMP6 (10 ng/ml) and IL-6 (50 ng/ml) appeared to induce hepcidin mRNA expression in a synergistic fashion, as compared to either treatment alone (n=4 per group, one-way ANOVA p<0.01, *p<0.05 vs. untreated control, #p<0.05 vs. cells treated with IL-6 or BMP6 alone).

Figure S2. IL-6 expression in cmlc-IL6 transgenic zebrafish embryos

(A) Double-transgenic fish expressing human IL-6 ectopically in the heart (cmlc-IL-6) were fixed at 48 hpf, and in situ hybridization was performed to detect human IL-6 mRNA. Abundant hIL-6 RNA was detected in the hearts of cmlc-IL-6 zebrafish embryos but not in those of WT embryos. (UB) Hepcidin mRNA levels were greater in adult (10 week old) cmlc-IL-6 zebrafish than in WT zebrafish (n=5 zebrafish per group, *p=0.01).

Figure S3. Serum iron levels in mice 6 days after challenge with turpentine

Mice were pretreated with LDN-193189 (3 mg/kg i.p.) or vehicle and then injected intrascapularly with turpentine (5 ml/kg) or saline. LDN-193189 and drug vehicle injections were continued every 12 h for 6 d, at which time serum iron levels were measured. Turpentine injection induced hypoferrremia which was evident at 6 d following a single treatment, while concurrent LDN-193189 treatment prevented the turpentine-induced decrease in serum iron levels (n≥5 mice per group, *p<0.05 vs. untreated control, †p<0.05 vs. turpentine-treated).

Figure S4. The number of hematopoietic stem cells are not affected by LDN-193189 injections for 14 and 28 days

Mice were injected with vehicle or LDN-193189 (3 mg/kg i.p.) daily for 14 d (A) or 28 d (B). Numbers of BM cells highly enriched for hematopoietic stem and progenitor cells based on the surface phenotype Lin⁻CD48⁻c-Kit⁺Sca⁺CD150⁺ (BM HSC) were measured as a percentage of total mononuclear cells by flow cytometry. No significant impact on the frequency of this BM HSC population were observed as a result of LDN-193189 treatment at 14 or 28 d (Un=5 mice per group). USimilar results were seen based on the analysis of absolute number of HSCs.

Figure S5. Treatment with LDN-193189 for 28 d decreases baseline hepatic hepcidin mRNA levels

Mice were injected with LDN-193189 (3mg/kg i.p.) or vehicle daily for 28 d. Hepatic hepcidin mRNA levels were less in LDN-193189-treated mice than in vehicle-treated animals (n=10, *p<0.05 vs. vehicle-treated mice).

Figure S6. Treatment of mice with LDN-193189 does not alter numbers of erythroid, myeloid, or granulocytic progenitors, T cells, B cells, or granulocytes

To evaluate the impact of inhibiting BMP signaling on erythropoiesis, we analyzed the abundance (10^3 cells/ μ l) of several erythroid progenitor cell populations, including the MEPs, the CMPs from which they are derived, as well as GMPs in the BM of mice treated with LDN-193189 (3 mg/kg i.p. daily) or vehicle for 21 d. BMP inhibition did not alter numbers of (A) CMPs, (B) GMPs, or (C) MEPs. Furthermore, LDN-193189 did not alter numbers of CD3⁺ T cells (D), B220⁺ B cells, (E) or Mac-1⁺ Gr-1⁺ granulocytes (F).

Figure S7. BM progenitor cells derived from LDN-193189 treated mice retain normal engraftment capacity following transplantation

Host CD45.2⁺ C57BL/6 mice were lethally irradiated and injected with a 1:1 mixture of mononuclear bone marrow cells isolated from CD45.1⁺ mice and from CD45.2⁺ mice that had received LDN-193189 (3 mg/kg i.p. daily) or vehicle for 14 d. The relative contribution to engraftment from different input populations was assessed by flow cytometry analysis of CD45.1⁺ and CD45.2⁺ mononuclear bone marrow cell populations. LDN-193189 treatment of CD45.2⁺ donors did not influence the functionality of their hematopoietic progenitor cells based upon engraftment efficiency. Results are expressed as percentage of BM cells that were CD45.2⁺ (U n=5 mice per group).

Figure S1

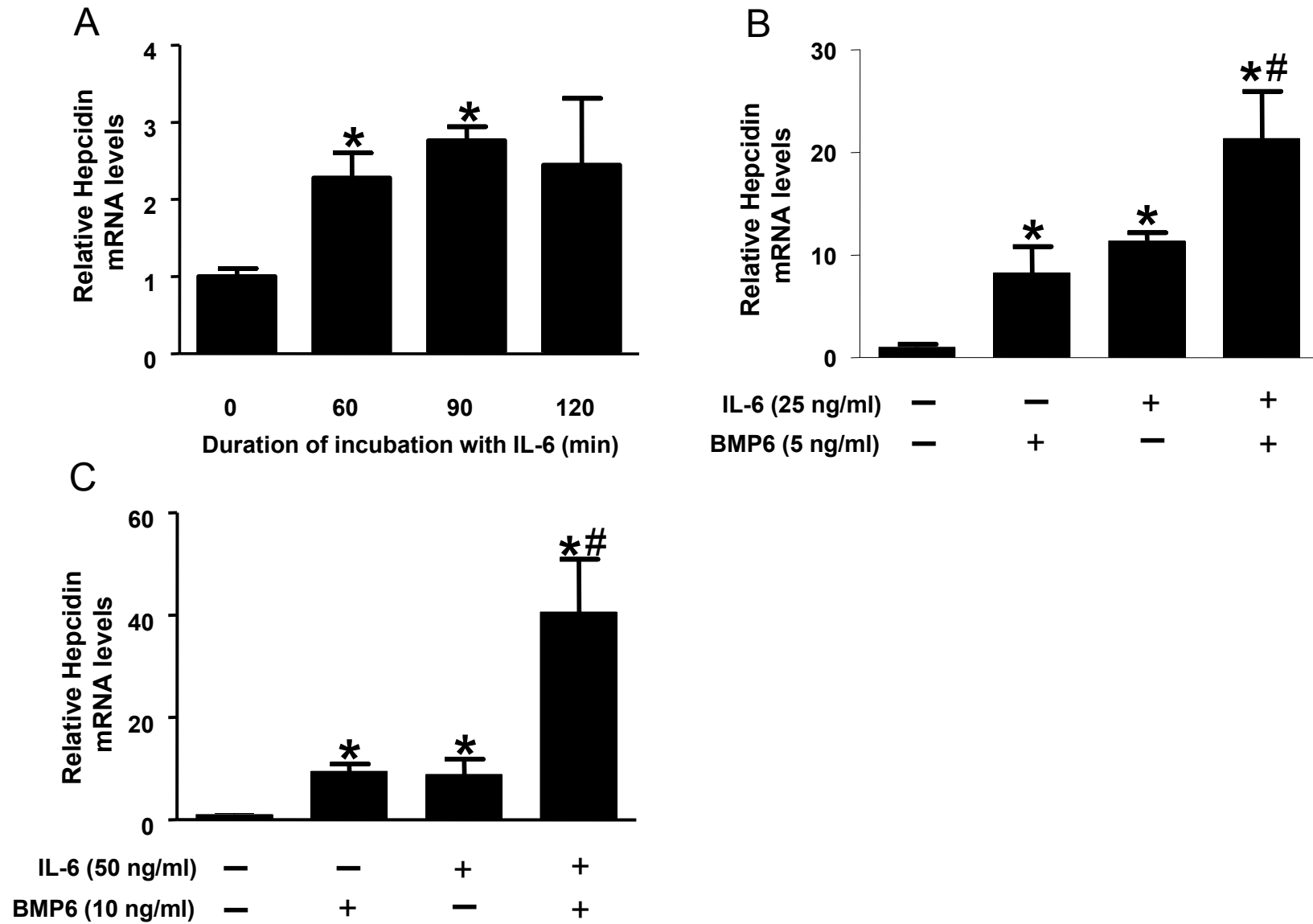


Figure S2

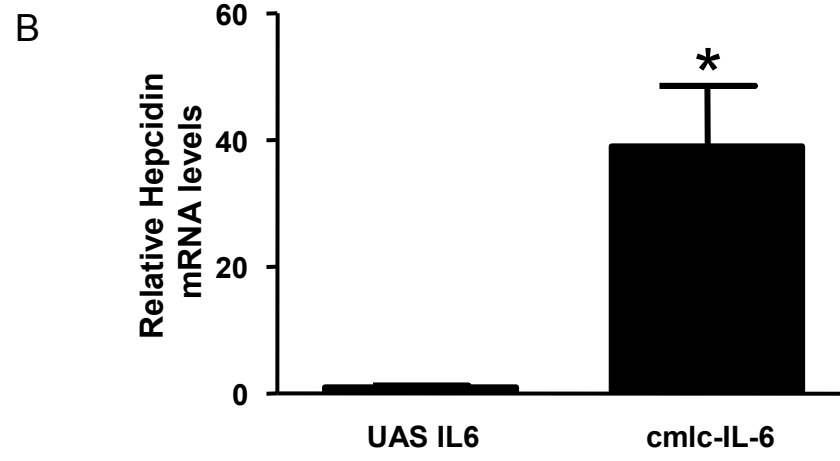
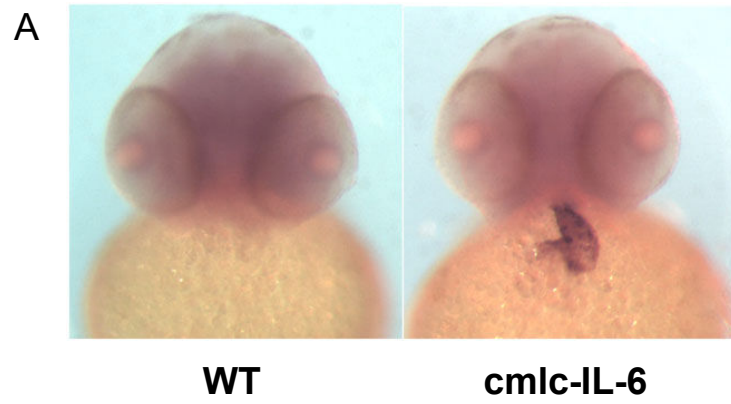


Figure S3

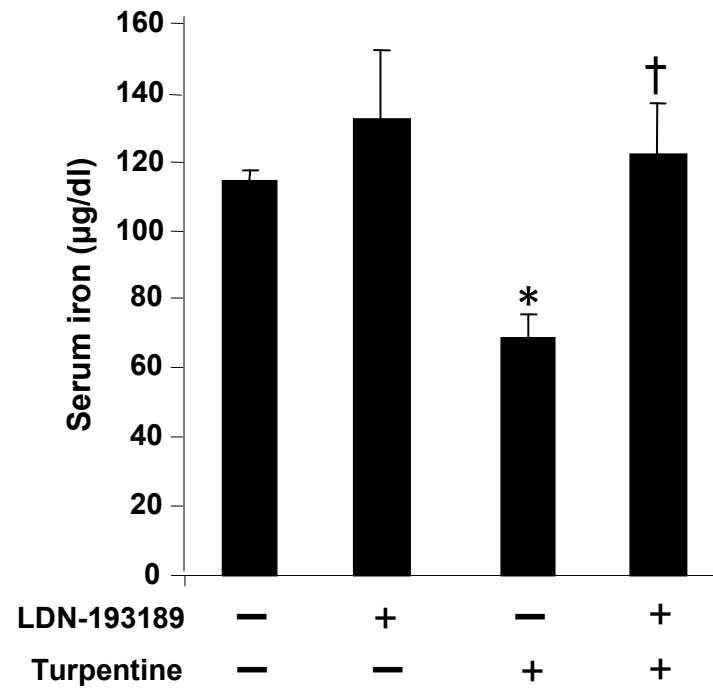


Figure S4

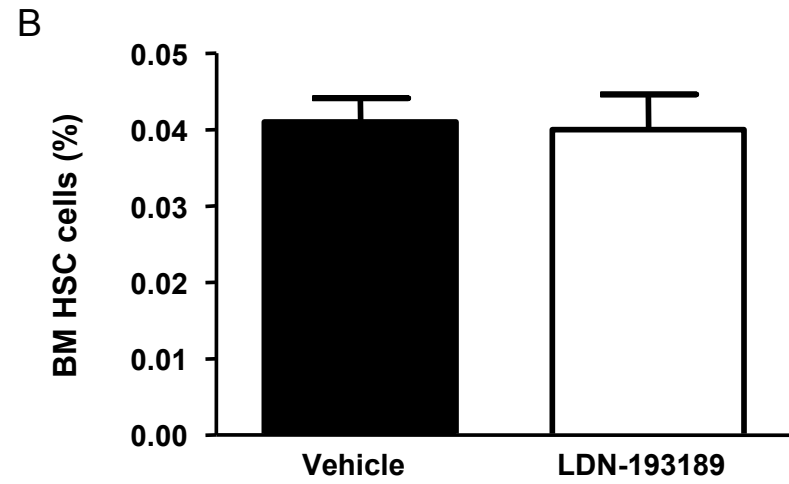
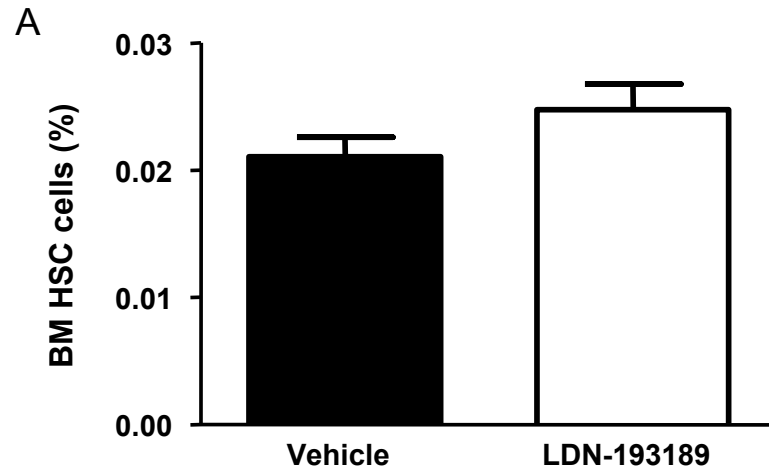


Figure S5

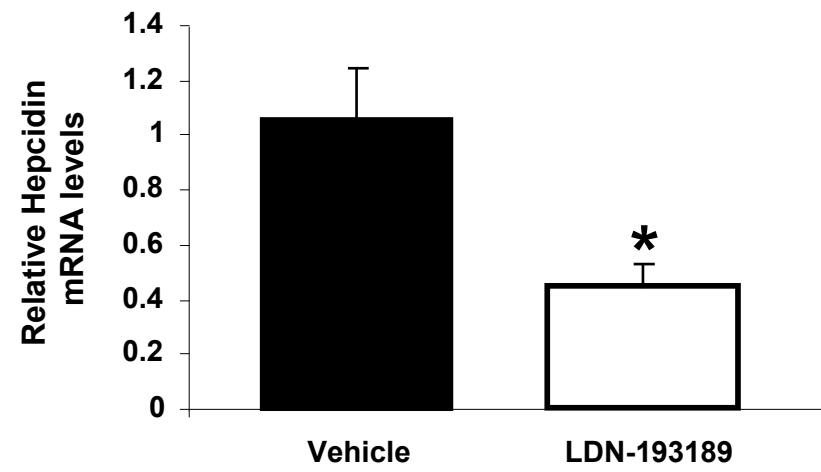


Figure S6

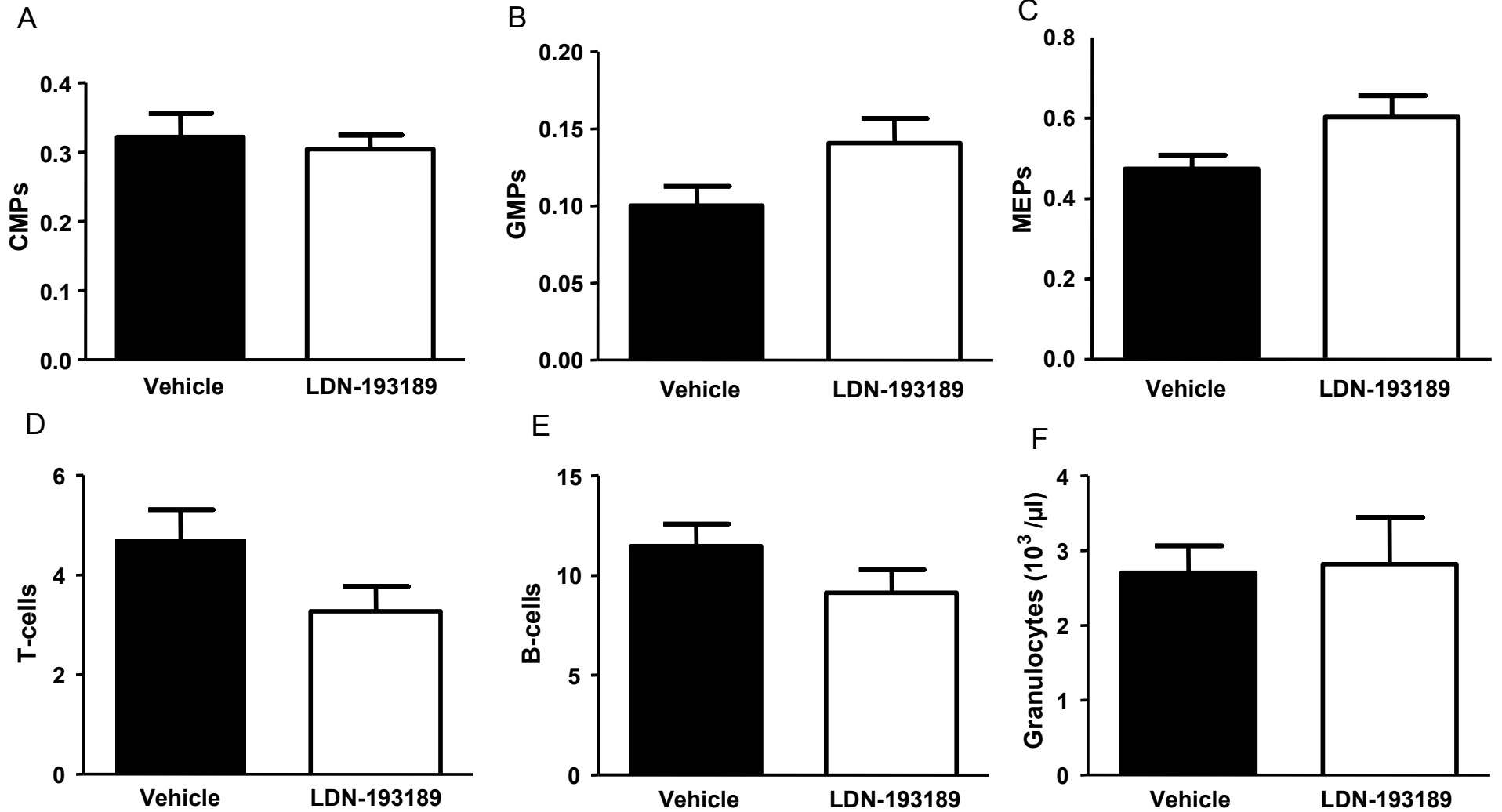


Figure S7

