Predicted editing of additional transfer RNAs in Acanthamoeba castellanii mitochondria

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Recently we demonstrated a novel form of RNA editing that alters the transcripts of four out of a cluster of five tRNA genes in the mitochondrion of *Acanthamoeba castellanii*, an amoeboid protozoan (1). Editing is localized to the 5'-half of the acceptor stem and is confined to the first three base pairs, where it effectively corrects mis-pairing at these sites. Editing occurs in a highly predictable pattern whereby the 5'-mis-matched nucleotide is altered to one that is complementary to its 3'-partner (which is always a pyrimidine), resulting in a standard G-C or A-U.

We describe here a second tightly linked cluster of five tRNA genes, immediately upstream of the large subunit rRNA gene (rnl) (see ref. 1), having the arrangement 5'-trnQ-[-1]-trnK-[4]trnE-[5]-trnI-[5]-trnL-[36]-rnl-3' (numbers in square brackets indicate the lengths of spacers separating the tRNA genes, with trnQ and trnK exhibiting a 1-bp overlap). The 3'-CCA terminus is not encoded by any of these genes, all of which are transcribed in the same direction as *rnl* (from left to right). Secondary structure modeling (see Figure 1) indicates that the transcripts of four of these five genes display the same pattern of mismatching in the acceptor stem as four previously characterized A. castellanii mitochondrial tRNAs that were shown to be edited (1). The predicted sites and nature of editing are indicated by arrows in Figure 1; only trnQ would encode a tRNA with a completely base-paired acceptor stem that would not need editing (secondary structure not shown). Thus, eight out of ten A. castellanii mitochondrial tRNAs characterized to date have been shown or are predicted to undergo RNA editing. In the case described here, the predicted editing comprises one C-to-A, four A-to-G and three U-to-G changes.

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Figure 1.

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