

Supporting Information

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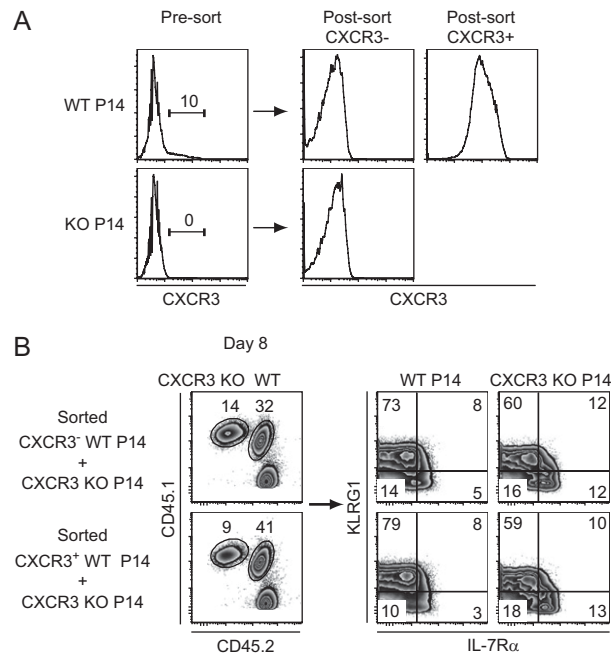


Fig. S1. Early expression of CXCR3 leads to generation of more CD8⁺ T cells at the peak of expansion. (A) Naive WT P14 T cells were sorted based on expression of CXCR3. Histograms are gated on CD8⁺ TCR V α 2⁺ P14 T cells and show expression of CXCR3 on presort and postsort populations. As a control, naive CXCR3-deficient cells were sorted using the same mixture of antibodies used for WT cells. (B) Equal numbers of either CXCR3⁻ or CXCR3⁺ CD45.1/CD45.2 WT P14 cells were cotransferred with CD45.1/CD45.1 CXCR3 KO P14 T cells into C57BL/6 mice that were then infected with LCMV. (Left) Proportion of each population within the CD8⁺ gate in spleen at day 8 postinfection. (Right) Proportion of cells within each P14 population that express KLRG1 and IL-7R α . Data shown are representative of two independent experiments ($n = 4$ for each time).

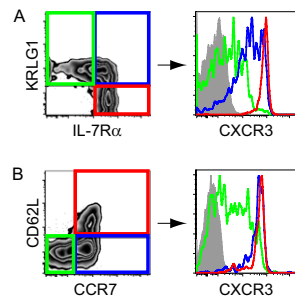


Fig. S2. CXCR3 is differentially expressed on subsets of memory P14 T cells. Memory-cell subsets defined by the expression of KLRG1 and IL-7R α (A) or CD62L and CCR7 (B) were gated for expression of CXCR3 in the spleen at day 60 postinfection. The colored histograms (Right) show CXCR3 expression within memory cell subsets designated by boxes of the same color (Left).

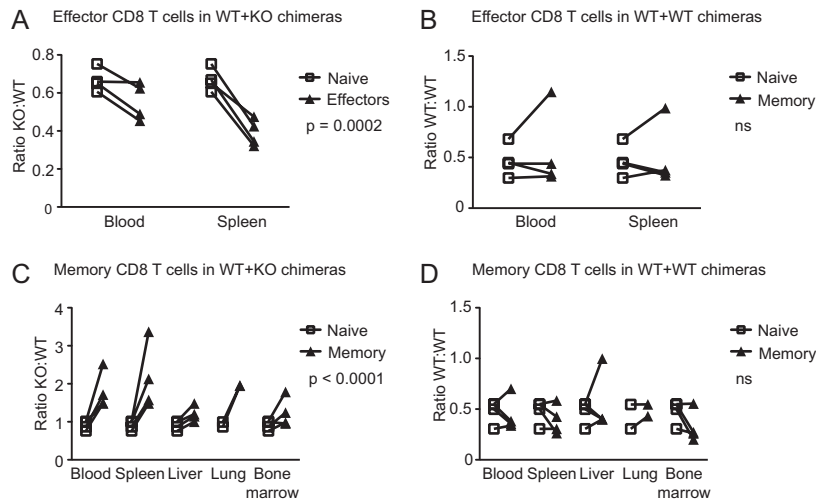


Fig. 53. Non-TCR Tg CXCR3-deficient CD8 T cells generate fewer antigen-specific effector cells and more memory cells after infection. (A–D) Irradiated C57BL/6 mice were reconstituted with a 1:1 mixture of bone marrow cells from CD45.1/CD45.2 WT and CD45.1/CXCR3 KO mice or from CD45.1/CD45.2 WT and CD45.1/CD45.1 WT mice as a control. Following reconstitution, chimeric mice were bled to determine the proportions of CD8⁺ T cells derived from each bone marrow donor population (labeled “Naive”). The mice were then infected with LCMV and analyzed at either day 8 (effectors) or day 220+ (memory) after infection to determine the ratio of tetramer-positive antigen-specific CD8⁺ T cells derived from each bone marrow donor population in the indicated tissues. The empty squares show data before infection, and the filled triangles show tetramer-positive (GP33, GP276, NP396) effector or memory CD8 T cells after infection. A line connects data from individual mice. Data from one set of chimeras are shown ($n = 4$ for each group). Statistics were done using two-way ANOVA.

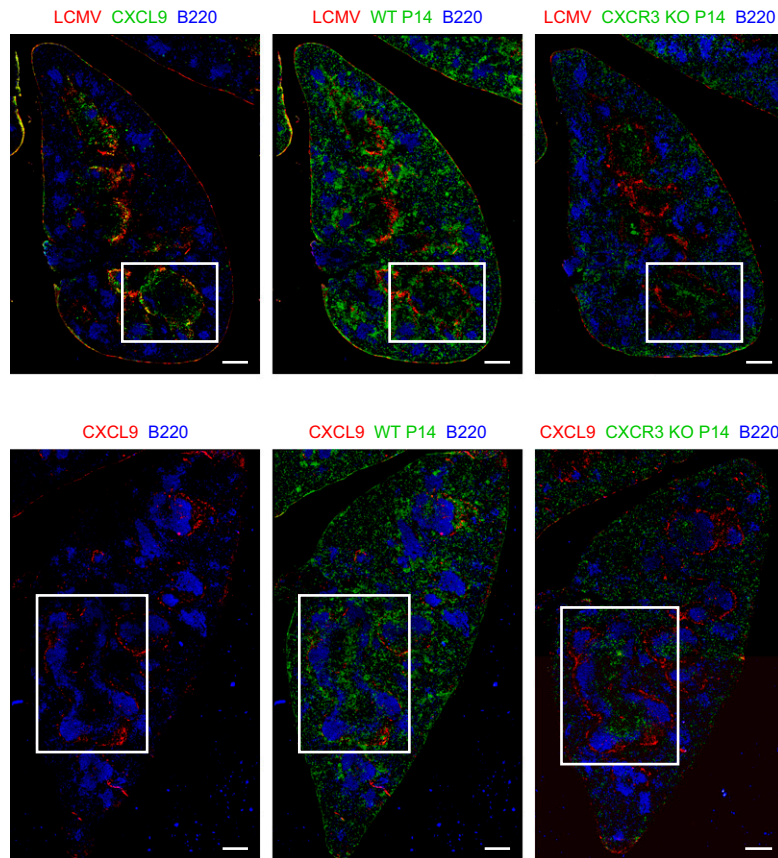


Fig. 54. Effector CD8 T cells colocalize with antigen and CXCL9 in a CXCR3-dependent manner. Equal numbers of Thy1.1⁺ CD45.2/CD45.2 WT and Thy1.2⁺ CD45.1/CD45.1 CXCR3 KO P14 T cells were cotransferred into C57BL/6 recipients that were then infected with LCMV. Consecutive sections were prepared from spleens taken on day 4 postinfection and were stained for viral antigen, CXCL9, WT P14 (Thy1.1), CXCR3 KO P14 (CD45.1), and B220. (Scale bars: 300 μm .) Data are representative of two independent experiments ($n = 4$ for each time). (Insets) Delineation of areas shown at higher magnification in Fig. 5.

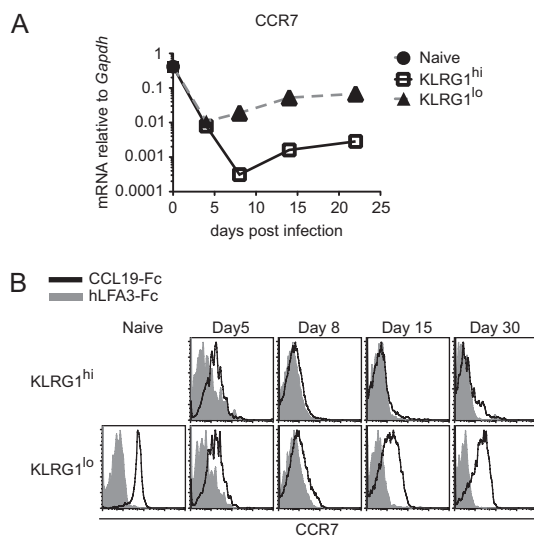


Fig. 55. CCR7 is differentially expressed on KLRG1^{hi} and KLRG1^{lo} effector CD8 T cells. (A) Expression of CCR7 mRNA in sorted KLRG1^{hi} and KLRG1^{lo} effector P14 CD8 T cells at the indicated times after infection was determined by quantitative RT-PCR and is expressed relative to *Gapdh* mRNA. Data shown are averages of three independently sorted samples. (B) Cell surface expression of CCR7 on KLRG1^{hi} and KLRG1^{lo} CD8⁺ effector P14 T cells at the indicated times after infection was determined by flow cytometry. Histograms are gated on P14 T cells or total CD8 T cells for the naive mouse, with isotype control shown as a shaded histogram. Data shown are representative of two independent experiments ($n = 4$ for each time).

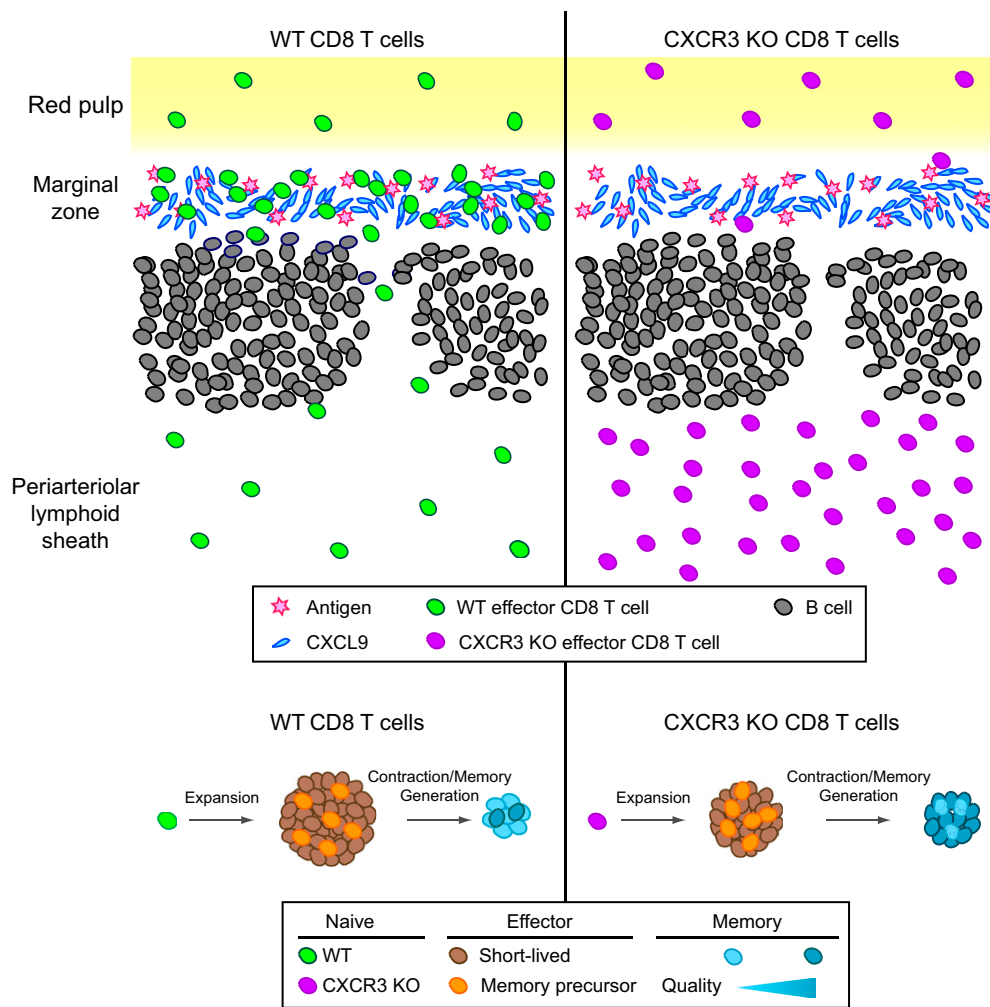


Fig. 56. Model of the contribution of CXCR3 to CD8 T-cell differentiation. In the spleen, marginal zone macrophages and marginal metallophilic macrophages are infected with LCMV and express the CXCR3 ligand CXCL9. Activated effector CD8 T cells express CXCR3 and move toward CXCL9 in marginal zone areas, where they are exposed to antigen. Consequently, WT CD8 T cells encounter more antigen and inflammation than CXCR3-deficient cells. As a result, CXCR3-expressing cells tend to become short-lived effector cells instead of long-lived memory precursors. In contrast, CD8 T cells that lack CXCR3 localize away from marginal zone areas and, as a result, are less exposed to antigen and inflammatory stimuli. This leads to generation of more long-lived memory CD8 T cells that are qualitatively better than WT cells.