

**Supplemental Material For:**

**Title:** Tailoring Enzyme-Rich Environmental DNA Clones: A Source of Enzymes for Generating Libraries of Unnatural Natural Products

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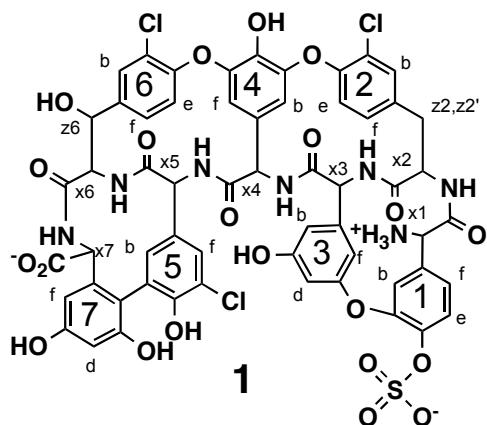
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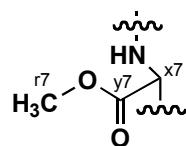
**Table S1:** Numbering Scheme and  $^1\text{H}$  Assignments for Compounds 1-5 Core

	$\delta$ (ppm) Compound 1	$\delta$ (ppm) Compound 2	$\delta$ (ppm) Compound 3	$\delta$ (ppm) Compound 4	$\delta$ (ppm) Compound 5
1b	7.01	7.02	7.00	7.02	6.97
1e	7.61	7.60	7.65	7.61	7.61
1f	7.38	7.35	7.38	7.38	7.33
2b	7.32	7.31	7.32	7.33	7.31
2e	7.21	7.19	7.35	7.22	7.20
2f	7.58	7.57	7.58	7.63	7.57
3b	6.49	6.49	6.49	6.50	6.46
3d	6.62	6.61	6.63	6.62	6.61
3f	6.24	6.27	6.22	6.26	6.21
4b	5.27	5.26	5.27	5.30	5.27
4f	5.68	5.68	5.82	5.70	5.68
5b	6.96	6.97	6.98	6.97	6.95
5f	7.01	6.98	7.02	7.01	7.01
6b	7.60	7.59	7.61	7.60	7.59
6e	7.21	7.18	7.21	7.21	7.20
6f	7.47	7.47	7.51	7.47	7.49
7d	6.52	6.58	6.53	6.52	6.51
7f	6.44	6.23	6.45	6.44	6.43
x1	5.40	5.40	5.40	5.40	4.94
x2	4.94	4.98	4.94	4.94	4.87
x3	5.49	5.49	5.49	5.49	5.43
x4	5.49	5.52	5.49	5.49	5.49
x5	4.35	4.35	4.36	4.36	4.35
x6	4.00	4.03	4.02	4.02	4.00
x7	4.53	4.72	4.53	4.53	4.53
z2	3.35	3.35	3.36	3.34	3.35
z2'	3.10	3.09	3.11	3.11	3.10
z6	5.47	5.43	5.48	5.47	5.47

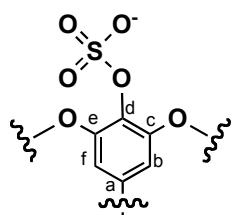
**Table S2:** Compound specific <sup>1</sup>H and <sup>13</sup>C Assignments for Compounds 2-5

The <sup>1</sup>H and <sup>13</sup>C chemical shifts reported in table S2 detail the specific chemical shifts used to make the structural assignments of compounds 2-5. For compound 2, these are the chemical shifts for both the parent compound (Compound 1) and the new compound at the C-terminus of the glycopeptide. For compounds 3 and 4, the chemical shifts reported are the key shifts vital to the assigning the position of the compound specific modification (sulfation and glycosylation, respectively) of the hydroxyl at the top of ring 4. For compound 5, the shifts reported are those new signals and the observed changes in the <sup>13</sup>C at the C- $\alpha$ , x1, at the N-terminus.

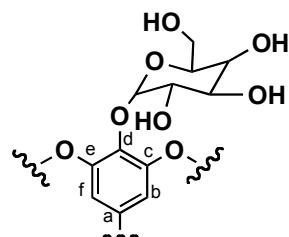
	Compound 1	Compound 2
r7 (1H)	-	3.85
r7 (13C)	-	53.21
y7 (13C)	176.79	172.32
x7 (13C)	59.09	57.11



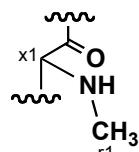
	Compound 1	Compound 3
4a (13C)	135.27	138.22
4b (13C)	107.5	108.93
4c (13C)	146.53	151.65
4d (13C)	132.98	128.41
4e (13C)	147.08	151.84
4f (13C)	104.16	104.22



	Compound 1	Compound 4
4a (13C)	135.27	134.29
4b (13C)	107.5	107.56
4c (13C)	146.53	151.38
4d (13C)	132.98	130.95
4e (13C)	147.08	151.78
4f (13C)	104.16	104.11

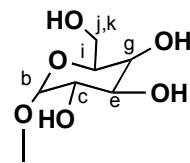


	Compound 1	Compound 5
r1 (1H)	-	2.48
r1 (13C)	-	31.34
x1 (13C)	54.16	64.25

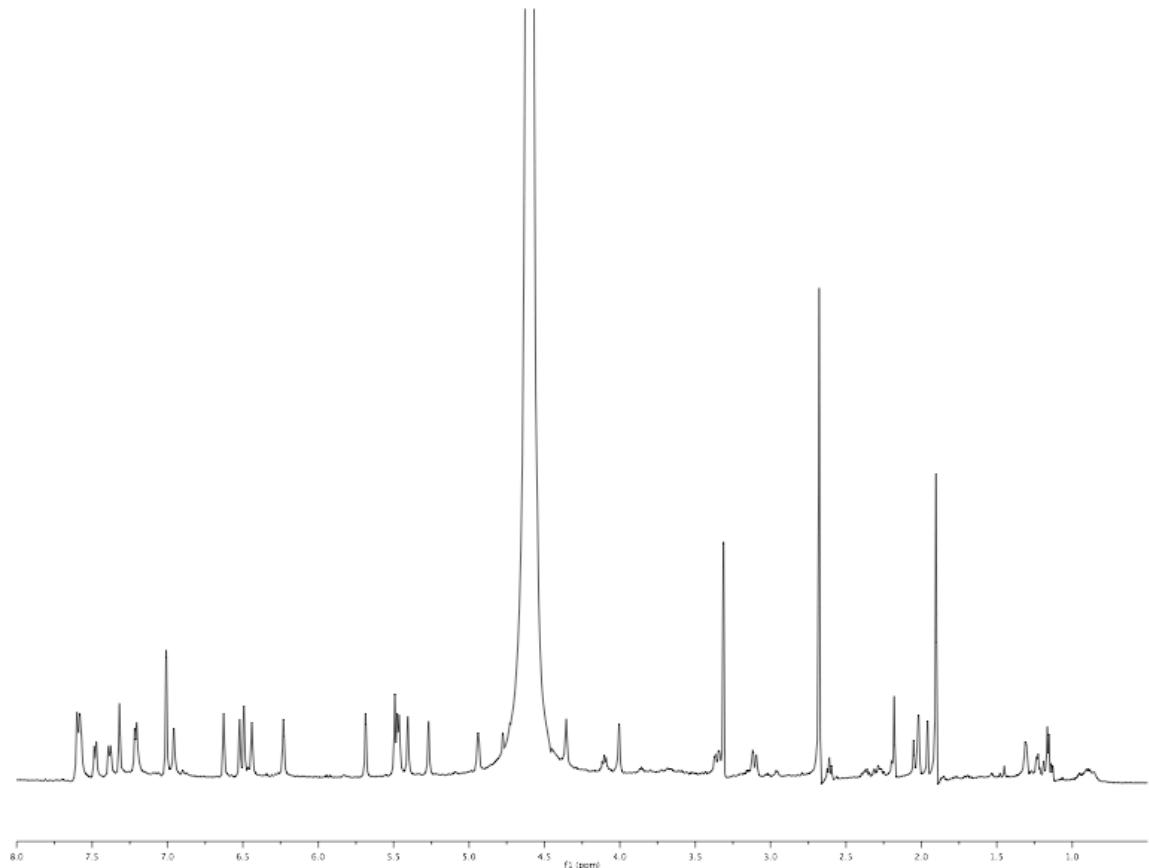


**Table S3:** Numbering scheme and  $^1\text{H}$  and  $^{13}\text{C}$  Assignments for Compound 4 - glucose

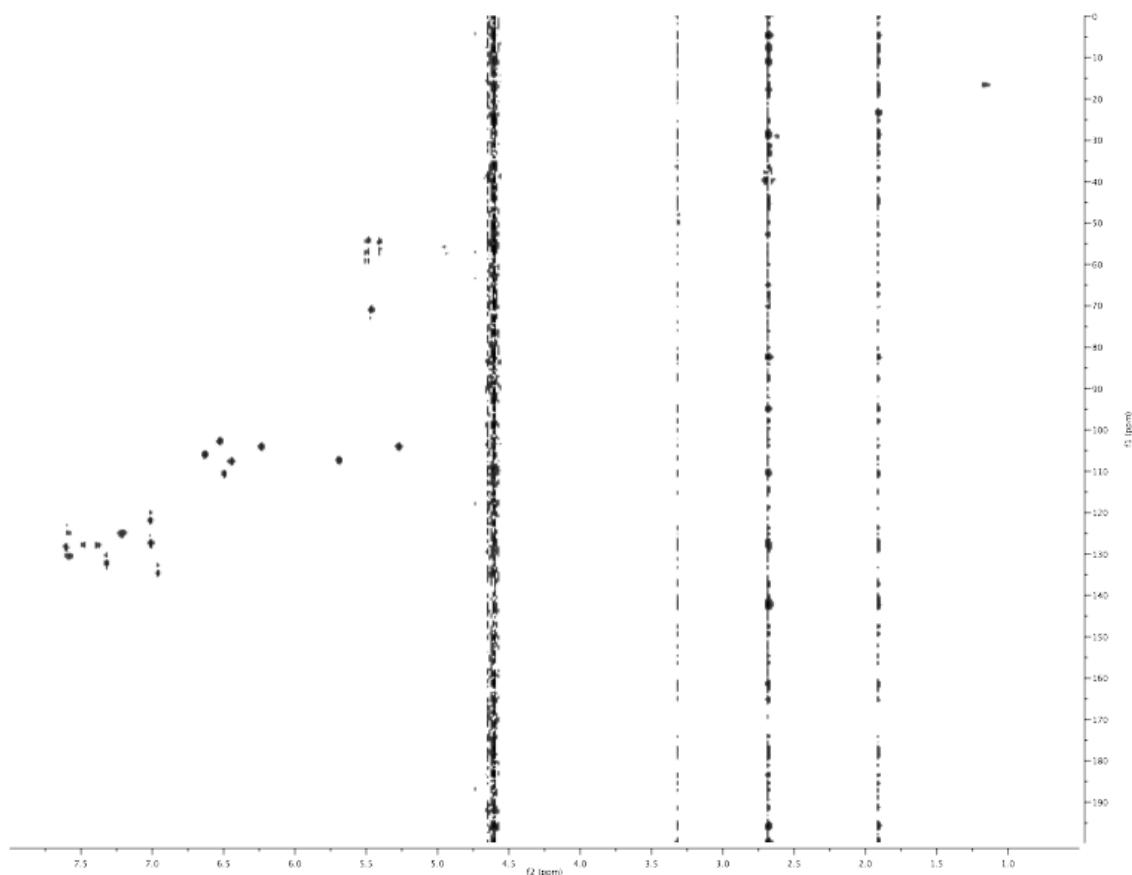
	$^1\text{H}$ (ppm)	$^{13}\text{C}$ (ppm)	Observed 3J (Hz)
b	5.33	104.49	7.80
c	3.75	73.20	8.60
e	3.68	75.28	9.50
g	3.63	68.69	9.50
I	3.56	76.06	-
j	3.88	59.98	5.18
k	3.80	59.98	-



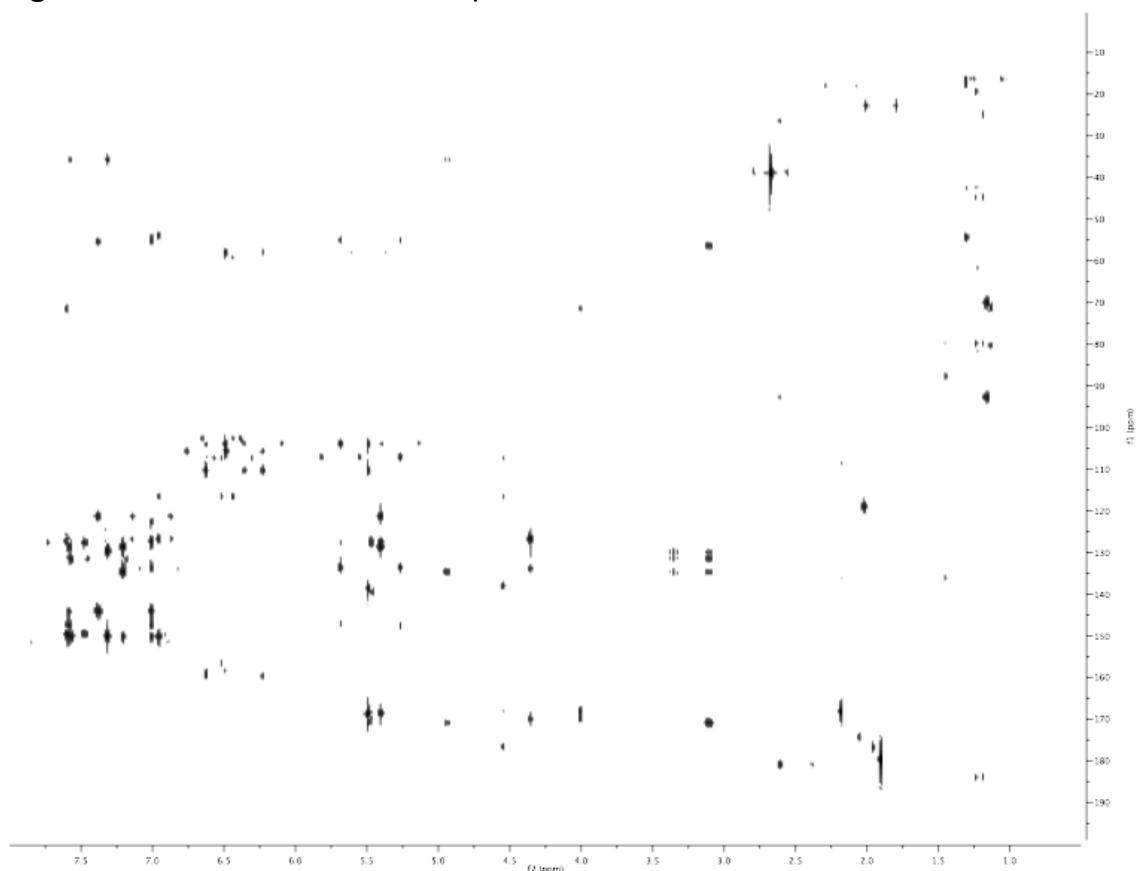
**Figure S1:**  $^1\text{H}$  NMR spectrum of Compound 1, 298K in 3:1  $\text{D}_2\text{O}:\text{CD}_3\text{CN}$



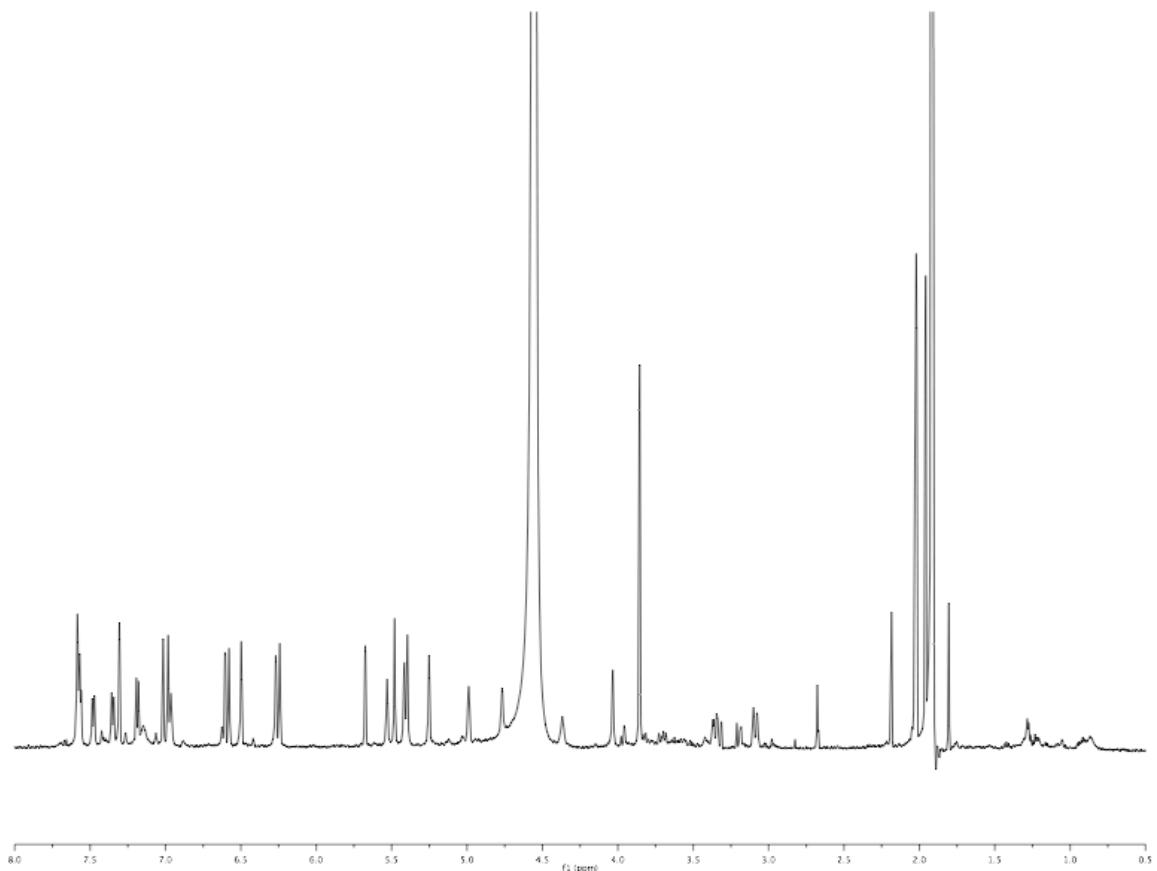
**Figure S2:**  $^1\text{H}$ - $^{13}\text{C}$  HMQC of Compound 1, 298K in 3:1 D<sub>2</sub>O:CD<sub>3</sub>CN



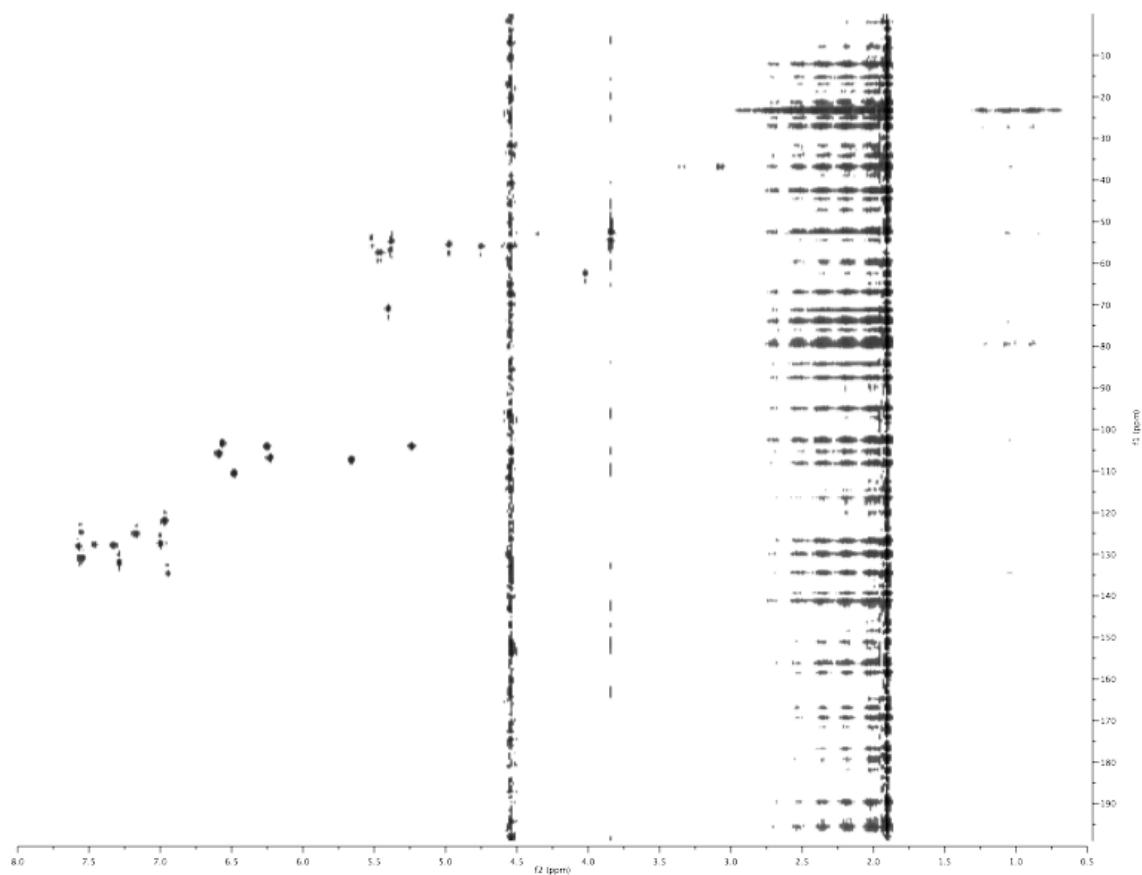
**Figure S3:**  $^1\text{H}$ - $^{13}\text{C}$  HMBC of Compound 1, 298K in 3:1  $\text{D}_2\text{O}:\text{CD}_3\text{CN}$



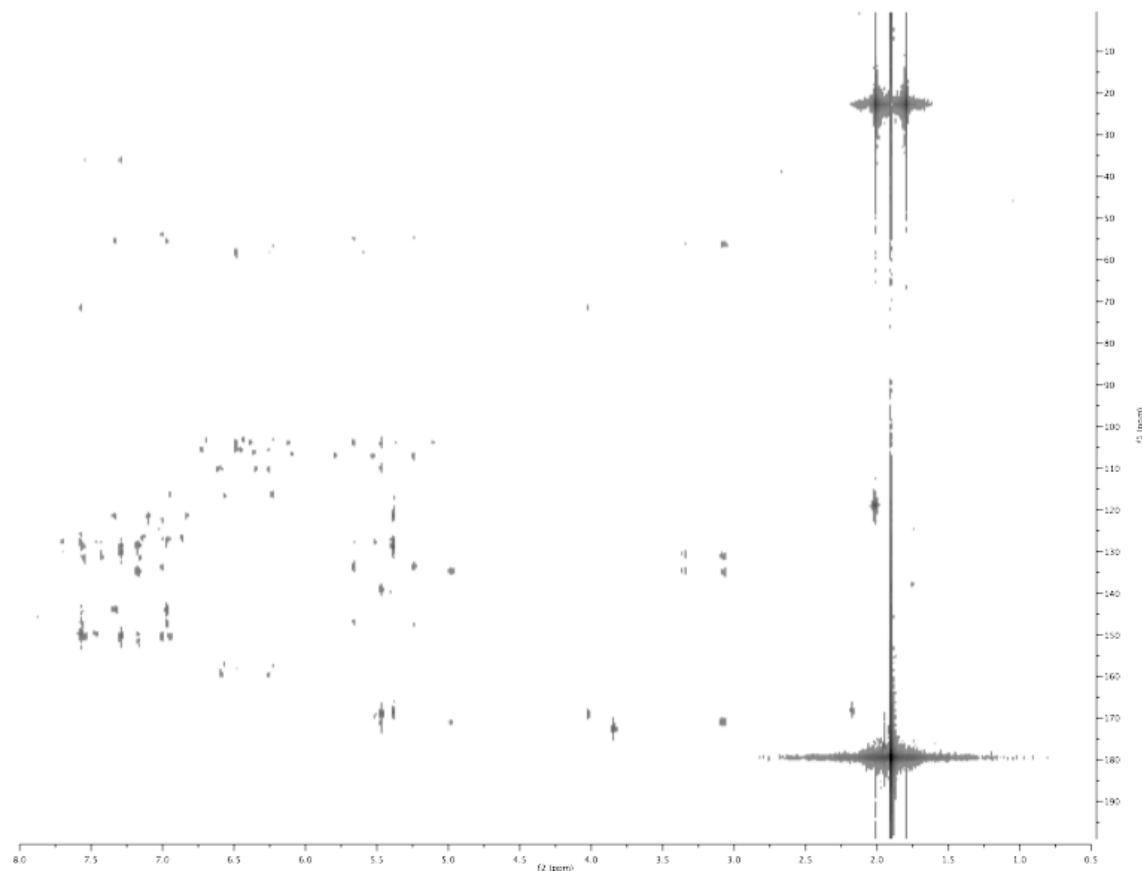
**Figure S4:**  $^1\text{H}$  NMR spectrum of Compound 2, 298K in 3:1  $\text{D}_2\text{O}:\text{CD}_3\text{CN}$



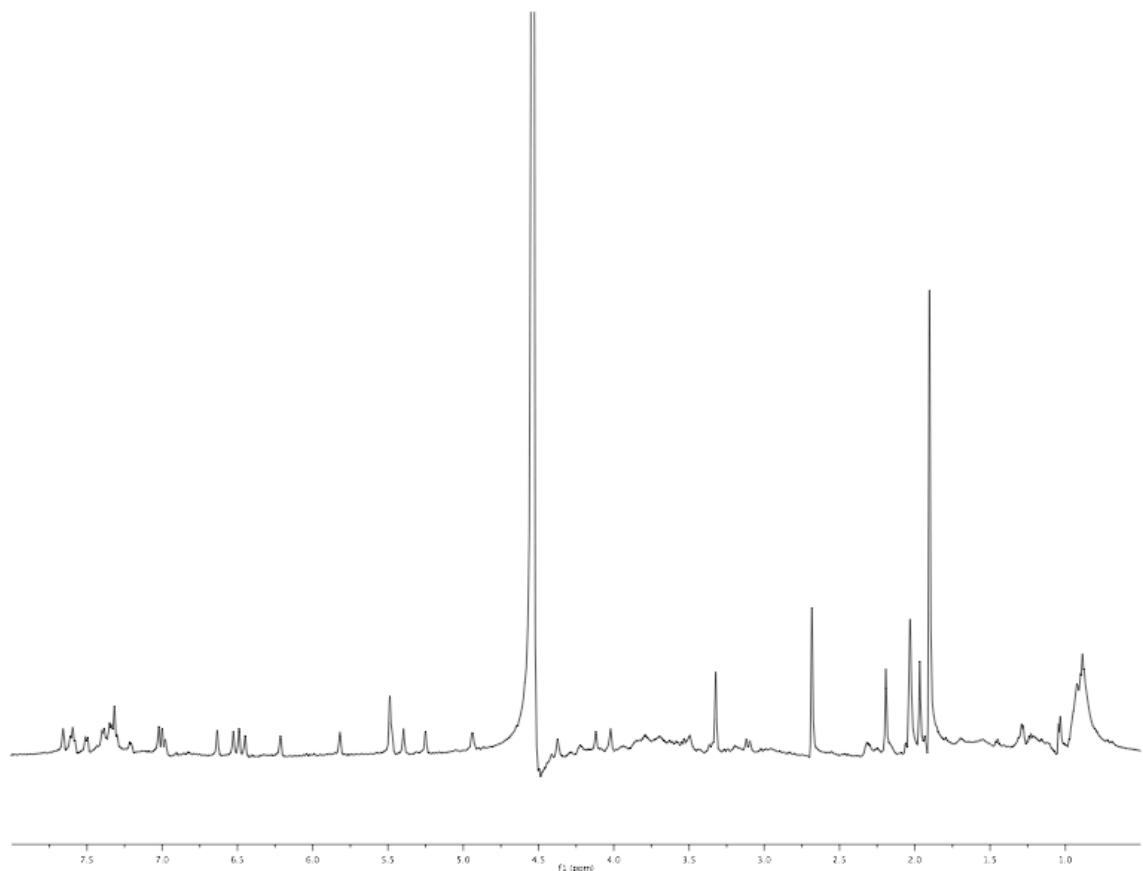
**Figure S5:**  $^1\text{H}$ - $^{13}\text{C}$  HMQC of Compound 2, 298K in 3:1  $\text{D}_2\text{O}:\text{CD}_3\text{CN}$



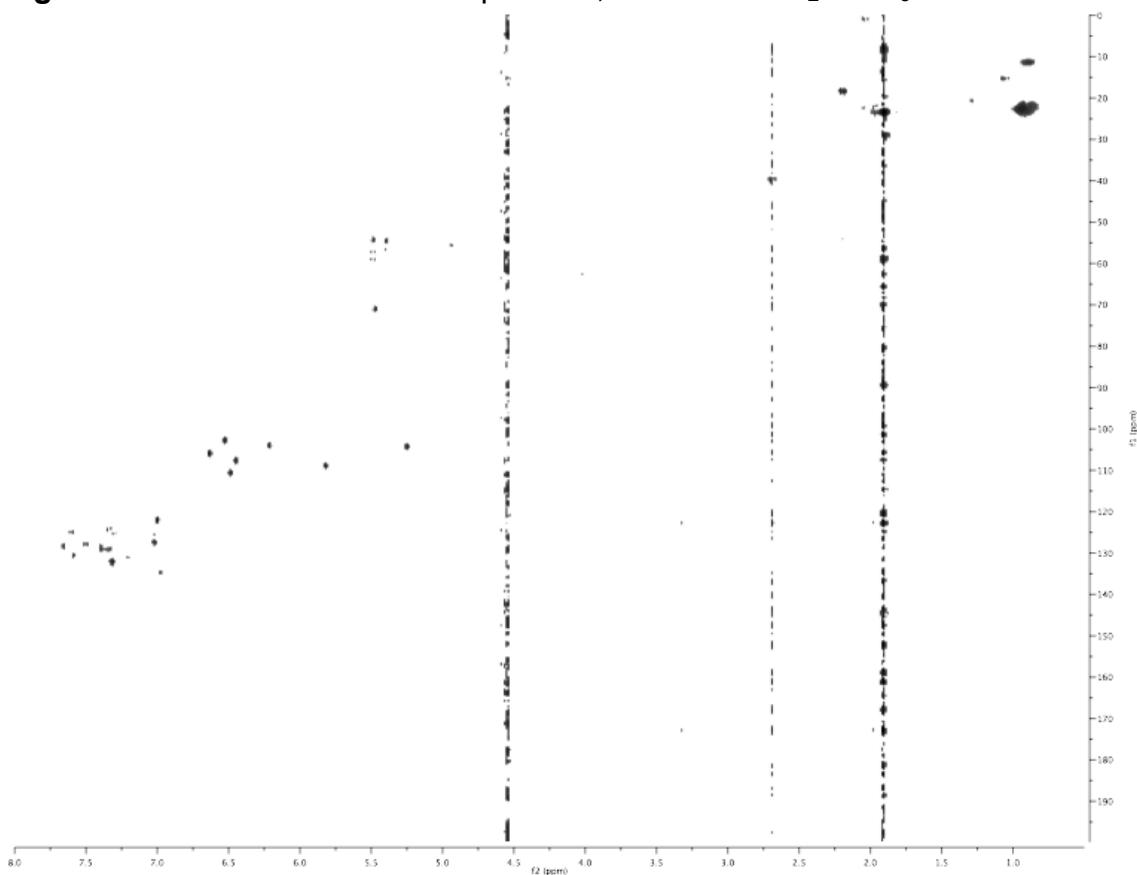
**Figure S6:**  $^1\text{H}$ - $^{13}\text{C}$  HMBC of Compound 2, 298K in 3:1 D<sub>2</sub>O:CD<sub>3</sub>CN



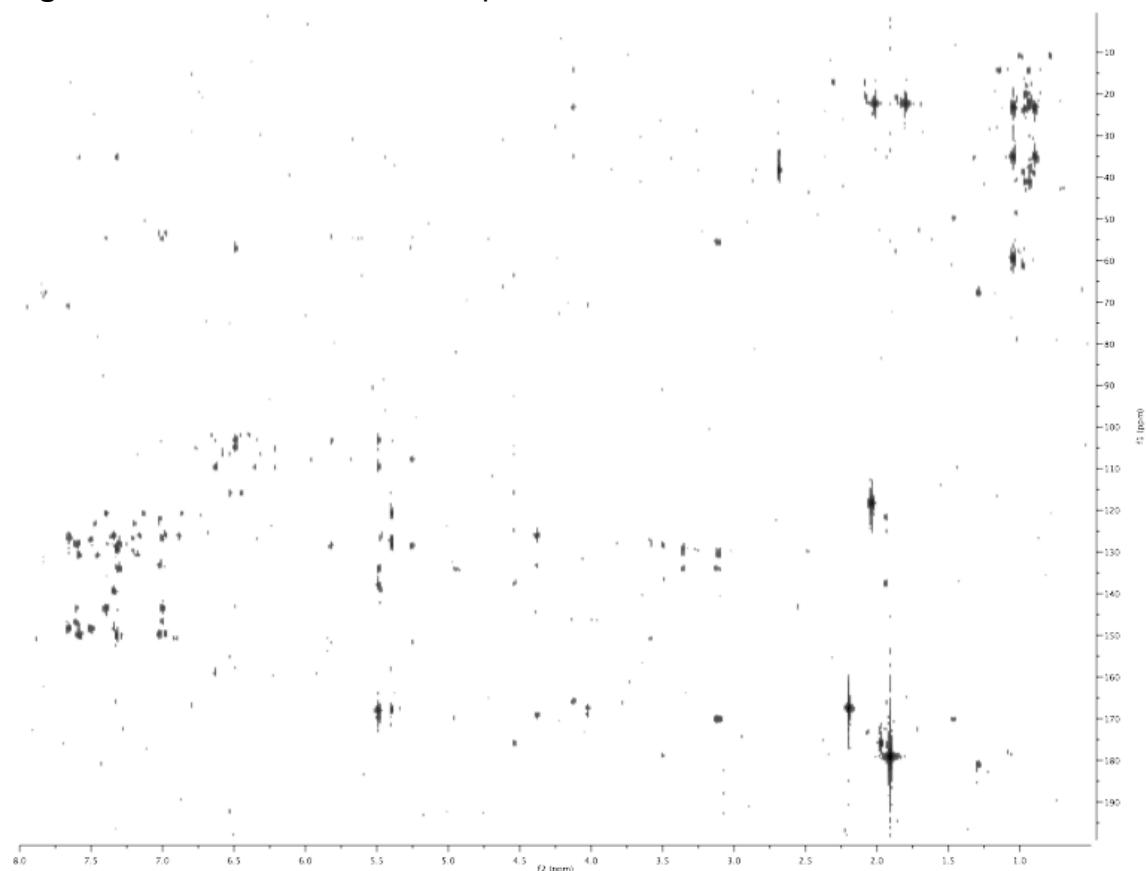
**Figure S7:**  $^1\text{H}$  NMR spectrum of Compound 3, 298K in 3:1  $\text{D}_2\text{O}:\text{CD}_3\text{CN}$



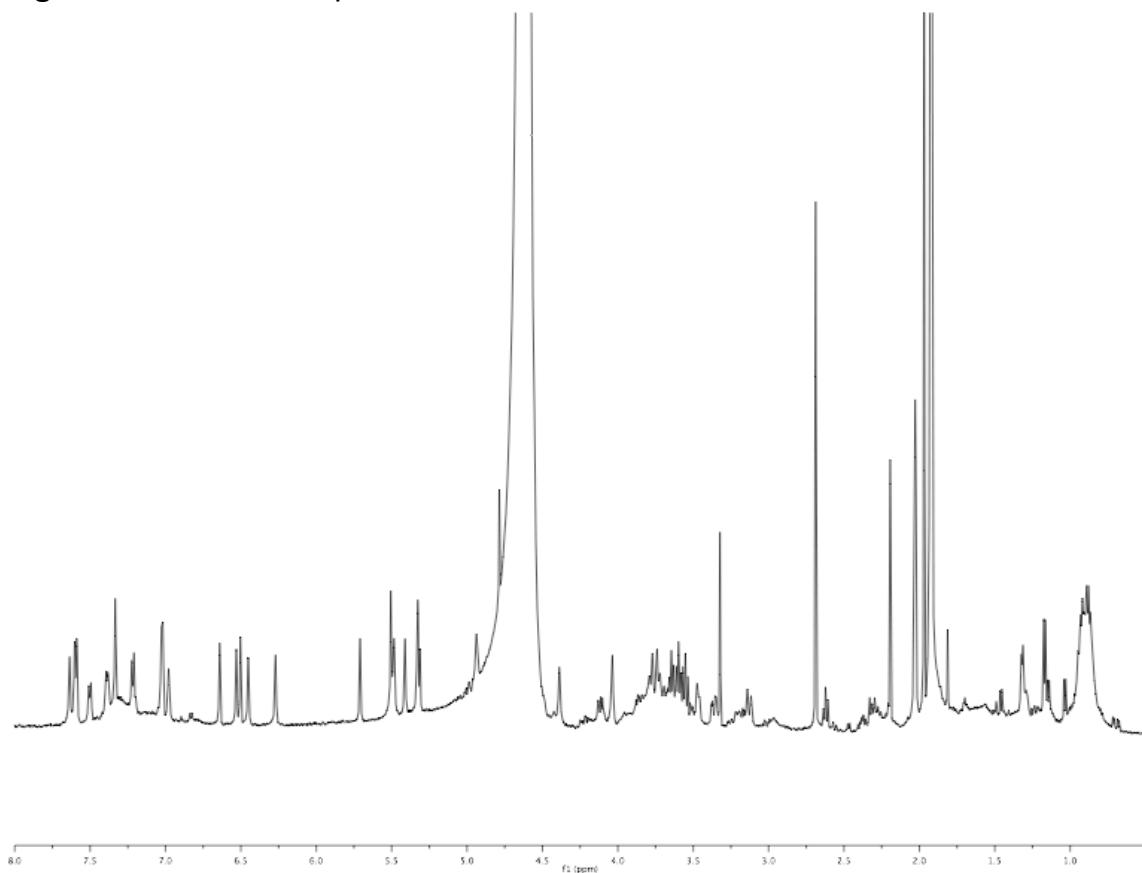
**Figure S8:**  $^1\text{H}$ - $^{13}\text{C}$  HMQC of Compound 3, 298K in 3:1  $\text{D}_2\text{O}:\text{CD}_3\text{CN}$



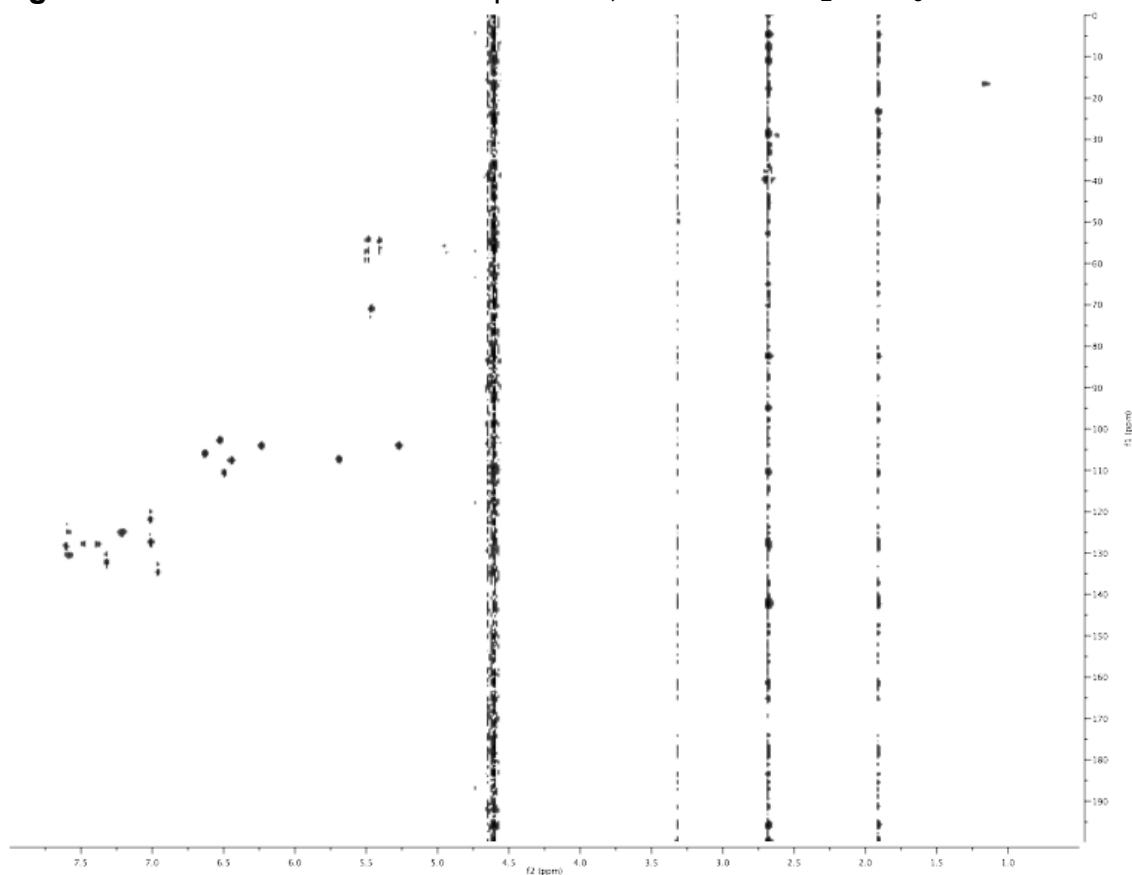
**Figure S9:**  $^1\text{H}$ - $^{13}\text{C}$  HMBC of Compound 3, 298K in 3:1  $\text{D}_2\text{O}:\text{CD}_3\text{CN}$



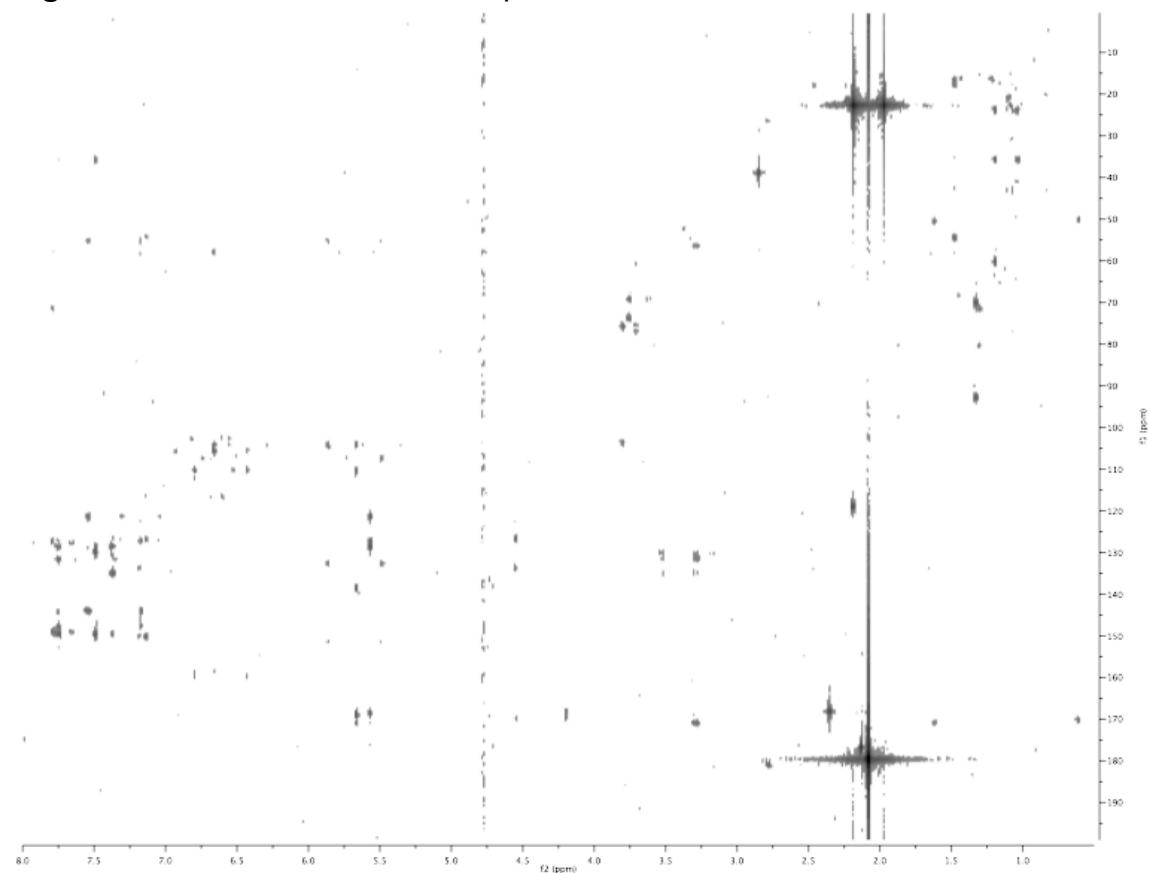
**Figure S10:**  $^1\text{H}$  NMR spectrum of Compound 4, 298K in 3:1  $\text{D}_2\text{O}:\text{CD}_3\text{CN}$



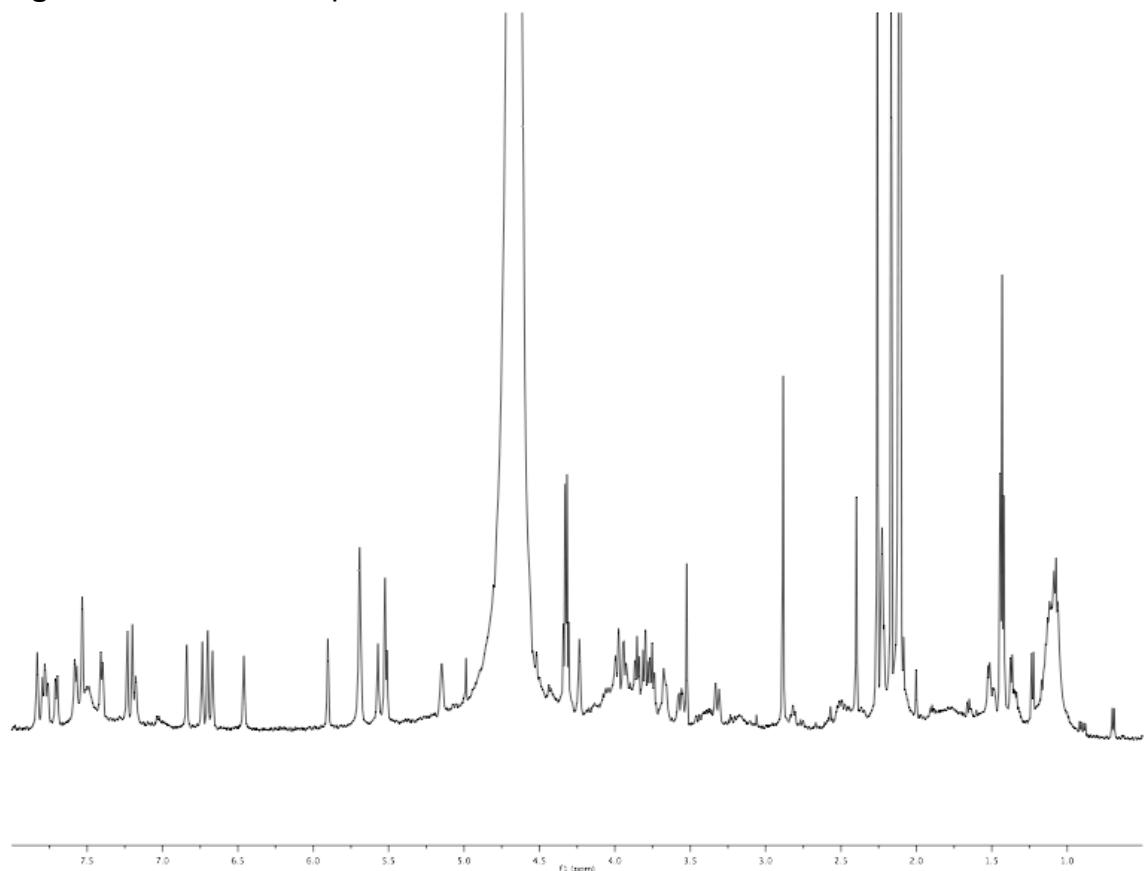
**Figure S11:**  $^1\text{H}$ - $^{13}\text{C}$  HMQC of Compound 4, 298K in 3:1 D<sub>2</sub>O:CD<sub>3</sub>CN



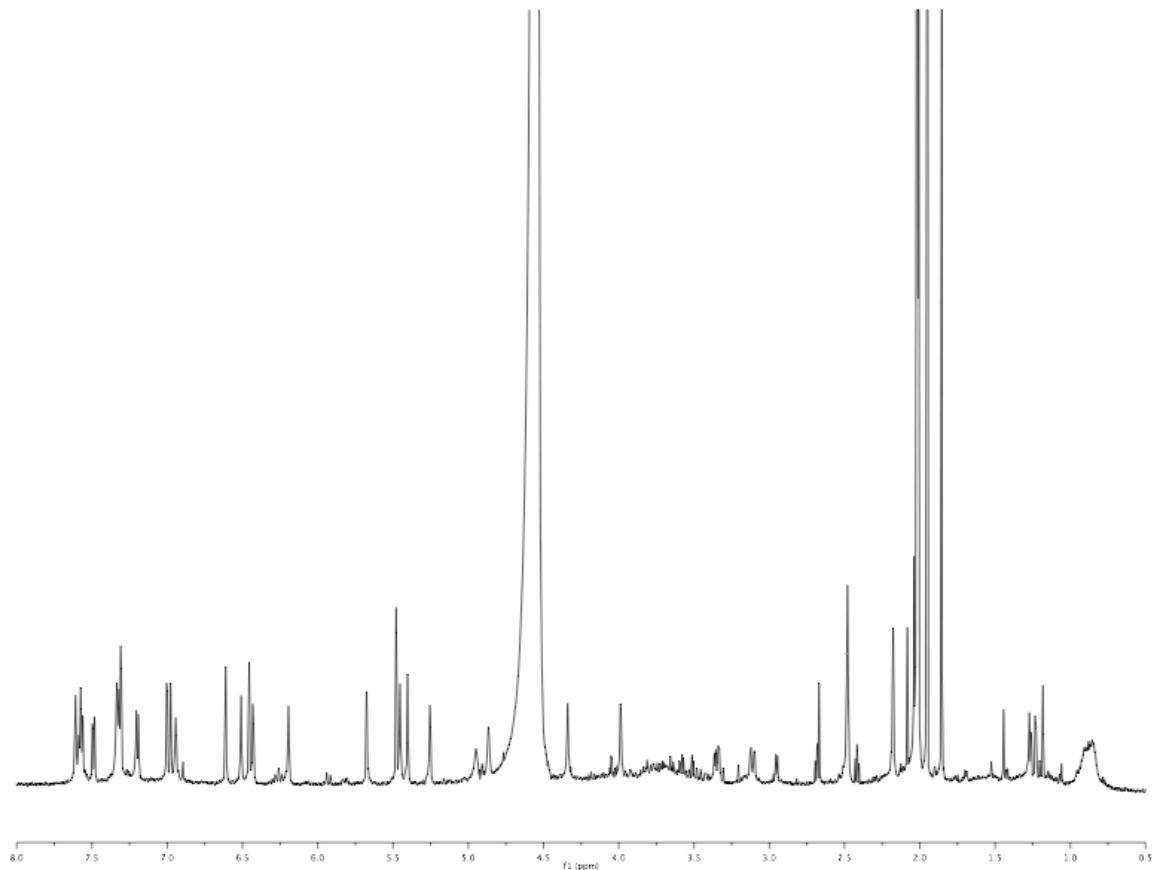
**Figure S12:**  $^1\text{H}$ - $^{13}\text{C}$  HMBC of Compound 4, 298K in 3:1 D<sub>2</sub>O:CD<sub>3</sub>CN



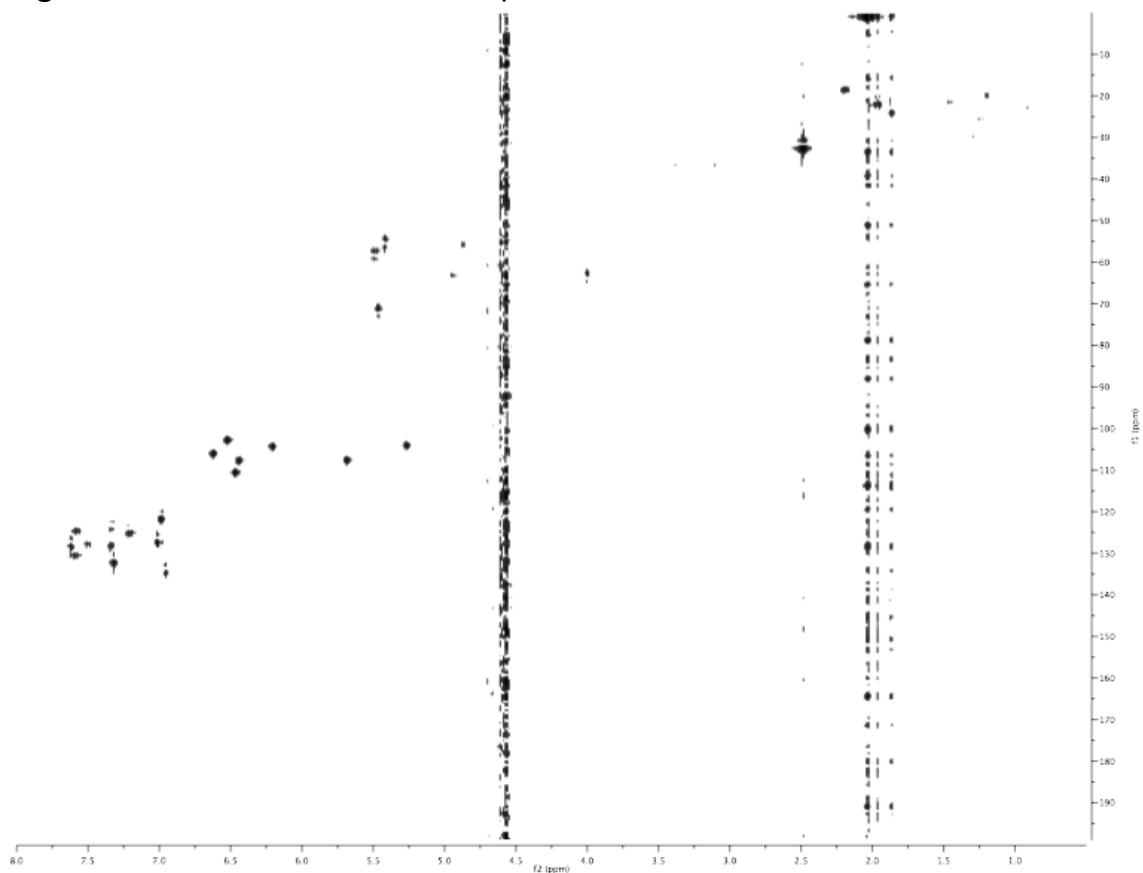
**Figure S13:**  $^1\text{H}$  of Compound 4, 313K in 3:1  $\text{D}_2\text{O}:\text{CD}_3\text{CN}$



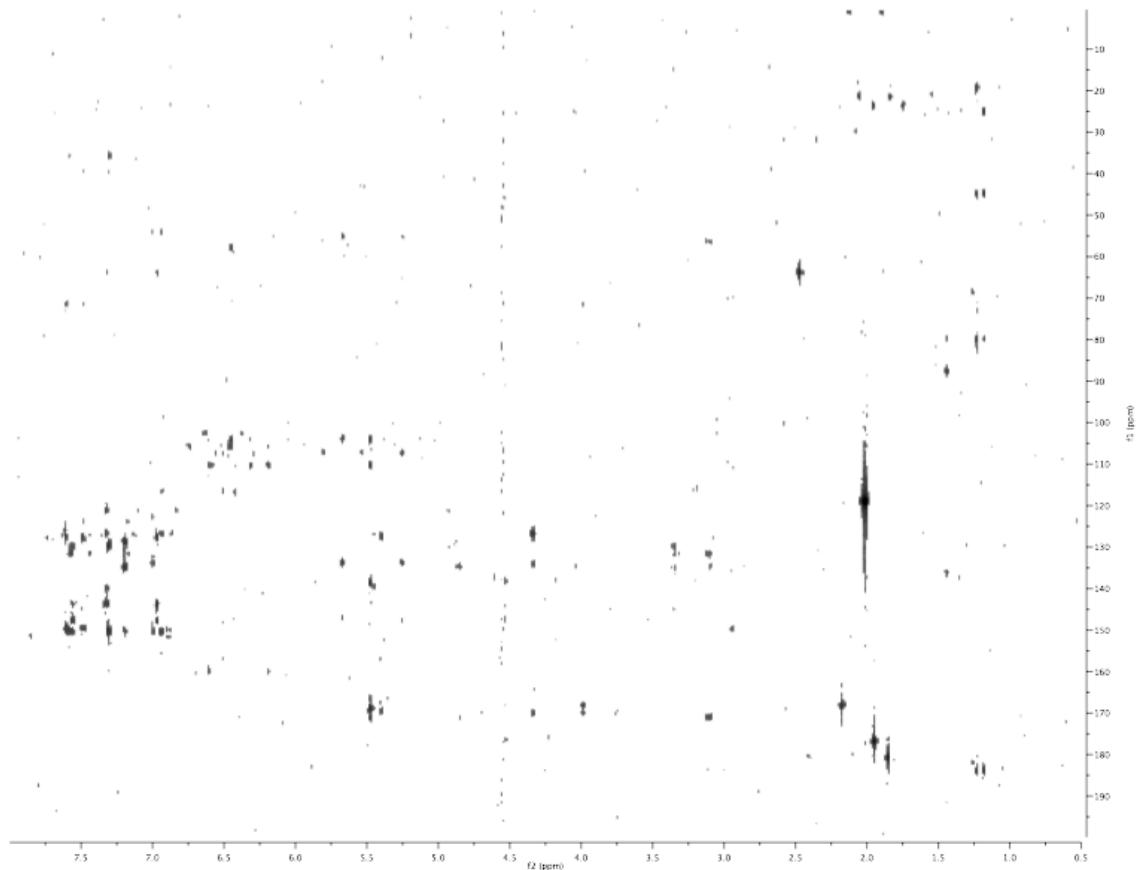
**Figure S14:**  $^1\text{H}$  NMR spectrum of Compound 5, 298K in 3:1  $\text{D}_2\text{O}:\text{CD}_3\text{CN}$



**Figure S15:**  $^1\text{H}$ - $^{13}\text{C}$  HMQC of Compound 5, 298K in 3:1  $\text{D}_2\text{O}:\text{CD}_3\text{CN}$



**Figure S16:**  $^1\text{H}$ - $^{13}\text{C}$  HMBC of Compound 5, 298K in 3:1  $\text{D}_2\text{O}:\text{CD}_3\text{CN}$



**Table S4:** Pathway CA37 ORFs

ORF	Length (AA)	Predicted Gene Product	Glycopeptide Homolog	AA% ID (Organism)	Accession #
1	202	TetR Family Transcriptional Regulator	None	62%( <i>K. flavigena</i> )	ZP_03865120
2	229	Short Chain Oxidoreductase	None	82%( <i>K. flavigena</i> )	ZP_03863529
3	291	Phytanoyl CoA Dioxygenase	None	43%( <i>M. carbonacea</i> )	ZP_04605278
4	217	D-ala-D-ala Peptidase	VanX	85%( <i>A. teichomyceticus</i> )	CAE53345
5	346	D-ala-D-lac Ligase	VanA	86%( <i>A. teichomyceticus</i> )	CAE53344
6	347	D-lactate Dehydrogenase	VanH	85%( <i>A. teichomyceticus</i> )	CAE53343
7	430	D-ala-D-ala Carboxypeptidase	VanY	65%( <i>A. balhimycinina</i> )	CAG25753
8	74	Two-Component Regulatory System	VanR	63% ( <i>A. balhimycinina</i> )	CAG25752
9	387	Two-Component Regulatory System	VanS	72%( <i>A. teichomyceticus</i> )	CAE53347
10	146	N-acetyltransferase	None	38%( <i>B. cepacia</i> )	YP_368498
11	448	UDP-N-acetylMuramoyltri peptide D-ala-Dala Ligase	MurF	78%( <i>A. teichomyceticus</i> )	CAE53342
12	369	DalaDala Ligase	None	62%( <i>T. fusca</i> YX)	YP_289698
13	370	UDP-muramoyl-pentapeptide transferase	MurG	88%( <i>K. flavigena</i> )	ZP_03862660
14	396	Ferredoxin Reductase	None	71%( <i>S. avermitilis</i> )	NP_822785
15	348	C/O-Methyltransferase	None	48%( <i>S. ghanaensis</i> )	ZP_04690473
16	2085	Nonribosomal Peptide Synthetase, modules 1-2	NRPS 1+ 2	79%( <i>A. teichomyceticus</i> )	CAE53350
17	1482	Nonribosomal Peptide Synthetase, module 3	NRPS3	83%( <i>A. teichomyceticus</i> )	CAE53351
18	4075	Nonribosomal Peptide Synthetase, modules 4-6	NRPS 4-6	82%( <i>A. teichomyceticus</i> )	CAE53352
19	1850	Nonribosomal Peptide Synthetase, module 7	NRPS 7	86%( <i>A. teichomyceticus</i> )	CAE53353
20	491	NDP-Hexose-2,3-Dehydratase	EvaA	56%( <i>A. balhimycinina</i> )	CAC48374
21	357	Glucose Thymidyltransferase	Glucose Thymidyltransferase	69%( <i>S. svceus</i> )	YP_002205669
22	378	Glycosyltransferase	GtfA	65%( <i>A. teichomyceticus</i> )	CAE53349
23	407	Glycosyltransferase	GtfB	72%( <i>A. teichomyceticus</i> )	CAE53364
24	572	Mannosyltransferase	MtfA	77%( <i>A. teichomyceticus</i> )	CAE53356
25	629	ABC Transporter	ABC transporter	83%( <i>A. teichomyceticus</i> )	CAE53357
26	392	P450 Monooxygenase OxyA	OxyA	88%( <i>A. teichomyceticus</i> )	CAE53359
27	385	P450 Monooxygenase OxyB	OxyB	86%( <i>A. teichomyceticus</i> )	CAE53360
28	399	P450 Monooxygenase OxyD	OxyD	86%( <i>A. teichomyceticus</i> )	CAE53361
29	411	P450 Monooxygenase OxyC	OxyC	87%( <i>A. teichomyceticus</i> )	CAE53363
30	441	Halogenase	Halogenase	69%( <i>A. teichomyceticus</i> )	CAE53362
31	507	Non-heme Iron Dioxygenase	Non-Heme Iron dioxygenase	93%( <i>A. teichomyceticus</i> )	CAE53366
32	356	DAHP Synthase	DAHP synthase	90%( <i>A. teichomyceticus</i> )	CAE53368
33	331	Str Family Transcriptional Regulator	Transcriptional Regulator	83%( <i>A. teichomyceticus</i> )	CAE53369
34	230	Response Regulator	Res. Regulator (2 comp)	88%( <i>A. teichomyceticus</i> )	CAE53348
35	836	LuxR Regulator	LuxR (2 comp)	78%( <i>A. teichomyceticus</i> )	CAE53370
36	368	Dihydroxyphenylacetic Acid Synthase	DpgA	93%( <i>A. teichomyceticus</i> )	CAE53371
37	222	Enoyl-CoA Hydratase/Iso merase	DpgB	84%( <i>A. teichomyceticus</i> )	CAE53372
38	434	Dihydroxyphenylacetyl-CoA Oxygenase	DpgC	88%( <i>A. teichomyceticus</i> )	CAE53373
39	239	Enoyl-CoA Hydratase/Iso merase	DpgD	91%( <i>A. teichomyceticus</i> )	CAE53374
40	199	GTP Cyclohydrolase	GTP Cyclohydrolase	91%( <i>A. teichomyceticus</i> )	CAE53376
41	420	Hydroxyphenylglycine Transaminase	HpgT	89%( <i>A. teichomyceticus</i> )	CAE53377
42	374	Prephenate Dehydrogenase	Prephenate Dehydrogenase	83%( <i>A. teichomyceticus</i> )	CAG15036
43	301	Transcriptional Regulator	None	50%( <i>S. kasugaensis</i> )	BAC53615
44	329	Transcriptional Regulator	None	50%( <i>S. griseus</i> )	YP_001827443
45	522	Non-heme Iron Dioxygenase	Non-heme Iron Dioxygenase	70%( <i>A. teichomyceticus</i> )	CAG15037
46	352	Hydroxymandelate Synthase	HmaS	92%( <i>A. teichomyceticus</i> )	CAE53378
47	365	Hydroxymandelate Oxygenase	Hmo	87%( <i>A. teichomyceticus</i> )	CAE53379

**Table S5: Pathway CA878 ORFs**

ORF	Length (AA)	Predicted Gene Product	Glycopeptide Homolog	AA %ID (Orgnaism)	Accession #
1	350	D-lactate Dehydrogenase	VanH	74% ( <i>A. teichomyceticus</i> )	CAG15002
2	346	D-ala-D-lac Ligase	VanA	77% ( <i>A. teichomyceticus</i> )	CAE53344
3	203	D-ala-D-ala Dipeptidase	VanX	81% ( <i>A. teichomyceticus</i> )	CAE53345
4	344	Two-Component Regulatory System	VanR	69% ( <i>S. toyocaensis</i> )	AAM80542
5	224	Two-Component Regulatory System	VanS	90% ( <i>S. toyocaensis</i> )	AAM80541
6	161	D-ala-D-ala Carboxypeptidase	VanY	73% ( <i>A. balhimycina</i> )	CAG25753
7	322	Str Family Transcriptional Regulator	Trans Reg	85% ( <i>A. balhimycina</i> )	CAG25754
8	358	Prephenate Dehydrogenase	Prephenate Dehydrogenase	83% ( <i>A. balhimycina</i> )	CAG25755
9	709	ABC Transporter	ABC	81% ( <i>A. teichomyceticus</i> )	CAE53357
10	1858	Nonribosomal Peptide Synthetase, modules 1-2	NRPS 1-2	77% ( <i>A. orientalis</i> )	CAA11794
11	1058	Nonribosomal Peptide Synthetase, modules 3	NRPS 3	80% ( <i>A. teichomyceticus</i> )	CAE53351
12	4078	Nonribosomal Peptide Synthetase, modules 4-6	NRPS 4-6	84% ( <i>A. orientalis</i> )	CAA11795
13	1835	Nonribosomal Peptide Synthetase, modules 7	NRPS 7	87% ( <i>A. orientalis</i> )	CAA11796
14	364	P450 Monooxygenase OxyA	OxyA	81% ( <i>A. teichomyceticus</i> )	CAE53359
15	386	P450 Monooxygenase OxyB	OxyB	78% ( <i>A. teichomyceticus</i> )	CAE53360
16	356	P450 Monooxygenase OxyD	OxyD	86% ( <i>A. orientalis</i> )	CAA11798
17	346	DAHP Synthase	DAHP Synthase	72% ( <i>A. teichomyceticus</i> )	CAE53368
18	411	P450 Monooxygenase OxyC	OxyC	81% ( <i>A. teichomyceticus</i> )	CAG15021
19	492	Halogenase	Halogenase	93% ( <i>A. orientalis</i> )	CAA11780
20	388	Glycosyltransferase	GtfA	70% ( <i>A. orientalis</i> )	AAB49292
21	409	Glycosyltransferase	GtfB	76% ( <i>A. orientalis</i> )	AAB49293
22	407	Glycosyltransferase	GtfC	72% ( <i>A. orientalis</i> )	AAB49294
23	278	Methyltransferase	None	54% ( <i>S. coelicolor</i> )	NP_628799
24	385	Glycosyltransferase	GtfA	69% ( <i>A. teichomyceticus</i> )	CAE53349
25	286	Deacetylase	Deacetylase	71% ( <i>A. teichomyceticus</i> )	CAE53355
26	577	Mannosyltransferase	Mannosyltransferase	79% ( <i>A. teichomyceticus</i> )	CAE53356
27	347	C/O-Methyltransferase	None	51% ( <i>S. ghanaensis</i> )	ZP_04690473
28	435	Phenylglycine Aminotransferase	HpgT	89% ( <i>A. balhimycina</i> )	CAC48367
29	277	BHT Perhydrolase	Bhp	92% ( <i>A. orientalis</i> )	CAA11784
30	581	BHT Peptide Synthetase	BpsD	92% ( <i>A. orientalis</i> )	CAA11773
31	397	BHT Oxygenase	OxyD*	89% ( <i>A. orientalis</i> )	CAA11772
32	354	Hydroxymandelate Synthase	HmaS	82% ( <i>A. orientalis</i> )	CAA11761
33	359	Hydroxymandelate Oxidase	Hmo	88% ( <i>A. orientalis</i> )	CAA11762
34	456	Na+/H+ Antiporter	Putative Antiporter	85% ( <i>A. balhimycina</i> )	CAC48373
35	477	NDP-Hexose-2,3-Dehydratase	EvaA	90% ( <i>A. balhimycina</i> )	CAC48374
36	326	NDP-Hexose-4-Ketoreductase	EvaE	86% ( <i>A. orientalis</i> )	CAA11764
37	370	NDP-Hexose-3-Aminotransferase	EvaB	91% ( <i>A. orientalis</i> )	CAA11782
38	206	NDP-Hexose-3,5-Epimerase	EvaD	82% ( <i>A. orientalis</i> )	CAA11781
39	608	Mannosyltransferase	None	64% ( <i>S. avermitilis</i> )	NP_822291
40	373	Dihydroxyphenylacetic Acid Synthase	DpgA	94% ( <i>A. orientalis</i> )	CAA11765
41	219	Enoyl-CoA Hydratase/Isoenzyme	DpgB	85% ( <i>A. balhimycina</i> )	CAC48379
42	379	Dihydroxyphenylacetyl-CoA Oxygenase	DpgC	87% ( <i>A. orientalis</i> )	CAA11787
43	268	Enoyl-CoA Hydratase/Isoenzyme	DpgD	89% ( <i>A. balhimycina</i> )	CAC48381

**Table S6:** Pathway CA915 ORFs

ORF	Length (AA)	Predicted Gene Product	Glycopeptide Homolog	
1	330	D-ala-D-ala Ligase	None	61% ( <i>T. fusca</i> YX) YP_289698
2	214	D-ala-D-ala Dipeptidase	VanX	81% ( <i>A. teichomyceticus</i> ) CAE53345
3	347	D-ala-D-lac-Ligase	VanA	79% ( <i>A. teichomyceticus</i> ) CAE53344
4	419	Betalactamase	None	51% ( <i>A. mirum</i> ) ZP_03818051
5	277	Phytanoyl CoA Dioxygenase	None	46% ( <i>M. carbonacea</i> ) ZP_04605278
6	448	UDP-N-acetyl muramoyl tripeptide-D-ala-D-ala Ligase	MurF	75% ( <i>A. teichomyceticus</i> ) CAE53342
7	333	D-lactate Dehydrogenase	VanH	70% ( <i>A. teichomyceticus</i> ) CAG15002
8	399	Two-component Regulatory System	VanS	67% ( <i>Frankia sp. EAN1pec</i> ) YP_001511361
9	230	Two-component Regulatory System	VanR	87% ( <i>S. toyocaensis</i> ) AAM80541
10	206	D-ala-D-ala Carboxypeptidase	VanY	54% ( <i>S. nassauensis</i> ) ZP_04486505
11	368	C/O-Methyltransferase	None	50% ( <i>S. ghanaensis</i> ) ZP_04690473
12	1054	Nonribosomal Peptide Synthetase Module 1	NRPS1	75% ( <i>S. toyocaensis</i> ) AAM80539
13	1106	Nonribosomal Peptide Synthetase Module 2	NRPS2	76% ( <i>A. teichomyceticus</i> ) CAE53350
14	1060	Nonribosomal Peptide Synthetase Module 3	NRPS3	77% ( <i>A. teichomyceticus</i> ) CAE53351
15	4068	Nonribosomal Peptide Synthetase Modules 4-6	NRPS4-6	78% ( <i>A. teichomyceticus</i> ) CAE53352
16	1872	Nonribosomal Peptide Synthetase Module 7	NRPS7	83% ( <i>A. teichomyceticus</i> ) CAE53353
17	501	NDP-Hexose-2,3-Dehydratase	EvaA	66% ( <i>S. cyanogenus</i> ) AAD13549
18	357	Glucose Thymidylyltransferase	Glucose Thymidylyltransferase	70% ( <i>S. svceus</i> ) YP_002205669
19	375	Glycosyltransferase	GtfA (glycosyltransferase)	61% ( <i>A. orientalis</i> ) AAB49292
20	408	Glycosyltransferase	GtfB (glycosyltransferase)	70% ( <i>Nonomuraea sp.</i> ) CAD91204
22	407	Glycosyltransferase	GtfC (glycosyltransferase)	67% ( <i>A. orientalis</i> ) AAB49294
21	274	Methyltransferase Type 11	None	57% ( <i>Frankia sp. EAN1pec</i> ) YP_001508433
23	381	Glycosyltransferase	GtfA (glycosyltransferase)	63% ( <i>A. teichomyceticus</i> ) CAE53349
24	374	NDP-Hexose-3-Aminotransferase	EvaB	80% ( <i>S. arenicola</i> ) YP_001537186
25	201	NDP-Hexose-3,5-Epimerase	EvaD	68% ( <i>S. arenicola</i> ) YP_001537187
26	334	NDP-Hexose-4-Ketoreductase	EvaE	64% ( <i>S. venezuelae</i> ) AAL14256
27	238	Methyltransferase Type 12	None	74% ( <i>S. arenicola</i> ) YP_001539299
28	579	Mannosyltransferase	Mannosyltransferase	71% ( <i>A. teichomyceticus</i> ) CAE53356
29	659	ABC Transporter	ABC Transporter	81% ( <i>A. teichomyceticus</i> ) CAE53357
30	231	Str Family Transcriptional Regulator	None	42% ( <i>S. griseus</i> ) YP_001822146
31	392	P450 Monooxygenase, OxyA	OxyA	84% ( <i>A. teichomyceticus</i> ) CAE53359
32	385	P450 Monooxygenase, OxyD	OxyD	80% ( <i>A. teichomyceticus</i> ) CAE53360
33	399	P450 Monooxygenase, OxyB	OxyB	78% ( <i>A. teichomyceticus</i> ) CAE53361
34	411	P450 Monooxygenase, OxyC	OxyC	86% ( <i>A. teichomyceticus</i> ) CAE53363
35	412	Halogenase	Halogenase	66% ( <i>A. teichomyceticus</i> ) CAE53362
36	531	Non-Heme Iron Dioxygenase	Non-Heme Iron Dioxygenase	84% ( <i>A. teichomyceticus</i> ) CAE53366
37	352	DAHP Synthase	DAHP Synthase	87% ( <i>A. teichomyceticus</i> ) CAE53368
38	339	Str Family Transcriptional Regulator	Str Family Trans Reg	77% ( <i>A. teichomyceticus</i> ) CAE53369
39	513	Non-Heme Iron Dioxygenase	Non-Heme Iron Dioxygenase	56% ( <i>S. toyocaensis</i> ) AAM80528
40	389	Hydroxymandelate Oxygenase	Hmo	70% ( <i>S. toyocaensis</i> ) AAM80552
41	806	Transcriptional Regulator	Transcriptional Regulator	58% ( <i>A. teichomyceticus</i> ) CAE53370
42	199	GTP Cyclohydrolase	GTP Cyclohydrolase	92% ( <i>A. teichomyceticus</i> ) CAE53376
43	420	Hydroxyphenylglycine Transaminase	HpgT	88% ( <i>A. teichomyceticus</i> ) CAG15035
44	122	Chorismate Mutase	None	68% ( <i>N. dassonvillei</i> ) ZP_04335576
45	354	Hydroxymandelate Synthetase	HmaS	64% ( <i>Frankia sp. Cc13</i> ) YP_481550
46	365	Prephenate Dehydrogenase	Prephenate Dehydrogenase	73% ( <i>A. teichomyceticus</i> ) CAG15036
47	369	Dihydroxyphenylacetic Acid Synthase	DpgA	84% ( <i>A. teichomyceticus</i> ) CAE53371
48	221	Enoyl-CoA Hydratase/Isoenzyme	DpgB	67% ( <i>A. teichomyceticus</i> ) CAE53372
49	434	Dihydroxyphenylacetyl-CoA Oxygenase	DpgC	78% ( <i>A. teichomyceticus</i> ) CAE53373
50	239	Enoyl-CoA Hydratase/Isoenzyme	DpgD	83% ( <i>A. teichomyceticus</i> ) CAE53374
51	395	UDP muramoyl pentapeptide transferase	MurG	60% ( <i>S. arenicola</i> ) YP_001535200

\*This is the fourth oxidative enzyme found in balhimycin biosynthesis, OxyD, which is essential for the biosynthesis of β-HT. This differs from the OxyD which forms one of the four macrocycles in the biosynthesis of type III and type IV glycopeptides.