

Figure S1. Timeline of revascularization in OIR and depiction of laser-capture microdissection

(A) Representative photo-micrographs of *Griffonia simplicifolia* lectin-stained retinal flatmounts illustrate the progression of vascular growth following the initial vaso-obliteration in response to oxygen exposure from P7-12. At P12, immediately after exposure to hyperoxia, retinas present maximal vascular loss. As the retina attempts to revascularize, there is an initial delay in regrowth followed by misdirected pre-retinal vascular tuft formation that peaks at P17. Subsequently at P19-21, vessels enter the avascular retina, as the pre-retinal neovascularization regresses. By P23, the vaso-obiterated zone is fully revascularized. (B) Representative retinal cross-section of lectin-stained normal vessels before laser-capture, compared to (pre-retinal) vascular tufts. Dotted lines represent the laser dissection.

Figure S2. VEGF is upregulated in the central avascular region of the OIR retina

Micro-dissection of avascular (A) and vascularized (V) areas of retinas from mice (P14) subjected to OIR, reveals a ~3-fold induction in VEGF protein (by Western blotting) specifically in the avascular center during pathological neovascularization (n=4). There is no significant difference in VEGF expression between peripheral (P) and central (C) normoxic retina at P14. Values are shown relative to peripheral and vascularized retinas \pm s.e.m. *p=0.019 compared to corresponding vascularized area (V).

Figure S3. Expression of IL-1R1 in RGC-5 and astrocytes. (A) Protein expression of IL-

1R1 in unstimulated rat RGC-5 (by Western blot); human Tf-1 cells were used as positive control (n=3).

(B) Confocal immunohistochemical imaging of retinal cross-sections from P14 mice subjected to OIR reveals expression of IL-1R in astrocytes (GFAP positive); note merged images. n=3. Scale bar: 25 μ m.

Figure S4. Intravitreal injection of lentivirus efficiently infects RGCs. (A) LV vectors containing a GFP transgene under a CMV promoter (Lv.GFP) were injected intravitreally into mouse pups at P2 and infected ~70% of RGCs as detected by co-localization with the RGC-specific marker Thy1.1. at P14 (n=3). (B) Intravitreal injections of Lv.shSema3A at P2 resulted in a ~38% reduction at P8 (n=3, *p=0.043) and ~54% reduction at P14 (n=3, *p=0.034) of Sema3A protein, relative to values for corresponding Lv.shGFP. (C, D) Specificity of shSema3A was confirmed as neither VEGF nor other related Semaphorins such as Sema3D, Sema3E or Sema3F were downregulated. n=3; ***p<0.005 compared to corresponding Lv.shGFP. Scale bar (A) 50 μ m.

Figure S5. Lv.shSema3A protects against vaso-obliteration and neovascularization

(A) Lectin-stained retinal flatmounts reveal that Lv.shSema3A treated retinas present significantly less vasoobliteration (67% relative to controls). n=13-15, **p=0.01 compared to control. (B) Neovascular areas shown in Figure 3B were quantified using Swift NV¹; representative quantification masks are presented.

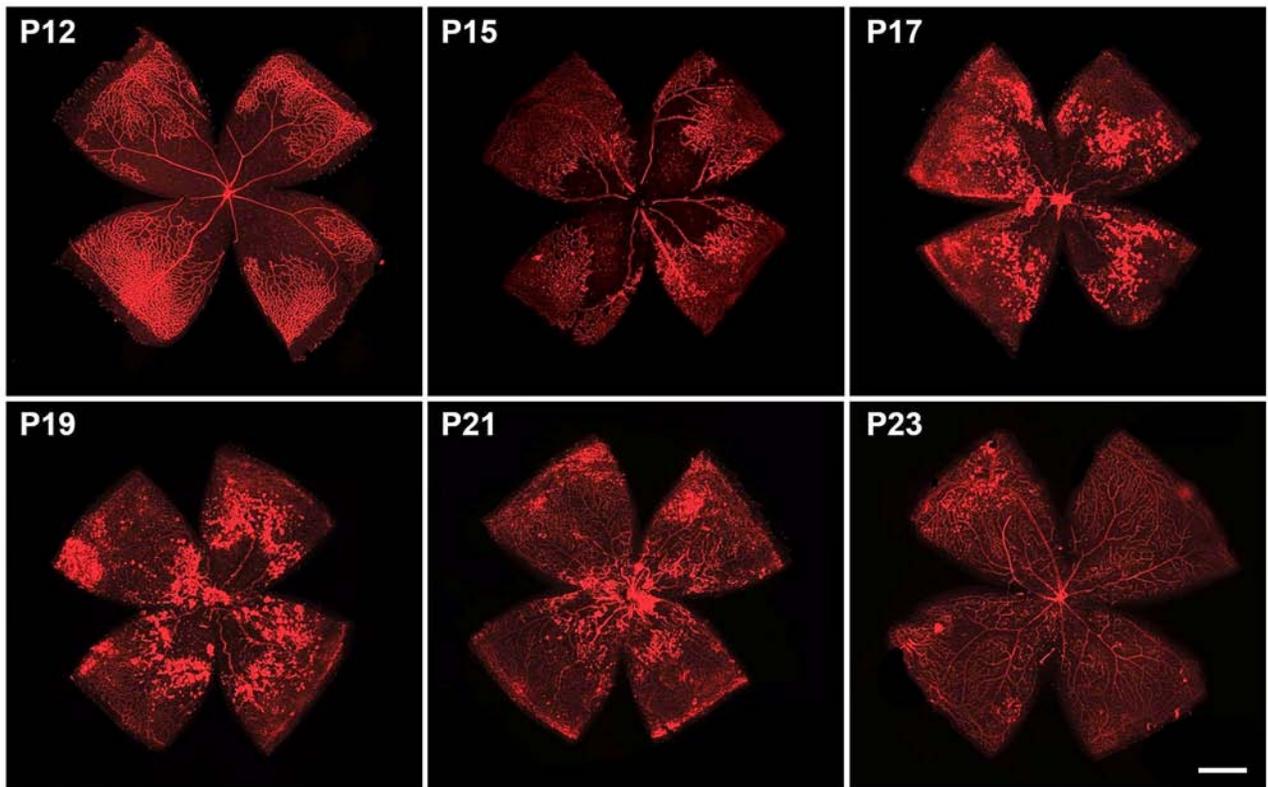
Figure S6. *In vitro* assessment of RGC-derived Sema3A. (A) Lentivirus efficiently infects cultured RGCs and shSema3A reduces Sema3A expression by ~50%. n=3; ***p<0.01 compared to control. (B) Time lapse images (0 to 45 min) of EC contraction upon stimulation with rSema3A (1 μ g/ml) or vehicle (control). (C) Pull-down of activated RhoA-GTP reveals that this permissive player in cytoskeletal growth and remodelling becomes activated when RGC-derived Sema3A is knocked down in CM.

REFERENCES

1. Stahl A, Connor KM, Sapienza P, Willett KL, Krah NM, Dennison RJ, Chen J, Guerin KI, Smith LE. Computer-aided quantification of retinal neovascularization. *Angiogenesis*. 2009;12(3):297-301.

Figure S1

A



B

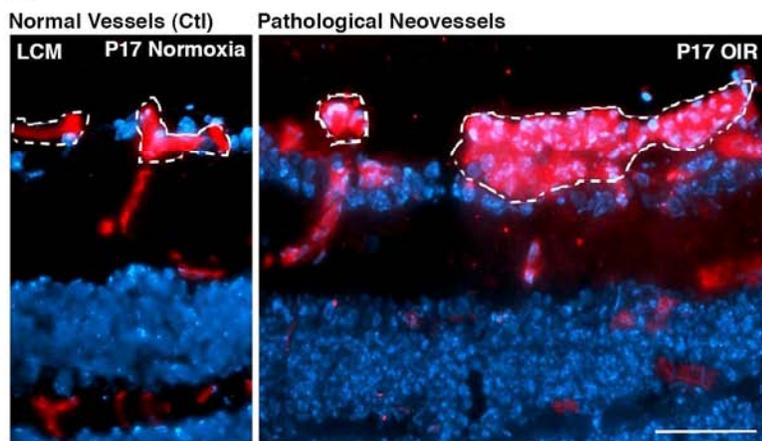


Figure S2

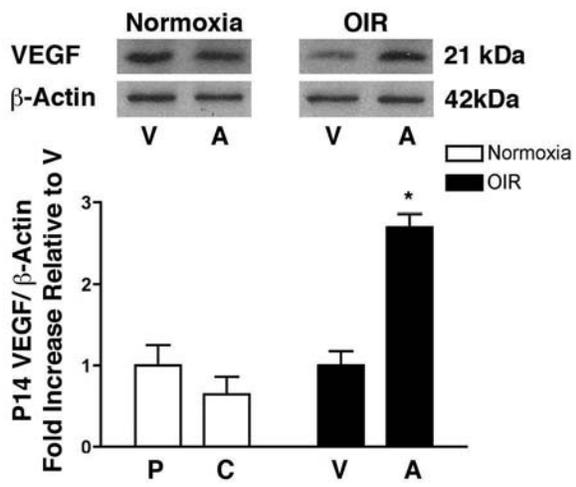
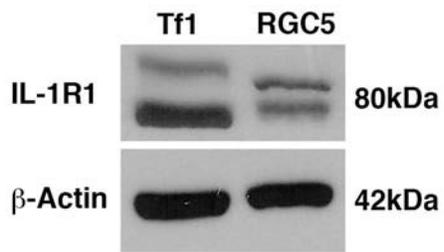


Figure S3

A



B

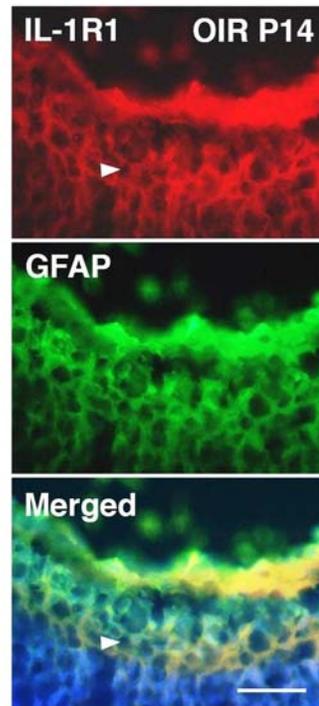
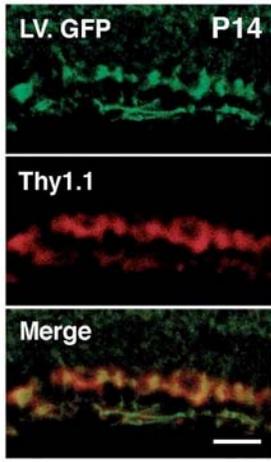
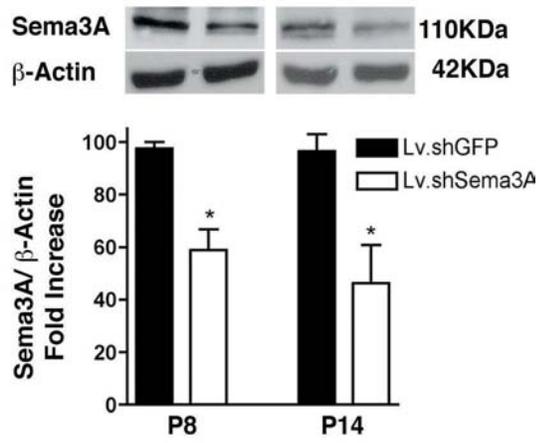


Figure S4

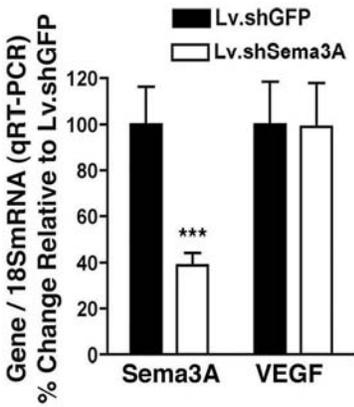
A



B



C



D

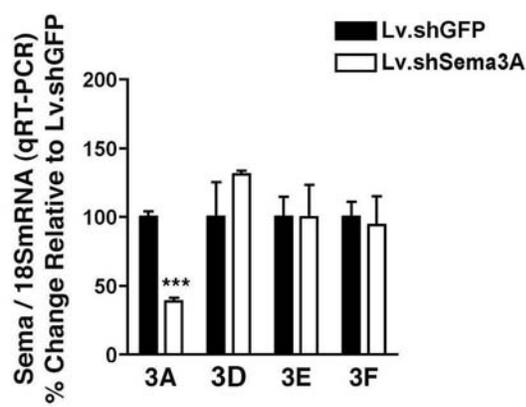


Figure S5

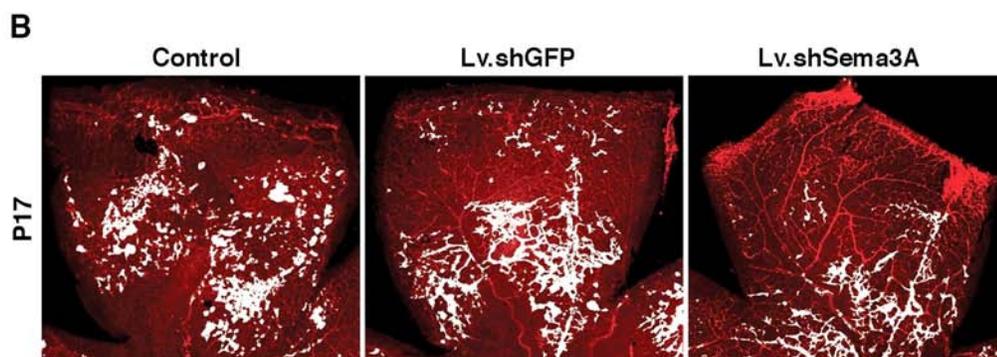
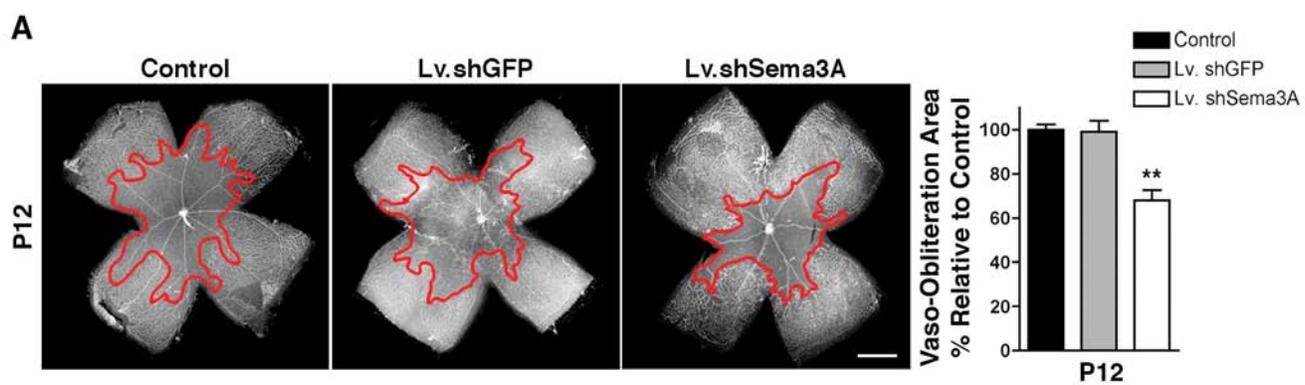


Figure S6

