Supporting Information

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SI Experimental Procedures

Drosophila Genetics. All crosses were performed according to standard procedures at 25 °C. Fly strains used in this study were *en-Gal4 UAS-GFP, ap-Gal4, omb-Gal4, UAS-flp, ey-FLP, dpp-lacZ, w¹¹¹⁸* (as wild type) (Bloomington *Drosophila* Stock Center), *UAS-hipk* (II), *UAS-hipk* (III), *UAS-hipk* ^{RNAi} (II), *UAS-hipk* ^{RNAi} (II), *UAS-hipk* ^{RNAi} (III) (Vienna *Drosophila* RNAi Center), *slimb*^{P1493} *FRT82* (2), *UAS-Daxin*^{A2-4} (3), *UAS-FLAG-Dcu1l* (4), *UAS-slimb* (5), *Tub > Myc-Slimb* (6), *hipk*⁴ *FRT79* (7), *Daxin*^{S044230} *FRT82/TM6B* (8), *Dcu11*^{EX} *FRT40* (9). In assays examining the interaction between two *UAS* transgenes, control crosses were performed with *UAS-lacZ* to rule out suppressive effects caused by titration of the Gal4 protein. MARCM clones were generated by crossing *y w hsflp UAS-GFP tubGal4; FRT42D tubGal80;* + or *y*

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w hsflpUAS-GFP tubGal4; +; FRT82B tubGal80 (gift from Bruce Edgar) females to males of the particular genotype for 24 h and the progeny were then heat-shocked at 38 °C for 90 min at 48 h AEL. Loss-of-function clones for *hipk* in the wing disk were generated by crossing *omb-Gal4/FM7; hipk⁴ FRT79/TM6B* females to *UAS-flp; GFP,FRT79/TM6B* males.

DNA Constructs. The following plasmids were used in this study: pUAST-Hipk (1), pGEX-GST-Hipk (10), pMK33-HA-Sgg (11), pMK33-HA-CK1 (12), β -TrCP-Myc-pCS2+ (13), pCDNA3-Hipk2^{WT}-FLAG and pCDNA3-Hipk2^{K221R}-FLAG (14), pCDNA3-Myc-Ubiquitin (15), pDA-FLAG-Ci, pDA-FLAG-Hh-N, pDA-RL (16), ptc\Delta136-Luc and ptc\Delta136-mut (17).

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α-Arm	GFP	merge	
A		-0	MARCM>Daxin ^{S044230}
B	*	2	MARCM>Daxin ^{S044230} + hipk ^w
C	18.5	19.5	MARCM>Daxin ^{S044230} + hipk ^{RI}
D			MARCM>Dcul1 ^{EX}
	Carl and		MARCM>Dcul1 ^{Ex} + hipk ^{wt}
F	· · · · · · · · · · · · · · · · · · ·	(F	MARCM>Dcul1 ^{Ex} + hipk ^{RNAi}
G	*	-	MARCM>slimb ^{P1493}
Н			MARCM>slimb ^{P1493} + hipk ^{wt}
			MARCM>slimb ^{P1493} + hipk ^{RNAI}

Fig. S1. Modulation of *hipk* has no effect on completely stabilized Arm. (A) The generation of positively GFP-marked loss-of-function MARCM clones for *Daxin* prevents the degradation of Arm and stabilizes the entire cytosolic pool. Once all cytosolic Arm is completely stabilized, the effects of overexpression of *hipk* (B) or *hipk*^{RNAi} (C) in this background are negligible. (D) Loss-of-function clones for *Dcul* also stabilizes all cytosolic Arm anywhere in the wing disk. Overexpression of *hipk* (E) or *hipk*^{RNAi} (F) in this context does not enhance or decrease the levels of stabilized Arm. (G) Similar to *Daxin* and *Dcul1*, loss-of-function clones for *slimb* result in the complete stabilization of Arm anywhere in the wing disk, which is affected by neither an increase (H) nor a decrease (I) in Hipk levels.



Fig. 52. Full-length Ci is reduced in loss-of-function clones for *hipk* in the wing disk. (A and B) Clonal analysis of *hipk* (indicated by a loss of GFP) shows a reduction in full-length Ci in certain parts of the wing disk.

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