

ATG14 induction

wt			wt rho ⁰			wt + AA			wt + O			∆atg7			time [h]
0	3	6	0	3	6	0	3	6	0	3	6	0	3	6	starvation
-	-	-	-	-	-	-	-	-				-	-	-	– GFP
-	_	_	_	_	-	~	_	~	~	_	_	-			– Pgk1





Glucose (-N) + rapamycin

Supplementary information

Fig. S1: Wild type, rho⁰, $\Delta npr2$, and $\Delta atg7$ cells harboring pr^{*ATG8*}-*GFP*-*ATG8* were exposed to amino acid starvation medium supplemented with amino acids and galactose. Samples were analyzed by whole cell extraction and western blot analysis using a α -GFP antibody.

Fig. S2: *ATG14* induction during amino acid starvation. Wild type, rho⁰, and $\Delta atg7$ cells harboring pr^{ATG14}-*GFP* were exposed to amino acid starvation medium supplemented with acetate in the presence or absence of antimycin A (AA) or oligomycin (O). Cells were analyzed at indicated time points by whole cell extraction and western blot analysis using α -GFP and α -Pgk1 antibodies.

Fig. S3: Rho⁰ cells display growth behavior consistent with intact vacuolar acidification. Wild type, rho⁰, and $\Delta vma2$ cells were grown on solid YP medium in the presence of glucose (YPD) or galactose (YPGal) at indicated pH and CaCl₂ concentration for 3-5 days at 30°C. Note that, in contrast to wild type and rho⁰ cells, the growth of cells lacking subunit B of the vacuolar H⁺-ATPase ($\Delta vma2$) is strongly impaired at pH 7.0 and in the presence of 100 mM CaCl₂ consistent with defects in vacuolar acidification described previously (Graham et al., 2000).

Fig. S4: Autophagic response during prolonged stationary phase. Log-phase wild type, rho⁰, and Δ*atg7* cells harboring pr^{*ATG8*}-*GFP*-*ATG8* (upper panels) or pr^{*ATG8*}-*GFP* (lower panels) were grown to stationary phase in galactose medium. Cells were analyzed at indicated time points by whole cell extraction and western blot analysis using α-GFP and α-Cdc11 antibodies.

Fig. S5: (**A**) Overexpression of cytosolic or mitochondrial superoxide dismutase does not affect the autophagic response in the presence of mitochondrial respiratory deficiency. Wild type and rho⁰ cells harboring pr^{*ATG8*}-*GFP*-*ATG8* were exposed to amino acid starvation medium supplemented with galactose in the presence of antimycin A (AA) or oligomycin (O) when indicated. Wild type and rho⁰ cells overexpressed cytosolic (Sod1) or mitochondrial (Sod2) superoxide dismutase when indicated. Samples were analyzed at indicated time points by whole cell extraction and western blot analysis using α -GFP and α -Cdc11 antibodies. (**B**) Effect of lowered membrane potential on the autophagic response during amino acid starvation. Wild type, rho⁰, and $\Delta atg7$ cells harboring pr^{*ATG8*}-

GFP-ATG8 were exposed to amino acid starvation medium supplemented with galactose. Wild type cells were exposed to CCCP at indicated concentrations during starvation. Samples were analyzed as described in (A).

Fig. S6: Autophagic response of respiratory deficient cells during nitrogen starvation. (**A**) Wild type, rho⁰, $\Delta npr2$, and $\Delta atg7$ cells were exposed to nitrogen starvation medium supplemented with glucose in the absence (upper panel) or presence (middle panel) of rapamycin (400 ng/ml). Wild type cells were exposed to antimycin (AA) or oligomycin (O) during nitrogen starvation as indicated (lower panel). Samples were analyzed by whole cell extraction and western blot analysis using a α-GFP antibody. (**B**) Wild type, rho⁰, $\Delta npr2$, and $\Delta atg7$ cells expressing pr^{*NPR1*}-NPR1-HA were exposed to nitrogen starvation medium supplemented with glucose in the absence (upper panels) or presence (lower panels) of rapamycin (400 ng/ml). The hyperphosphorylated (Npr1-P) and dephosphorylated (Npr1) forms of Npr1 are indicated. Cells were analyzed at indicated time points by whole cell extraction and western blot analysis using α-HA and α-Cdc11 antibodies.

Fig. S7: Mitochondrial respiratory deficiency impairs mitophagy under amino acid starvation. Wild type, Δ*atg7*, Δ*atg11*, and Δ*atg32* cells expressing the outer mitochondrial membrane protein and established mitophagy reporter pr^{*OM45*}-*OM45-GFP* (Kanki and Klionsky, 2008) were grown in YPlactate medium (log-phase) and exposed to amino acid starvation medium (starvation) supplemented with indicated carbon sources for 24 h. To impair mitochondrial function, wild type cells were exposed to antimycin A (AA) or oligomycin (O) during starvation. Om45-GFP (upper panels) and free GFP levels (lower panels) were monitored by whole cell extraction and western blot analysis using a α-GFP antibody. In wild type cells, a significant amount of free GFP was detected by western blot analysis under all tested amino acid starvation conditions, indicating that mitochondria were degraded. Consistent with published work, Om45-GFP degradation was blocked in Δ*atg7*, Δ*atg11*, and Δ*atg32* cells, demonstrating that mitochondrial degradation depends on both the general autophagy (Atg7) and mitophagy-specific machinery (Atg11 and Atg32) (Kanki and Klionsky, 2008; Kanki et al., 2009; Okamoto et al., 2009). Significantly, mitochondrial turnover was completely blocked in wild type cells in the presence of antimycin A or oligomycin under all carbon source conditions. (*) indicates an unspecific α-GFP signal. Quantification of the ratio of free GFP to total GFP (Om45-GFP and free GFP) relative to untreated wild type (wt=1) after starvation is shown as means and standard deviations of three (n=3) independent experiments.