Supporting Information

Temperature dependence of charge separation and recombination in porphyrin oligomer-fullerene donor-acceptor systems

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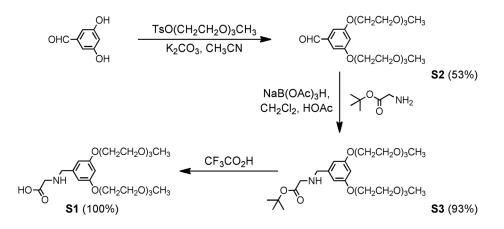
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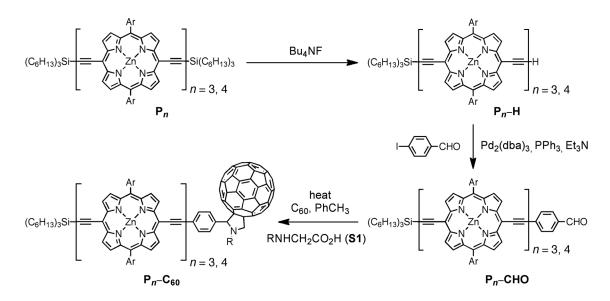
Section S1: Synthetic Strategy and Schemes

The sarcosine derivative **S1** was designed to overcome solubility problems with the longer fullerene-terminated porphyrin oligomers. It was synthesized in three steps from 3,5-dihydroxybenzaldehyde, as shown in Scheme S1.1.

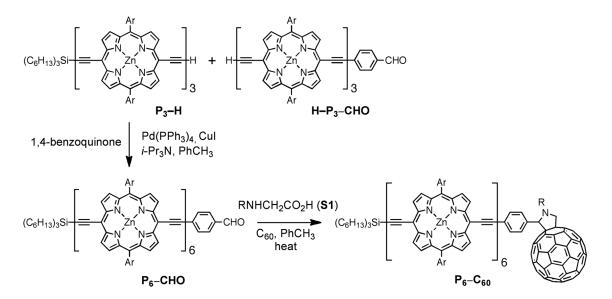


Scheme S1.1. The synthesis of sarcosine derivative S1.

The fullerene-terminated trimer and tetramer, P_3 - C_{60} and P_4 - C_{60} , were prepared using Prato coupling, as shown in Scheme S1.2. The hexamer derivative P_6 - C_{60} was synthesized by the route shown in Scheme S1.3 which involves statistical cross-coupling of two trimers P_3 -H and H-P₃-CHO.



Scheme S1.2. The synthesis of fullerene-terminated porphyrin oligomers P₃-C₆₀ and P₄-C₆₀.



Scheme S1.3. The synthesis of hexamer P₆-C₆₀.

Section S2: Synthetic Procedures

Materials, Instrumentation and General Synthetic Procedures. Solvents were dried by passing through alumina under N₂ pressure. Triethylamine was distilled from CaH₂. Manipulation of all air and/or water sensitive compounds was carried out using standard high vacuum techniques. Porphyrin oligomers P₃, P₄ and P₄-CHO were synthesized using published procedures.¹⁻² Column chromatography was carried out on Merck® silica gel 60 using a positive pressure of nitrogen. TLC was carried out on aluminum backed silica gel 60 F254 plates. Size exclusion chromatography was carried out using Bio-Beads S-X1, 200-400 mesh (Bio-Rad). Where mixtures of solvents were used, ratios reported are by volume. NMR spectra were recorded on a Bruker DPX-400 spectrometer, unless otherwise stated. Chemical shifts are quoted as parts per million (ppm) relative to tetramethylsilane and coupling constants (J) are quoted in Hertz (Hz). MALDI-TOF mass spectra were acquired by the EPSRC Mass Spectrometry Service, Swansea, UK, using a DCTB matrix. UV-visible spectra were recorded on a Perkin-Elmer Lambda 20 spectrometer. HPLC purification was performed using HPLC grade solvents with an ACE column (dimensions 250×10 mm, 100 Å pore size, 5 µm phase silica) on a VWR-Hitachi LaChrom Elite system, equipped with an L-2455 Diode Array Detector, L-2200 Autosampler, L-2350 Column Oven, L-2130 Pump and a Foxy Jr. Fraction Collector.

Synthesis of 3,5-Bis(2-(2-(2-methoxy)ethoxy)ethoxy)benzaldehyde S2. 3,5-Dihydroxybenzaldehyde (0.50 g, 3.6 mmol) and K₂CO₃ (1.00 g, 7.7 mmol) were dried under

vacuum for 2 hr then acrylonitrile (3 cm³) and triethyleneglycol monotosylate (2.3 g, 7.2 mmol) were added, and the mixture was heated to reflux under nitrogen for 16 hr. The solvent was removed and the product was purified by silica gel column chromatography (EtOAc, $R_f = 0.3$) to yield **S2** (827 mg, 53%) as a colorless liquid. ¹H NMR (400 MHz CDCl₃): $\delta = 9.89$ (s, 1H), 7.03 (d, J = 2.2, 2H), 6.77 (t, J = 2.3, 1H), 4.18 (t, J = 4.4, 4H), 3.89 (t, J = 5.0, 4H), 3.75–3.55 (m, 16H), 3.39 (s, 6H) (This compound has been prepared previously using similar procedures).³⁻⁵

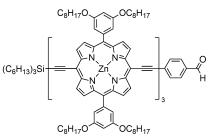
Synthesis of Aryl-sarcosine tert-Butyl ester S3. 3,5-Bis(2-(2-(2-methoxyethoxy)ethoxy)benzaldehydeS2 (650 mg, 1.51

mmol) was dissolved in CH₂Cl₂ (64 cm³) under argon and acetic acid ^{MeO}(γ_3^{M}) (γ_3^{OMe}) was added (1.28 cm³). Glycine *tert*-butyl ester (2 eq., 413 µl, 3.02 mmol) was added, the mixture was stirred for 20 min, sodium triacetoxyborohydride (2.5 eq., 801 mg, 3.78 mmol) was then added and the mixture was stirred for a further 1 hr. A saturated aqueous solution of sodium carbonate (25 cm³) was then added, the product extracted in to CH₂Cl₂ (3 × 20 cm³), the organic layer washed with brine (25 cm³), dried (MgSO₄), filtered and the solvent removed to give **S3** as a pale yellow oil (765 mg, 93 %). ¹H NMR (400 MHz CDCl₃): δ = 6.50 (d, *J* = 2.2, 2H, ArH), 6.40 (t, *J* = 2.2, 1H, ArH), 4.10 (t, *J* = 4.7, 4H, OCH₂), 3.86–3.83 (m, 4H, OCH₂), 3.76–3.72 (m, 6H, 4 OCH₂, 2 CH₂) 3.70–3.65 (m, 8H, OCH₂), 3.58–3.55 (m, 4H, OCH₂), 3.39 (s, 6H, OCH₃), 3.28 (s, 2H, CH₂), 1.47 (s, 9H, OCH₃); *m/z* (ESI) 546.3 [(M + 1)⁺ C₂₇H₄₈NO₁₀ calc. 546.3], 489.3 [(M – ^tBu)⁺ C₂₃H₃₉NO₁₀ calc. 489.3].

Synthesis of Aryl-sarcosine S1. Aryl-sarcosine *tert*-butyl ester S3 (202 mg, 0.307 mmol) was dissolved in trifluoroacetic acid (11 eq., 0.409 mmol, 2.75 cm³) and the solution was stirred for 1 hr. The solvent was meo(-0_{3}) -0_{3}) -0_{3}) -0_{3} meo(-0_{3}) -0_{3} meo(-0_{3}) -0_{3}) -0_{3} meo(-0_{3}) meo(-0_{3}) -0_{3} meo(-0_{3}) meo

Synthesis of P₃-CHO. The protected porphyrin trimer P₃ (50.9 mg, 0.0132 mmol) was

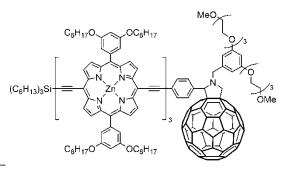
dissolved in dry CH_2Cl_2 (1.6 cm³) and $CHCl_3$ (1.6 cm³). TBAF solution (1.0 M in THF, 1.5 eq., 0.027 mmol, 27 µl) was added and the mixture was stirred under argon for 23 min, monitored by TLC [40–60 petroleum ether/pyridine (85:15)]. Acetic acid (50 µl, 0.87 mmol)



was added, the mixture was passed though a short column of silica (CH₂Cl₂) and the solvent was removed. This mixture was used immediately in the next step without further purification. Deprotected trimer mixture (0.0132 mmol), para-iodobenzaldehyde (10 eq., 0.133 mmol, 30.9 mg), Pd₂(dba)₃ (10 mol%, 0.0013 mmol, 1.2 mg) and PPh₃ (40 mol%, 0.0053 mmol, 1.4 mg) were dried under vacuum for 40 min, and the vessel was purged with argon. Triethylamine (4.3 cm^3) was added, the mixture was treated to three freeze-pump-thaw cycles and then heated to 50 °C under argon for 14 hr. The mixture was passed though a short column of silica (CH₂Cl₂), the solvent removed and the residue was separated using column chromatography with 40-60 petroleum ether/pyridine (85:15) as eluent. The first eluting band was unreacted P_3 , followed by the desired product P_3 -CHO. The product was purified by recrystallization from CH₂Cl₂/MeOH to give a dark brown powder (17.6 mg, 36 % over 2 steps). ¹H NMR (400 MHz CDCl₃ / 1 % C₅D₅N): δ = 10.15 (s, 1H, CHO_a), 9.89–9.86 (m, 8H, βH_{b2-5}), 9.70 (d, J = 4.4, 2H, βH_{b1}), 9.64 (d, J = 4.6, 2H, βH_{b6}), 9.08–9.06 (m, 8H, βH_{c2-5}), 9.03 (d, J = 4.4, 2H, β H_{c1}), 8.97 (d, J = 4.6, 2H, β H_{c6}), 8.18 (d, J = 8.0, 2H, ArH_d), 8.09 (d, J = 4.6, 2H, β H_{c6}), 8.18 (d, J = 8.0, 2H, ArH_d), 8.09 (d, J = 8.0, 2H, Ar $= 8.0, 2H, ArH_e$, 7.42 (d, $J = 2.2, 4H, ArH_{f2}$), 7.39 (d, $J = 2.0, 4H, ArH_{f1}$), 7.38 (d, $J = 2.2, 4H, ArH_{f2}$), 7.39 (d, $J = 2.2, 4H, ArH_{f1}$), 7.38 (d, $J = 2.2, 4H, ArH_{f2}$), 7.39 (d, $J = 2.2, 4H, ArH_{f1}$), 7.38 (d, $J = 2.2, 4H, ArH_{f2}$), 7.39 (d, $J = 2.0, 4H, ArH_{f1}$), 7.38 (d, $J = 2.2, 4H, ArH_{f2}$), 7.39 (d, $J = 2.0, 4H, ArH_{f1}$), 7.38 (d, $J = 2.2, 4H, ArH_{f2}$), 7.39 (d, $J = 2.0, 4H, ArH_{f1}$), 7.38 (d, $J = 2.2, 4H, ArH_{f2}$), 7.39 (d, $J = 2.0, 4H, ArH_{f1}$), 7.38 (d, $J = 2.2, 4H, ArH_{f2}$), 7.39 (d, $J = 2.0, 4H, ArH_{f1}$), 7.38 (d, $J = 2.2, 4H, ArH_{f2}$), 7.39 (d, $J = 2.0, 4H, ArH_{f1}$), 7.38 (d, $J = 2.2, 4H, ArH_{f2}$), 7.39 (d, $J = 2.2, 4H, ArH_{f2}$), 7.39 (d, $J = 2.2, 4H, ArH_{f2}$), 7.39 (d, $J = 2.2, 4H, ArH_{f1}$), 7.38 (d, $J = 2.2, 4H, ArH_{f2}$), 7.39 (d, $J = 2.2, 4H, ArH_{f2}$), 7.39 (d, $J = 2.2, 4H, ArH_{f2}$), 7.39 (d, $J = 2.2, 4H, ArH_{f1}$), 7.38 (d, $J = 2.2, 4H, ArH_{f2}$), 7.39 (d, $J = 2.2, 4H, ArH_{f2$ 4H, ArH_{f3}), 6.93–6.91 (m, 6H, ArH_g), 4.19–4.16 (m, 24H, octyl-CH_{2h}), 1.95–1.89 (m, 24H, octyl-CH_{2h}), 1.83–1.75 (m, 6H, hexyl-CH_{2i}), 1.59–1.52 (m, 36 H, 12 hexyl-CH_{2i}, 24 octyl-CH_{2h}), 1.42–1.28 (m, 102H, 6 hexyl-CH_{2i}, 96 octyl-CH_{2h}), 1.06–1.02 (m, 6H, hexyl-CH_{2i}), 0.95–0.86 (m, 45H, 9 hexyl-CH_{3i}, 36 octyl-CH_{3h}); m/z (MALDI) 3643.0 (M⁺, $C_{229}H_{290}N_{12}O_{13}SiZn_3$ calc. 3643.0); UV/vis (CHCl₃/pyridine 99:1): λ_{max}/nm ($\epsilon/10^3$ M⁻¹ cm⁻¹) 468 (523), 500 (227), 590 (29.0), 699 (sh 93.4), 759 (181); UV/vis (THF): $\lambda_{max}/nm (\epsilon/10^3 \text{ M}^{-1})$ cm⁻¹) 464 (556), 496 (237), 585 (32.1), 697 (sh 111), 743 (175); UV/vis (THF/pyridine 99:1): $\lambda_{max}/nm (\epsilon/10^3 \text{ M}^{-1} \text{ cm}^{-1}) 466 (567), 497 (224), 589 (30.5), 698 (sh 106), 749 (174).$

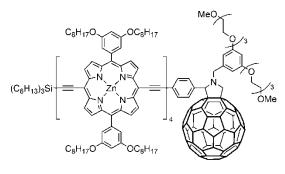
-S6-

Synthesis of P₃-C₆₀. C₆₀ (10 eq., 0.0274 mmol, 19.7 mg) was sonicated in dry toluene (6.7 cm³) for 1 hr and this solution was added to vacuumdried P₃-CHO (10.0 mg, 0.00274 mmol) and



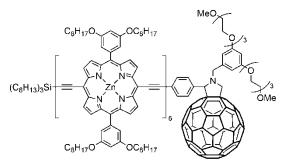
aryl-sarcosine S1 (20 eq., 0.0549 mmol, 26.9 mg). The mixture was degassed and refluxed under argon for 13 hr, then allowed to cool, purified on silica (toluene, then CH₂Cl₂, then CH₂Cl₂/MeOH 1 %) and recrystallized from CH₂Cl₂/MeOH to give P₃-C₆₀ as a dark brown powder (10.6 mg, 81 %). ¹H NMR (400 MHz CDCl₃ / 1 % C₅D₅N): δ = 9.88–9.86 (m, 8H, βH_{a2-5} , 9.71 (d, $J = 4.4, 2H, \beta H_{a1}$), 9.64 (d, $J = 4.4, 2H, \beta H_{a2}$), 9.07–9.05 (m, 8H, βH_{b2-5}), 8.98 (d, $J = 4.6, 2H, \beta H_{b1}$), 8.97 (d, $J = 4.6, 2H, \beta H_{b6}$), 8.12 (br, 4H, ArH_c), 7.41 (d, J = 2.0, 4H, ArH_{d2}), 7.39 (m, 8H, ArH_{d1.3}), 6.94–6.91 (m, 8H, 6 ArH_e, 2 ArH_f), 6.58–6.57 (m, 1H, ArH_g), 5.29 (br, 1H, CH_h), 4.94–4.91 (m, 1H, CH_{2i}), 4.65–4.60 (br, 1H, CH_{2i}), 4.26–4.24 (m, 4H, OCH_{2k}), 4.20–4.16 (m, 25H, 24 octyl-CH_{2l}, 1 CH_{2i}), 3.95–3.93 (m, 4H, OCH_{2k}), 3.80– 3.78 (m, 4H, OCH_{2k}), 3.73–3.71 (m, 4H, OCH_{2k}), 3.68–3.66 (m, 5H, 4 OCH_{2k}, 1 CH_{2i}), 3.57– 3.55 (m, 4H, OCH_{2k}), 3.38 (s, 6H, OCH_{3m}), 1.93–1.75 (m, 36H, 24 octyl-CH_{2l}, 6 hexyl-CH_{2n}), 1.58-1.51 (m, 36H, 12 hexyl-CH_{2n}, 24 octyl-CH_{2l}), 1.41-1.26 (m, 102H, 6 hexyl-CH_{2n}, 96 octyl-CH₂₁), 1.06–1.00 (m, 6H, hexyl-CH_{2n}), 0.93–0.85 (m, 45H, 9 hexyl-CH_{3n}, 36 octyl-CH₃₁); m/z (MALDI) 3791.3 (M⁺, C₃₁₁H₃₂₇N₁₃O₂₀SiZn₃ calc. 3791.3); UV/vis (CHCl₃/pyridine 99:1): λ_{max}/nm ($\epsilon/10^3$ M⁻¹ cm⁻¹) 466 (505), 500 (215), 590 (29.6), 699 (sh 88.4), 759 (172); UV/vis (THF): λ_{max}/nm ($\varepsilon/10^3$ M⁻¹ cm⁻¹) 462 (535), 496 (228), 586 (31.3), 688 (sh 101), 743 (167); UV/vis (THF/pyridine 99:1): λ_{max}/nm ($\epsilon/10^3$ M⁻¹ cm⁻¹) 465 (554). 498 (218), 587 (30.7), 698 (sh 103), 749 (168). Nierengarten and co-workers studied the restricted rotation of aryl-substituted fullerene derivatives.⁶ Our spectral assignment around the C_{60} group is in agreement with that reported.

Synthesis of P_4 - C_{60} . C_{60} (0.0226 mmol, 16.3 mg) was sonicated in dry toluene (10 cm³) for 1 hr and this solution was added to vacuum-dried P_4 -CHO (21.4 mg, 0.00453 mmol) and aryl-sarcosine S1 (0.0453 mmol, 22.2 mg). The mixture was degassed and refluxed under nitrogen overnight.



The mixture was allowed to cool, purified on silica (toluene, then CH₂Cl₂, then CH₂Cl₂/MeOH 1 %) and then recrystallized from CH₂Cl₂/MeOH to give **P**₄-**C**₆₀ as a dark brown powder (25.2 mg, 94 %). ¹H NMR (250 MHz CDCl₃ / 1 % C₅D₅N, 298K): δ = 9.86–9.89 (m, 12H, β H_{a2-7}), 9.70 (d, *J* = 4.5, 2H, β H_{a1}), 9.64 (d, *J* = 4.5, 2H, β H_{a8}), 9.08–9.04 (m, 12H, β H_{b2-7}), 8.97 (d, *J* = 4.5, 2H, β H_{b1}), 8.96 (d, *J* = 4.5, 2H, β H_{b8}), 8.12 (br, 4H, ArH_c), 7.41 (d, *J* = 2, 8H, ArH_{d2-3}), 7.37 (m, 8H, ArH_{d1,4}), 6.94–6.91 (m, 10H, 8 ArH_c, 2 ArH_f), 6.67 (br, 1H, ArH_g), 5.13 (br, 1H, CH_b), 4.92 (br, 1H, CH_{2i}), 4.60 (br, 1H, CH_{2j}), 4.21–4.14 (m, 37H, 4 CH_{2k}, 32 octyl-CH_{2l}, 1 CH_{2i}), 3.96–3.92 (m, 4H, OCH_{2k}), 3.81–3.77 (m, 4H, OCH_{2k}), 3.74–3.71 (m, 4H, OCH_{2k}), 3.68–3.64 (m, 5H, 4 OCH_{2k}, 1 CH_{2j}), 3.57–3.53 (m, 4H, OCH_{2k}), 3.37 (s, 6H, OCH_{3m}), 1.93–184 (m, 32H, octyl-CH_{2l}), 1.80–1.72 (m, 6H, hexyl-CH_{2n}), 1.59–1.48 (m, 44H, 12 hexyl–CH_{2n}, 32 octyl-CH_{2l}), 0.93–0.83 (m, 63H, 9 hexyl-CH_{3n}, 54 octyl-CH_{3l}); *m/z* (MALDI + NaOAc) 5877.4 (M⁺, C₃₇₉H₄₀₉N₁₇O₂₄SiZn₄ calc. 5876.05), 5211.3 ([M – C₆₀ + AcO⁻]⁺, calc. 5216.8; UV/vis (CHCl₃/pyridine 99:1): λ_{max}/nm ($\varepsilon/10^3$ M⁻¹ cm⁻¹) 467 (511), 769 (210).

Synthesis of P_6 - C_{60} . The protected porphyrin trimer P_3 (49.6 mg, 0.0130 mmol) was dissolved in dry CH₂Cl₂ (1.6 cm³) and CHCl₃ (1.6 cm³) and mixture was placed under argon. TBAF solution (1.0 M in THF, 2 eq., 0.026 mmol, 26 µl) was added and the mixture was stirred under



argon for 35 min, monitored by TLC (40–60 petroleum ether/pyridine/THF 85:10:5). The mixture was passed though a short column of silica (CH₂Cl₂) and the solvent was removed. The residue was separated using column chromatography with 40–60 petroleum ether/pyridine (85:15) as eluent. The first eluting band was unreacted **P**₃ (29.2 mg, 59 %), followed by the desired intermediate **P**₃-**H** (14.1 mg, 31 %).

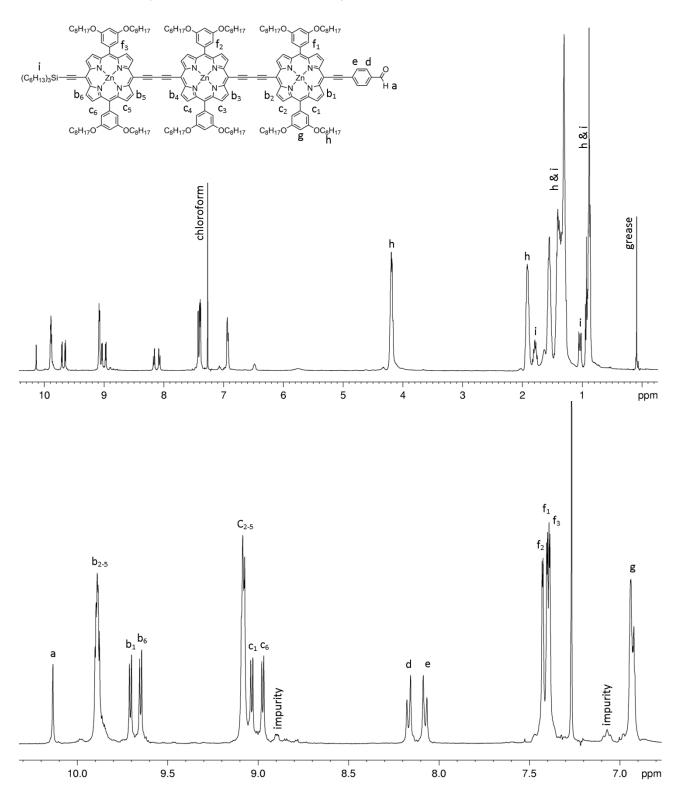
P₃-CHO (14.5 mg, 0.00398 mmol) was dissolved in CH_2Cl_2 (2 cm³) under argon, TBAF solution (1.0 M in THF, 10 eq., 0.0398 mmol, 40 µl) was added and the mixture was stirred at

room temperature for 30 min. Mixture was passed through a short column of silica $(CH_2Cl_2/pyridine 99:1)$ to give the desired **H-P₃-CHO**, and this product was used immediately in the next step without further purification.

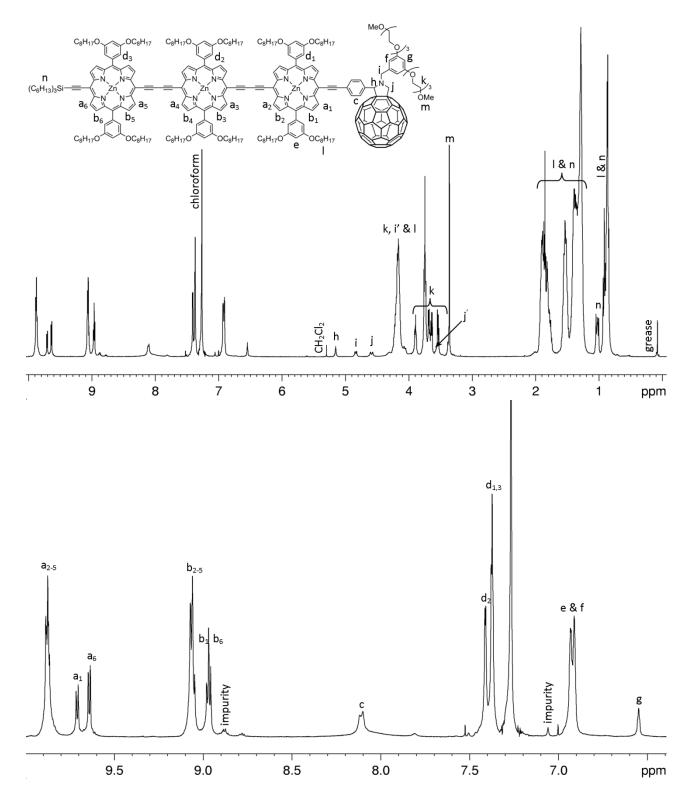
P₃-H (14.1 mg, 0.00398 mmol), **H-P₃-CHO** (0.00398 mmol), Pd(PPh₃)₄ (5 mol%, 0.000398 mmol, 0.46 mg), CuI (50 mol%, 0.00398 mmol, 0.76 mg) and 1,4-benzoquinone (2 eq., 0.0159 mmol, 1.7 mg) were dried for 25 min, the vessel was purged with argon. Diisopropylamine (0.77 cm³) and toluene (2.0 cm³) were added and the mixture was heated to 50 °C for 15 hr. The statistical mixture was passed through a short silica column (CH₂Cl₂/pyridine 99:1), and purified by size exclusion chromatography (toluene with 0.02 % pyridine). C₆₀ (10 eq., 0.0332 mmol, 23.9 mg) was sonicated in dry toluene (11 cm³) and pyridine (1%, 110 µl) for 30 min and this solution was added to vacuum-dried hexamer mixture (22.9 mg, 0.00332 mmol) and aryl-sarcosine S1 (20 eq., 0.0664 mmol, 32.5 mg). The mixture was degassed, refluxed under argon for 15 hr and allowed to cool. The mixture was purified using a silica chromatography (toluene/pyridine 99:1, then toluene/THF/pyridine 97:2:1, then toluene/THF/pyridine 89:10:1). The first eluting band was P_6 , followed by the desired product P_6 - C_{60} . This fraction was purified using semi-preparative HPLC, and the product was then recrystallized from $CH_2Cl_2/MeOH$ to give a dark brown powder (P₆-C₆₀, 4.1 mg, 15 %, from 3 steps). ¹H NMR (400 MHz CDCl₃ / 1 % C₅D₅N): δ = 9.90–9.87 (m, 20H, β H_{a2-11}), 9.71 (d, J = 4.4, 2H, β H_{a1}), 9.64 (d, J = 4.5, 2H, β H_{a12}), 9.09–9.07 (m, 20H, β H_{b2-11}), 8.98 (d, J = 4.4, 2H, β H_{b1}), 8.96 (d, J = 4.5, 2H, β H_{b12}), 8.13 (br, 4H, ArH_c), 7.44–7.42 (m, 16H, ArH_{d2-5}), 7.39–7.37 (m, 8H, ArH_{d1.6}) 6.95–6.91 (m, 14H, 12 ArH_e, 2 ArH_f), 6.58–6.57 (m, 1H, ArH_g), 5.28 (br, 1H, CH_h), 4.92–4.89 (m, 1H, CH_{2i}), 4.65–4.60 (br, 1H, CH_{2i}), 4.32– 4.08 (m, 53H, 4 OCH_{2k}, 48 octyl-CH_{2l}, 1 CH_{2i}), 3.94–3.91 (m, 4H, OCH_{2k}), 3.79–3.76 (m, 4H, OCH_{2k}), 3.73–3.70 (m, 4H, OCH_{2k}), 3.68–3.65 (m, 5H, 4 OCH_{2k}, 1 CH_{2i}), 3.57–3.54 (m, 4H, OCH_{2k}), 3.37 (s, 6H, OCH_{3m}), 1.94–1.75 (m, 54H, 48 octyl-CH_{2l}, 6 hexyl-CH_{2n}), 1.59– 1.50 (m, 60H, 12 hexyl-CH_{2n}, 48 octyl-CH_{2l}), 1.44–1.25 (m, 198H, 6 hexyl-CH_{2n}, 192 octyl-CH₂₁), 1.05–1.00 (m, 6H, hexyl-CH_{2n}), 0.94–0.84 (m, 81H, 9 hexyl-CH_{3n}, 72 octyl-CH_{3l}); UV/vis (CHCl₃/pyridine 99:1): λ_{max}/nm ($\epsilon/10^3$ M⁻¹ cm⁻¹) 468 (666), 500 (sh 445), 592 (48.8), 795 (299); UV/vis (THF): λ_{max}/nm ($\epsilon/10^3$ M⁻¹ cm⁻¹) 465 (724), 495 (466), 587 (52.1), 766 (305); UV/vis (CHCl₃/pyridine 99:1): λ_{max}/nm ($\epsilon/10^3$ M⁻¹ cm⁻¹) 466 (737), 497 (sh 450), 589 (50.7), 776 (307).

Section S3: NMR Spectra

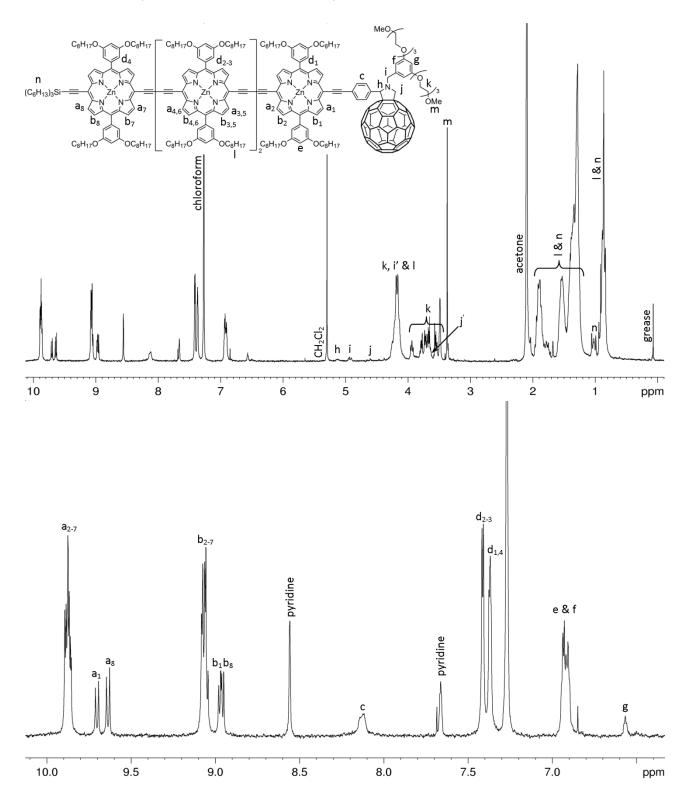
P₃-CHO; ¹H NMR (400 MHz, CDCl₃/1 % C₅H₅N)



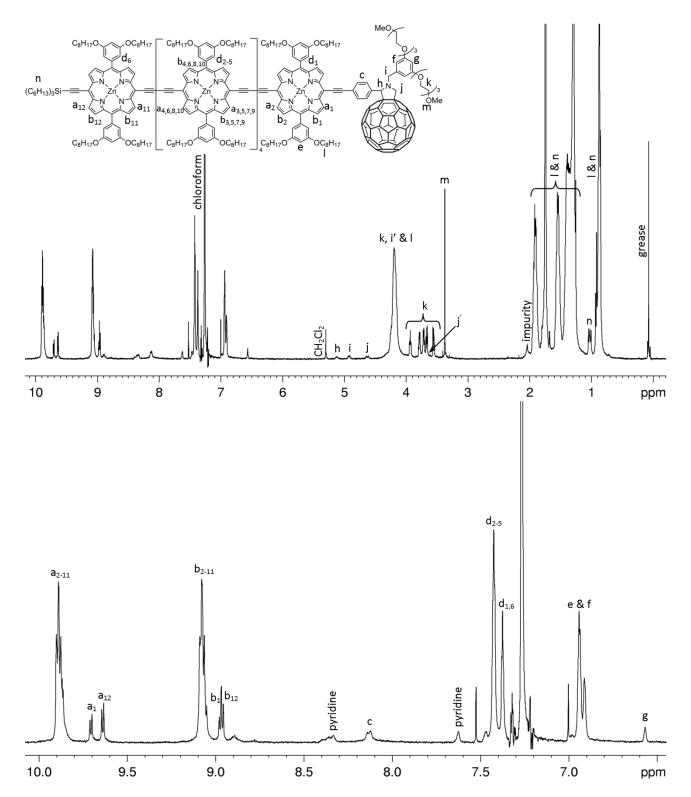


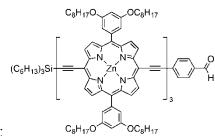


P₄-**C**₆₀; ¹H NMR (250 MHz, CDCl₃/1 % C₅H₅N)



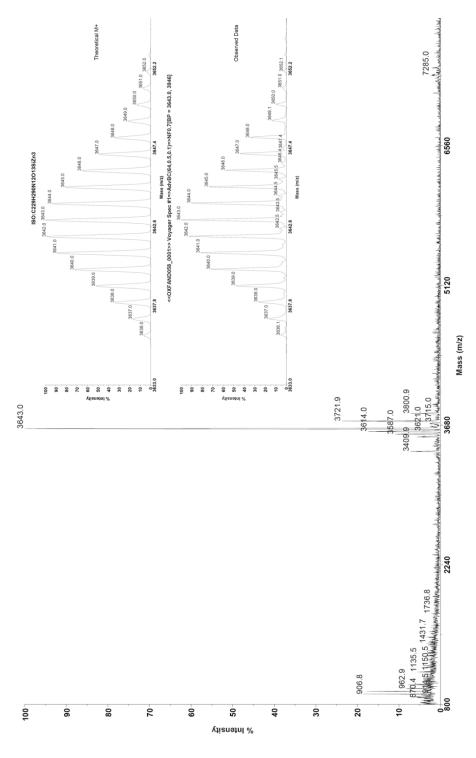




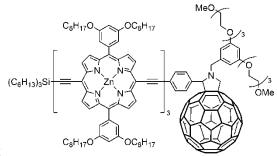


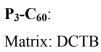


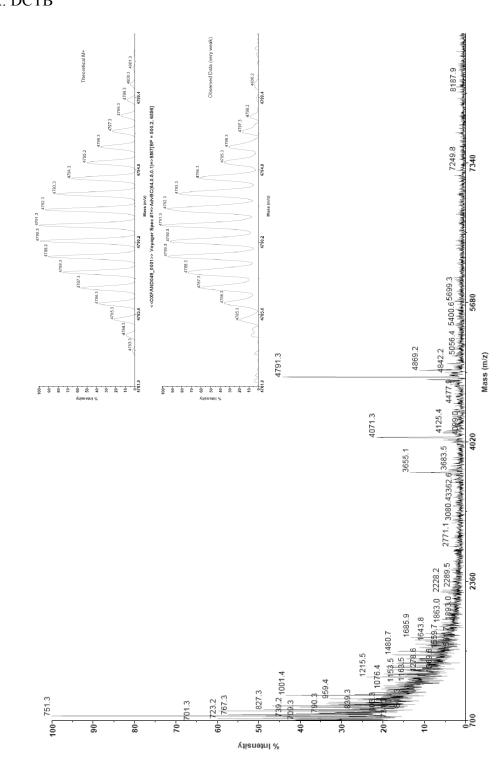
Matrix: DCTB

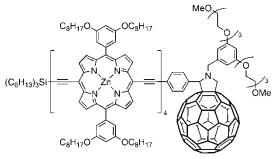


—S14—

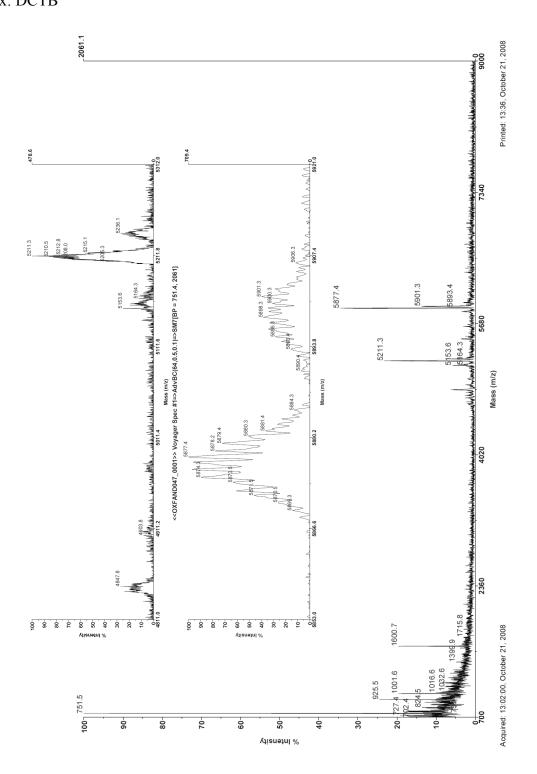








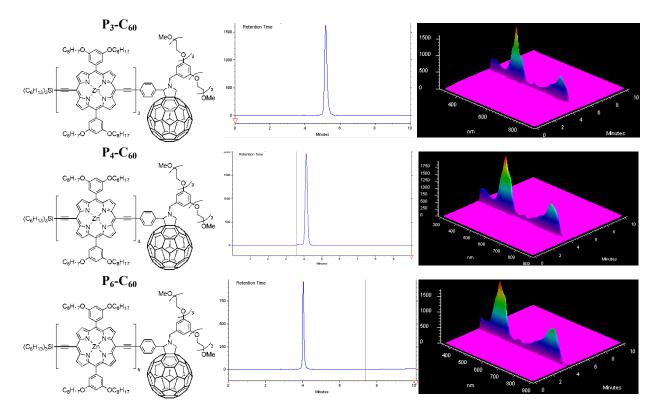
P₄-C₆₀: Matrix: DCTB

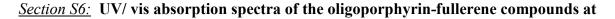


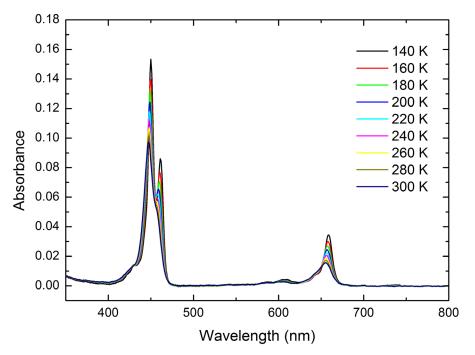
Section S5: HPLC traces

The HPLC purification of compounds **P3-C60**, **P4-C60** and **P6-C60** was performed using a flow rate of 4 mL/min with the following method:

| Time (min) | % Toluene (with 1% pyridine) | % THF |
|------------|------------------------------|-------|
| 0 | 80 | 20 |
| 8 | 65 | 35 |
| 9 | 35 | 65 |
| 10 | 80 | 20 |
| 14 | 80 | 20 |







different temperatures

Figure S6.1. Ground state absorption of P_1 - C_{60} in 2MTHF + 1% pyridine at different temperatures.

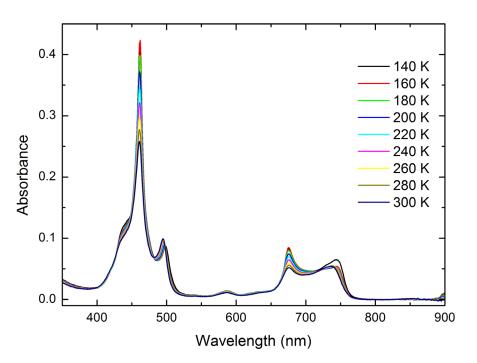


Figure S6.2. Ground state absorption of P_2 - C_{60} in 2MTHF + 1% pyridine at different temperatures.

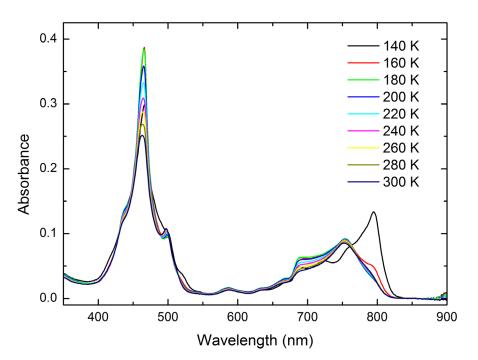


Figure S6.3. Ground state absorption of P_3 - C_{60} in 2MTHF + 1% pyridine at different temperatures.

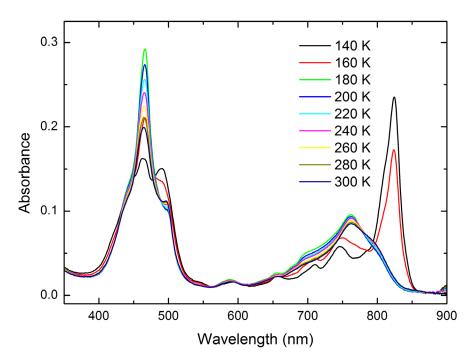


Figure S6.4. Ground state absorption of P_4 - C_{60} in 2MTHF + 1% pyridine at different temperatures.

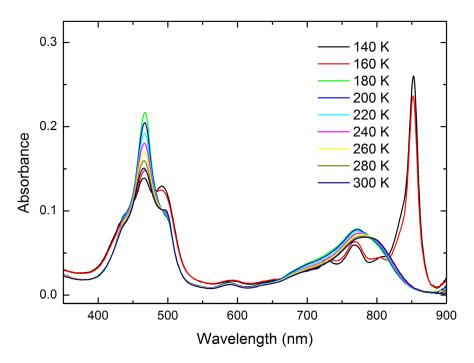
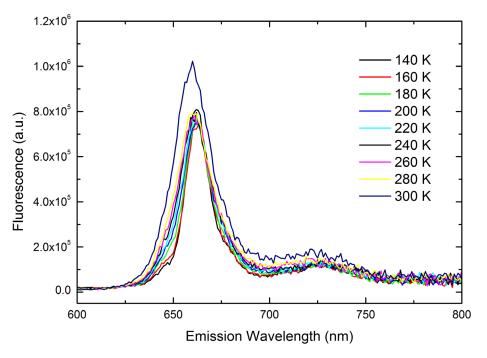


Figure S6.5. Ground state absorption of P_6 - C_{60} in 2MTHF + 1% pyridine at different temperatures.

Section S7: Fluorescence spectra of the oligoporphyrin-fullerene compounds at



different temperatures

Figure S7.1. Steady state emission of P_1 - C_{60} in 2MTHF + 1% pyridine at different temperatures excited at 450 nm.

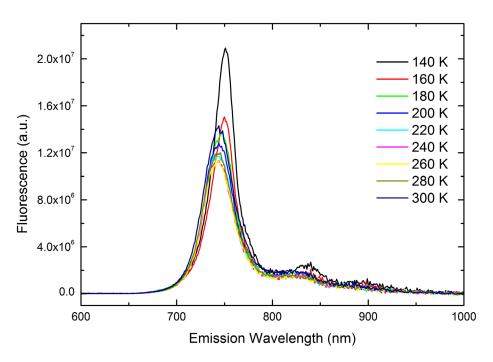


Figure S7.2. Steady state emission of P_2 - C_{60} in 2MTHF + 1% pyridine at different temperatures, excited at 495 nm.

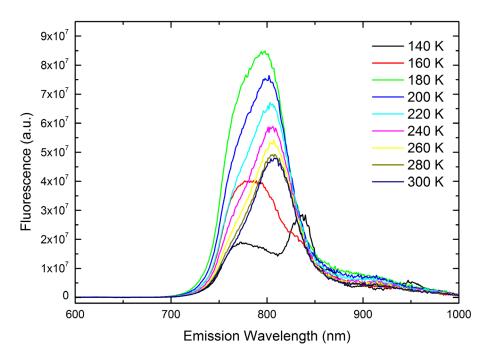


Figure S7.3. Steady state emission of P_4 - C_{60} in 2MTHF + 1% pyridine at different temperatures excited at 495 nm.

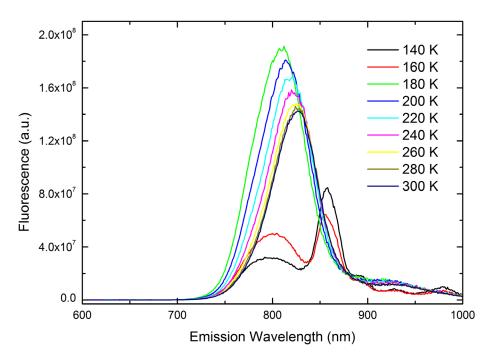
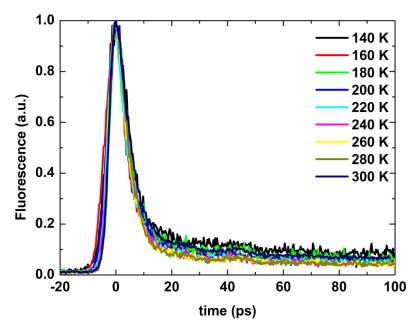


Figure S7.4. Steady state emission of P_6 - C_{60} in 2MTHF + 1% pyridine at different temperatures excited at 495 nm.

Section S8: Fluorescence time profiles of the oligoporphyrin-fullerene compounds at



different temperatures

Figure S8.1. Fluorescence time profiles for P_1 - C_{60} in 2MTHF + 1% pyridine at different temperatures.

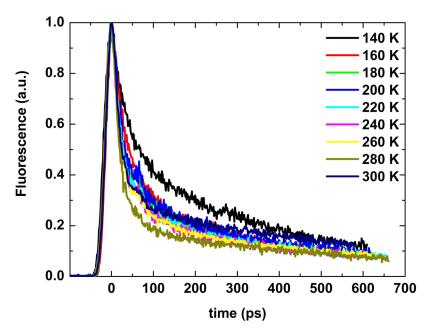


Figure S8.2. Fluorescence time profiles for P_2 - C_{60} in 2MTHF + 1% pyridine at different temperatures.

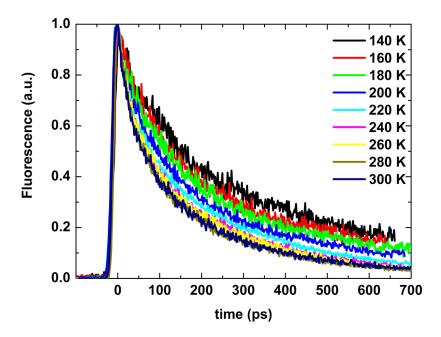


Figure S8.3. Fluorescence time profiles for P_4 - C_{60} in 2MTHF + 1% pyridine at different temperatures.

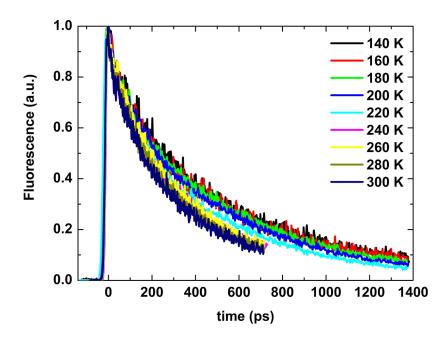


Figure S8.4. Fluorescence time profiles for P_6 - C_{60} in 2MTHF + 1% pyridine at different temperatures.

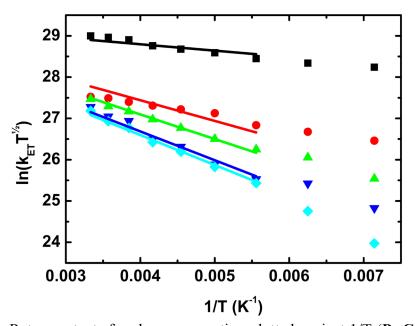


Figure S8.5. Rate constants for charge separation plotted against 1/T (P₁-C₆₀ (black), P₂-C₆₀ (red), P₃-C₆₀ (green), P₄-C₆₀ (blue) and P₆-C₆₀ (cyan)). The lines are the global fit to the linearized form of Eq.2 assuming a global matrix coupling element V.

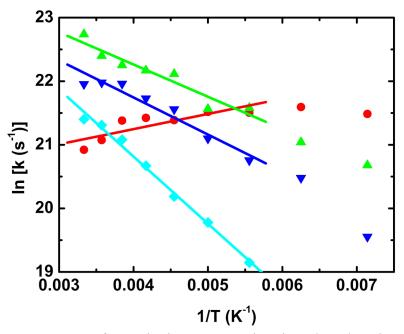
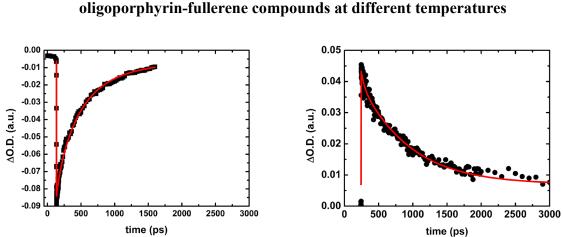


Figure S8.6. Rate constants for excitation energy migration plotted against 1/T, P_2 - C_{60} (red), P₃- C_{60} (green), P₄- C_{60} (blue) and P₆- C_{60} (cyan)). The activation energies estimated from the slopes are: P₃- C_{60} = 1.01 kcal/mol, P₄- C_{60} = 1.16 kcal/mol and P₆- C_{60} = 2.11 kcal/mol.

| T / K | | P ₂ | -C ₆₀ | | | P ₃ -C ₆₀ | | | |
|-------|------|----------------|------------------|---------------|----------------|---------------------------------|------|---------------|--|
| | α1 | τ_1 / ps | α2 | τ_2 / ps | α ₁ | τ_1 / ps | α2 | τ_2 / ps | |
| 140 | 0.59 | 37 | 0.41 | 337 | 0.47 | 88 | 0.53 | 536 | |
| 160 | 0.71 | 32 | 0.29 | 311 | 0.54 | 58 | 0.46 | 438 | |
| 180 | 0.74 | 29 | 0.26 | 332 | 0.60 | 51 | 0.40 | 306 | |
| 200 | 0.76 | 23 | 0.24 | 329 | 0.68 | 42 | 0.32 | 310 | |
| 220 | 0.80 | 22 | 0.20 | 361 | 0.62 | 35 | 0.38 | 203 | |
| 240 | 0.81 | 21 | 0.18 | 352 | 0.63 | 29 | 0.37 | 194 | |
| 260 | 0.83 | 20 | 0.17 | 363 | 0.66 | 25 | 0.34 | 181 | |
| 280 | 0.88 | 19 | 0.12 | 445 | 0.68 | 23 | 0.32 | 160 | |
| 300 | 0.81 | 19 | 0.19 | 489 | 0.69 | 20 | 0.30 | 119 | |

Table 8.1. Temperature dependence of the fitted lifetimes for P_2 - C_{60} , P_3 - C_{60} , P_4 - C_{60} , and P_6 - C_{60} excited at 495 nm, including the normalized pre-exponential factors.

| T / K | | P ₄ | -C ₆₀ | | | P ₆ -C ₆₀ | | | |
|-------|------|-----------------------|------------------|---------------|------|---------------------------------|------|---------------|--|
| | α1 | τ_1 / ps | α2 | τ_2 / ps | α1 | τ_1 / ps | α2 | τ_2 / ps | |
| 140 | 0.59 | 158 | 0.41 | 660 | 0.43 | 269 | 0.57 | 707 | |
| 160 | 0.57 | 101 | 0.43 | 503 | 0.37 | 167 | 0.63 | 650 | |
| 180 | 0.56 | 97 | 0.44 | 447 | 0.29 | 102 | 0.71 | 573 | |
| 200 | 0.54 | 74 | 0.46 | 376 | 0.28 | 76 | 0.72 | 519 | |
| 220 | 0.47 | 52 | 0.53 | 285 | 0.34 | 57 | 0.66 | 471 | |
| 240 | 0.49 | 46 | 0.51 | 254 | 0.25 | 48 | 0.75 | 402 | |
| 260 | 0.45 | 31 | 0.55 | 215 | 0.22 | 36 | 0.78 | 337 | |
| 280 | 0.51 | 29 | 0.49 | 212 | 0.26 | 32 | 0.74 | 300 | |
| 300 | 0.52 | 24 | 0.48 | 216 | 0.23 | 26 | 0.77 | 285 | |



<u>Section S9:</u> Tables of the fitting parameters of the time absorption profiles of the oligoporphyrin-fullerene compounds at different temperatures

Figure S9.1. Time absorption profiles of P_4 - C_{60} at 220 K. Left: Time absorption profile at 730 nm showing the recovery of the Q-band absorption. **Right**: Transient absorption decay at 1015 nm where the C_{60} ⁻⁻ and P_4 ⁺⁺ species dominate the absorption. The red solid lines show typical fits using the parameters stated in tables 8.4a and 8.4b.

Table S9.1. Fitting parameter for the transient absorption time profiles for P_1 - C_{60} taken at 650 nm representing recovery of the Q-band bleaching.

| T / K | α1 | τ_1 / ps | α2 | τ_2 / ps | Io |
|-------|----------|---------------|----------|---------------|----------|
| 140 | -0.04681 | 6.4 | -0.03306 | 455 | -0.03113 |
| 160 | -0.00716 | 6.2 | -0.00750 | 349 | -0.00204 |
| 180 | -0.00510 | 5.9 | -0.00632 | 309 | -0.00155 |
| 200 | -0.02348 | 5.4 | -0.03191 | 274 | -0.01371 |
| 220 | -0.02646 | 5.2 | -0.03316 | 226 | -0.01363 |
| 240 | -0.00723 | 5.0 | -0.01295 | 208 | -0.00307 |
| 260 | -0.01760 | 4.5 | -0.03571 | 201 | -0.01912 |
| 280 | -0.00901 | 4.4 | -0.01636 | 191 | -0.00969 |
| 300 | -0.00452 | 4.4 | -0.02172 | 189 | -0.01213 |

| T / K | α1 | τ_1 / ps | α2 | τ_2 / ps | α3 | τ_3 / ps | I ₀ |
|-------|----------|---------------|----------|---------------|----------|---------------|----------------|
| 140 | -0.0077 | 37 | -0.0006 | 337 | -0.02671 | 732 | -0.00153 |
| 160 | -0.00933 | 32 | -0.00051 | 311 | -0.0434 | 692 | -0.00459 |
| 180 | -0.03533 | 29 | -0.00094 | 332 | -0.03727 | 640 | -0.00576 |
| 200 | -0.00919 | 23 | -0.00013 | 329 | -0.00516 | 584 | -0.00021 |
| 220 | -0.02531 | 22 | -0.00127 | 361 | -0.06235 | 493 | -0.00912 |
| 240 | -0.01408 | 21 | -0.00087 | 352 | -0.02113 | 454 | -0.00283 |
| 260 | -0.01205 | 20 | -0.00097 | 363 | -0.03214 | 373 | -0.0047 |
| 280 | -0.00401 | 19 | -0.00177 | 445 | -0.04834 | 290 | -0.00911 |
| 300 | -0.00306 | 19 | -0.00020 | 489 | -0.02061 | 276 | -0.0005 |

Table S9.2a. Fitting parameter for the transient absorption time profiles for P_2 - C_{60} taken at720 nm representing recovery of the Q-band bleaching.

Table S9.2b. Fitting parameter for the transient absorption time profiles for P_2 - C_{60} taken at 1015 nm.

| T / K | α1 | τ_1 / ps | α ₂ | τ_2 / ps | α3 | τ_3 / ps | I ₀ |
|-------|---------|---------------|----------------|---------------|---------|---------------|----------------|
| 140 | 0.00076 | 37 | -0.00048 | 337 | 0.00481 | 732 | 0.00067 |
| 160 | 0.00196 | 32 | -0.00092 | 311 | 0.00551 | 692 | 0.00053 |
| 180 | 0.00181 | 29 | -0.00051 | 332 | 0.00537 | 640 | 0.00016 |
| 200 | 0.00271 | 23 | -0.00024 | 329 | 0.00305 | 584 | 0.00043 |
| 220 | 0.00159 | 22 | -0.00032 | 361 | 0.00371 | 493 | 0.00058 |
| 240 | 0.00212 | 21 | -0.00012 | 352 | 0.00321 | 454 | 0.00032 |
| 260 | 0.00102 | 20 | -0.00071 | 363 | 0.00416 | 373 | 0.00010 |
| 280 | 0.00212 | 19 | -0.00097 | 445 | 0.00558 | 290 | 0.00059 |
| 300 | 0.00108 | 19 | -0.00031 | 489 | 0.00449 | 276 | 0.00071 |

| T / K | α1 | τ_1 / ps | α2 | τ_2 / ps | α3 | τ_3 / ps | I ₀ |
|-------|----------|---------------|----------|---------------|----------|---------------|----------------|
| 140 | -0.0043 | 88 | -0.01373 | 536 | -0.00066 | 904 | -0.012 |
| 160 | -0.01715 | 58 | -0.01073 | 438 | -0.0105 | 794 | -0.025 |
| 180 | -0.00119 | 51 | -0.0048 | 306 | -0.06186 | 710 | -0.0216 |
| 200 | -0.02776 | 42 | -0.02335 | 310 | -0.03643 | 696 | -0.026 |
| 220 | -0.01525 | 35 | -0.01774 | 203 | -0.0529 | 627 | -0.021 |
| 240 | -0.02092 | 29 | -0.03724 | 194 | -0.0212 | 554 | -0.031 |
| 260 | -0.04388 | 25 | -0.04044 | 181 | -0.04029 | 504 | -0.027 |
| 280 | -0.00845 | 23 | -0.01275 | 160 | -0.06033 | 483 | -0.0162 |
| 300 | -0.00503 | 20 | -0.00076 | 119 | -0.08282 | 463 | -0.02571 |

Table S9.3a. Fitting parameter for the transient absorption time profiles for P_3 - C_{60} taken at 730 nm representing recovery of the Q-band bleaching.

Table S9.3b. Fitting parameter for the transient absorption time profiles for P_3 - C_{60} taken at 1015 nm.

| T / K | α1 | τ_1 / ps | α ₂ | τ_2 / ps | α3 | τ_3 / ps | I ₀ |
|-------|---------|---------------|----------------|----------------------|---------|---------------|----------------|
| 140 | 0.00105 | 88 | -0.00126 | 536 | 0.00668 | 904 | 0.0010 |
| 160 | 0.00071 | 58 | -0.00065 | 438 | 0.00455 | 794 | 0.0005 |
| 180 | 0.00088 | 51 | -0.00199 | 306 | 0.00769 | 710 | 0.0010 |
| 200 | 0.00075 | 42 | -0.00201 | 310 | 0.01161 | 696 | 0.0010 |
| 220 | 0.00038 | 35 | -0.00147 | 203 | 0.00989 | 627 | 0.00075 |
| 240 | 0.00046 | 29 | -0.00192 | 194 | 0.00952 | 554 | 0.00065 |
| 260 | 0.00266 | 25 | -0.00480 | 181 | 0.01259 | 504 | 0.00064 |
| 280 | 0.00552 | 23 | -0.00212 | 160 | 0.02051 | 483 | 0.002 |
| 300 | 0.00012 | 20 | -0.00022 | 119 | 0.00332 | 463 | 0.001 |

| T / K | α1 | τ_1 / ps | α2 | τ_2 / ps | α3 | τ_3 / ps | I ₀ |
|-------|----------|---------------|----------|---------------|----------|---------------|----------------|
| 140 | -0.0035 | 158 | -0.00122 | 660 | -0.0127 | 917 | -0.0045 |
| 160 | -0.00781 | 101 | -0.01178 | 503 | -0.00554 | 846 | -0.0079 |
| 180 | -0.01337 | 97 | -0.00181 | 447 | -0.01908 | 734 | -0.0097 |
| 200 | -0.02109 | 74 | -0.00766 | 376 | -0.03657 | 708 | -0.0050 |
| 220 | -0.00697 | 52 | -0.04257 | 285 | -0.0285 | 689 | -0.0062 |
| 240 | -0.02319 | 46 | -0.01182 | 254 | -0.02293 | 604 | -0.0019 |
| 260 | -0.02999 | 31 | -0.01972 | 215 | -0.01231 | 567 | -0.0012 |
| 280 | -0.01143 | 29 | -0.01096 | 212 | -0.01101 | 538 | -0.0017 |
| 300 | -0.00207 | 24 | -0.00319 | 216 | -0.03318 | 499 | -0.0021 |

Table S9.4a. Fitting parameter for the transient absorption time profiles for P_4 - C_{60} taken at 750 nm representing recovery of the Q-band bleaching.

Table S9.4b. Fitting parameter for the transient absorption time profiles for P_4 - C_{60} taken at 1015 nm.

| T / K | α ₁ | τ_1 / ps | α2 | τ_2 / ps | α3 | τ_3 / ps | I ₀ |
|-------|----------------|---------------|----------|---------------|---------|---------------|----------------|
| 140 | 0.01638 | 158 | -0.00918 | 660 | 0.03414 | 917 | 0.0020 |
| 160 | 0.00902 | 101 | -0.00814 | 503 | 0.04898 | 846 | 0.0054 |
| 180 | 0.00355 | 97 | -0.00091 | 447 | 0.0354 | 734 | 0.0075 |
| 200 | 0.00293 | 74 | -0.00419 | 376 | 0.03192 | 708 | 0.0085 |
| 220 | 0.00472 | 52 | -0.00269 | 285 | 0.03421 | 689 | 0.0070 |
| 240 | 0.00142 | 46 | -0.00582 | 254 | 0.03859 | 604 | 0.0071 |
| 260 | 0.00045 | 31 | -0.00041 | 215 | 0.01273 | 567 | 0.00255 |
| 280 | 0.00034 | 29 | -0.02201 | 212 | 0.11566 | 538 | 0.01154 |
| 300 | 0.00939 | 24 | -0.04715 | 216 | 0.14728 | 499 | 0.00972 |

| T / K | α1 | τ_1 / ps | α ₂ | τ_2 / ps | α3 | τ_3 / ps | I ₀ |
|-------|----------|---------------|----------------|---------------|----------|---------------|----------------|
| 140 | -0.00514 | 269 | -0.00245 | 707 | -0.00194 | 1112 | -0.001 |
| 160 | -0.00107 | 167 | -0.00139 | 650 | -0.00188 | 1005 | -0.003 |
| 180 | -0.00593 | 102 | -0.00724 | 573 | -0.00392 | 951 | -0.0044 |
| 200 | -0.00334 | 76 | -0.00471 | 519 | -0.00556 | 903 | -0.0035 |
| 220 | -0.00588 | 57 | -0.00651 | 471 | -0.0071 | 821 | -0.0095 |
| 240 | -0.00528 | 48 | -0.00525 | 402 | -0.00349 | 786 | -0.0165 |
| 260 | -0.00302 | 36 | -0.0032 | 337 | -0.00498 | 748 | -0.0058 |
| 280 | -0.00482 | 32 | -0.00383 | 300 | -0.00372 | 658 | -0.005 |
| 300 | -0.00177 | 26 | -0.00446 | 285 | -0.00342 | 643 | -0.0031 |

Table S9.5a. Fitting parameter for the transient absorption time profiles for P_{6} - C_{60} taken at 785 nm representing recovery of the Q-band bleaching.

Table S9.5b. Fitting parameter for the transient absorption time profiles for P_6 - C_{60} taken at 1015 nm.

| T / K | α1 | τ_1 / ps | α2 | τ_2 / ps | α3 | τ_3 / ps | I ₀ |
|-------|---------|---------------|---------|----------------------|---------|---------------|----------------|
| 140 | 0.00316 | 269 | 0.00075 | 707 | 0.00315 | 1112 | 0.0007 |
| 160 | 0.00621 | 167 | 0.09045 | 650 | 0.04249 | 1005 | 0.00947 |
| 180 | 0.00068 | 102 | 0.01503 | 573 | 0.00182 | 951 | 0.00191 |
| 200 | 0.00145 | 76 | 0.00618 | 519 | 0.00449 | 903 | 0.00081 |
| 220 | 0.00072 | 57 | 0.00587 | 471 | 0.0126 | 821 | 0.00108 |
| 240 | 0.00060 | 48 | 0.00334 | 402 | 0.01125 | 786 | 0.00145 |
| 260 | 0.00089 | 36 | 0.00199 | 337 | 0.01373 | 748 | 0.00108 |
| 280 | 0.00044 | 32 | 0.00141 | 300 | 0.01801 | 658 | 0.0016 |
| 300 | 0.00088 | 26 | 0.00247 | 285 | 0.0148 | 643 | 0.00177 |

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