Table S1. Summary of the bioinformatic analysis targeting potential new substrates for PC5/6. Listed are the most relevant protein candidates (mouse) selected based on the following screening criteria: i) presence of a signal peptide; ii) presence of a PC5/6 specific cleavage site (R/K)- X_n -R \downarrow N (n = 0, 2, 4, or 6 and X is any aa, except Cys); iii) the proteins/peptides have cardiac relevance when their gene is knocked out; iv) the cleavage site motif is conserved in both human and mouse. In bold are depicted the candidates considered for in vitro digestion experiments with the four constitutively secreted PCs: furin, PACE4, PC5/6, and PC7. The synthetic peptides designed for this purpose are also listed in bold. TM, transmembrane domains

UniProt #	Symbol	Protein	Length (aa)	Motif Studied peptide	ТМ
Q9R229	BMP10	Bone morphogenetic protein 10	421	DSSARIRR ₃₁₃ NAKG	-
Q62226	Shh	Sonic hedgehog	437	SGRYEG <u>K</u> IT <u>R₆₉NSE</u>	-
P29319-1	Epha3	Ephrin type-A receptor 3	983	TIKKDRTSR449NSISL	1
O08852	Pkd1	Polycystin-1	4293	QVLMRRSQR ₂₁₆₂ NYLE	> 1
O88393	Tgfbr3	Transforming growth factor, beta receptor III	850	TELKIAR ₁₈₃ NIYI	1
Q62181	Sema3c	Semaphorin-3C	751	<u>K</u> AY <u>R</u> 573N	-
Q01279	Egfr	Epidermal growth factor receptor	1210	<u>r</u> elpm <u>r₁₃₈n</u>	1
P11276	Fn1	Fibronectin 1	2477	<u>R</u> LPVN <u>R₉₅₇N</u>	-
				<u>r</u> vpps <u>r</u> ₁₅₀₀ n	
Q62009	Postn	Periostin	838	<u>K</u> QG <u>R</u> 483N	-

Supplemental Table S1

Table S2. Comparison of the in vitro processing of 12-mer mouse BMP10 peptides, WT-mBMP10 and N315D-mBMP10, at the predicted $\underline{R}_{311}IR\underline{R}_{314}\downarrow$ cleavage site, and the kinetic constants of the furin cleavage of the WT peptide. (A) Each synthetic peptide (200 µM) was incubated for 2h in vitro with 2U of purified soluble furin, PC5/6, PACE4, or PC7, as described in Materials and Methods. The products were separated by RP-HPLC and detected at 214 nm. The % cleavage was calculated as the ratio of the normalized peak areas (peak area/number of peptide bonds) of the C-terminal fragment (NAKG for WT-mBMP10, or DAKG for N315-DmBMP10) and of the intact 12-mer peptide (at time zero). n.d., not determined. (B) Calculated kinetic constants from the digestion of the 12-mer WT peptide. Fitted values \pm SD to the fit.

Α	Peptide (200 μM)	Sequence	hFurin	hPACE4	hPC5	rPC7
	WT- mBMP10	DSSA <u>R₃₁₀IRR₃₁₃</u> ↓NAKG	86%	67%	68%	4%
	N314D- mBMP10	DSSA <u>R₃₁₀IR<u>R</u>₃₁₃↓ DAKG</u>	42%	30%	23%	n.d.

B	K _{m(app)}	V _{max(app)}	V _{max(app)} /K _{m(app)}	
	(μΜ)	(μM·h⁻¹)	(h ⁻¹)	
	34.9 ± 8.1	65.5 ± 5.4	1.9	

Supplemental Table S2

Fig. S1. Schematic representation of the mouse preproBMP10 and its processed forms. Upon removal of the signal peptide (SP), the resulting inactive BMP10 precursor (proBMP10; ~ 60 kDa) is proteolytically processed at the motif RIRR \downarrow into a 108 aa mature growth factor. Mature BMP10 contains the conserved pattern of 7 cysteines (depicted in ball-stick representation) out of which one is engaged in an interchain disulfide bond. Also shown are the predicted N-glycosylation sites (N67, N131) and the sequence of the 12-mer peptide (mBMP10 peptide) encompassing the predicted cleavage site of mouse proBMP10, which was used for the in vitro digestion studies.



Fig. S2. Endo H and PNGase F sensitivity of proBMP10 and processed BMP10 (prosegment). (A) Cell lysates and 20h conditioned media from COS-1 cells transiently transfected with (ProtC)-BMP10 were subjected to digestion with either PNGase F or endo H. The digestion mixtures were analyzed by Western blotting using a mouse ProtC Ab. The blot shows that both secreted proBMP10 and prosegment are PNGase F sensitive and endo H resistant. In the cells, proBMP10 is sensitive to both PNGase F and endo H. Note the presence in the cells of a non-specific band (*) just below and partially overlapping with the specific band resulting from the digestion with either of the two glycosidases. A control from cells expressing an empty vector is shown for reference. (B) Western blot of PNGase F digestion product of cell lysates from the furin-deficient CHO-FD11 cells expressing (ProtC)-BMP10 (proBMP10), as compared to control undigested protein. Note that in these cells the non-specific band seen in COS-1 cells (* in panel A) is not present.



Fig. S3. Effects of RVKR-cmk and D6R on the TGN processing of the neuroendocrine secretory precursor protein pro7B2 into 7B2. Western blot analyses of 20h conditioned media from COS-1 cells transiently expressing either no protein (vector) or mouse pro7B2 and collected after no treatment (DMSO) or treatment with either the cell permeable convertase inhibitor RVKR-cmk (30 μ M) or the cell surface inhibitor D6R (20 μ M). Proteins were revealed with a rabbit 7B2 Ab (1).

(1) Seidah, N. G., Hsi, K. L., De Serres, G., Rochemont, J., Hamelin, J., Antakly, T., Cantin, M., and Chretien, M. (1983) *Arch. Biochem. Biophys.* **225**, 525-534



Supplemental Figure S3

Fig. S4. The transmembrane domain of furin is required for its efficient processing of proBMP10. (A) Schematic diagram depicting human proFurin and proPC7 and chimeras of human furin generated by the swapping of its transmembrane (TM) and/or cytopolasmic (CT) domains with the corresponding human PC7 ones (furin_{CT-PC7}; furin_{TMCT-PC7}) (33). (B) Western blot analyses of the 20h conditioned media from CHO-FD11 cells transiently transfected with either empty vector (vector; lane 6), or with a vector expressing (ProtC)-BMP10 and vectors expressing furin_{CT-PC7} (lane 1), furin_{TMCT-PC7} (lane 2), soluble furin (sFurin; lane 3), furin (lane 4), or empty vector (vector; lane 5). Proteins were revealed with a rabbit ProtC Ab. Please note that the two panels of the figure come from different parts of the same blot.



Β



Supplemental Figure S4

Fig. S5. In mouse heart, PACE4 does not significantly contribute to the processing of proBMP10. (A) Tissue extracts from right atria isolated from adult male mice, WT and total PACE4 knockout (PACE4-KO), were analyzed by Western blotting using mouse BMP10 or β -actin antibodies. β -actin was used as a loading control. Proteins were resolved on SDS-PAGE 8% Tris-tricine gels under non-reducing conditions. The migration position of mature BMP10 is emphasized. (B) The bands from Western blot in (A) were quantitated using ImageJ software (National Institutes of Health) and the relative band intensities of mature BMP10 and β -actin were plotted for WT (patterned bar) and total PACE4-KO (grey bar) mice. Means \pm SDEV are given and n = 5-7 mice per group. There is no significant difference between the two groups.



Supplemental Figure S5