Online supplemental material

S1 Figure Legends

FIGURE S1. GSK3 inhibitors did not affect survival and proliferation of CD4⁺T cells. *A-B*, CD4⁺ T cells were polarized toward Th17 cells for 5 days cultured on irradiated lymph node cells and splenocytes in medium supplemented with 20 ng/mL IL-6 and 5 ng/mL TGF β , with the indicated GSK3 inhibitors 10 mM LiCl, 1 µM CHIR99021, 1 µM TDZD-8, or 10 µM SB415286. Cells were washed and stimulated for 4 hr with PMA/ionomycin and CD4⁺T cells were analyzed for live-dead staining by flow cytometry. C, CD4⁺T cells were polarized toward Th17 cells for 5 days cultured on anti-CD3/CD28 beads in medium supplemented with 20 ng/mL IL-6 and 5 ng/mL TGF β , with the indicated GSK3 inhibitors 10 mM LiCl, 1 µM CHIR99021, 1 µM TDZD-8, or 10 µM SB415286. Cells were washed and stimulated for 4 hr with PMA/ionomycin and CD4⁺T cells were analyzed by TUNNEL staining by flow cytometry. D, CD4⁺ T cells were cultured on anti-CD3/CD28 beads for 5 days with the indicated GSK3 inhibitors 10 mM LiCl, 1 µM CHIR99021, 1 uM TDZD-8, or 10 uM SB415286. Cells were washed and stimulated for 4 hr with PMA/ionomycin and CD4⁺ T cells were analyzed by TUNNEL staining by flow cytometry. E, CD4⁺T cells were polarized toward Th17 cells for 5 days cultured on irradiated lymph node cells and splenocytes in medium supplemented with 20 ng/mL IL-6 and 5 ng/mL TGF β , with the indicated GSK3 inhibitors 10 mM LiCl, 1 µM CHIR99021, 1 µM TDZD-8, or 10 µM SB415286. Cells were washed and stimulated for 4 hr with PMA/ionomycin and CD4⁺T cells were analyzed for CFSE staining by flow cytometry gated on IL-17A⁺CD4⁺T cells. F, CD4⁺T cells were cultured for 5 days on irradiated lymph node cells and splenocytes with the indicated GSK3 inhibitors 10 mM LiCl, 1 µM CHIR99021, 1 µM TDZD-8, or 10 µM SB415286. Cells were washed and stimulated for 4 hr with PMA/ionomycin and CD4⁺ T cells were analyzed for CFSE staining by flow cytometry. G, GSK3 α and GSK3 β were knocked down by siRNA with a double-stranded

RNA oligomer labeled with Alexa-Fluor-555 in CD4⁺T cells for 48 h, cells were differentiated on feeder cells for 5 days, and cells were restimulated with ionomycin/PMA for 4 h and analyzed by flow cytometry to evaluate the percentage of siRNA-transfected cells.

FIGURE S2. GSK3 inhibitors reduce IL-17A-producing CD4⁺ T cells. *A-B*, CD4⁺ T cells were polarized toward Th17 cells for 5 days (A) with varying concentrations of CHIR99021, or (B) with 10 mM lithium added on different days of culture. T cells were recovered and stimulated with PMA/ionomycin for 4 h and IL-17A in the media was measured by ELISA. n=3, *p<0.05. C, CD4⁺ T cells were polarized towards Th17 for 5 days, cells were restimulated with ionomycin/PMA/BFA with or without 10 mM LiCl or 1 µM CHIR99021 for 4 h, and IL-17A-producing CD4⁺ T cells were analyzed by flow cytometry (n=2). D, $CD4^+T$ cells were polarized towards Th17 cells during culture with anti-CD3/anti-CD28 beads for 5 days with GSK3 inhibitors, followed by measurement of IL-17A-producing CD4⁺T cells by flow cytometry (n=3). E, mRNA levels of GSK3 β and IL-17A were measured by qRT-PCR after 4 days of polarization towards Th17, with or without 1 µM CHIR99021. F, CD4⁺T cells from wild-type and ROR $\gamma T^{+/-}$ mice were polarized toward Th17 cells for 5 days and analyzed on different days of culture for their content in IL-17A⁺ CD4⁺ T cells. T cells were recovered and stimulated with PMA/ionomycin for 4 h and IL-17A⁺CD4⁺ T cells were analyzed by flow cytometry (n=3), Insert: ROR γ T was immunoblotted in CD4⁺T cells from wildtype (WT) and ROR $\gamma T^{+/-}$ mice after 3 days of polarization, or not, towards Th17 cells. Blots were reblotted for the loading control β -actin. G, CD4⁺T cells isolated from GSK3 knockin mice were polarized toward Th17 cells for 5 days cultured on irradiated lymph node cells and splenocytes in medium supplemented with 20 ng/mL IL-6 and 5 ng/mL TGFβ with or without 10 μM SB415286. Cells were washed and stimulated for 4 hr with PMA/ionomycin/BFA and IL-17A-producing CD4⁺ T cells were analyzed by flow cytometry.

FIGURE S3. GSK3 inhibitors reduce responses to IL-6, IL-23, and IL-21. *A*, Chromatin immunoprecipitation of STAT3 on the promoter of *IL-17A* in response to 30 min treatment with 20 ng/mL IL-6 with or without 10 mM LiCl. Q-PCR was applied to quantify the ChIP and input DNA. Values were normalized to input values (*p<0.05, n=3). *B-C*, CD4⁺T cells were polarized towards Th17 cells for 5 days cultured on irradiated lymph node cells and splenocytes in medium supplemented with (*B*) 10 ng/mL IL-23, 20 ng/mL IL-6 and 0.25 ng/mL TGFβ or (*C*) 75 ng/mL IL-21 and 5 ng/mL TGFβ with the GSK3 inhibitors 1 μM CHIR99021 or 1 μM TDZD-8 (n=5). IL-17A-producing CD4⁺T cells were analyzed after 4 h restimulation with PMA/ionomycin/BFA by flow cytometry (n=2-3). *D-E*, CD4⁺T cells were pretreated for 2 h with 10 mM LiCl (Li), 1 μM CHIR99021 (CH), or 1 μM TDZD-8 (TDZD) and then stimulated for 30 min with (*D*) 10 ng/mL IL-23 or (*E*) 75 ng/mL IL-21, and cell lysates were immunoblotted for phospho-Tyr705-STAT3, STAT3 and β-actin.

FIGURE S4. Effects of lithium treatment after Ft LVS or EAE. *A*, The percents of CD4⁺T cells, CD8⁺ T cells, CD11c⁺ dendritic cells and activated macrophages (CD45^{high} F4/80⁺) were determined in spleen cell preparations from lithium-treated or non-treated mice, 6 days after infection with Ft LVS. *B*, The percents of spleen IL-17A-producing, IFN γ -producing or FoxP3-expressing CD4⁺ T cells were analyzed by flow cytometry after restimulation with PMA/ionomycin/BFA for 4 h or no restimulation (FoxP3). *C*, Spinal cord lysates were immunoblotted for phospho-Tyr705-STAT3, phospho-Tyr701-STAT1, total STAT3, STAT1, and phospho-Ser9-GSK3β (n=2).





Figure S2



Figure S3





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+ Lithium	
<u> </u>	Phospho-Tyr701-STAT1
	STAT1
	Phospho-Tyr705-STAT3
	STAT3
	Phospho-Ser9-GSK3β

Figure S4