

Supporting Information

**Systematic Variation in the Temperature Dependence of
Physiological and Ecological Traits**

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SI Materials and Methods

Body Size Estimation. Wet mass estimates for each species are required to calculate mass-corrected scaling coefficients and to interconvert between mass-specific and per capita trait measurements. The diversity of taxa in our data and the large number of mass estimates necessitated automation of this process, so for cases when wet body mass is not given in the original source, we create and use an algorithm that assigns a wet mass estimate to species in each data series. This algorithm is based largely on the taxonomic relatedness to published size estimates and length-mass regressions, and allows us to rapidly obtain estimates of wet body mass that well match published measurements. Our algorithm comprises four main steps: 1) body size measurements (mass, length, or otherwise) are acquired from the original data source when available; 2) When no size estimate is given in the original source (38% of responses), body size is assigned using measurements compiled from the literature into a database; 3) All non-mass estimates of size are converted to mass (wet, dry, or ash-free-dry) using 364 published size-mass regressions. To be conservative, we do not extrapolate outside the non-mass size range of individuals that were used to construct these regressions; 4) All non-wet masses are converted to wet mass using 10 published taxon-specific conversion ratios. This algorithm relies on a richer set of literature data and regressions than previous studies.

Analysis of Intercept Coefficients for Monotonic Rise and Fall Responses. Random error in fitted regression parameters will typically result in a normal distribution. Our measured intercept coefficients (Eq. 1) are indeed distributed approximately normally. Thus the right-skewness observed in activation energies (Fig. 2 & 3) is most likely biologically significant. For analysis of effect of motivation on body velocity (in our test of the life-dinner principle, see main text) we calculate intercept coefficients at 20°C across all traits (standardized intercept). The exponentials of the resulting intercepts are the predicted trait values at 20°C. We correct these standardized values for the effect of body mass by multiplying each value by $m^{-1/4}$, where m is the mass of the consumer species (resource mass was used for traits that had negative motivation). Choosing any other standardization temperature between 0 and 50°C, or an allometric scaling exponent of 1/3, does not qualitatively affect our results.

Estimation of T_{opt} . We compare the T_{opt} values estimated as the temperature at which the maximum trait value occurs (main text), with those obtained by fitting a unimodal function (Fig. S1). We choose the Johnson & Lewin (1) model, a unimodal extension of the Boltzmann-Arrhenius function (Eq. 1) for trait rises (2-7):

$$h(T) = ce^{-\frac{E}{kT}} / \left(1 + e^{-\frac{1}{kT} \left(E_D - \left(\frac{E_D}{T_{opt}} + k \ln \left(\frac{E}{E_D - E} \right) \right) T \right)} \right) \quad (2)$$

Here the additional thermodynamic parameter E_D determines the steepness of decline of the trait values at temperatures higher than T_{opt} , while c is a constant. All responses classified as being unimodal are fitted to this model using nonlinear least-squares regression (8). We use the Levenberg-Marquardt algorithm with a maximum of 2000 iterations and error tolerance of 1×10^{-30} . Reasonable response-specific initial values are allocated for the parameters to improve algorithm convergence and parameter estimation. We find that the overall mean T_{opt} value obtained from the unimodal model fits (26.0°C) is comparable with that from the direct method described above (25.3°C). We use the direct method for T_{opt} estimation because 38% of the fits to the Johnson & Lewin model have very large confidence intervals (bounds > 25% away from the mean).

Treatment of Pseudoreplicates. We define pseudoreplicates as responses that share taxa (or combinations of taxa for species interaction traits) and experimental conditions. From each pseudoreplicate group, we obtain a single value of the parameters E , T_{opt} and scaling coefficient by taking the weighted average of their estimates across the individual responses in that group. The weights are a linear function of the mean number of data points across responses within each pseudoreplicate group.

SI References

1. Johnson FH & Lewin I (1946) The growth rate of *E. coli* in relation to temperature, quinine and coenzyme. *Journal of Cellular and Comparative Physiology* 28(1):47-75.
2. Farquhar GD, Caemmerer S, & Berry JA (1980) A biochemical model of photosynthetic CO₂ assimilation in leaves of C₃ species. *Planta* 149(1):78-90.
3. Sharpe P & DeMichele D (1977) Reaction kinetics of poikilotherm development. *Journal of Theoretical Biology* 64:649-670.
4. Schoolfield RM, Sharpe PJH, & Magnuson CE (1981) Non-linear regression of biological temperature-dependent rate models based on absolute reaction-rate theory. *Journal of Theoretical Biology* 88(4):719-731.
5. Johnson FH, Eyring H, & Stover BJ (1977) Reaction rate theory in bioluminescence and other life phenomena. *Annual Review of Biophysics and Bioengineering* 6(1):111-133.
6. Ratkowsky DA, Olley J, & Ross T (2005) Unifying temperature effects on the growth rate of bacteria and the stability of globular proteins. *Journal of Theoretical Biology* 233(3):351-362.
7. Johnson FH, Eyring H, & Stover BJ (1974) *The theory of rate processes in biology and medicine* (Wiley, New York,).
8. Seber GAF & Wild CJ (2003) *Nonlinear regression* (Wiley-Interscience, Hoboken, N.J.).
9. Fargues J, Goettel MS, Smits N, Ouedraogo A, & Rougier M (1997) Effect of temperature on vegetative growth of *Beauveria bassiana* isolates from different origins. *Mycologia* 89(3):383-392.
10. Gillooly JF, Brown JH, West GB, Savage VM, & Charnov EL (2001) Effects of size and temperature on metabolic rate. *Science* 293(5538):2248-2251.
11. Denny MW (1990) Terrestrial versus aquatic biology - the medium and its message. *American Zoologist* 30(1):111-121.

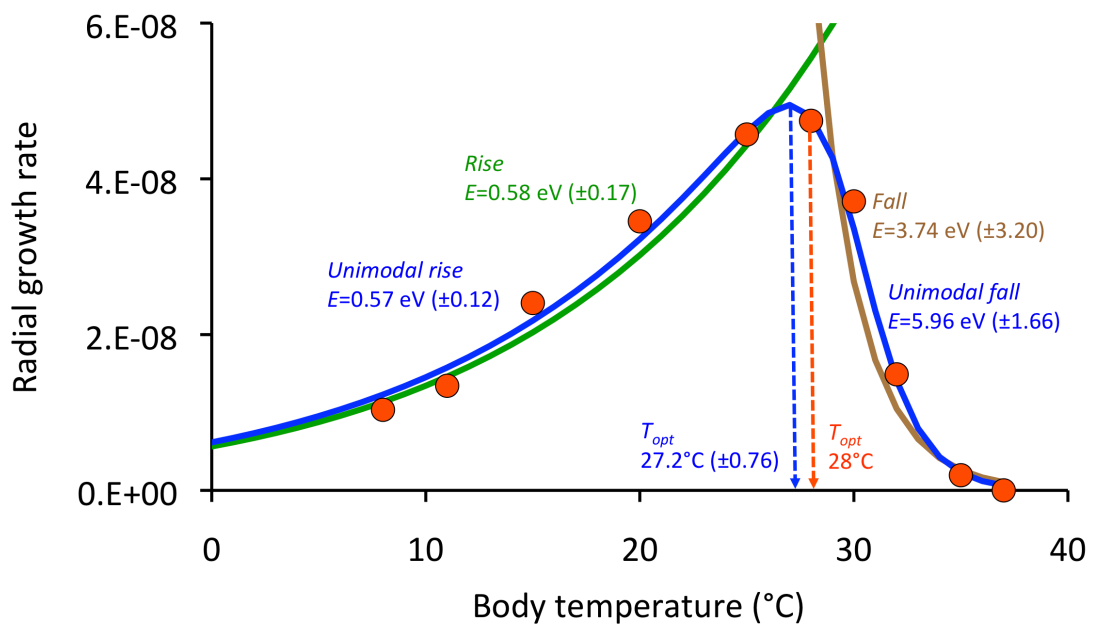


Fig. S1. The unimodal thermal response of radial growth rate of sac fungi (m / (colony * s)). Green and brown curves are OLS regressions to the Boltzmann-Arrhenius model (Eq. 1) for the subset of data that are the rise and fall components, respectively. These components were extracted by the algorithm described in the Materials and Methods. For this particular response, the rise was obtained by the algorithm through removal of the measurements at the 4 highest temperatures and the fall through removal of measurements at the 5 lowest temperatures. The blue curve is the best fit to the Johnson & Lewin model (SI Methods) (1). Values shown are estimated activation energies with 95% confidence intervals for the respective response components. Dotted vertical arrows are estimated temperatures for T_{opt} —the temperature at which the trait value is optimal—calculated from the direct method (red) and Johnson & Lewin model (blue). See SI for more details. Data are from Fargues et al. (9).

Table S1. Trait definitions. Traits are listed alphabetically within level of organization. Unless stated otherwise, traits are measured per capita (i.e., per individual). All measurements are listed in SI units. C_n (R_n) is the number of individual consumers (resources). When the trait involves a single species, C_n is used as the default. Times are denoted as ‘organism * s’ so that rates are interpreted as per individual, because rate is our focal unit. A (arena size) is measured as area or volume depending on the dimensionality of the habitat, as determined by the original authors. We standardize by resource and consumer density whenever possible, and this is the default unit of all traits listed below. Weights are wet mass unless otherwise stated. Original trait definitions and units are given in the original sources (Table S3 & S6).

<u>Trait name</u>	<u>Trait unit</u>	<u>Motivation</u>	<u>Trait definition</u>
<u>Internal</u>			
Ammonia Excretion Rate	kg / (C_n * s)	autonomic	Rate of ammonia (NH ₃) mass excretion per consumer.
Digestion Rate	R_n / (C_n * s)	autonomic	Rate at which resources are digested per consumer.
Faecal Excretion Rate	kg / (C_n * s)	autonomic	Rate of faecal mass excretion per consumer.
Feeding Heart Beat Rate	event / (C_n * s)	positive	Rate of heartbeats per consumer while filter feeding.
Filtration Metabolic Efficiency	m ³ / (m ³ * C_n)	autonomic	Metabolic efficiency of the filtration process expressed as water volume per oxygen volume per consumer.
Food Energy Assimilation Efficiency	proportion / C_n	autonomic	Efficiency of digesting ingested energy per consumer expressed as the amount of energy that is digested in proportion to that which is ingested.
Food Mass Conversion Efficiency	proportion / C_n	autonomic	Efficiency of converting food mass to body mass per consumer expressed as growth in tissue mass in proportion to total mass of resource consumed.
Gut Clearance Rate	event / (C_n * s)	autonomic	Rate food moves through a consumer from initial ingestion to evacuation (i.e., faeces).
Gut Loading Rate	event / (C_n * s)	autonomic	Rate at which the gut physically fills with food.
In Vitro Heart Beat Rate	event / s	autonomic	Rate of heartbeats measured in a heart removed from a living organism.

<u>Trait name</u>	<u>Trait unit</u>	<u>Motivation</u>	<u>Trait definition</u>
In Vitro Muscle Isometric Tension	N / m^2	autonomic	Isometric tension of muscle measured in muscle removed from a living organism.
In Vitro Muscle Optimal Phase	proportion	autonomic	Phase at which the power output of the muscle is maximum in muscle removed from a living organism.
In Vitro Muscle Optimal Rate	event / s	autonomic	Optimal frequency corresponding to the maximum power output of the muscle removed from a living organism.
In Vitro Muscle Optimal Strain	proportion	autonomic	Strain at which the power output of the muscle is maximum in muscle removed from a living organism.
In Vitro Muscle Power Output	W	autonomic	Power output of muscle measured in muscle removed from a living organism.
In Vitro Muscle Shortening Velocity	m / s	autonomic	Velocity of muscle shortening measured in muscle removed from a living organism.
In Vitro Muscle Work Per Cycle	J / event	autonomic	Muscle work per cycle at optimal frequency measured in muscle removed from a living organism.
Log-Linear Gut Clearance Rate	event / (C_n * s)	autonomic	The slope of the regression of log gut content mass per consumer individual per time.
Oxygen Mass Scope For Activity	$\text{kg} / (C_n * \text{s})$	voluntary	Amount of oxygen available for use for activity measured as rate of oxygen mass production per consumer.
Photosynthetic Oxygen Production Rate	$\text{kg} / (C_n * \text{s})$	autonomic	Organic oxygen mass production rate per consumer through photosynthesis.
POC Photosynthetic Oxygen Production Rate	$\text{kg} / (\text{kg} * \text{s})$	autonomic	Carbon-specific (POC) oxygen mass production rate through photosynthesis.
Respiration Rate	$\text{kg} / (C_n * \text{s})$	autonomic	Organic oxygen mass consumption rate per consumer during respiration.
Square Root-Linear Gut Clearance Rate	event / (C_n * s)	autonomic	The slope of the regression of square root gut content mass versus time.

<u>Trait name</u>	<u>Trait unit</u>	<u>Motivation</u>	<u>Trait definition</u>
Surface Area-Specific Dark Respiration Rate	kg / (m ² * s)	autonomic	Surface area-specific CO ₂ production during dark respiration.
Surface Area-Specific Maximum Photosynthesis Rate	kg / (m ² * s)	positive	Surface area-specific maximum photosynthesis rate.
Surface Area-Specific Mitochondrial Respiration Rate	kg / (m ² * s)	autonomic	Surface-area specific respiration rate in leaf mitochondria during photosynthesis.
Surface Area-Specific Photosynthetic Oxygen Production Rate	kg / (m ² * s)	autonomic	Surface area-specific oxygen production rate during photosynthesis.
Voluntary Heart Beat Rate	event / (C _n * s)	voluntary	Rate of heartbeats measured in an organism that is voluntarily stationary.
<u>Individual</u>			
48-hr Hatching Probability	proportion / C _n	autonomic	Probability of an egg having hatched at 48 hrs.
Avoidance Body Velocity	m / (R _n * s)	voluntary	Velocity of the body during movement in avoidance of a weak stimulus (differs from Escape Body Velocity because the stimulus is not an immediate threat).
Bite Rate	event / (C _n * s)	positive	Rate of bites or analogue (e.g., radular scrape) per consumer.
Critical Holding Velocity	m / (R _n * s)	negative	Velocity at which animal failed to hold position on the substrate when placed in a multi-speed flow chamber for a set time at sequentially increasing speeds.
Critical Travel Velocity	m / (R _n * s)	negative	Velocity at which an individual fails to maintain when placed in a multi-speed flow chamber for a set time at sequentially increasing speeds.
Critical Upright Time	R _n * s	negative	Time taken for animal to become completely exhausted from repeated up-righting of body.

<u>Trait name</u>	<u>Trait unit</u>	<u>Motivation</u>	<u>Trait definition</u>
Development Rate	$1 / (C_n * s)$	autonomic	Rate at which individuals complete development of one or more life stages.
Endurance Time	$R_n * s$	negative	Time maintained on a single-speed treadmill or flow chamber until exhaustion during escape locomotion.
Escape Angle of Body Turning	rad / R_n	negative	The sum of the absolute angles of turning of the head relative to the body during escape burst locomotion.
Escape Angular Rate of Body Turning	$rad / (R_n * s)$	negative	Velocity of the turning of the front of the resource relative to the mid-point throughout movement during escape burst locomotion.
Escape Body Acceleration	$m / (R_n * s^2)$	negative	Acceleration of the whole body during escape burst locomotion.
Escape Body Deceleration	$m / (R_n * s^2)$	negative	Deceleration of the whole body during escape burst locomotion.
Escape Body Power Production	W / R_n	negative	Power production of the whole body during escape burst locomotion.
Escape Body Response Rate	$event / (R_n * s)$	negative	Rate of response of a resource to an attacking consumer or otherwise negative stimulus.
Escape Body Velocity	$m / (R_n * s)$	negative	Velocity of the whole body during escape burst locomotion.
Escape Body Velocity Probability	$proportion / R_n$	negative	Velocity of the whole body during escape burst locomotion expressed as the ratio of sprint velocity of an individual to the maximum velocity of that individual in all trials at all temperatures.
Escape Gait Change Velocity	$m / (R_n * s)$	negative	Velocity at which resource changes gait during escape burst locomotion.
Escape Jump Contact Rate	$event / (R_n * s)$	negative	Rate of time the resource exerts force on substrate during an escape jump.
Escape Jump Distance	m / R_n	negative	Distance animal travels in a single escape jump.
Escape Jump Force	N / R_n	negative	Force exerted on the substrate by a resource during an escape jump.
Escape Jump Rate	$event / (R_n * s)$	negative	Jump rate of a resource during escape locomotion.

<u>Trait name</u>	<u>Trait unit</u>	<u>Motivation</u>	<u>Trait definition</u>
Escape Stroke Length	m / R_n	negative	Distance covered by a resource in a single locomotory stroke during escape.
Escape Stroke Peak Force	N / R_n	negative	Force attained on the substrate by the resource during a locomotory stroke during escape.
Escape Stroke Peak Force Rate	$N / (R_n * s)$	negative	Rate of force attained on the substrate by the resource during a locomotory stroke during escape.
Escape Stroke Rate	$event / (R_n * s)$	negative	Rate of locomotory strokes of a resource during escape.
Escape Tail Beat Rate	$event / (R_n * s)$	negative	Rate of tail beats of a resource during escape burst locomotion.
Flee Distance	m / R_n	negative	Distance moved by a resource when fleeing a predator before stopping (includes escape burst component and other slower movement, if present).
Foraging Body Undulation Rate	$event / (C_n * s)$	positive	Rate of undulating body strokes used for feeding.
Foraging Gill Beat Rate	$event / (C_n * s)$	positive	Rate of beating cilia on gill of living consumer measured by direct examination of cilia.
Foraging Submersion Rate	$event / (C_n * s)$	voluntary	Rate consumer swims underwater while foraging.
Foraging Velocity	$m / (C_n * s)$	voluntary	Velocity of the whole consumer when foraging for food.
In Vitro Gill Beat Rate	$event / s$	autonomic	Rate of cilia beating on gill fragments removed from a living organism measured by direct examination of cilia.
In Vitro Gill Particle Transport Velocity	m / s	autonomic	Velocity of particles in grooves of gill fragments removed from a living organism.
Individual Length Growth Rate	$m / (C_n * s)$	autonomic	Rate of increase in length of an individual.
Individual Mass Growth Rate	$kg / (C_n * s)$	autonomic	Rate of increase in mass of an individual.
Population Voluntary Activity Probability	proportion	voluntary	Proportion of individuals in a population that are active (i.e., awake, not sleeping) at time of observation.

<u>Trait name</u>	<u>Trait unit</u>	<u>Motivation</u>	<u>Trait definition</u>
Population Voluntary Movement Probability	proportion	voluntary	Proportion of individuals in a population that are physically moving through space at time of observation.
Rattle Rate	event / ($R_n * s$)	negative	Rate of a rattlesnake's rattle.
Strike Acceleration	m / ($C_n * s^2$)	positive	Acceleration of a consumer's head during a strike at a resource.
Strike Completion Rate	event / ($C_n * s$)	positive	Rate a consumer completes a strike.
Strike Distance	m / C_n	positive	Distance between a consumer and resource when consumer initiates a strike per consumer.
Strike Velocity	m / ($C_n * s$)	positive	Velocity of a consumers strike per consumer (averaged over strike distance).
Subjugation-Consumption Body Contraction Rate	event / ($C_n * s$)	positive	Rate of body contractions for locomotion of a consumer during subjugation and consumption of resources.
Surface Area-Specific Foraging Gill Filtration Rate	$m^3 / (m^2 * s)$	positive	Area-specific volumetric flow rate of water across the surface-area of a gill of a filter feeding organism (flow rate measured directly, or by the clearance method where flow rate is estimated by the volume of water cleared of material per time).
Voluntary Activity Probability	proportion / C_n	voluntary	Probability that an organism is active at time of observation.
Voluntary Body Contraction Rate	event / ($C_n * s$)	voluntary	Rate of body contractions for locomotion in an organism with no apparent stimulus.
Voluntary Body Velocity	m / ($C_n * s$)	voluntary	Velocity of the whole organism during voluntary locomotion.
Voluntary Eye Saccade Angular Velocity	rad / ($C_n * s$)	voluntary	Angular velocity of fast eye movements, or saccade velocity, of an organism during voluntary activity.
Voluntary Movement Rate	event / ($C_n * s$)	voluntary	Rate organism physically moves through space.
Voluntary Stroke Rate	event / ($C_n * s$)	voluntary	Stroke rate of an organism during voluntary locomotion.
Voluntary Tail Beat Rate	event / ($C_n * s$)	voluntary	Rate of tail-beat cycles of an organism during voluntary locomotion.

<u>Trait name</u>	<u>Trait unit</u>	<u>Motivation</u>	<u>Trait definition</u>
Voluntary Tongue Flick Cycle Number	event / C_n	voluntary	Number of up-and-down motions or cycles of the tongue per flick in an organism with no obvious stimulus.
Voluntary Tongue Flick Cycle Rate	event / ($C_n * s$)	voluntary	Rate of up-and-down motions or cycles of the tongue per flick in an organism with no obvious stimulus.
Voluntary Tongue Flick Non-Cycle Rate	event / ($C_n * s$)	voluntary	Rate tongue is outside mouth and not moving in an organism with no obvious stimulus.
Voluntary Wing Beat Rate	event / ($C_n * s$)	voluntary	Rate of wing beating in a flying organism with no apparent stimulus.

Population

Chlorophyll-a-Specific Carbon Production Rate	kg / ($kg * s$)	autonomic	Production rate of carbon by a primary producer, measured as mass of carbon produced per mass of chlorophyll-a.
Fecundity	1 / ($C_n * s$)	autonomic	Number of offspring produced by a female per time.
Mortality Rate	1 / ($C_n * s$)	autonomic	Number of deaths scaled by population size per time.
Population Density	C_n / A	autonomic	Number of individuals in the population per arena size.
Population Growth Rate	1 / ($C_n * s$)	autonomic	Intrinsic rate of population growth measured as individuals per individuals per time.
Radial Growth Rate	m / ($C_n * s$)	autonomic	Rate of increase in size (length, mass, or volume) of a population over time.

Interaction

Attack Density Rate	event / ($(R_n * C_n * s) / A$)	positive	Rate of the completion of one attack to the start of the next attack per consumer standardized by arena size.
Attack Rate	event / ($C_n * s$)	positive	Rate of the completion of one attack to the start of the next attack per consumer.

<u>Trait name</u>	<u>Trait unit</u>	<u>Motivation</u>	<u>Trait definition</u>
Consumption Probability	proportion / C_n	positive	Probability that an active consumer will consume food offered to it.
Consumption Rate	event / ($R_n * C_n * s$)	positive	Rate of resources consumed per consumer.
Filtration Rate	$m^3 / (C_n * s)$	positive	Volumetric flow rate of water through a filter feeding consumer (flow rate measured directly, or by the clearance method as the volume of water cleared of material per time).
Foraging Behavior Probability	proportion / C_n	voluntary	Proportion of foraging time a consumer spends undertaking a particular foraging behavior within a subset of a single foraging bout.
Foraging Rate	event / ($C_n * s$)	voluntary	Length of a single foraging bout of a consumer.
Grazing Rate	$m^2 / (C_n * s)$	voluntary	The area grazed per consumer per time.
Handling Rate	$R_n / (C_n * s)$	positive	Rate at which consumer pursues, subjugates, and ingests resources (differs from attack rate because includes ingestion).
Host-Per-Parasitoid Parasitization Rate	event / ($(R_n * C_n * s) / A$)	positive	Rate of host parasitization per parasitoid standardized by arena size.
Intraspecific Confrontation Probability Density	proportion / (C_n / A)	voluntary	Proportion of time organism spends in intraspecific confrontations out of total observation time standardized by arena size.
Line Encounter Rate	event / s	voluntary	Encounter rate of individuals in a population moving past a fixed line.
Nest Provisioning Rate	$R_n / (C_n * s)$	positive	Rate resources bought back to the nest for consumption, by itself or its young, per consumer.
Point Encounter Density Rate	event / ($(C_n * s) / A$)	voluntary	Encounter rate of individuals in a population with an arbitrary point or line per consumer standardized by arena size.
Point Encounter Number Rate	event / ($C_n * s$)	voluntary	Encounter rate per consumer of individuals in a population with a fixed point or line.
Population Catchability	$C_n / (R_n * s)$	voluntary	Rate of resources caught in baited fishing traps per number of traps set.

<u>Trait name</u>	<u>Trait unit</u>	<u>Motivation</u>	<u>Trait definition</u>
Population Foraging Probability	proportion	voluntary	Proportion of consumer population that are foraging at time of observation.
Refuge Distance	m / R_n	negative	Distance to refuge for resource when initially spotted by a consumer (e.g., bush, rock, clump of grass).
Resource Habitat Encounter Density Rate	$\text{event} / ((C_n * s) / A)$	voluntary	Rate consumer encounters its resource habitat within the larger landscape per consumer standardized by arena size.
Resource Reaction Distance	m / R_n	negative	Distance from resource to consumer when resource apparently first reacts to approaching consumer (i.e., stops and/or moves away).
Resource Size Capture Intent Acceptance Probability	proportion / C_n	positive	Proportion of times that a 6 mg resource item was accepted when presented to a consumer after a captured 32 mg resource was taken.
Sediment Mass Processing Rate	$\text{kg} / (C_n * s)$	positive	Rate sediment mass is ingested and processed for food by a deposit feeder (most of what is ingested in non-digestible inorganic sediment).
Subjugation-through-Consumption Rate	$\text{event} / (C_n * s)$	positive	Rate resources are subdued and consumed per consumer.

Table S2. Q_{10} values associated with different 10°C ranges (10). Q_{10} 's are calculated over four temperature ranges (0-10, 0-20, 20-30, and 30-40°C) and then averaged across the full 0-40°C range for four activation energies, representing small (0.2 eV), median (0.55 eV), mean (0.65 eV), and large (1.2 eV) activation energies.

Temp. Range (°C)	0.2 eV	0.55 eV	0.65 eV	1.2 eV
0-10	1.34	2.28	2.65	6.05
10-20	1.32	2.15	2.48	5.35
20-30	1.29	2.05	2.33	4.79
30-40	1.27	1.95	2.21	4.33
0-40	1.31	2.11	2.42	5.13

Table S3. Summary of intraspecific thermal responses used in our analysis. Data are listed in alphabetical order by traits and then taxa. Trait names correspond to those in Table S1. Taxa names represent the lowest level of taxonomy given in the original source (typically species), followed by life stage and sex when present (in square parenthesis), and trophic designation (P, producer; H, herbivore; O, omnivore; C, carnivore; D, detritivore; S, non-feeding organisms, such as eggs, pupae, etc). For interaction traits, consumer and resource are separated by an arrow (consumer is on the left). Artificial “taxa”, such as light as a resource for photosynthesis or pressure waves as a consumer for escape body velocity, are appropriately designated and are not assigned a trophic group. **Cit** is citation code (Table S6); **H** is habitat (M, marine; F, freshwater; T, terrestrial); **LF** is whether the setting was laboratory (L) or field (F); **Temp** is minimum and maximum temperatures over which the response was measured; **P** is the number of distinct temperature points. E_R and E_F are estimated activation energies of the rise and fall components of the temperature responses, calculated from fits to the Boltzmann-Arrhenius model. Q_{10R} and Q_{10F} are estimated Q_{10} values of the rise and fall components of the temperature responses. NS are non-significant fits; T_{opt} are estimates of optimal temperatures. See Materials and Methods (main text) and SI for details on how these values were calculated. Blank cells in the Trait/Consumer-Resource columns signify that the trait and taxa are the same as the last non-blank cell above in the same column. Blank cells in all other columns indicate that the quantity could not be calculated for that response or was not measurable.

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
48-hr Hatching Probability (individual)										
Acartia sinjiensis [adult] O → microalgae P	125	M	L	10–38	8	0.66	2.42			34
Ammonia Excretion Rate (internal)										
Dreissena polymorpha [adult] O	2	F	L	20–32	4	1.1	4.17			
Attack Density Rate (species interaction)										
Perca flavescens [juvenile] O → Coregonus clupeaformis [juvenile] C	190	F	L	5–18	4					10
	190	F	L	5–18	4					15
Perca fluviatilis [adult] O → Chaoborus obscuripes [juvenile] C	140	F	L	12–21	4			NS	NS	
Rutilus rutilus [adult] O → Chaoborus obscuripes [juvenile] C	140	F	L	12–21	4			NS	NS	
Attack Rate (species interaction)										
Cicindela hybrida [adult] C → cursorial insects	42	T	F	23–40.3	8	0.66	2.33			34.7
Avoidance Body Velocity (individual)										
gravity → Aphidius ervi [adult] C	60	T	L	12–36	7	0.9	3.37			28
	60	T	L	12–36	7	0.92	3.47			28
light → Homarus americanus [adult] C	120	M	L	10–28	5	NS	NS			25
	120	M	L	2–15	4					5
	120	M	L	2–20	5					10

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
	120	M	L	2–25	6					12.5
	120	M	L	2–25	6	NS	NS			20
	120	M	L	5–25	5					20
Bite Rate (individual)										
Cellana ornata [adult] H → microalgae P	21	M	F	7.3–17.3	12	0.52	2.08			
Hyles lineata [juvenile] H → desert plants P	27	T	F	13.7–34	5	0.43	1.77			
Littorina littorea [adult] H → epiphytic micro-organisms O	133	M	L	5–25	5	0.4	1.76			
Manduca sexta [juvenile] H → Datura inoxia P	27	T	F	13.1–31.8	7	0.51	1.96			
Chlorophyll-a-Specific Carbon Production Rate (population)										
Periphyton P	93	F	L	3–21	7	0.49	1.99			
Consumption Probability (species interaction)										
Pituophis catenifer affinis [adult] C → Mus musculus [adult] O	65	T	L	18–33	4	NS	NS			
Consumption Probability (species interaction)										
Uta stansburiana [adult] C → Acheta sp. A [adult] O	178	T	L	20–36	6	NS	NS			32
	178	T	L	20–36	6	NS	NS			32
Consumption Rate (species interaction)										
Acroneuria californica [juvenile] O → Hydropsyche spp. [juvenile] O	75	F	L	14–28	8	0.33	1.57			26
	75	F	L	16–28	7	0.42	1.74			
	75	F	L	6–20	7	0.62	2.42			18
Acroneuria californica [juvenile] O → Simulium spp. [juvenile] O	75	F	L	10–24	8			0.12	0.85	12
	75	F	L	10–24	8	0.15	1.22			
	75	F	L	18–30	6	NS	NS	NS	NS	
Agonum dorsale [adult] C → Sitobion avenae [juvenile] H	162	T	L	12.3–23.6	4	1.15	4.79			
Bembidion lampros [adult] C → Rhopalosiphum padi [adult] H	31	T	L	10–25	4	0.72	2.7			
Bembidion lampros [adult] C → Rhopalosiphum padi [juvenile] H	31	T	L	10–25	4					25
Bembidion lampros [adult] C → Sitobion avenae [juvenile] H	162	T	L	12.3–23.6	4					20.6
Bembidion obtusum [adult] C → Sitobion avenae [juvenile] H	162	T	L	12.3–23.6	4	NS	NS			
Carcinops pumilio [adult] C → Musca domestica [juvenile] D	59	T	L	15–33	4	0.8	2.87			
Carcinus maenas [adult] O → Mytilus edulis [adult] O	179	M	L	3.8–17.7	4	0.77	3.05			
	179	M	L	6.9–18.4	6	NS	NS			16.3
Celithemis fasciata [juvenile] C → Chironomus tentans [juvenile] O	66	F	L	10–25	4					15
	66	F	L	10–25	4					20

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
	66	F	L	10–25	4	0.85	3.2			
	66	F	L	10–25	4	NS	NS			
	66	F	L	10–25	4	NS	NS			
Chaoborus americanus [juvenile] C → Diaptomus kenai [adult] O	50	F	L	5–20	4	0.68	2.63			
	50	F	L	5–20	4	NS	NS			
Chaoborus americanus [juvenile] C → Diaptomus tyrelli [adult] O	50	F	L	5–20	4	NS	NS			
Chaoborus trivittatus [juvenile] C → Diaptomus kenai [adult] O	50	F	L	5–20	4					15
	50	F	L	5–20	4	NS	NS			
	50	F	L	5–20	4	NS	NS			
Chaoborus trivittatus [juvenile] C → Diaptomus tyrelli [adult] O	50	F	L	5–20	4	0.69	2.66			
Cicindela hybrida [adult] C → cursorial insect	42	T	F	19.9–39.9	5					29.9
Dreissena polymorpha [adult] O → Chlorella spp. P	2	F	L	20–32	4					24
Gymnocephalus cernuus [adult] C → Chaoborus obscuripes [juvenile] C	18	F	L	4–20	5	0.16	1.25			
	18	F	L	4–20	5	0.4	1.76			
	18	F	L	4–20	5	NS	NS			
	18	F	L	4–20	5	NS	NS			
Harpalus rufipes [adult] C → Sitobion avenae [juvenile] H	162	T	L	12.3–23.6	4	0.89	3.38			
Ischnura elegans elegans [juvenile] C → Daphnia magna [adult] O	171	F	L	5–27.5	6			NS	NS	12
	171	F	L	5–27.5	6	0.59	2.33			16
	171	F	L	5–27.5	6	NS	NS			16
	171	F	L	5–27.5	6	NS	NS			16
	171	F	L	5–27.5	6	NS	NS			16
	171	F	L	5–27.5	6	NS	NS			16
	171	F	L	5–27.5	6	NS	NS			16
	171	F	L	5–27.5	7	0.97	4			16
	171	F	L	5–27.5	8	NS	NS			
Macrocheles muscaedomesticae [adult] C → Musca domestica [juvenile] D	59	T	L	15–33	4	0.85	3.06			
Naucoris congrex [adult] C → Culicidae spp. [juvenile] O	118	F	L	5–25	4					20
	118	F	L	5–25	4	0.52	2.07			
	118	F	L	5–25	4	0.53	2.1			
	118	F	L	5–25	4	0.57	2.23			
	118	F	L	5–25	4	0.58	2.24			

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
	118	F	L	5–25	4	0.6	NS			
	118	F	L	5–25	4	NS	NS			
Notonecta glauca [adult] C → Asellus aquaticus [adult] O	34	F	L	5–20	4					10
	34	F	L	5–25	5					15
Notonecta glauca [adult] C → Culex pipiens [juvenile] O	34	F	L	5–25	5	0.76	2.86			
	34	F	L	5–25	5	1.1	4.76			20
Notonecta hoffmani [adult] C → Culex pipiens [juvenile] O	129	F	L	10–25	4	1.1	4.54			
Nucella lapillus [adult] C → Mytilus edulis [adult] O	99	M	L	3–25	8	0.91	3.65			
Orius insidiosus [adult] C → Panonychus ulmi [adult] H	117	T	L	18.3–35	4					29.4
	117	T	L	18.3–35	4					29.4
	117	T	L	18.3–35	4	0.72	2.55			
	117	T	L	18.3–35	4	NS	NS			
	117	T	L	18.3–35	4	NS	NS			
Parus major [adult] O → Zygliella x-notata [adult] C	7	T	L	2.9–12.7	6	NS	NS			
Perca flavescens [juvenile] O → Coregonus clupeaformis [juvenile] C	190	F	L	5–18	4					15
	190	F	L	5–18	4					18
Perca fluviatilis [adult] O → Chaoborus obscuripes [juvenile] C	18	F	L	4–20	5					16
	18	F	L	4–20	5	0.8	3.11			
	18	F	L	4–20	5	0.99	4.21			16
	18	F	L	4–20	5	NS	NS			16
	18	F	L	4–20	5	NS	NS			16
	18	F	L	4–20	5	NS	NS			16
	140	F	L	12–21	4	0.46	1.88			
	140	F	L	12–21	4	0.53	NS			
	140	F	L	12–21	4	NS	NS			
	140	F	L	12–21	4	NS	NS			
	140	F	L	12–21	4	NS	NS			
	140	F	L	12–21	4	NS	NS			
Phytoseiulus persimilis [adult] C → Tetranychus urticae [adult] H	48	T	L	15–30	4					25
	48	T	L	15–30	4	0.46	1.83			
	48	T	L	15–30	4	0.63	2.31			
	48	T	L	15–30	4	0.65	2.37			

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
	48	T	L	15–30	4	0.74	2.67			
	48	T	L	15–30	4	0.85	3.07			
Polinices duplicatus [adult] C → Mya arenaria [adult] O	45	M	F	9.5–23	4	1.14	5.26			22.8
Porotermes adamsoni [adult] D → Eucalyptus regnans [adult] P	105	T	L	11.5–24	4	0.74	2.75			
Porotermes adamsoni [adult] D → Eucalyptus viminalis [adult] P	105	T	L	11.5–24	4	0.65	2.42			
Porotermes adamsoni [adult] D → Pinus radiata [adult] P	105	T	L	9–26	5	0.77	2.86			
Ranatra dispar [adult] C → Anisops deanei [adult] C	10	F	L	15–30	4	0.51	1.97			
	10	F	L	15–30	4	0.56	NS			
	10	F	L	15–30	4	NS	NS			
	10	F	L	15–30	4	NS	NS			
Rutilus rutilus [adult] O → Chaoborus obscuripes [juvenile] C	140	F	L	12–21	4	0.77	2.89			
	140	F	L	12–21	4	0.86	3.28			
	140	F	L	12–21	4	0.86	3.27			
	140	F	L	12–21	4	0.92	3.55			
	140	F	L	12–21	4	0.93	3.62			
	140	F	L	12–21	4	NS	NS			
Salvelinus malma [juvenile] C → dead Euphausia superba [adult]	93	F	L	3–21	7	NS	NS	2.4	0.04	12
Stethorus punctum [adult] O → Panonychus ulmi [adult] H	82	T	L	21–32.5	5	0.53	1.99			
	82	T	L	21–32.5	5	0.58	2.13			
	82	T	L	21–32.5	5	0.61	2.21			31
	82	T	L	21–32.5	5	0.93	3.36			31
	82	T	L	21–32.5	5	NS	NS			31
Tachyporus hypnorum [adult] C → Sitobion avenae [juvenile] H	162	T	L	12.3–23.6	4					20.6
Thais haemastoma [adult] C → Crassostrea virginica [juvenile] O	58	M	L	10–30	6	NS	NS			
Urosalpinx cinerea [adult] C → Crassostrea virginica [juvenile] O	70	M	L	8.3–30	5	1.18	5.1			25
Urosalpinx cinerea [adult] C → Mytilus edulis [juvenile] O	70	M	L	10–30	5	NS	NS			25
Critical Holding Velocity (individual)										
electric shock → Salmo salar [juvenile] C	63	F	L	3.1–14.6	7	0.31	1.6			9.6
	63	F	L	3.1–14.6	7	0.56	2.29			9.6
Critical Travel Velocity (individual)										
electric shock → Barbus barbus [adult] O	135	F	L	7–25	4					19
	135	F	L	7–25	4	NS	NS			

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
electric shock → <i>Cyprinella spiloptera</i> [juvenile] O	78	F	L	15–35	5	0.53	2.01			30
electric shock → <i>Exodon paradoxus</i> [adult] C	15	F	L	20–35	4					30
electric shock → <i>Leporinus fasciatus</i> [adult] O	15	F	L	25–37	4					35
electric shock → <i>Micropterus salmoides</i> [juvenile] C	78	F	L	15–35	5	0.24	1.37			30
electric shock → <i>Oncorhynchus nerka</i> [juvenile] C	22	F	L	5–27.5	7			NS	NS	20
electric shock → <i>Puntius schwanenfeldii</i> [adult] O	135	F	L	17–30	4					25
	135	F	L	20–33	4					30
light → <i>Micropterus dolomieu</i> [juvenile] C	100	F	L	10–30	5	0.31	1.52			
	100	F	L	10–30	5	0.43	1.77			
	100	F	L	10–30	5	0.56	2.16			25
	100	F	L	5–20	4	0.53	2.13			
	100	F	L	5–25	5	0.53	2.12			20
retaining screen → <i>Ictalurus punctatus</i> [juvenile] O	78	F	L	15–35	5	NS	NS			30
Critical Upright Time (individual)										
<i>Homo sapiens</i> [adult] O → <i>Natrix maura</i> [adult] C	68	T	L	10–35	6	0.48	1.91			30
Development Rate (individual)										
<i>Aphis gossypii</i> H → <i>Cucumis sativus</i> P	191	T	L	10–30	5	0.6	2.26			
<i>Bactrocera correcta</i> [egg] S	112	T	L	18–36	5	0.48	1.88			33
<i>Bactrocera correcta</i> [pupae] S	112	T	L	18–36	5	0.45	1.78			
	112	T	L	18–36	5	0.44	1.78			33
<i>Cherax quadricarinatus</i> [juvenile] O → crayfish ration	121	F	L	16–32	9	0.8	2.89			28
<i>Chinemys reevesii</i> [egg] S	43	F	L	24–34	6	0.29	1.44			
<i>Cydia pomonella</i> [egg] S	1	T	L	14–33	6	0.76	2.74			30
<i>Cydia pomonella</i> [juvenile] H → artificial diet	1	T	L	14–33	6	0.76	2.8			27
<i>Cydia pomonella</i> [pupae] S	1	T	L	14–33	6	0.77	2.77			30
<i>Euplectrus ronnai</i> [juvenile] C → <i>Pseudaletia sequax</i> [juvenile] H	189	T	L	15–29	5	0.79	2.88			
<i>Euplectrus ronnai</i> [pupae] S → <i>Pseudaletia sequax</i> [juvenile] H	189	T	L	15–29	5	0.91	3.34			
<i>Glyptapanteles muesebecki</i> [juvenile] C → <i>Pseudaletia sequax</i> [juvenile] H	54	T	L	14–29	6	0.72	2.64			26
<i>Glyptapanteles muesebecki</i> [pupae] S → <i>Pseudaletia sequax</i> [juvenile] H	54	T	L	14–29	6	0.82	2.98			
<i>Macrocentrus iridescens</i> [juvenile] C → <i>Choristoneura rosaceana</i> [juvenile] H	97	T	L	13.9–31	6	0.74	2.72			25.8
<i>Planococcus citri</i> [egg] S → <i>Solenostemon scutellarioides</i> P	61	T	L	15–32	8	0.72	2.59	NS	NS	25
	61	T	L	18–32	7	1.2	4.96	NS	NS	25

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
Planococcus citri [juvenile] H → Solenostemon scutellarioides P	61	T	L	15–32	8	0.93	3.51	0.49	0.53	25
	61	T	L	18–32	7	0.46	1.83			
Planococcus citri [pupae] H → Solenostemon scutellarioides P	61	T	L	18–32	7	0.84	3.02	NS	NS	25
	61	T	L	18–32	7	1.03	3.86	NS	NS	25
Procambarus clarkii [adult] O → uncooked mixed vegetables	30	F	L	15–30	6	0.65	2.38			
Procambarus clarkii [juvenile] O → uncooked mixed vegetables	30	F	L	15–30	6	0.54	2.05			
Sitona discoideus [egg] S	5	T	L	8.5–30	8	0.67	2.48			28
Telenomus chrysopae [juvenile] C → Chrysoperla rufilabris [egg] S	155	T	L	15.6–26.7	5	0.85	3.14			
	155	T	L	15.6–26.7	5	0.91	3.36			
Telenomus isis [juvenile] C → Busseola fusca [egg] S	25	T	L	18–32	6	0.61	2.21			30
	25	T	L	18–32	6	0.64	2.31			30
Telenomus isis [juvenile] C → Sesamia calamistis [egg] S	25	T	L	18–32	6	0.57	2.13			30
	25	T	L	18–32	6	0.59	2.18			30
Telenomus isis [juvenile] C → Sesamia nonagrioides [egg] S	25	T	L	18–32	6	0.59	2.16			30
	25	T	L	18–32	6	0.64	2.32			30
Telenomus lobatus [juvenile] C → Chrysoperla rufilabris [egg] S	155	T	L	15.6–26.7	5	0.89	3.31			
Tetraneura nigri abdominalis [juvenile] H → Oryza sativa P	98	T	L	10–35	6	0.64	2.39			30
Theocolax elegans [juvenile] C → Sitophilus zea-mais [egg] S	84	T	L	20–35	6	0.79	2.79			32
	84	T	L	20–35	6	0.81	2.86			32
Trichogramma brunii [juvenile] C → Corcyra cephalonica [egg] S	90	T	L	15–32	6	0.6	2.23			30
	90	T	L	20–32	5	NS	NS			30
Trichogramma sp. nr. lutea [juvenile] C → Corcyra cephalonica [egg] S	90	T	L	15–32	6	0.47	1.84			
	90	T	L	15–32	6	0.56	2.09			30
	90	T	L	15–32	6	0.57	2.13			30
	90	T	L	15–32	6	0.65	2.38			28
	90	T	L	15–35	7	0.41	1.71			
	90	T	L	15–35	7	0.43	1.75			
	90	T	L	15–32	6	0.59	2.17			30
	90	T	L	15–32	6	0.64	2.34			30
Trichogramma sp. nr. mwanzai [juvenile] C → Corcyra cephalonica [egg] S	90	T	L	15–32	6	0.59	2.17			30
	90	T	L	15–32	6	0.64	2.34			30
	90	T	L	15–35	7	0.42	1.73			
90	T	L	15–35	7	0.48	1.86				

Digestion Rate (internal)

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
Perca fluviatilis [adult] O → Gammarus pulex [adult] O	139	F	L	4–21.7	7	1.07	4.64			
Ptychocheilus oregonensis [adult] C → Oncorhynchus mykiss [juvenile] C	163	F	L	6–24	5	0.88	3.4			
	163	F	L	6–24	5	1.05	4.33			
	163	F	L	6–24	5	1.26	5.8			
Salmo trutta [adult] C → dead Hydropsyche spp. [juvenile]	47	F	L	5.2–15	4	0.77	3.06			
	47	F	L	5.2–15	4	0.77	3.06			
	47	F	L	5.2–15	4	0.77	3.05			
	47	F	L	5.2–15	4	0.78	3.07			
Salmo trutta [adult] C → dead invertebrate	47	F	L	5.2–15	5	0.77	3.06			
	47	F	L	5.2–15	5	0.77	3.07			
	47	F	L	5.2–15	5	0.78	3.07			
	47	F	L	5.2–15	5	0.78	3.1			
Salmo trutta [adult] C → dead Protonemura meyeri [juvenile]	47	F	L	5.2–15	5	0.73	2.88			
	47	F	L	5.2–15	5	0.74	2.91			
	47	F	L	5.2–15	5	0.74	2.9			
	47	F	L	5.2–15	5	0.74	2.91			
Salmo trutta [adult] C → Tenebrio molitor [juvenile] H	47	F	L	5.2–15	5	0.77	3.06			
	47	F	L	5.2–15	5	0.77	3.06			
	47	F	L	5.2–15	5	0.77	3.06			
	47	F	L	5.2–15	5	0.78	3.07			
Thamnophis elegans vagrans [adult] C → Mus musculus [adult] O	167	T	L	9.8–35	10	1.06	4.19	NS	NS	24.8
Endurance Time (individual)										
Homo sapiens [adult] O → Sceloporus undulatus [adult] O	4	T	L	11.4–40.6	9	0.33	1.54			33
Escape Angle of Body Turning (individual)										
prodding with a probe → Myoxocephalus scorpius [adult] C	170	M	L	0.8–20	4					5
	170	M	L	0.8–20	4			NS	NS	
	170	M	L	0.8–20	4			NS	NS	
prodding with a probe → Taurulus bubalis [adult] C	170	M	L	0.8–20	4					5
Escape Angular Rate of Body Turning (individual)										
pressure waves → Carassius auratus [adult] O	87	F	L	10–40	5					40
	87	F	L	5–30	5					25
pressure waves → Fundulus heteroclitus [adult] O	87	F	L	5–30	5					25

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
prodding with a probe → <i>Myoxocephalus scorpius</i> [adult] C	170	M	L	0.8–20	4	NS	NS			
prodding with a probe → <i>Taurulus bubalis</i> [adult] C	170	M	L	0.8–20	4					5
	170	M	L	0.8–20	4					5
Escape Body Acceleration (individual)										
electric shock → <i>Danio rerio</i> [juvenile] O	56	F	L	21.1–30	7	NS	NS			
<i>Homo sapiens</i> [adult] O → <i>Phelsuma dubia</i> [adult] O	19	T	L	15–35	5	0.39	1.69			30
prodding with a probe → <i>Myoxocephalus scorpius</i> [adult] C	170	M	L	0.8–20	4					15
	170	M	L	0.8–20	4					15
prodding with a probe → <i>Taurulus bubalis</i> [adult] C	170	M	L	0.8–20	4					15
	170	M	L	0.8–20	4	NS	NS			
Escape Body Deceleration (individual)										
<i>Homo sapiens</i> [adult] O → <i>Phelsuma dubia</i> [adult] O	19	T	L	15–35	5	0.36	1.61			30
Escape Body Power Production (individual)										
electric shock → <i>Rana pipiens</i> [adult] C	77	T	L	14–30	4					25
<i>Homo sapiens</i> [adult] O → <i>Phelsuma dubia</i> [adult] O	19	T	L	15–35	5	0.5	1.91			
Escape Body Response Rate (individual)										
pressure waves → <i>Calanus finmarchicus</i> [adult] O	106	M	L	3.7–15.4	11	0.33	1.62			14.5
Escape Body Velocity (individual)										
electric shock → <i>Danio rerio</i> [juvenile] O	56	F	L	21–30	4	0.18	1.26			
	56	F	L	21–30	4	0.25	1.38			
electric shock → <i>Necturus maculosus</i> [adult] C	126	F	L	5–25	5					15
electric shock → <i>Xenopus laevis</i> [adult] C	126	F	L	10–30	7	0.25	1.4			27
<i>Homo sapiens</i> [adult] O → <i>Acanthodactylus erythrurus</i> [adult] C	14	T	L	25.4–40.4	6	0.21	1.3			
<i>Homo sapiens</i> [adult] O → <i>Agama savignyi</i> [adult] C	76	T	L	18–42	7	0.5	1.92			34
<i>Homo sapiens</i> [adult] O → <i>Bufo boreas</i> [adult] C	143	T	L	3.8–27.9	5	0.61	2.33			
<i>Homo sapiens</i> [adult] O → <i>Bufo woodhousii woodhousii</i> [adult] C	113	T	L	15–30	4					30
	113	T	L	15–30	4					30
<i>Homo sapiens</i> [adult] O → <i>Coleonyx brevis</i> [adult] C	81	T	L	20–40	6	0.24	1.35			37.5
<i>Homo sapiens</i> [adult] O → <i>Coleonyx variegatus</i> [adult] C	81	T	L	15–40	7	0.28	1.43			
<i>Homo sapiens</i> [adult] O → <i>Hemidactylus frenatus</i> [adult] C	81	T	L	15–39.5	7	0.46	1.83			34
<i>Homo sapiens</i> [adult] O → <i>Hemidactylus turcicus</i> [adult] C	81	T	L	20–40	6	0.25	1.37			38.8
<i>Homo sapiens</i> [adult] O → <i>Iberolacerta monticola</i> [adult] C	14	T	L	26.6–40	6	NS	NS			34.9

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
Homo sapiens [adult] O → Lacerta agilis [adult] C	14	T	L	25.9–39.5	6	0.38	1.61			37.7
Homo sapiens [adult] O → Lacerta schreiberi [adult] C	14	T	L	24.8–40.4	6	0.3	1.47			35.9
Homo sapiens [adult] O → Lepidodactylus lugubris [adult] O	81	T	L	15–36.5	6	0.33	1.53			
Homo sapiens [adult] O → Natrrix maura [adult] C	68	T	L	4.2–34.1	7	0.63	2.35			
	68	T	L	5.7–35.6	7	0.62	2.28			
Homo sapiens [adult] O → Phelsuma dubia [adult] O	19	T	L	15–35	5	0.33	1.53			
Homo sapiens [adult] O → Podarcis bocagei [adult] O	14	T	L	26.2–39.7	6	0.42	1.7			35
Homo sapiens [adult] O → Podarcis hispanica [adult] O	14	T	L	26.1–40.2	6	0.5	1.88			37.5
	14	T	L	26–39.5	6	0.47	1.8			37.3
Homo sapiens [adult] O → Podarcis lilfordi [adult] O	14	T	L	26.1–39.9	6	0.32	1.48			37.3
Homo sapiens [adult] O → Podarcis muralis [adult] O	14	T	L	25.1–39.6	6	0.3	1.45			35.2
Homo sapiens [adult] O → Podarcis tiliguerta [adult] O	175	T	L	20–37.5	7	0.3	1.46			
	175	T	L	20–37.5	7	0.31	1.49			
Homo sapiens [adult] O → Psammodromus algirus [adult] C	14	T	L	24.5–39.8	6	0.36	1.57			34.7
Homo sapiens [adult] O → Psammodromus hispanicus [adult] C	14	T	L	25.8–39.5	6	0.48	1.83			34.5
Homo sapiens [adult] O → Rana pipiens [adult] C	143	T	L	4.4–29.1	5	0.33	1.57			
Homo sapiens [adult] O → Sceloporus occidentalis [adult] O	116	T	L	9.9–39.5	9	0.63	2.27			35.3
Homo sapiens [adult] O → Sceloporus undulatus [adult] O	4	T	L	11.3–40.5	9	0.44	1.76			36
Homo sapiens [adult] O → Scincella lateralis [adult] C	159	T	L	19.5–38	5	0.29	1.45			33.5
Homo sapiens [adult] O → Thamnophis elegans vagrans [adult] C	167	F	L	5.4–35.6	7	0.36	1.63			30.5
	167	F	L	5.6–35.7	7	0.38	1.68			30.7
	167	T	L	3.9–34	7	0.71	2.61			
	167	T	L	3.9–34.1	7	0.55	2.1			
Homo sapiens [adult] O → Thamnophis sirtalis [adult] C	73	T	L	15.5–30.6	5	0.57	2.12			
	73	T	L	16.2–32.4	11	0.51	1.94			
	73	T	L	16.8–30.6	5	0.6	2.23			26.3
	73	T	L	16.8–31.5	5	0.57	2.11			
	73	T	L	16.9–31.2	5	NS	NS			
	73	T	L	17.1–31.2	5	0.68	2.48			31.2
pressure waves → Barbus barbus [adult] O	135	F	L	7–25	4					25
	135	F	L	7–25	4	0.19	1.31			
pressure waves → Carassius auratus [adult] O	87	F	L	10–40	5					35

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
	87	F	L	10–40	5	NS	NS			35
	87	F	L	5–30	5					15
	87	F	L	5–30	5	0.2	NS			25
pressure waves → <i>Fundulus heteroclitus</i> [adult] O	87	F	L	5–30	5					20
	87	F	L	5–30	5					20
prodding with a probe → <i>Clupea harengus</i> [juvenile] H	12	M	L	4.7–17.5	22	0.2	1.34			
	12	M	L	4.9–17.2	13	0.29	1.52			
prodding with a probe → <i>Cnemidophorus murinus</i> [adult] C	17	T	L	20–43.5	6	NS	NS			40
	17	T	L	20–44	6	0.51	NS			40
	17	T	L	20–44	6	NS	NS			40
prodding with a probe → <i>Dipsosaurus dorsalis</i> [adult] H	17	T	L	15–43.5	7	0.89	3.1			40
	17	T	L	15–44	7	0.85	3			35
	17	T	L	15–44	7	0.96	3.48			35
prodding with a probe → <i>Elgaria multicarinata</i> [adult] C	17	T	L	10–37.5	7	0.44	1.82			30
	17	T	L	10–37.5	7	0.5	1.92			
	17	T	L	10–37.5	7	0.52	2.02			30
prodding with a probe → <i>Myoxocephalus scorpius</i> [adult] C	170	M	L	0.8–20	4					15
	170	M	L	0.8–20	4					15
prodding with a probe → <i>Sceloporus occidentalis</i> [adult] O	17	T	L	10–40	5	0.54	2.03			
	17	T	L	10–40	7	0.38	1.64			35
	17	T	L	10–40	7	0.46	1.84			35
	17	T	L	10–40	7	0.51	1.92			
prodding with a probe → <i>Taurulus bubalis</i> [adult] C	170	M	L	0.8–20	4					15
	170	M	L	0.8–20	4	NS	NS			
prodding with a probe → <i>Uma inornata</i> [adult] C	17	T	L	20–43.5	7	0.24	1.35			40
Escape Body Velocity Probability (individual)										
prodding with a probe → <i>Conolophus pallidus</i> [juvenile] O	32	T	F	15–39.3	5	NS	NS			33.5
	32	T	F	17–39.5	10	0.72	2.53			34.4
	32	T	F	20.4–39.8	4					39.8
prodding with a probe → <i>Uta stansburiana</i> [adult] C	177	T	L	15–38.5	4					33.5
	177	T	L	15–41.9	9	0.39	1.65			37.9
	177	T	L	19.9–39.9	6	1.27	5			38

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
Escape Gait Change Velocity (individual) electric shock → Uromastix aegyptia [adult] H	41	T	L	35.1–44	7			4.28	0.01	
Escape Jump Contact Rate (individual) electric shock → Rana pipiens [adult] C	77	T	L	14–30	4	0.29	1.47			
Escape Jump Distance (individual) electric shock → Rana pipiens [adult] C	77	T	L	14–30	4					18
	77	T	L	14–30	4					25
	77	T	L	14–30	4					25
	77	T	L	14–30	4	0.35	1.61			
	77	T	L	14–30	4	NS	NS			
	77	T	L	14–30	4	NS	NS			
Homo sapiens [adult] O → Bufo woodhousii woodhousii [adult] C	113	T	L	15–30	4					30
	113	T	L	15–30	4	0.24	1.38			
Homo sapiens [adult] O → Rana clamitans [adult] C	80	T	L	5–30	7	0.25	1.43			20
prodding with a probe → Acris crepitans [adult] C	96	T	L	5–30	6	0.2	1.31			
prodding with a probe → Bufo americanus [adult] C	146	T	L	5–25	5	0.13	1.2			
	146	T	L	5–25	5	NS	NS			
prodding with a probe → Bufo woodhousii woodhousii [adult] C	113	T	L	15–30	4					25
	113	T	L	15–30	4	0.21	1.31			
prodding with a probe → Hyla femoralis [adult] C	96	T	L	5–30	6	0.2	1.32			
prodding with a probe → Limnodynastes tasmaniensis [adult] C	184	T	L	4.4–33.2	6	NS	NS			29.5
	184	T	L	4.4–35.7	7	0.33	1.58			29.5
prodding with a probe → Pseudacris triseriata [adult] C	96	T	L	5–30	6	0.13	1.29			
	96	T	L	5–30	6	0.15	1.23			
prodding with a probe → Rana clamitans [adult] C	96	T	L	5–30	6	0.33	1.59			25
prodding with a probe → Rana pipiens [adult] C	146	T	L	5–25	5	0.38	1.71			
	146	T	L	5–25	5	NS	NS			
prodding with a probe → Rana sylvatica [adult] C	96	T	L	5–30	6	0.26	1.43			
	96	T	L	5–30	6	0.29	1.49			
Escape Jump Force (individual) electric shock → Rana pipiens [adult] C	77	T	L	14–30	4					25
Escape Jump Rate (individual)										

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
Homo sapiens [adult] O → Bufo woodhousii woodhousii [adult] C	113	T	L	15–30	4					20
	113	T	L	15–30	4					25
Escape Stroke Length (individual)										
Homo sapiens [adult] O → Phelsuma dubia [adult] O	19	T	L	15–35	5	0.18	1.27			
Homo sapiens [adult] O → Sceloporus occidentalis [adult] O	116	T	L	9.9–39.6	9	0.11	1.16			
Escape Stroke Peak Force (individual)										
pressure waves → Calanus finmarchicus [adult] O	106	M	L	3.8–15.4	11	0.23	1.41			
Escape Stroke Peak Force Rate (individual)										
	106	M	L	4.5–14.6	8	0.44	1.89			
Escape Stroke Rate (individual)										
Homo sapiens [adult] O → Atelopus muisca [adult] C	130	F	F	5–25	5					10
Homo sapiens [adult] O → Atelopus sp. nov. [adult] C	130	F	F	5–25	5					15
Homo sapiens [adult] O → Atelopus varius [adult] C	130	F	F	5–30	6	0.37	1.68			27.5
Homo sapiens [adult] O → Colostethus flotator [adult] C	130	F	F	14.6–30	4					20
Homo sapiens [adult] O → Colostethus subpunctatus [adult] C	130	F	F	5–25	5					15
Homo sapiens [adult] O → Colostethus talamancae [adult] C	130	F	F	10–30	5					25
Homo sapiens [adult] O → Eleutherodactylus bogotensis [adult] C	130	F	F	5–25	5					15
Homo sapiens [adult] O → Eleutherodactylus diastema [adult] C	130	F	F	10–30	5					25
Homo sapiens [adult] O → Hyla ebraccata [adult] C	130	F	F	5–30	6	NS	NS			25
Homo sapiens [adult] O → Hyla labialis [adult] C	130	F	F	5–25	5	NS	NS			
Homo sapiens [adult] O → Hyla microcephala [adult] C	130	F	F	9.7–30	5	NS	NS			25
Homo sapiens [adult] O → Phelsuma dubia [adult] O	19	T	L	15–35	5	0.26	1.41			
Homo sapiens [adult] O → Sceloporus occidentalis [adult] O	116	T	L	10.3–39.7	9	0.48	1.87			35.4
pressure waves → Calanus finmarchicus [adult] O	106	M	L	3.6–15.3	11	0.23	1.4			
	106	M	L	4.9–14.8	13	0.38	1.75			
Escape Tail Beat Rate (individual)										
prodding with a probe → Clupea harengus [juvenile] H	12	M	L	4.6–17.4	22	0.4	1.78			
	12	M	L	5–17.1	11	0.38	1.72			17
Faecal Excretion Rate (internal)										
Hexagenia limbata [juvenile] H → sediment 'a'	192	F	L	5–25	5	0.31	1.55			
	192	F	L	5–25	5	0.37	1.69			
	192	F	L	5–25	5	0.43	1.84			

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
Fecundity (population)										
Tetraneura nigri abdominalis [adult] H → Oryza sativa P	98	T	L	10–35	6	1.37	6.27			30
Feeding Heart Beat Rate (internal)										
Crassostrea virginica [adult] O	51	M	L	10–21.9	4	0.75	2.84			
	51	M	L	10–21.9	4	0.89	3.6			
	51	M	L	10–22	4					17.9
	51	M	L	10–22	4					17.9
	51	M	L	10–22	4	0.92	3.58			
	51	M	L	10–22	4	1.26	5.73			
	51	M	L	4.8–39.5	6	0.5	1.97			34.7
	51	M	L	4.8–40.1	6	NS	NS			29.8
	51	M	L	4.9–39.7	6	NS	NS			29.5
	51	M	L	4.9–40.1	6	0.56	2.14			29.8
	51	M	L	5.1–39.9	6	NS	NS			30
	51	M	L	5.2–39.9	6	0.24	1.38			35
	51	M	L	5.2–39.9	6	0.6	2.2			
	51	M	L	5.3–40	6	0.79	NS			30
	51	M	L	5.3–40.1	6	0.66	NS			29.9
	51	M	L	9.8–22	4	0.67	2.54			
	51	M	L	9.9–21.9	4	0.88	3.4			
	51	M	L	9.9–22	4	0.85	3.24			
	51	M	L	9.9–22	4	0.92	3.57			
Mytilus edulis [adult] O	185	M	L	10–25	4	0.45	1.85			
	185	M	L	10–25	4	0.45	1.87			
	185	M	L	10–25	4	0.46	1.87			
	185	M	L	5–25	5	0.44	1.84			
	185	M	L	5–25	5	0.71	2.7			
	185	M	L	5–30	6	0.65	2.47			25
Filtration Metabolic Efficiency (internal)										
Cardium lamarcki [adult] O → yeast & sediment	23	M	L	4–20	5			NS	NS	
	23	M	L	4–20	5			NS	NS	
	23	M	L	4–20	5			NS	NS	

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
Cerastoderma edule [adult] O → yeast & sediment	23	M	L	4–28	7			0.32	0.64	
	23	M	L	4–28	7			NS	NS	
	23	M	L	4–28	7	NS	NS			
	23	M	L	4–20	5			0.15	0.81	
	23	M	L	4–20	5			0.25	0.7	
	23	M	L	4–28	7			0.57	0.45	8
	23	M	L	4–28	7			0.76	0.35	
Ostrea edulis [adult] O → Phaeodactylum tricornutum P	23	M	L	4–28	7			1.04	0.24	
	132	M	L	10–30	6			0.71	0.38	
	132	M	L	10–30	6			0.79	0.34	
	132	M	L	10–30	6			NS	NS	
	132	M	L	10–30	6			NS	NS	
	132	M	L	10–30	6			NS	NS	
Filtration Rate (species interaction)										
Cardium lamarcki [adult] O → yeast & sediment	23	M	L	4–20	5	0.71	2.78			
	23	M	L	4–20	5	NS	NS			
	23	M	L	4–20	5	NS	NS			
	23	M	L	4–28	7	0.57	2.23			24
	23	M	L	4–28	7	1.09	4.74			20
Cerastoderma edule [adult] O → yeast & sediment	23	M	L	4–20	5	0.42	1.83			
	23	M	L	4–20	5	0.56	2.23			
	23	M	L	4–20	5	NS	NS			
	23	M	L	4–28	7			0.65	0.42	12
	23	M	L	4–28	7	NS	NS			
Ciona intestinalis [adult] O → Rhodomonas spp. P	141	M	L	4.3–21.6	5	0.59	2.33			18.1
	141	M	L	5.3–20.7	4	0.65	2.52			
	141	M	L	5–22	5	NS	NS			19.5
	141	M	L	6.3–21.5	5	0.79	3.09			18.1
	141	M	L	6.3–21.6	5	0.64	2.46			
Conopeum reticulum [adult] O → Cryptomonas spp. P	123	M	L	6–22	4					18
	123	M	L	6–22	4					18
	123	M	L	6–22	4					18

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
	123	M	L	6–22	4					18
	123	M	L	6–22	4					18
	123	M	L	6–22	4					18
	123	M	L	6–22	4					18
	123	M	L	6–22	4					18
	123	M	L	6–22	4	0.35	1.65			
	123	M	L	6–22	4	0.42	1.81			
	123	M	L	6–22	4	0.44	1.86			
	123	M	L	6–22	4	0.55	2.16			
Daphnia magna [adult] O → Scenedesmus acutus P	119	F	L	10–25	4					20
	119	F	L	5–25	5			0.29	0.67	10
	119	F	L	5–25	5	0.67	2.57			
	119	F	L	5–25	5	0.84	3.31			20
	119	F	L	5–25	5	NS	NS			
Daphnia rosea [adult] O → Chlamydomonas spp. P	91	M	L	5–25.2	9	0.29	1.52	0.16	0.81	14.1
Electra crustulenta [adult] O → Rhodomonas spp. P	111	M	L	6–24	6	0.6	2.32			22
Electra pilosa [adult] O → Cryptomonas spp. P	123	M	L	6–22	4					18
	123	M	L	6–22	4					18
	123	M	L	6–22	4					18
	123	M	L	6–22	4					18
	123	M	L	6–22	4	0.55	2.18			
	123	M	L	6–22	4	0.45	1.89			
	123	M	L	6–22	4	0.26	1.43			
	123	M	L	6–22	4	0.4	1.76			
	123	M	L	6–22	4	0.49	1.99			
Halichondria panicea [adult] O → Rhodomonas spp. P	151	M	L	6.1–14.9	10	1.54	9.19			
	151	M	L	6.1–15	10	0.72	2.84			
	151	M	L	6–14.9	9	1.04	4.45			
	151	M	L	7–13.9	4	NS	NS			
Hiatella arctica [adult] O → Phaeodactylum tricornutum P	3	M	L	1.6–24.9	8	0.97	4.16	3.42	0.01	15.4
	3	M	L	3.3–22.4	4					16.3
Mya arenaria [adult] O → Rhodomonas spp. P	149	M	L	4–22.3	6	0.32	1.58			18

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
	149	M	L	5.6–22	5	0.35	1.63			
Mytilus californianus [adult] O → colloidal graphite	145	M	L	5–20	4	NS	NS			
Mytilus edulis [adult] O → Phaeodactylum tricornutum P	186	M	L	5–25	5			NS	NS	10
Mytilus edulis [adult] O → Rhodomonas spp. P	89	M	L	2.7–15	6	0.41	1.81			
	89	M	L	4.9–14.8	4	0.29	1.51			
	89	M	L	5.6–21.4	7	0.33	1.59			
	94	M	L	4.8–19.6	7	0.3	1.54			
	94	M	L	8.6–19.4	6	0.33	1.6			17.3
Nereis diversicolor [adult] C → phytoplankton P	152	M	L	11–24	6			NS	NS	
Nereis diversicolor [adult] C → Rhodomonas spp. P	152	M	L	5.1–25	6	0.36	1.67			16
	152	M	L	7.9–26.7	4					14.9
	152	M	L	8.1–26.3	11	0.39	1.7			23.4
	152	M	L	8.4–21.6	4					18.9
Ostrea edulis [adult] O → Phaeodactylum tricornutum P	132	M	L	10–30	6			NS	NS	
	132	M	L	5–30	7	1.15	NS			
	132	M	L	5–30	7	1.56	9.25			20
	132	M	L	5–30	7	2.42	31.24	NS	NS	
	132	M	L	5–30	7	NS	NS	NS	NS	
Paraphysomonas imperforata C → Phaeodactylum tricornutum P	26	M	L	14–26	4	NS	NS			
Rutilus penicillus [adult] O → Rhodomonas spp. P	150	M	L	13.7–21.8	6	0.18	1.28			
	150	M	L	6.2–21.6	8	0.31	1.56			
Flee Distance (individual)										
Homo sapiens [adult] O → Holbrookia propinqua [adult] C	35	T	F	26.6–49.3	12	0.6	2.03			
	35	T	F	29.3–49.6	7	NS	NS			
Homo sapiens [adult] O → Sceloporus mucronatus [adult] O	160	T	F	26.1–34.1	8	1.58	7.48			
Homo sapiens [adult] O → Urosaurus bicarinatus [adult] C	160	T	F	35.2–40.6	4			NS	NS	
Food Energy Assimilation Efficiency (internal)										
Acroneuria californica [juvenile] O → Hydropsyche spp. [juvenile] O	75	F	L	10–26	5			0.58	0.45	
	75	F	L	8–20	4			0.58	0.44	
Acroneuria californica [juvenile] O → Simulium spp. [juvenile] O	75	F	L	10–24	7			NS	NS	
	75	F	L	11–23	4			NS	NS	
	75	F	L	13–27	4					17

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
Food Mass Conversion Efficiency (internal)										
Cherax quadricarinatus [juvenile] O → crayfish ration	121	F	L	16–32	9			0.22	0.75	20
Dicentrarchus labrax [juvenile] C → commercial extruded dry pellet	138	M	L	13.4–28.8	6	0.39	1.71			21.9
Paraphysomonas imperforata C → Halomonas marina D	154	M	L	0–20	5	0.23	1.4			15
Foraging Behaviour Probability (species interaction)										
Calidris mauri [adult] C → mudflat invertebrate	131	T	F	25–35	8			NS	NS	
	131	T	F	25–35	8	NS	NS			
Foraging Body Undulation Rate (individual)										
Nereis diversicolor [adult] C → Rhodomonas spp. P	152	M	L	6.1–16.9	7	0.48	1.98			
	152	M	L	7.6–27.8	8	0.41	1.75			
	152	M	L	7.9–26.7	4	0.42	1.77			
Foraging Gill Beat Rate (individual)										
Ciona intestinalis [adult] O → Rhodomonas spp. P	137	M	L	7.4–20.1	4	NS	NS			
Foraging Rate (species interaction)										
Bembidion lampros [adult] C	31	T	L	10–30	4					20
	31	T	L	5–30	5	NS	NS			
	31	T	L	5–30	5	NS	NS			
Cicindela hybrida [adult] C	42	T	F	22.4–43.5	8	0.68	2.35			37.4
Nereis diversicolor [adult] C → Rhodomonas spp. P	152	M	L	5.5–28	11	0.45	1.85			
Pterostichus cupreus [adult] C	31	T	L	5–30	5					15
	31	T	L	5–30	5			NS	NS	
	31	T	L	5–30	5	NS	NS			
	31	T	L	5–30	5	NS	NS			
Foraging Submersion Rate (individual)										
Notonecta glauca [adult] C	34	F	L	5–25	5	1.09	4.65			
	34	F	L	5–25	5	1.45	7.72			
	34	F	L	5–25	5	1.69	10.75			
	34	F	L	5–25	5	1.93	14.91			
Foraging Velocity (individual)										
Acromyrmex versicolor [adult] H	83	T	F	17.2–32.4	8	0.53	2			30.3
Aphaenogaster senilis [adult] H	102	T	F	6–40	14	0.7	2.5			
Cicindela hybrida [adult] C	42	T	F	17.5–42.6	15	0.27	1.42	NS	NS	33

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
Dorymyrmex goetschi [adult] O	172	T	F	18.9–37.4	5	0.2	1.29			
Formica rufa [adult] C	79	T	F	9.1–20.8	5	0.52	2.05			
	79	T	F	9.8–17.5	6	0.6	2.32			
Gymnocephalus cernuus [adult] C	18	F	L	4–20	5	0.18	1.3			
Kinixys spekii [adult] H	67	T	F	21.5–38.5	8			0.97	0.29	
Leptogenys intermedia [adult] C	44	T	L	20–35	4	0.54	1.99			
Leptogenys schwabi [adult] C	44	T	L	20–35	4	0.41	1.7			
Linepithema humile [adult] C	33	T	F	5.9–35.6	10	0.7	2.54			
	33	T	F	8.4–35.3	7	0.4	1.71			
	158	T	F	25.5–33.8	6	0.46	1.8			
Liometopum apiculatum [adult] O	157	T	F	9.7–38.3	10	0.61	2.23			
Messor pergandei [adult] H	83	T	F	21.5–36.3	5	0.72	2.51			34.1
	108	T	F	21.4–43.2	11	0.4	1.71			40
	153	T	F	17–37.4	6	0.44	1.77			34.5
Ocymyrmex barbiger [adult] C	115	T	F	28.6–61.2	12	0.44	1.65			
Perca fluviatilis [adult] O	18	F	L	4–20	5	0.72	2.79			
Pogonomyrmex barbatus [adult] H	128	T	F	25–51.7	10	0.32	1.48			
Pogonomyrmex desertorum [adult] H	128	T	F	27.2–48.4	8	0.42	1.65			45.9
Pogonomyrmex maricopa [adult] H	182	T	F	26.1–45.8	10	0.69	2.32			43.8
Pogonomyrmex occidentalis [adult] H	128	T	F	34.1–50	15	NS	NS			
Pogonomyrmex rugosus [adult] H	182	T	F	24.5–44	11	0.55	1.99			41.1
Rutilus rutilus [adult] O → Chaoborus obscuripes [juvenile] C	140	F	L	12–21	4	NS	NS			
Solenopsis invicta [adult] O	148	T	F	10.5–32.3	10	0.54	2.05			
Tapinoma sessile [adult] O	158	T	F	20.4–36.9	10	0.49	1.87			
Grazing Rate (species interaction)										
Glossosoma spp. [juvenile] H → periphyton P	93	F	L	3–21	7			0.57	0.45	9
Gut Clearance Rate (internal)										
Aurelia aurita [juvenile] C → Clupea harengus [juvenile] H	74	M	L	5–22	4	0.66	2.55			
Centropages hamatus [adult] O → Ditylum brightwellii P	92	M	L	1–15	4	0.92	3.84			
Conopeum reticulum [adult] O → Cryptomonas spp. P	123	M	L	6–22	4	0.36	1.67			
Electra pilosa [adult] O → Cryptomonas spp. P	123	M	L	6–22	4	0.5	2.03			
Gadus morhua [juvenile] C → dead Pandalus montagui [adult]	174	M	L	2–19	5	0.73	2.92			15

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
Hexagenia limbata [juvenile] H → sediment 'a'	174	M	L	2–19	5	0.75	2.99			15
	192	F	L	5–25	5	0.43	1.86			20
	192	F	L	5–25	5	0.44	1.88			20
Pleuronectes platessa [adult] C → Arenicola marina [adult] O	46	M	L	1–20	5	0.48	1.99			
	46	M	L	1–20	5	1.15	5.25			
Pleuronectes platessa [juvenile] C → fish-paste	85	M	L	5–21	5	0.49	2			
Uta stansburiana [adult] C → Acheta sp. A [adult] O	178	T	L	22–32	5	NS	NS			
	178	T	L	22–32	5	NS	NS			
Gut Loading Rate (internal)										
Hexagenia limbata [juvenile] H → sediment 'a'	192	F	L	5–25	5	0.51	2.09			20
Handling Rate (species interaction)										
Gymnocephalus cernuus [adult] C → Chaoborus obscuripes [juvenile] C	18	F	L	4–20	5	0.28	1.49			
Perca fluviatilis [adult] O → Chaoborus obscuripes [juvenile] C	18	F	L	4–20	5	0.83	3.24			
Host-Per-Parasitoid Parasitization Rate (species interaction)										
Anisopteromalus calandreae [adult] C → Rhyzopertha dominica [juvenile] H	122	T	L	20–35	4					30
	122	T	L	20–35	4					35
	122	T	L	20–35	4					35
	122	T	L	20–35	4	NS	NS			
	122	T	L	20–35	4	NS	NS			
Anisopteromalus calandreae [adult] C → Sitophilus zea-mais [juvenile] H	161	T	L	20.2–35.3	4	NS	NS			
Cardiochiles philippinensis [adult] C → Cnaphalocrocis medinalis [juvenile] H	156	T	L	25–35	5			0.55	0.5	
	156	T	L	25–35	5			0.74	0.39	
	156	T	L	25–35	5			0.84	0.35	28
	156	T	L	25–35	5			1.08	0.26	28
	156	T	L	25–35	5			NS	NS	
	156	T	L	25–35	5			NS	NS	28
Cephalonomia waterstoni [adult] C → Cryptolestes ferrugineus [juvenile] H	53	T	L	20–38	5					25
	53	T	L	20–38	5					30
	53	T	L	20–38	5					30
	53	T	L	20–38	5			NS	NS	25
	53	T	L	20–38	5	NS	NS			35
Praon exsoletum [adult] C → Therioaphis trifolii [adult] H	124	T	L	10–23.9	5					23.9

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
	124	T	L	10–23.9	5	NS	NS			
	124	T	L	10–26.7	4					12.8
	124	T	L	10–26.7	4					23.9
	124	T	L	10–26.7	6					15.6
	124	T	L	10–26.7	6	NS	NS			21.1
	124	T	L	10–26.7	6	NS	NS			23.9
	124	T	L	10–26.7	6	NS	NS	NS	NS	23.9
	124	T	L	10–26.7	7	0.72	2.7			23.9
	124	T	L	10–26.7	7	NS	NS			23.9
	124	T	L	11.4–25.8	4					21.1
	124	T	L	11.4–25.8	4	NS	NS			
Theocolax elegans [adult] C → Rhyzopertha dominica [juvenile] H	52	T	L	20–32.5	4					30
	52	T	L	20–32.5	4					30
In Vitro Gill Beat Rate (individual) Dreissena polymorpha [adult] O	103	F	L	8–22	4	0.5	2.01			
	103	F	L	8–22	4	0.5	2			
In Vitro Gill Particle Transport Velocity (individual) Mytilus edulis [adult] O → yeast D	88	M	L	5–20	4	0.36	1.67			
	88	M	L	5–20	4	0.45	1.91			
	88	M	L	5–20	4	0.47	1.96			
	88	M	L	5–20	4	0.49	2.01			
Mytilus sp. A [adult] O	64	M	L	0–35	10	0.61	2.34			32.5
In Vitro Heart Beat Rate (internal) Oceanites oceanicus [egg] S	187	T	L	10.1–40.4	15	0.86	3.05			
Oceanodroma leucorhoa [egg] S	187	T	L	12–41	14	1.15	4.4			
In Vitro Muscle Isometric Tension (internal) Myoxocephalus scorpius [adult] C	11	M	L	0–20	5			NS	NS	15
	11	M	L	0–20	5	0.3	1.56			15
In Vitro Muscle Optimal Phase (internal) Manduca sexta [adult] H	166	T	L	20.2–40.3	5			NS	NS	
In Vitro Muscle Optimal Rate (internal) Manduca sexta [adult] H	166	T	L	20–39.9	5	0.32	1.5			

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
In Vitro Muscle Optimal Strain (internal) Manduca sexta [adult] H	166	T	L	19.9–39.9	5	0.13	1.17			
In Vitro Muscle Power Output (internal) Manduca sexta [adult] H	166	T	L	20–40	5	0.6	2.14			
In Vitro Muscle Shortening Velocity (internal) Myoxocephalus scorpius [adult] C	11	M	L	0–20	5	0.23	1.39			
	11	M	L	0–20	5	0.41	1.81			
In Vitro Muscle Work Per Cycle (internal) Manduca sexta [adult] H	166	T	L	20–40	5	0.28	1.42			
Individual Length Growth Rate (individual) Moina macrocopa [adult] O → Chlorella sorokiniana P	16	F	L	15–30	5	0.99	3.7			
Moina macrocopa [juvenile] O → Chlorella sorokiniana P	16	F	L	15–30	5	0.86	3.14			
Individual Mass Growth Rate (individual) Caulerpa serrulata [adult] P → light	107	M	L	5–40	8	0.6	2.27			30
Cherax quadricarinatus [juvenile] O → crayfish ration	121	F	L	16–32	9	2.57	30.3			28
Dicentrarchus labrax [juvenile] C → commercial extruded dry pellet	138	M	L	13.4–28.8	6	0.94	3.58			24.9
Intraspecific Confrontation Probability Density (species interaction) Bembidion lampros [adult] C	31	T	L	5–30	5					20
	31	T	L	5–30	5					20
Pterostichus cupreus [adult] C	31	T	L	5–30	5	0.61	2.31			
	31	T	L	5–30	5	0.61	2.34			
	31	T	L	5–30	5	NS	NS			
	31	T	L	5–30	5	NS	NS			
Line Encounter Rate (species interaction) Pogonomyrmex occidentalis [adult] H	36	T	F	23.5–52.2	5					41.6
	36	T	F	23.7–57.3	11	1.16	4.13	NS	NS	
	36	T	F	24.3–51.4	9	1.35	5.15			47.1
	36	T	F	25.7–55.8	7	0.83	2.74			46.3
	36	T	F	26.5–51.4	9	1.02	3.34			
	36	T	F	26.6–57.9	13	1.52	6.41	1.95	0.11	42.9
	36	T	F	26.9–58.3	10	1.27	4.5	NS	NS	51.8
	36	T	F	27.3–51.1	11	0.59	2.01			48.3

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
	36	T	F	27.7–57.1	11	1.12	3.75	0.87	0.38	42.6
	36	T	F	27.9–57.9	8	1.88	9.86	NS	NS	42.1
	36	T	F	28.2–50.5	9	NS	NS	NS	NS	42.7
	36	T	F	28.6–56.7	9	1.13	3.71	NS	NS	
	36	T	F	28.9–58.1	11	1.52	6.31	2.37	0.07	43.6
	36	T	F	29.5–55.3	10	1.21	4.19	NS	NS	47.2
	36	T	F	30.7–59	11	1.1	3.71	1.38	0.22	47.9
	36	T	F	32.3–53.6	6	NS	NS			44.6
	36	T	F	32.9–51.4	5			NS	NS	39
	36	T	F	32.9–52.8	6	NS	NS			49.7
	36	T	F	33.5–53.2	6	NS	NS			45
	36	T	F	36.1–55	7	NS	NS			
	36	T	F	37–56.4	9	NS	NS			
	36	T	F	45.7–56.5	4					49.8
Log-Linear Gut Clearance Rate (internal)										
Temora longicornis [adult] O → Thalassiosira weissflogii [adult] P	37	M	L	1–17	10	0.88	3.65			13
Mortality Rate (population)										
Aphis gossypii [adult] H → Cucumis sativus P	191	T	L	10–30	5	0.54	2.08			
	191	T	L	10–30	5	NS	NS			
Aphis gossypii [juvenile] H → Cucumis sativus P	191	T	L	10–30	5	NS	NS			
Euplectrus ronnai [juvenile] C → Pseudaletia sequax [juvenile] H	189	T	L	15–29	5	NS	NS			
Moina macrocopa [adult] O → Chlorella sorokiniana P	16	F	L	15–30	5	0.43	1.78			
Planococcus citri [adult] H → Solenostemon scutellarioides P	61	T	L	18–32	7	0.32	1.52			
	61	T	L	18–32	7	0.47	1.86			30
	61	T	L	18–32	7	NS	NS	0.32	0.66	25
Procambarus clarkii [adult] O → uncooked mixed vegetables	30	F	L	15–30	6	0.68	2.45			
Procambarus clarkii [juvenile] O → uncooked mixed vegetables	30	F	L	15–30	6	0.66	2.4			
Telenomus chrysopae [juvenile] C → Chrysoperla rufilabris [egg] S	155	T	L	15.6–26.7	5	NS	NS			
Telenomus isis [adult] C → Busseola fusca [egg] S	25	T	L	18–32	6	0.6	2.19			
	25	T	L	18–32	6	0.8	2.87			
Telenomus isis [adult] C → Sesamia calamistis [egg] S	25	T	L	18–32	6	0.94	3.44			
	25	T	L	18–32	6	1.03	3.86			

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
Telenomus isis [adult] C → Sesamia nonagrioides [egg] S	25	T	L	18–32	6	0.54	2.03			30
	25	T	L	18–32	6	1.12	4.33			
Telenomus isis [juvenile] C → Busseola fusca [egg] S	25	T	L	18–32	6	0.55	2.06			30
	25	T	L	18–32	6	0.74	2.65			
Telenomus isis [juvenile] C → Sesamia calamistis [egg] S	25	T	L	18–32	6	0.54	2.04			
	25	T	L	18–32	6	NS	NS			
Telenomus isis [juvenile] C → Sesamia nonagrioides [egg] S	25	T	L	18–32	6	0.47	1.87			
	25	T	L	18–32	6	0.57	2.12			
Tetraneura nigri abdominalis [adult] H → Oryza sativa P	98	T	L	10–30	5	0.56	2.13			
	98	T	L	10–35	6	0.24	1.38			
Theocolax elegans [adult] C	84	T	L	20–35	6	0.58	2.12			
	84	T	L	20–35	6	0.86	3.04			
Nest Provisioning Rate (species interaction)										
Buteo jamaicensis [adult] C → Serpentes spp. [adult] C	168	T	F	15.3–30.2	12			0.43	0.57	
Oxygen Mass Scope For Activity (internal)										
Oncorhynchus mykiss [adult] C	39	F	L	5–25	5					15
	39	F	L	5–25	5					20
Photosynthetic Oxygen Production Rate (internal)										
Caulerpa serrulata [adult] P → light	107	M	L	5–40	8	0.66	2.48			30
Lithophyllum margaritae P → light	164	M	L	10–30	5	NS	NS			25
POC Photosynthetic Oxygen Production Rate (internal)										
Phaeodactylum tricornutum P → light	69	M	L	0–30	7	0.38	1.71			25
Prorocentrum minimum P → light	69	M	L	0–30	7	0.51	2.07			25
Prymnesium patelliferum O → light	69	M	L	0–30	7	0.4	1.75			25
Point Encounter Density Rate (species interaction)										
Phytoseiulus persimilis [adult] C	48	T	L	15–30	4					25
Tetranychus urticae [adult] H	48	T	L	15–30	4	NS	NS			
Point Encounter Number Rate (species interaction)										
Homarus americanus [adult] C	147	M	L	11–28.5	9	0.76	2.81			
Salmo salar [juvenile] C	142	F	L	6–21	6	1.2	5.36			
	142	F	L	6–27	8	0.78	2.95			
Population Catchability (species interaction)										

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
Homarus americanus [adult] C	120	M	F	2.8–11.5	6	2.28	29.18			
Population Density (population)										
Acartia sinjiensis [adult] O → microalgae P	125	M	L	10–38	8	1.68	9.45			30
Acarus siro [adult] H → wheat germs, oat flakes and baker's yeast	6	F	L	5–35	12	1.71	10.5	2.19	0.06	27.5
Aleuroglyphus ovatus [adult] H → wheat germs, oat flakes and baker's yeast	6	T	L	5–35	12	2.23	21.15	NS	NS	27.5
Tyrophagus putrescentiae [adult] H → wheat germs, oat flakes and baker's yeast	6	T	L	10–35	11	1.33	6.26			20
Population Foraging Probability (species interaction)										
Brachycentrus americanus [juvenile] H → dead Artemia spp. [adult]	57	F	F	8.6–30	5	NS	NS			24.6
	57	F	F	8.8–30.9	6					13.1
	57	F	F	15.4–26.3	7			0.24	0.73	
	57	F	F	15.4–26.3	7			0.26	0.7	
	57	F	F	15.4–26.3	7			0.39	0.59	15.4
	57	F	F	4–16	4	NS	NS			
	57	F	F	4–16	4	NS	NS			
Hyles lineata [juvenile] H → desert plant P	27	T	F	14.8–36.1	6					27.5
Manduca sexta [juvenile] H → Datura inoxia P	27	T	F	18.9–35.8	6			0.36	0.63	22.6
Population Growth Rate (population)										
Acarus siro [adult] H → wheat germs, oat flakes and baker's yeast	6	T	L	5–35	12	1.93	13.86	0.76	0.38	26.3
Aleuroglyphus ovatus [adult] H → wheat germs, oat flakes and baker's yeast	6	T	L	5–35	12	1.74	10.73	NS	NS	26.3
Aphis gossypii [adult] H → Cucumis sativus P	191	T	L	10–30	5	0.91	3.47			25
Chlorella vulgaris P → light + mineral medium	38	F	L	10–35	6	0.39	1.69			30
Coelastrum microporum P → light	20	F	L	15–35	5	0.5	1.92			
Cosmarium subprotumidum P → light	20	F	L	15–35	5	NS	NS			
Escherichia coli O → luria broth	24	T	L	10–45	11	0.45	1.81	NS	NS	37
Fragilaria crotonensis P → light + mineral medium	38	F	L	10–35	6	0.47	1.9			25
Lemna minor P → aqueous growth medium	101	F	L	5–35	12	1.09	4.49	1.35	0.18	27
Moerisia lyonsi [adult] C → Acartia tonsa O	114	M	L	10–29	5	1.56	8.24			
Paraphysomonas imperforata C → Halomonas marina D	154	M	L	0–20	5	1.04	4.58			15
Paraphysomonas imperforata C → Phaeodactylum tricornutum P	26	M	L	14–26	4	0.65	2.41			
Planococcus citri [adult] H → Solenostemon scutellarioides P	61	T	L	18–32	7	1.18	4.84	NS	NS	25
Salmonella enterica C → tetrathionate broth	24	T	L	10–43	10	0.56	2.09			36
Selenastrum minutum P → light	20	F	L	15–35	5	0.34	1.56			

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
Staurastrum pingue P → light + mineral medium	38	F	L	10–35	6	0.56	2.15			25
Synechocystis minima P → light + mineral medium	38	F	L	10–35	6	0.36	1.61			
Telenomus isis [adult] C → Busseola fusca [egg] S	25	T	L	18–32	6	0.74	2.64			30
	25	T	L	18–32	6	0.95	3.53			27
Telenomus isis [adult] C → Sesamia calamistis [egg] S	25	T	L	18–32	6	0.75	2.67			30
	25	T	L	18–32	6	0.98	3.67			27
Telenomus isis [adult] C → Sesamia nonagrioides [egg] S	25	T	L	18–32	6	0.87	3.15			27
	25	T	L	18–32	6	NS	NS			27
Tetraneura nigri abdominalis [adult] H → Oryza sativa P	98	T	L	10–30	5	0.87	3.21			
Tyrophagus putrescentiae [adult] H → wheat germs, oat flakes and baker's yeast	6	T	L	10–35	11	1.37	6.63			25.6
Urotricha farcta H → Cryptomonas spp. P	183	F	L	9–24	6	0.6	2.28			
Population Voluntary Activity Probability (individual)										
Crangonyx richmondensis [adult] O	180	F	L	3.9–22.1	8	0.97	4.02			20.2
Hyallolela azteca [adult] O	180	F	L	8–23.3	6	1.34	6.53			20.4
Population Voluntary Movement Probability (individual)										
Crangonyx richmondensis [adult] O	180	F	L	5–20	5					12.5
Hyallolela azteca [adult] O	180	F	L	8.9–20	4			NS	NS	
Pomacea paludosa [adult] H → lettuce, spinach, bladderwort P	165	F	L	14.5–21.3	6	NS	NS			
	165	F	L	14.5–24	6	1.81	11.72			
	165	F	L	17–22	5	2.06	16.33			
Radial Growth Rate (population)										
Beauveria bassiana [adult] C → agar medium	49	T	L	8–32	8	0.91	NS			20
	49	T	L	8–35	9	0.55	2.13	NS	NS	25
	49	T	L	8–35	9	0.63	2.42	NS		20
	49	T	L	8–35	9	0.65	2.46			28
	49	T	L	8–35	9	0.8	3.03			28
	49	T	L	8–35	9	0.83	3.1			28
	49	T	L	8–35	9	0.83	3.15	NS	NS	25
	49	T	L	8–35	9	0.86	3.21	NS	NS	25
	49	T	L	8–35	9	0.88	3.37	NS	NS	25
	49	T	L	8–35	9	0.94	3.67	NS	NS	25
	49	T	L	8–35	9	1	3.97			28

Trait / Consumer [stage] trophic group → Resource [stage] trophic group

	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
	49	T	L	8-35	9	1.12	4.79			20
	49	T	L	8-35	9	NS	NS	NS	NS	25
	49	T	L	8-37	10	0.49	1.96	4.08	0.01	28
	49	T	L	8-37	10	0.51	2.04	3.85	0.01	25
	49	T	L	8-37	10	0.56	2.16	3.32	0.02	25
	49	T	L	8-37	10	0.57	2.17	NS	NS	28
	49	T	L	8-37	10	0.58	2.2	3.74	0.01	28
	49	T	L	8-37	10	0.61	2.29	NS	NS	28
	49	T	L	8-37	10	0.62	2.35	NS	NS	25
	49	T	L	8-37	10	0.64	2.42	NS	NS	25
	49	T	L	8-37	10	0.65	2.43			25
	49	T	L	8-37	10	0.66	2.49	3.47	0.01	25
	49	T	L	8-37	10	0.66	2.47	5.14	0	28
	49	T	L	8-37	10	0.66	2.46			30
	49	T	L	8-37	10	0.68	2.49	NS	NS	28
	49	T	L	8-37	10	0.7	2.6	NS	NS	28
	49	T	L	8-37	10	0.73	2.73	2.83	0.03	25
	49	T	L	8-37	10	0.73	2.73	NS	NS	25
	49	T	L	8-37	10	0.75	2.82	NS	NS	25
	49	T	L	8-37	10	0.76	2.82	NS	NS	28
	49	T	L	8-37	10	0.77	2.9	2.89	0.03	25
	49	T	L	8-37	10	0.77	2.88	NS	NS	25
	49	T	L	8-37	10	0.78	2.95	NS	NS	25
	49	T	L	8-37	10	0.79	2.99	3.97	0.01	25
	49	T	L	8-37	10	0.8	3.03	NS	NS	25
	49	T	L	8-37	10	0.83	3.1	NS	NS	25
	49	T	L	8-37	10	0.83	3.12	NS	NS	25
	49	T	L	8-37	10	0.83	3.05	NS	NS	28
	49	T	L	8-37	10	0.85	3.2	2.15	0.07	25
	49	T	L	8-37	10	0.85	3.22	NS	NS	25
	49	T	L	8-37	10	0.85	3.2	NS	NS	28
	49	T	L	8-37	10	0.86	3.29	NS	NS	28

Trait / Consumer [stage] trophic group → Resource [stage] trophic group

	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
	49	T	L	8-37	10	0.88	3.37	NS	NS	25
	49	T	L	8-37	10	0.89	3.433	NS	NS	25
	49	T	L	8-37	10	0.91	2.4	4.28	0	25
	49	T	L	8-37	10	0.93	3.56	NS	NS	25
	49	T	L	8-37	10	0.93	3.55	NS	NS	28
	49	T	L	8-37	10	0.95	3.71	NS	NS	25
	49	T	L	8-37	10	0.96	3.78	NS	NS	28
	49	T	L	8-37	10	0.98	3.86	NS	NS	25
	49	T	L	8-37	10	1	3.93	0.83	0.35	25
	49	T	L	8-37	10	1	3.95	NS	NS	25
	49	T	L	8-37	10	1.01	4.03	NS	NS	25
	49	T	L	8-37	10	1.03	4.13	NS	NS	20
	49	T	L	8-37	10	1.03	4.1	NS	NS	25
	49	T	L	8-37	10	1.04	4.19	NS	NS	25
	49	T	L	8-37	10	1.05	4.19	NS	NS	25
	49	T	L	8-37	10	1.05	4.21	NS	NS	28
	49	T	L	8-37	10	1.07	4.34	NS	NS	25
	49	T	L	8-37	10	1.11	4.62	0.84	0.35	25
	49	T	L	8-37	10	1.31	6.1	NS	NS	25
	49	T	L	8-37	10	1.36	6.47	NS	NS	25
	49	T	L	8-37	10	1.54	8.26	NS	NS	25
	49	T	L	8-37	10	NS	NS	NS	NS	25
Metarhizium anisopliae [adult] C → agar medium	136	T	L	11-37	9			NS	NS	25
	136	T	L	11-37	9			NS	NS	25
	136	T	L	11-37	9			NS	NS	25
	136	T	L	11-37	9	NS	NS			29
	136	T	L	8-35	9	0.69	NS	1.94	0.08	25
	136	T	L	8-35	9	1.08	4.44	NS	NS	25
	136	T	L	8-35	9	1.24	5.51	NS	NS	25
	136	T	L	8-35	9	1.62	9.26			25
	136	T	L	8-35	9	NS	NS	NS	NS	25
	136	T	L	8-37	10	0.76	2.82	NS	NS	25

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
	136	T	L	8-37	10	0.82	3.02	1.78	0.11	28
	136	T	L	8-37	10	0.88	3.25	2	0.08	28
	136	T	L	8-37	10	0.89	NS	1.59	0.13	25
	136	T	L	8-37	10	0.89	3.33	NS	NS	28
	136	T	L	8-37	10	1	3.92	2.47	0.05	26.5
	136	T	L	8-37	10	1.01	3.94	0.71	0.41	25
	136	T	L	8-37	10	1.02	4.01	NS	NS	28
	136	T	L	8-37	10	1.03	4.1	NS	NS	28
	136	T	L	8-37	10	1.07	4.34	1.89	0.09	25
	136	T	L	8-37	10	1.16	NS			30
	136	T	L	8-37	10	NS	NS	NS	NS	25
	136	T	L	8-37	10	NS	NS	NS	NS	25
Metarhizium flavoviride [adult] C → agar medium	136	T	L	11-37	9	0.66	NS			30
	136	T	L	11-37	9	0.72	NS			30
	136	T	L	11-37	9	0.79	NS			30
	136	T	L	11-37	9	NS	NS	NS	NS	28
	136	T	L	11-37	9	NS	NS	NS	NS	28
	136	T	L	11-37	9	NS	NS	NS	NS	28
	136	T	L	8-37	10	0.78	2.92	NS	NS	25
	136	T	L	8-37	10	0.78	2.93	NS	NS	28
	136	T	L	8-37	10	1.08	4.3	NS	NS	28
	136	T	L	8-37	10	1.28	NS			30
	136	T	L	8-37	10	1.31	5.88	NS	NS	28
	136	T	L	8-37	10	1.38	6.55	NS	NS	28
	136	T	L	8-37	10	1.43	7.02	NS	NS	26.5
	136	T	L	8-37	10	1.75	11.14	NS	NS	25
Paecilomyces fumosoroseus C → agar medium	176	T	L	8-30	7	0.83	3.14			25
	176	T	L	8-30	7	0.88	3.36			25
	176	T	L	8-32	8	0.54	NS			25
	176	T	L	8-32	8	0.56	NS			25
	176	T	L	8-32	8	0.63	2.41	NS	NS	20
	176	T	L	8-32	8	0.69	2.66	NS	NS	20

Trait / Consumer [stage] trophic group → Resource [stage] trophic group

Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
176	T	L	8-32	8	0.74	NS			25
176	T	L	8-32	8	0.78	2.99	NS	NS	20
176	T	L	8-32	8	0.93	3.66	NS	NS	20
176	T	L	8-35	9	0.55	NS			28
176	T	L	8-35	9	0.61	2.29			30
176	T	L	8-35	9	0.64	2.4	NS	NS	25
176	T	L	8-35	9	0.72	NS			28
176	T	L	8-35	9	0.76	2.89	NS	NS	20
176	T	L	8-35	9	0.79	NS			28
176	T	L	8-35	9	0.8	NS			28
176	T	L	8-35	9	0.8	NS			28
176	T	L	8-35	9	0.83	NS			28
176	T	L	8-35	9	0.85	NS			28
176	T	L	8-35	9	0.86	NS			28
176	T	L	8-35	9	0.86	NS			28
176	T	L	8-35	9	0.86	3.27	NS	NS	25
176	T	L	8-35	9	0.9	NS			28
176	T	L	8-35	9	0.9	NS			28
176	T	L	8-35	9	0.91	NS			28
176	T	L	8-35	9	0.91	3.49	NS	NS	25
176	T	L	8-35	9	0.92	3.56	NS	NS	25
176	T	L	8-35	9	0.95	3.64	NS	NS	25
176	T	L	8-35	9	0.96	3.74	NS	NS	25
176	T	L	8-35	9	0.96	3.76	NS	NS	25
176	T	L	8-35	9	0.97	3.79	NS	NS	25
176	T	L	8-35	9	0.98	3.86	NS	NS	25
176	T	L	8-35	9	1.03	4.13	NS	NS	25
176	T	L	8-35	9	1.04	4.21	1.97	0.08	25
176	T	L	8-35	9	1.04	4.16	NS	NS	25
176	T	L	8-35	9	1.05	4.23	NS	NS	25
176	T	L	8-35	9	1.1	NS	NS		28

Rattle Rate (individual)

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
Homo sapiens [adult] O → Crocalus viridis viridis [adult] C	29	T	L	8–36	7	0.42	1.75			
Refuge Distance (species interaction)										
Homo sapiens [adult] O → Holbrookia propinqua [adult] C	35	T	F	26.6–49.3	12	NS	NS			
	35	T	F	29.3–49.6	7	NS	NS			
Resource Habitat Encounter Density Rate (species interaction)										
Bembidion lampros [adult] C	31	T	L	9–30	4		NS			20
Pterostichus cupreus [adult] C	31	T	L	5–30	5	0.47	1.91			
Resource Reaction Distance (species interaction)										
Homo sapiens [adult] O → Holbrookia propinqua [adult] C	35	T	F	26–42.5	13	NS	1.68			
	35	T	F	27.9–41.6	10	0.61	2.11			
Homo sapiens [adult] O → Norops lineatopus [adult] O	144	T	F	24.4–31.1	6			NS	NS	
Homo sapiens [adult] O → Sceloporus anahuacus [adult] O	160	T	F	23.1–32.8	4			NS	NS	
Homo sapiens [adult] O → Sceloporus gadoviae [adult] O	160	T	F	27.6–36.9	4			1.2	0.23	
Homo sapiens [adult] O → Sceloporus mucronatus [adult] O	160	T	F	26.1–34.1	8	NS	NS			
Homo sapiens [adult] O → Scincella lateralis [adult] C	159	T	F	23.5–33.8	8			0.38	0.62	
Homo sapiens [adult] O → Urosaurus bicarinatus [adult] C	160	T	F	35.2–40.6	4	NS	NS			
Resource Size Capture Intent Acceptance Probability (species interaction)										
Formica schaufussi [adult] C → dead Nauphoeta cinerea [adult]	173	T	F	16.5–33.6	7	0.89	3.18			
Respiration Rate (internal)										
Cherax quadricarinatus [juvenile] O → crayfish ration	121	F	L	16–32	9	3.05	57.99			28
Lithophyllum margaritae P → light	164	M	L	10–30	5	0.57	2.15			
Sediment Mass Processing Rate (species interaction)										
Pectinaria gouldii [adult] O → fine sediment	62	M	L	13–19	5	1.27	5.81			
	62	M	L	13–19	5	2.84	51.7			
	62	M	L	13–19	5	NS	NS			
Square Root-Linear Gut Clearance Rate (internal)										
Pleuronectes platessa [juvenile] C → fish-paste	86	M	L	5–15.5	4	0.55	2.2			
Strike Acceleration (individual)										
Pituophis catenifer affinis [adult] C → Mus musculus [adult] O	65	T	L	18–33	4					27
	65	T	L	18–33	4					27
Strike Completion Rate (individual)										
Pituophis catenifer affinis [adult] C → Mus musculus [adult] O	65	T	L	18–33	4					27

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
Strike Distance (individual)										
Pituophis catenifer affinis [adult] C → Mus musculus [adult] O	65	T	L	18–33	4			NS	NS	
Strike Velocity (individual)										
Pituophis catenifer affinis [adult] C → Mus musculus [adult] O	65	T	L	18–33	4					22
	65	T	L	18–33	4					27
	65	T	L	18–33	4					27
	65	T	L	18–33	4					27
	65	T	L	18–33	4					27
	65	T	L	18–33	4					27
	65	T	L	18–33	4					27
	65	T	L	18–33	4					27
	65	T	L	18–33	4			NS	NS	
	65	T	L	18–33	4	NS	NS			
Subjugation-Consumption Body Contraction Rate (individual)										
Aurelia aurita [juvenile] C → Clupea harengus [juvenile] H	74	M	L	5–22	4	NS	NS			
Subjugation-through-Consumption Rate (species interaction)										
Cicindela hybrida [adult] C → cursorial insect	42	T	F	25.3–40.3	6			NS	NS	
Notonecta hoffmani [adult] C → Culex pipiens [juvenile] O	129	F	L	10.8–25	4	0.72	2.67			
Zootoca vivipara [adult] C → Acheta domesticus [juvenile] O	8	T	L	11.2–32.2	7	0.76	2.77			
	8	T	L	11–32.1	9	0.61	2.26			
	8	T	L	14.6–32.1	7	0.83	3.01			
	8	T	L	14.6–32.2	6	0.88	3.21			
	8	T	L	8.2–32.2	10	1.18	4.88			
	8	T	L	8.3–32.1	9	0.96	3.62			
Surface Area-Specific Dark Respiration Rate (internal)										
Betula pendula [adult] P → light	188	T	L	-5–40	7	0.58	2.23			
Fagus sylvatica [adult] P → light	188	T	L	-5–40	8	0.41	1.77			
Surface Area-Specific Foraging Gill Filtration Rate (individual)										
Mytilus edulis [adult] O → Rhodomonas spp. P	89	M	L	5.9–16.9	8	0.38	1.72			
Surface Area-Specific Maximum Photosynthesis Rate (internal)										
Betula pendula [adult] P → light	188	T	L	-5–40	7	0.55	2.19			30
Eucalyptus regnans [juvenile] P → light	181	T	L	10–35	6	0.14	1.21			
	181	T	L	10–35	6	0.16	1.24			30

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
Fagus sylvatica [adult] P → light	188	T	L	-5-40	8	0.31	1.56			30
Surface Area-Specific Mitochondrial Respiration Rate (internal)										
Eucalyptus regnans [juvenile] P → light	181	T	L	10-35	6	0.74	2.66			
Surface Area-Specific Photosynthetic Oxygen Production Rate (internal)										
Embothrium coccineum [adult] P → light	28	T	L	5-40	8					
	28	T	L	5-40	8	0.52	2.02			30
Gevuina avellana P → light	28	T	L	5-40	8	0.79	2.98			25
	28	T	L	5-40	8	NS	NS			
Lomatia ferruginea P → light	28	T	L	5-40	8	0.27	1.45			25
	28	T	L	5-40	8	0.39	NS			25
Voluntary Activity Probability (individual)										
Uta stansburiana [adult] C → Acheta sp. A [adult] O	178	T	L	20-36	6	0.03	1.04			
Voluntary Body Contraction Rate (individual)										
Aurelia aurita [juvenile] C	40	M	L	10-35	6					25
Voluntary Body Velocity (individual)										
Anisops deanei [adult] C	9	F	L	15-28	4					20
	9	F	L	15-28	4					20
	9	F	L	15-28	4					20
	9	F	L	15-28	4					20
	9	F	L	15-28	4					20
	9	F	L	15-28	4					20
	9	F	L	15-28	4					25
	9	F	L	15-28	4					25
	9	F	L	15-28	4					25
	9	F	L	15-28	4			0.84	0.33	
	9	F	L	15-28	4			0.9	NS	
	9	F	L	15-28	4			NS	NS	
	9	F	L	15-28	4			NS	NS	
	9	F	L	15-28	4	1.03	3.94			
	9	F	L	15-28	4	NS	NS			
	9	F	L	15-28	4	NS	NS			
	9	F	L	15-28	4	NS	NS			

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
	9	F	L	15–28	4	NS	NS			
	9	F	L	15–28	4	NS	NS			
	9	F	L	15–28	4	NS	NS			
	9	F	L	15–28	4	NS	NS			
	9	F	L	15–28	4	NS	NS			
	9	F	L	15–28	4	NS	NS			
Astacus astacus [adult] O	95	F	L	0.6–34.4	6	NS	NS	NS	NS	26.1
	95	F	L	-1.3–32	8	NS	NS			20.2
Barbus barbus [adult] O	135	F	L	7–25	4	0.74	2.81			
	135	F	L	7–25	4	NS	NS			
Bembidion lampros [adult] C	31	T	L	5–30	4	NS	NS			
Cataglyphis bicolor [adult] D	71	T	F	29.2–60.1	12	0.16	1.2			
Chionoecetes opilio [adult] C	55	M	L	0–18	7	0.39	1.76			
Culicoides variipennis [juvenile] C	109	F	L	6–36	6	0.52	2.01			
Diaptomus kenai [adult] O	169	F	L	4–22	4	0.19	1.31			
Diaptomus kenai [juvenile] O	169	F	L	4–22	4					14
Dorymyrmex goetschi [adult] O → sugar microspheres	172	T	F	18.6–37.4	5	0.28	NS			
Gymnocephalus cernuus [adult] C	18	F	L	4–20	5	0.36	1.68			16
Homarus americanus [adult] C	147	M	L	10–25	4	NS	NS			
Micropterus salmoides [adult] C	104	F	L	3–17	8					15
Nucella lapillus [adult] C	99	M	L	5–25	5	0.53	2.12			20
Perca fluviatilis [adult] O	18	F	L	4–20	5	1.28	6.18			
Pterostichus cupreus [adult] C	31	T	L	5–30	5	0.43	1.8			
Solenopsis invicta [adult] O	148	T	F	10.5–32.3	10	0.54	2.05			
Thamnophis sirtalis [adult] C	73	T	L	15.3–33	4	0.07	1.1			
Zygiella x-notata [adult] C	7	T	L	2–20	5	1.15	5.15			
Voluntary Eye Saccade Angular Velocity (individual)										
Girella tricuspidata [adult] H	127	M	L	6.9–14	8	1.24	6			
Voluntary Heart Beat Rate (internal)										
Rana temporaria [adult] C	72	T	L	15.5–29.4	8	0.47	1.88			
	72	T	L	16–29.4	7	0.4	1.69			
	72	T	L	16–29.7	7	0.54	2.06			

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
	72	T	L	17.5–29.7	5	0.54	2.03			
Voluntary Movement Rate (individual)										
Anisops deanei [adult] C	9	F	L	15–28	4					15
	9	F	L	15–28	4					15
	9	F	L	15–28	4					20
	9	F	L	15–28	4					25
Bembidion lampros [adult] C	31	T	L	10–30	4					15
	31	T	L	5–30	5			NS	NS	
	31	T	L	5–30	5	NS	NS			
Chionoecetes opilio [adult] C	55	M	L	0–18	7	1.11	5.06	NS	NS	9
Pterostichus cupreus [adult] C	31	T	L	5–30	5			NS	NS	
	31	T	L	5–30	5	0.3	1.53			20
	31	T	L	5–30	5	NS	NS			
Voluntary Stroke Rate (individual)										
Anisops deanei [adult] C	9	F	L	15–28	4					25
	9	F	L	15–28	4					25
	9	F	L	15–28	4					25
	9	F	L	15–28	4					25
	9	F	L	15–28	4					25
	9	F	L	15–28	4					28
	9	F	L	15–28	4			NS	NS	
	9	F	L	15–28	4			NS	NS	
	9	F	L	15–28	4	0.62	2.29			
	9	F	L	15–28	4	NS	NS			
	9	F	L	15–28	4	NS	NS			
	9	F	L	15–28	4	NS	NS			
	9	F	L	15–28	4	NS	NS			
	9	F	L	15–28	4	NS	NS			
	9	F	L	15–28	4	NS	NS			
	9	F	L	15–28	4	NS	NS			
	9	F	L	15–28	4	NS	NS			
	9	F	L	15–28	4	NS	NS			
	9	F	L	15–28	4	NS	NS			
	9	F	L	15–28	4	NS	NS			

Trait / Consumer [stage] trophic group → Resource [stage] trophic group	Cit	Hab	L/F	Temp	N	E_R	Q_{10R}	E_F	Q_{10F}	T_{opt}
	9	F	L	15–28	4	NS	NS			
	9	F	L	15–28	4	NS	NS			
	9	F	L	15–28	4	NS	NS			
	9	F	L	15–28	4	NS	NS			
	9	F	L	15–28	4	NS	NS			
	9	F	L	15–28	4	NS	NS			
Voluntary Tail Beat Rate (individual)										
Culicoides variipennis [juvenile] C	109	F	L	6–36	6	0.42	1.74			
Dendrodoa grossularia [juvenile] O	13	M	L	9.5–18.1	4	0.64	2.47			
Voluntary Tongue Flick Cycle Number (individual)										
Thamnophis elegans vagrans [adult] C	167	T	L	4.9–35	7			0.21	0.76	
Voluntary Tongue Flick Cycle Rate (individual)										
Thamnophis elegans vagrans [adult] C	167	T	L	5.1–34.8	7	0.74	2.75			29.8
	167	T	L	5–34.9	7	0.42	1.76			
Voluntary Tongue Flick Non-Cycle Rate (individual)										
Thamnophis elegans vagrans [adult] C	167	T	L	5.6–35.7	7	0.5	1.98			30.5
Voluntary Wing Beat Rate (individual)										
Coleomegilla fuscilabris [adult] O	134	T	F	28.2–35.9	7	0.11	1.15			
Popillia japonica [adult] H	134	T	F	30.7–40.3	10	0.14	1.19			

Table S4. Results of ANOVA to test effects on mean activation energy, E , of rise responses. We do not include motivation because of the strong overlap in some categories in motivation and organization level, for example all responses that are autonomic (motivation) are also internal (level of organization). The effect of taxonomy is examined separately because most traits above the internal and individual organization level consist of multiple taxa. A similar problem arises with trophic level categorization. Because activation energies in most categories are right skewed (Fig. 2b, main text), we log-transformed E 's to render them approximately normal (two-tailed, one-sample Kolmogorov-Smirnov test, $p < 0.05$) across category combinations. To mitigate imbalances in sample sizes across categories, we combine data from freshwater and marine habitats into a single aquatic category and used Type III sums of squares. This merging of data from marine and freshwater habitats is reasonable because there is no significant difference in mean activation energy between them, and because marine and freshwater environments share many physical properties (11). Figure 2 (main text) shows that the significant effect of organization partially arises from the fact that E 's of population traits tend to have higher values than those in other categories. This pattern may also be weakly determined by habitat, as seen by the significant organization x habitat interaction.

Source	Sum Sq.	d.f.	Mean Sq.	F	P-value
Organization	11.238	3	3.746	11.91	0
Habitat	0.025	1	0.02506	0.08	0.7779
Organization × Habitat	2.631	3	0.87692	2.79	0.0408
Error	98.446	313	0.31452		
Total	116	320			

Table S5. Results of ANOVA to test effects on mean optimum temperature, T_{opt} . Methods for this analysis largely follow those for activation energies (Table S4), except that transformation is unnecessary because distributions are approximately normal (two-tailed one-sample Kolmogorov-Smirnov test, $p < 0.05$) across category combinations. Results show a significant effect of habitat and organization, as well as significant interactions between them.

Source	Sum Sq.	d.f.	Mean Sq.	F	P-value
Organization	614.8	3	204.94	5.75	0.0008
Habitat	1780.5	1	1780.48	49.92	0
Organization \times Habitat	324.5	3	108.16	3.03	0.0301
Error	8239.8	231	35.67		
Total	14652.7	238			

Table S6. Data sources. Numbers on left correspond to citation codes (Cit) in Table S3.

1. Aghdam H. R., Fathipour Y., Radjabi G. & Rezapanah M. 2009. Temperature-dependent development and temperature thresholds of codling moth (Lepidoptera: Tortricidae) in Iran. *Environmental Entomology* 38, 885-895.
2. Aldridge D.W., Payne B.S., & Miller, A.C. 1995. Oxygen consumption, nitrogenous excretion, and filtration rates of *Dreissena polymorpha* at acclimation temperatures between 20 and 32C. *Canadian Journal of Fisheries and Aquatic Science* 52, 1761-1767.
3. Ali R.M. 1970. The influence of suspension and temperature on the filtration rate of *Hiattella arctica*. *Marine Biology* 6, 291-302.
4. Angilletta M.J. Jr., Hill, T. & Robson, M.A. 2002. Is physiological performance optimized by thermoregulatory behaviour? A case study of the eastern fence lizard, *Sceloporus undulatus*. *Journal of Thermal Biology* 27, 199-204.
5. Arbab A., Kontodimas D. C. & McNeill M. R. 2008. Modelling embryo development of *Sitona discoideus* Gyllenhal (Coleoptera: Curculionidae) under constant temperature. *Environmental Entomology* 37, 1381-1388.
6. Aspaly G., Stejskal V., Pekar S. & Hubert J. 2007. Temperature-dependent population growth of three species of stored product mites (Acari : Acaridida). *Experimental and Applied Acarology* 42, 37-46.
7. Avery M.I. & Krebs J.R. 1984. Temperature and foraging success of Great Tits *Parus major* hunting for spiders. *IBIS* 126, 33-38.
8. Avery R.A., Bedford J.D. & Newcombe C.P. 1982. The role of thermoregulation in lizard biology: predatory efficiency in a temperate diurnal basker. *Behavioural Ecology and Sociobiology* 11, 261-267.
9. Bailey P.C.E. 1988. The effect of density and temperature on the swimming and aggregating behaviour of the backswimmer, *Anisops deanei* (Heteroptera: Notonectidae) and subsequent encounter rate with a sit-and-wait predator. *Ethology* 77, 44-57.
10. Bailey P.C.E. 1989. The effect of water temperature on the functional response of the water stick insect *Ranatra dispar* (Heteroptera: Nepidae). *Australian Journal of Ecology* 14, 381-386.
11. Ball D. & I.A. Johnston. 1996. Molecular mechanisms underlying the plasticity of muscle contractile properties with temperature acclimation in the marine fish *Myoxocephalus scorpius*. *The Journal of Experimental Biology* 199, 1363-1373.
12. Batty R.S., J.H.S. Blaxter & K. Fretwell. 1993. Effect of temperature on the escape responses of larval herring, *Clupea harengus*. *Marine Biology* 115, 523-528.
13. Batty R.S., Blaxter, J.H.S. & Bone Q. 1991. The effect of temperature on the swimming of a teleost (*Clupea harengus*) and an ascidian larva (*Dendrodoa grossularia*). *Comparative Biochemistry and Physiology. A, Comparative Physiology* 100(2), 297-300.
14. Bauwens D., Garland T., Castilla A.M. & van Damme R. 1995. Evolution of sprint speed in lacertid lizards: morphological, physiological and behavioural co-variation. *Evolution* 49(5), 848-863.
15. Beamish F.W.H. 1981. Swimming performance and metabolic rate of three tropical fishes in relation to temperature. *Hydrobiologia* 83, 245-254.
16. Benider, A., Tifnouti A. & Pourriot R. 2002. Growth of *Moina macrocopa* (Straus 1820) (Crustacea, Cladocera): influence of trophic conditions, population density and temperature. *Hydrobiologia* 468, 1-11.
17. Bennett A.F. 1980. The thermal dependence of lizard behaviour. *Animal Behaviour* 28, 752-762.
18. Bergman E. 1987. Temperature-dependent differences in foraging ability of two percids, *Perca fluviatilis* and *Gymnocephalus cernuus*. *Environmental Biology of Fishes* 19(1), 45-53.

19. Bergmann P. & Irschick D.J. 2006. Effects of temperature on maximum acceleration, deceleration and power output during vertical running in geckos. *The Journal of Experimental Biology* 209, 1404-1412.
20. Bouterfas R., Belkoura M. & Dauta A. 2002. Light and temperature effects on the growth rate of three freshwater algae isolated from a eutrophic lake. *Hydrobiologia* 489, 207-217.
21. Boyden C.R. & Zeldis J.R. 1979. Preliminary observations using an attached microphonic sensor to study feeding behaviour of an intertidal limpet. *Estuarine and Coastal Marine Science* 9, 759-769.
22. Brett J.R. 1967. Swimming performance of sockeye salmon (*Oncorhynchus nerka*) in relation to fatigue time and temperature. *Journal of the Fisheries Research Board of Canada* 24(8), 1731-1741.
23. Brock V. & Kofoed L.H. 1987. Species specific irrigatory efficiency in *Cardium (Cerastoderm) edule* (L.) and *C. lamarcki* (Reeve) responding to different environmental temperatures. *Biological Oceanography* 4(3), 211-226.
24. Bronikowski A.M., Bennett A.F. & Lenski R.E. 2001. Evolutionary adaptation to temperature. VII. Effects of temperature on growth rate in natural isolates of *Escherichia coli* and *Salmonella enterica* from different thermal environments. *Evolution* 55, 33-40.
25. Bruce A.Y., Schulthess F. & Mueke J. 2009. Host acceptance, suitability, and effects of host deprivation on the West African egg parasitoid *Telenomus isis* (Hymenoptera: Scelionidae) reared on East African stem-borers under varying temperature and relative humidity regimes. *Environmental Entomology* 38(3), 904-919.
26. Caron D.A., Goldman J.C. & Dennett M.R. 1986. Effect of temperature on growth, respiration, and nutrient regeneration by an omnivorous microflagellate. *Applied and Environmental Microbiology* 52(1340), 1347.
27. Casey T.M. 1976. Activity patterns, body temperature and thermal ecology in two desert caterpillars (Lepidoptera: Sphingidae). *Ecology* 57(3), 485-497.
28. Castro-Arevalo M., Reyes-Diaz M., Alberdi M., Jara-Rodriguez V., Sanhueza C., Corcuera L. J. & Bravo L. A. 2008. Effects of low temperature acclimation on photosynthesis in three Chilean Proteaceae. *Revista Chilena De Historia Natural* 81, 321-333.
29. Chadwick L.E. & H. Rahn. 1954. Temperature dependence of rattling frequency in the rattlesnake, *Crotalus v. viridis*. *Science* 119(3092), 442-443.
30. Chen S.L., Wu J.W., Huner J. V. & Malone R. F. 1995. Effects of temperature upon ablation-to-molt interval and mortality of red swamp crawfish (*Procambarus clarkii*) subjected to bilateral eyestalk ablation. *Aquaculture* 138, 191-204.
31. Chiverton P.A. 1988. Searching behaviour and cereal aphid consumption by *Bembidion lampros* and *Pterostichus cupreus*, in relation to temperature and prey density. *Entomologia experimentalis et applicata* 47, 173-182.
32. Christian K.A. & Tracy R. 1981. The effect of the thermal environment on the ability of hatchling Galapagos land iguanas to avoid predation during dispersal. *Oecologia* 49, 218-223.
33. Clopton J.R. 2007. Temperature: human regulating, ants conforming. *American Biology Teacher* 69(5), 59-63.
34. Cockrell B.J. 1984. Effects of temperature and oxygenation on predator-prey overlap and prey choice of *Notonecta glauca*. *Journal of Animal Ecology* 53, 519-532.
35. Cooper W.E. Jr. 2000. Effect of temperature on escape behaviour by an ectothermic vertebrate, the keeled earless lizard (*Holobrookia propinqua*). *Behaviour* 137, 1299-1315.
36. Crist T.O. & MacMahon J.A. 1991. Foraging patterns of *Pogonomyrmex occidentalis* (Hymenoptera: Formicidae) in a shrub-steppe ecosystem: the roles of temperature, trunk trails, and seed resources. *Environmental Entomology* 20(1), 265-275.

37. Dam H.G. & Peterson W.T. 1988. The effect of temperature on the gut clearance rate constant of planktonic copepods. *Journal of Experimental Marine Biology and Ecology* 123, 1-14.
38. Dauta, A., Devaux, J., Piquemal, F. & Boumnic, L. 1990. Growth-rate of 4 fresh-water algae in relation to light and temperature. *Hydrobiologia* 207, 221-226.
39. Dickson I.W. & Kramer R.H. 1971. Factors influencing scope for activity and active standard metabolism of rainbow trout (*Salmo gairdneri*). *Journal of the Fisheries Research Board of Canada* 28, 587-596.
40. Dillon T.M. 1977. Effects of acute changes in temperature and salinity on pulsation rates in ephyrae of the Scyphozoan *Aurelia aurita*. *Marine Biology* 42, 31-35.
41. Dmi'el R. & Rapoport D. 1976. Effect of temperature on metabolism during running in the lizard *Uromastix aegyptius*. *Physiological Zoology* 49, 77-84.
42. Dreisig H. 1981. The rate of predation and its temperature dependence in a tiger beetle, *Cicindela hybrida*. *Oikos* 36, 196-202.
43. Du W.G., Hu L.J., Lu J.L. & Zhu L.J. 2007. Effects of incubation temperature on embryonic development rate, sex ratio and post-hatching growth in the Chinese three-keeled pond turtle, *Chinemys reevesii*. *Aquaculture* 272, 747-753.
44. Duncan F.D. & Crewe R.M. 1993. A comparison of the energetics of foraging of three species of *Leptogenys* (Hymenoptera, Formicidae). *Physiological Entomology* 18, 372-378.
45. Edwards D.C & Huebner J.D. 1977. Feeding and growth rates of *Polinies duplicatus* preying on *Mya arenaria* at Barnstable Harbor, Massachusetts. *Ecology* 58(6), 1218-1236.
46. Edwards D.J. 1971. Effect of temperature on rate of passage of food through the alimentary canal of the plaice *Pleuronectes platessa* L. *Journal of Fish Biology* 3, 433-439.
47. Elliott J.M. 1972. Rates of gastric evacuation in brown trout, *Salmo trutta* L. *Freshwater Biology* 2, 1-18.
48. Everson P. 1980. The relative activity and functional response of *Phytoseiulus persimilis* (Acarina: Phytoseiidae) and *Tetranychus urticae* (Acarina: Tetranychidae): the effect of temperature. *The Canadian Entomologist* 112, 17-24.
49. Fargues J., Goettel M. S., Smits N., Ouedraogo A. & Rougier M. 1997. Effect of temperature on vegetative growth of *Beauveria bassiana* isolates from different origins. *Mycologia* 89, 383-392.
50. Fedorenko A.Y. 1975. Feeding characteristic and predation impact of Chaoborus (Diptera, Chaoboridae) larvae in a small lake. *Limnology and Oceanography* 20(2), 250-258.
51. Feng S.Y. & W. van Winkle. 1975. The effect of temperature and salinity on the heart beat of *Crassostrea virginica*. *Comparative Biochemistry and Physiology. A, Comparative Physiology* 50, 473-476.
52. Flinn P.W. & Hagstrum D.W. 2002. Temperature-mediated functional response of *Theocolax elegans* (Hymenoptera: Pteromalidae) parasitizing *Rhyzopertha dominica* (Coleoptera: Bostrichidae) in stored wheat. *Journal of Stored Products Research* 38, 185-190.
53. Flinn P.W. 1991. Temperature-dependent functional response of the parasitoid *Cephalonomia waterstoni* (Gahan) (Hymenoptera: Bethyridae) attacking rusty grain beetle larvae (Coleoptera: Cucujidae). *Environmental Entomology* 20(3), 872-876.
54. Foerster L.A., Avanci M.R.F. & Doetzer A.K. 1999. Effect of temperature on the development and progeny production of *Glyptapanteles muesebecki* (Blanchard) (Hymenoptera: Braconidae) parasitizing larvae of *Pseudaletia sequax* Franclemont (Lepidoptera: Noctuidae). *Anais da Sociedade Entomologica do Brasil* 28, 243-249.
55. Foyle T.P., O'Dor R.K., & Elnor R.W. 1989. Energetically defining the thermal limits of the snow crab. *Journal of Experimental Biology* 145, 371-393.
56. Fuiman L.A. 1986. Burst-swimming performance of larval zebra danios and the effects of diel temperature fluctuations. *Transactions of the American Fisheries Society* 115, 143-148.

57. Gallepp G.W. 1977. Responses of caddisfly larvae (*Brachycentrus* spp.) to temperature, food availability and current velocity. *American Midland Naturalist* 98(1), 59-84.
58. Garton D. & Stickle W.B. 1980. Effects of salinity and temperature on the predation rate of *Thais haemastoma* on *Crassostrea virginica* spat. *Biological Bulletin* 158, 49-57.
59. Geden C.J. & Axtell R.C. 1988. Predation by *Carcinops pumilio* (Coleoptera: Histeridae) and *Macrocheles muscaedomesticae* (Acarina: Macrochelidae) on the house fly (Diptera: Muscidae): Functional response, effects of temperature, and availability of alternative prey. *Environmental Entomology* 17(4), 739-744.
60. Gilchrist G.W. 1996. A quantitative genetic analysis of thermal sensitivity in the locomotor performance curve of *Aphidius ervi*. *Evolution* 50(4), 1560-1572.
61. Goldasteh S., Talebi A.A., Fathipour Y., Ostovan H., Zamani A. & Shoushtari R. V. 2009. Effect of temperature on life history and population growth parameters of *Planococcus citri* (Homoptera, Pseudococcidae) on *Coleus (Solenostemon scutellarioides* (L.) Codd). *Archives of Biological Sciences* 61, 329-336.
62. Gordon D.C. Jr. 1966. The effects of the deposit feeding polychaete *Pectinaria gouldii* on the intertidal sediments of Barnstable Harbor. *Limnology and Oceanography* 11(3), 327-332.
63. Graham W.D., Thorpe J.E. & Metcalfe N.B. 1996. Seasonal current holding performance of juvenile Atlantic salmon in relation to temperature and smolting. *Canadian Journal of Fisheries and Aquatic Science* 53, 80-86.
64. Gray J. 1923. The mechanism of ciliary movement. III. the effect of temperature. *Proceedings of the Royal Society of London, Series B* 95(664), 6-15.
65. Greenwald O.E. 1974. Thermal dependence of striking and prey capture by gopher snakes. *Copeia* 1974(1), 141-148.
66. Gresens S.E., Cothran M.L. & Thorp. J.H. 1982. The influence of temperature on the functional response of the dragonfly *Celithemis fasciata* (Odonata: Libellulidae). *Oecologia* 53, 281:284.
67. Hailey A. & Coulson I.M. 1996. Temperature and the tropical tortoise *Kinixys spekii*: constraints on activity level and body temperature. *Journal of Zoology* 240, 523-536.
68. Hailey A. & Davies P.M.C. 1986. Effects of size, sex, temperature and condition on activity metabolism and defence behaviour of the viperine snake, *Natrix maura*. *Journal of Zoology* 208, 541-558.
69. Hancke K., Hancke T. B., Olsen L. M., Johnsen G. & Glud R. N. 2008. Temperature effects on microalgal photosynthesis-light responses measured by O-2 production, pulse-amplitude-modulated fluorescence, and C-14 assimilation. *Journal of Phycology* 44, 501-514.
70. Hanks J.E. 1957. The rate of feeding of the common Oyster (bivalve) drill, *Urosalpinx cinera* (Say), at controlled water temperatures. *Biological Bulletin* 112, 330-335.
71. Harkness R.D. 1979. The speed of walking of *Cataglyphis bicolor* (F.) (Hym., Formicidae). *Entomologist's Monthly Magazine* 114, 203-209.
72. Harri M.N.E. & Talo A. 1975. Effect of season and temperature acclimation on the heart rate-temperature relationship in the frog, *Rana temporaria*. *Comparative Biochemistry and Physiology. A, Comparative Physiology* 50, 469-472.
73. Heckrotte C. 1967. Relations of body-temperature, size, and crawling speed of the common garter snake, *Thamnophis s. sirtalis*. *Copeia* 1967(4), 759-763.
74. Heeger T. & Moller H. 1987. Ultrastructural observations on prey capture and digestion in the scyphomedusa *Aurelia aurita*. *Marine Biology* 96, 391-400.
75. Heiman D.R. & Knight A.W. 1975. The influence of temperature on the bioenergetics of the carnivorous stonefly nymph, *Acroneuria californica* Banks (Plecoptera: Perlidae). *Ecology* 56, 105-116.

76. Hertz P.E., Huey R.B. & Nevo E. 1982. Flight versus fight: body temperature influences defensive responses of lizards. *Animal Behaviour* 30, 676-679.
77. Hirano M. & Rome L.C. 1984. Jumping performance of frogs (*Rana pipiens*) as a function of muscle temperature. *Journal of Experimental Biology* 108, 429-439.
78. Hocutt C.H. 1973. Swimming performance of three warm-water fishes exposed to a rapid temperature change. *Chesapeake Science* 14(1), 11-16.
79. Holt S.J. 1955. On the foraging activity of the wood ant. *Journal of Animal Ecology* 24, 1-34.
80. Huey R.B. & Stevenson R.D. 1979. Integrating thermal physiology and ecology of ectotherms: a discussion of approaches. *American Zoologist* 19, 357-366.
81. Huey R.B., Niewiarowski P.H., Kaufmann J. & Herron J.C. 1989. Thermal biology of nocturnal ectotherms; is sprint performance of geckos maximal at low body temperatures? *Physiological Zoology* 62(2), 488-504.
82. Hull L.A., Asquith D. & Mowery P.D. 1977. The functional response of *Stethorus punctum* to densities of the European red mite. *Environmental Entomology* 6(1), 85-90.
83. Hurlbert A., Ballantyne F. & Powell S. 2008. Shaking a leg and hot to trot: the effects of body size and temperature on running speed in ants. *Ecological Entomology* 33, 144-154.
84. Imamura T., Uraichuen J., Visarathanonth P., Morimoto S. & Miyanoshita A. 2004. Effect of temperature on development of *Theocolax elegans* (Westwood) (Hymenoptera : Pteromalidae) parasitizing larvae of the maize weevil *Sitophilus zeamais* (Coleoptera : Curculionidae) in brown rice. *Applied Entomology and Zoology* 39, 497-503.
85. Jobling M. & Davies P.S. 1979. Gastric evacuation in plaice, *Pleuronectes platessa* L.: effects of temperature and meal size. *Journal of Fish Biology* 14, 539-546.
86. Jobling M. 1980. Gastric evacuation in plaice, *Pleuronectes platessa* L.: effects of temperature and fish size. *Journal of Fish Biology* 17, 547-551.
87. Johnson T.P. & Bennett A.F. 1995. The thermal acclimation of burst escape performance in fish: an integrated study of molecular and cellular physiology and organismal performance. *The Journal of Experimental Biology* 198, 2165-2175.
88. Jorgensen C.B. 1975. On gill function in the mussel *Mytilus edulis* L. *Ophelia* 13, 187-232.
89. Jorgensen C.B., Larsen P.S. & Riisgard H.U. 1990. Effects of temperature on the mussel pump. *Marine Ecology Progress Series* 64, 89-97.
90. Kalyebi A., Overholt W.A., Schulthess F., Mueke J.M. & Sithanatham S. 2006. The effect of temperature and humidity on the bionomics of six African egg parasitoids (Hymenoptera : Trichogrammatidae). *Bulletin of Entomological Research* 96, 305-314.
91. Kibby H.V. 1971. Effect of temperature on the feeding behaviour of *Daphnia rosea*. *Limnology and Oceanography* 16(3), 580-581.
92. Kiorboe T., Mohlenberg F. & Nicolajsen H. 1982. Ingestion rate and gut clearance in the planktonic copepod *Centropages hamatus* (Lilljeborg) in relation to food concentration and temperature. *Ophelia* 21(2), 181-194.
93. Kishi D., Murakami M., Nakano S. & Maekawa K. 2005. Water temperature determines strength of top-down control in a stream food web. *Freshwater Biology* 50, 1315-1322.
94. Kittner C. & Riisgard H.U. 2005. Effect of temperature on filtration rate in the mussel *Mytilus edulis*: no evidence for temperature compensation. *Marine Ecology Progress Series* 305, 147-152.
95. Kivivuori L. 1983. Temperature acclimation of walking in the crayfish *Astacus astacus* L. *Comparative Biochemistry and Physiology. A, Comparative Physiology* 75A(3), 375-378.
96. Knowles T.W. & Weigl P.D. 1990. Thermal dependence of anuran burst locomotor performance. *Copeia* 1990(3), 796-802.

97. Krugner R., Daane K.M., Lawson A.B. & Yokota G.Y. 2007. Temperature-dependent development of *Macrocentrus iridescens* (Hymenoptera : Braconidae) as a parasitoid of the oblique banded leafroller (Lepidoptera : Tortricidae): Implications for field synchrony of parasitoid and host. *Biological Control* 42, 110-118.
98. Kuo M.H., Lu W.N., Chu M.C., Kuo Y.H. & Havang S.H. 2006. Temperature-dependent development and population growth of *Tetraneura nigriabdominalis* (Homoptera : Pemphigidae) on three host plants. *Journal of Economic Entomology* 99, 1209-1213.
99. Largen M.J. 1967. The influence of water temperature upon the life of the dog-whelk *Thais lapillus* (Gastropoda: Prosobranchia). *Journal of Animal Ecology* 36(1), 207-214.
100. Larimore R.W. & Duever M.J. 1968. Effects of temperature acclimation on the swimming ability of smallmouth bass fry. *Transactions of the American Fisheries Society* 97, 175-184.
101. Lasfar S., Monette F., Millette L. & Azzouz A. 2007. Intrinsic growth rate: A new approach to evaluate the effects of temperature, photoperiod and phosphorus-nitrogen concentrations on duckweed growth under controlled eutrophication. *Water Research* 41, 2333-2340.
102. Ledoux P.A. 1967. Action de la temperature sur l'activited' *Aphaenogaster senilis* (Testaceo-Pilosa) Mayr (Hym. Formicoidea). *Insectes Sociaux* 14, 131-156.
103. Lei J., Payne B.S. & Wang. S.Y. 1996. Filtration dynamics of the zebra mussel, *Dreissena polymorpha*. *Canadian Journal of Fisheries and Aquatic Science* 53, 29-37.
104. Lemons D.E. & Crawshaw L.I. 1985. Behavioural and metabolic adjustments to low temperatures in the largemouth bass (*Micropterus salmoides*). *Physiological Zoology* 58(2), 175-180.
105. Lenz M., Barrett R.A. & Williams E.R.. 1982. Influence of diet on the survival and wood consumption of *Porotermes adamsoni* (Froggatt) (Isoptera: Termopsidae) at different temperatures. *Bulletin of Entomological Research* 72, 423-435.
106. Lenz P.H., Hower A.E. & Hartline D.K. 2005. Temperature compensation in the escape response of a marine copepod, *Calanus finmarchicus* (Crustacea). *Biological Bulletin* 209, 75-85.
107. Li D.M., Wang G.C., Chen L.M., Lu F. & Shen Z.G. 2009. Effects of irradiance and temperature on the photosynthesis and vegetative propagation of *Caulerpa serrulata*. *Journal of Integrative Plant Biology* 51, 147-154.
108. Lighton J.R.B. & Duncan F.D. 2002. Energy cost of locomotion: validation of laboratory data by in situ respirometry. *Ecology* 83, 3517-3522.
109. Linley J.R. 1986. Swimming behaviour of the larva of *Culicoides variipennis* (Diptera: Ceratopogonidae) and its relationship to temperature and velocity. *Journal of Medical Entomology* 23(5), 473-483.
110. Lisbjerg D. & Petersen J.K. 2000. Clearance capacity of *Electra bellula* (Bryozoa) in seagrass meadows of Western Australia. *Journal of Experimental Marine Biology and Ecology* 244, 285-296.
111. Lisbjerg D. & Petersen J.K. 2001. Feeding activity, retention efficiency, and effects of temperature and particle concentration on clearance rate in the marine bryozoan *Electra crustulenta*. *Marine Ecology Progress Series* 215, 133-141.
112. Liu X. F. & Ye H. 2009. Effect of temperature on development and survival of *Bactrocera correcta* (Diptera: Tephritidae). *Scientific Research and Essays* 4, 467-472.
113. Londos P.L. & Brooks R.J. 1988. Effect of temperature acclimation on locomotory performance curves in the toad, *Bufo woodhousii woodhousii*. *Copeia* 1988(1), 26-32.
114. Ma X.P., & Purcell J.E. 2005. Effects of temperature, salinity, and predators on mortality of and colonization by the invasive hydrozoan *Moerisia lyonsi*. *Marine Biology* 147, 215-224.
115. Marsh A.C. 1985. Microclimatic factors influencing foraging patterns and success of the thermophilic desert ant, *Ocymyrmex barbiger*. *Insectes Sociaux* 32, 286-296.

116. Marsh R.L. & Bennett A.F. 1986. Thermal dependence of sprint performance of the lizard *Sceloporus occidentalis*. *Journal of Experimental Biology* 126, 79-87.
117. McCaffrey J.P. & Horsburgh R.L. 1986. Functional response of *Orius insidiosus* (Hemiptera: Anthocoridae) to the European red mite, *Panonychus ulmi* (Acari: Tetranychidae), at different constant temperatures. *Environmental Entomology* 15, 532-535.
118. McCoull C.J., Swain R. & Barnes R.W. 1998. Effect of temperature on the functional response and components of attack rate in *Naucoris congrex* Stal (Hemiptera: Naucoridae). *Australian Journal of Entomology* 37, 323-327.
119. McKee D. 1995. Long-term temperature acclimation in *Daphnia magna*: effects on filtering rates. *Journal of Plankton Research* 17(5), 1095-1103.
120. McLeese D.W. & Wilder D.G. 1958. The activity and catchability of the lobster (*Homarus americanus*) in relation to temperature. *Journal of the Fisheries Research Board of Canada* 15(6), 1345-1354.
121. Meade M.E., Doeller J.E., Kraus D.W. & Watts S.A. 2002. Effects of temperature and salinity on weight gain, oxygen consumption rate, and growth efficiency in juvenile red-claw crayfish *Cherax quadricarinatus*. *Journal of the World Aquaculture Society* 33, 188-198.
122. Menon A., Flinn P.W. & Dover B.A. 2002. Influence of temperature on the functional response of *Anisopteromalus calandrae* (Hymenoptera: Pteromalidae), a parasitoid of *Rhyzopertha dominica* (Coleoptera: Bostrichidae). *Journal of Stored Products Research* 38, 463-469.
123. Menon N.R. 1974. Clearance rates of food suspension and food passage rates as a function of temperature in two North-Sea bryozoans. *Marine Biology* 24, 65-67.
124. Messenger P.S. 1968. Bioclimatic studies of the aphid parasite *Praon exsoletum* 1. Effects of temperature on the functional response of females to varying host densities. *The Canadian Entomologist* 100, 728-741.
125. Milione M. & Zeng C.S. 2008. The effects of temperature and salinity on population growth and egg hatching success of the tropical calanoid copepod, *Acartia sinjiensis*. *Aquaculture* 275, 116-123.
126. Miller K. 1982. Effect of temperature on sprint performance in the frog *Xenopus laevis* and the salamander *Necturus maculosus*. *Copeia* 1982(3), 695-698.
127. Montgomery J.C., McVean A.R. & McCarthy D. 1983. The effects of lowered temperature on spontaneous eye movements in a teleost fish. *Comparative Biochemistry and Physiology. A, Comparative Physiology* 3, 363-368.
128. Morehead S.A. & Feener D.H. 1998. Foraging behaviour and morphology: seed selection in the harvester ant genus, *Pogonomyrmex*. *Oecologia* 114, 548-555.
129. Murdoch W.W., Scott M.A. & Ebsworth P. 1984. Effects of the general predator, *Notonecta* (Hemiptera) upon a freshwater community. *Journal of Animal Ecology* 53, 791-808.
130. Navas C.A. 1996. Metabolic physiology, locomotor performance, and thermal niche breadth in neotropical anurans. *Physiological Zoology* 69(6), 1481-1501.
131. Nebel S. & Thompson G.J. 2005. Foraging behaviour of western sandpipers changes with sediment temperature: implications for their hemispheric distribution. *Ecological Research* 20, 503-507.
132. Newell R.C., Johnson L.G. & Kofoed L.H. 1977. Adjustment of the components of energy balance in response to temperature change in *Ostrea edulis*. *Oecologia* 30, 97-110.
133. Newell R.C., Pye V.I. & Ahsanullah M. 1971. Factors affecting the feeding rate of the winkle *Littorina littorea*. *Marine Biology* 9, 138-144.
134. Oertli J.J. 1989. Relationship of wing beat frequency and temperature during take-off flight in temperate-zone beetles. *Journal of Experimental Biology* 145, 321-338.

135. O'Steen S. & Bennett A.F. 2003. Thermal acclimation effects differ between voluntary, maximum, and critical swimming velocities in two cyprinid fishes. *Physiological and Biochemical Zoology* 76(4), 484-496.
136. Ouedraogo A., Fargues J., Goettel M.S. & Lomer C.J. 1997. Effect of temperature on vegetative growth among isolates of *Metarhizium anisopliae* and *M. flavoviride*. *Mycopathologia* 137, 37-43.
137. Patersen J.K., Mayer S. & Knudsen M.A. 1999. Beat frequency of cilia in the branchial basket of the ascidian *Ciona intestinalis* in relation to temperature and algal cell concentration. *Marine Biology* 133, 185-192.
138. Person-Le Ruyet J., Mahe K., Le Bayon N. & Le Delliou H. 2004. Effects of temperature on growth and metabolism in a Mediterranean population of European sea bass, *Dicentrarchus labrax*. *Aquaculture* 237, 269-280.
139. Persson L. 1979. The effects of temperature and different food organisms on the rate of gastric evacuation in perch (*Perca fluviatilis*). *Freshwater Biology* 9, 99-104.
140. Persson L. 1986. Temperature-induced shift in foraging ability in two fish species, roach (*Rutilus rutilus*) and perch (*Perca fluviatilis*): implications for coexistence between poikilotherms. *Journal of Animal Ecology* 55, 829-839.
141. Petersen J.K. & Riisgard H.U. 1992. Filtration capacity of the ascidian *Ciona intestinalis* and its grazing impact in a shallow fjord. *Marine Ecology Progress Series* 88, 9-17.
142. Peterson R.H. & Anderson J.M. 1969. Influence of temperature change on spontaneous locomotor activity and oxygen consumption of Atlantic salmon, *Salmo salar*, acclimated to two temperatures. *Journal of the Fisheries Research Board of Canada* 26, 93-109.
143. Putnam R.W. & Bennett A.F. 1981. Thermal dependence of behavioural performance of anuran amphibians. *Animal Behaviour* 29, 502-509.
144. Rand A.S. 1964. Inverse relationship between temperature and shyness in the lizard *Anolis lineatopus*. *Ecology* 45(4), 863-864.
145. Rao K.P. 1953. Rate of water propulsion in *Mytilus californianus* as a function of latitude. *Biological Bulletin* 104(2), 171-181.
146. Renaud J.M. & Stevens E.D. 1983. The extent of long-term temperature compensation for jumping distance in the frog, *Rana pipiens*, and the toad, *Bufo americanus*. *Canadian Journal of Zoology* 61, 1284-1287.
147. Reynolds W.W. & Casterlin M.E. 1979. Behavioural thermoregulation and activity in *Homarus americanus*. *Comparative Biochemistry and Physiology. A, Comparative Physiology* 64, 25-28.
148. Rigatuso R., Bertoluzzo S.M.R., Quattrin F.E. & Bertoluzzo M.G. 2000. Ant activity associated with a chemical compound. *Journal of Chemical Education* 77, 183-185.
149. Riisgard H.U. & Seerup D.F. 2003. Filtration rates in the soft clam *Mya arenaria*: effects of temperature and body size. *Sarsia* 88, 415-428.
150. Riisgard H.U. & Ivarsson N.M. 1990. The crown-filament pump of the suspension-feeding polychaete *Sabella penicillus*: filtration, effects of temperature, and energy cost. *Marine Ecology Progress Series* 62, 249-257.
151. Riisgard H.U., Thomassen S., Jakobsen H., Weeks J.M. & Larsen P.S. 1993. Suspension feeding in marine sponges *Halichondria panicea* and *Haliclona urseolus*: effects of temperature on filtration rate and energy cost of pumping. *Marine Ecology Progress Series* 96, 177-188.
152. Riisgard H.U., Vedel A., Boye H. & Larsen P.S. 1992. Filter-net structure and pumping activity in the polychaete *Nereis diversicolor*: effects of temperature and pump-modeling. *Marine Ecology Progress Series* 83, 79-89.
153. Rissing S.W. 1982. Foraging velocity of seed-harvester ants, *Veromessor pergandei* (Hymenoptera: Formicidae). *Environmental Entomology* 11, 905-907.

154. Rose J.M., Vora N.M., Countway P.D., Gast R.J. & Caron D.A. 2009. Effects of temperature on growth rate and gross growth efficiency of an Antarctic bacterivorous protist. *Isme Journal* 3, 252-260.
155. Ruberson J.R., Tauber C.A. & Tauber M.J. 1995. Developmental effects of host and temperature on *Telenomus* spp (Hymenoptera, Scelionidae) parasitizing Chrysopid eggs. *Biological Control* 5, 245-250.
156. Runjie Z, Heong K.H. & Domingo I.T. 1996. Relationship between temperature and functional response in *Cardiochiles philippinensis* (Hymenoptera: Braconidae), a larvae parasitoid of *Cnaphalocrocis medinalis* (Lepidoptera: Pyralidae). *Environmental Entomology* 25(6), 1321-1324.
157. Shapley H. 1920. Thermokinetics of *Liometopum apiculatum* Mayr. *Proceedings of the National Academy of Sciences of the United States of America* 6, 204-211.
158. Shapley H. 1924. Note on the thermokinetics of Dolichoderine ants. *Proceedings of the National Academy of Sciences of the United States of America* 10, 436-439.
159. Smith D.G. 1997. Ecological factors influencing the antipredator behaviours of the ground skink, *Scincella lateralis*. *Behavioural Ecology* 8(6), 622-629.
160. Smith G.R. & Lemos-Espinal J.A. 2005. Comparative escape behaviour of four species of Mexican phrynosomatid lizards. *Herpetologia* 61(3), 225-232.
161. Smith L. 1994. Temperature influences functional response of *Anisopteromalus calandrae* (Hymenoptera: Pteromalidae) parasitizing maize weevil larvae in shelled corn. *Annals of the Entomological Society of America* 87(6), 849-855.
162. Sopp P. & Wratten S.D. 1986. Rates of consumption of cereal aphids by some polyphagous predators in the laboratory. *Entomologia experimentalis et applicata* 41, 69-73.
163. Steigenberger L.W. & Larkin P.A. 1974. Feeding activity and rates of digestion of northern squawfish (*Ptychocheilus oregonensis*). *Journal of the Fisheries Research Board of Canada* 31, 411-420.
164. Steller D.L., Hernandez-Ayon J.M., Riosmena-Rodriguez R. & Cabello-Pasini A. 2007. Effect of temperature on photosynthesis, growth and calcification rates of the free-living coralline alga *Lithophyllum margaritae*. *Ciencias Marinas* 33, 441-456.
165. Stevens A.J., Welch Z.C., Darby P.C. & Percival H.F. 2002. Temperature effects on Florida applesnail activity: implications for snail kite foraging success and distribution. *Wildlife Society Bulletin* 30(1), 75-81.
166. Stevenson R.D. & Josephson R.K. 1990. Effects of operating frequency and temperature on mechanical power output from moth flight muscle. *Journal of Experimental Biology* 149, 61-78.
167. Stevenson R.D., Peterson C.R. & Tsuji J. 1985. The thermal dependence of locomotion, tongue flicking, digestion, and oxygen consumption in the wandering garter snake. *Physiological Zoology* 58(1), 46-57.
168. Stinson C.H. 1980. Weather-dependent foraging success and sibling aggression in Red-Tailed Hawks in central Washington. *The Condor* 82(1), 76-80.
169. Swift M.C. & Fedorenko A.Y. 1975. Some aspects of prey capture by *Chaoborus* larvae. *Limnology and Oceanography* 20(3), 418-425.
170. Temple G.K. & Johnston I.A. 1998. Testing hypotheses concerning the phenotypic plasticity of escape performance in fish of the family Cottidae. *The Journal of Experimental Biology* 201, 317-331.
171. Thompson D.J. 1978. Towards a realistic predator-prey model: the effect of temperature on the functional response and life history of larvae of the damselfly, *Ischnura elegans*. *Journal of Animal Ecology* 47, 757-767.

172. Torres-Contreras H. & Vasquez R.A. 2004. A field experiment on the influence of load transportation and patch distance on the locomotion velocity of *Dorymyrmex goetschi* (Hymenoptera, Formicidae). *Insectes Sociaux* 51, 265-270.
173. Traniello J.F.A., Fujita M.S. & Bowen R.V. 1984. Ant foraging behaviour: ambient temperature influences prey selection. *Behavioural Ecology and Sociobiology* 15, 65-68.
174. Tyler A.V. 1970. Rates of gastric emptying in young cod. *Journal of the Fisheries Research Board of Canada* 27, 1177-1189.
175. van Damme R., Bauwens D., Castilla A.M. & Verheyen R.F. 1989. Altitudinal variation of the thermal biology and running performance in the lizard *Podarcis tiliguerta*. *Oecologia* 80, 516-524.
176. Vidal C., Fargues J. & Lacey L. A. 1997. Intraspecific variability of *Paecilomyces fumosoroseus*: Effect of temperature on vegetative growth. *Journal of Invertebrate Pathology* 70, 18-26.
177. Waldschmidt S. & Tracy C.R. 1983. Interactions between a lizard and its thermal environment: implications for sprint performance and space utilization in the lizard *Uta stansburiana*. *Ecology* 64(3), 476-484.
178. Waldschmidt S., Jones S.M. & Porter W.R. 1986. The effect of body temperature and feeding regime on activity, passage time, and digestive coefficient in the lizard *Uta stansburiana*. *Physiological Zoology* 59(3), 376-383.
179. Walne P.R. & Dean G.J. 1972. Experiments on predation by the shore crab, *Carcinus maenas* L., on *Mytilus* and *Mercenaria*. *Journal de Conseil International pour l'Exploration de la Mer* 34(2), 190-199.
180. Ware D.M. 1973. Risk of epigenetic prey to predation by rainbow trout (*Salmo gairdneri*). *Journal of the Fisheries Research Board of Canada* 30, 787-797.
181. Warren, C.R. 2008. Does growth temperature affect the temperature responses of photosynthesis and internal conductance to CO₂? A test with *Eucalyptus regnans*. *Tree Physiology* 28, 11-19.
182. Weier J.A. & Feener D.H. 1995. Foraging in the seed-harvester ant genus *Pogonomyrmex*: are energy costs important? *Behavioral Ecology and Sociobiology* 36, 291-300.
183. Weisse T., Stadler P., Lindstrom E.S., Kimmance S.A. & Montagnes D.J.S. 2002. Interactive effect of temperature and food concentration on growth rate: A test case using the small freshwater ciliate *Urotricha farcta*. *Limnology and Oceanography* 47, 1447-1455.
184. Whitehead P.J., Puckridge J.T., Leigh C.M. & Seymour R.S. 1989. Effect of temperature on jump performance of the frog *Limnodynastes tasmaniensis*. *Physiological Zoology* 62(4), 937-949.
185. Widdows J. 1973. Effect of temperature and food on the heart beat, ventilation rate and oxygen uptake of *Mytilus edulis*. *Marine Biology* 20, 269-276.
186. Widdows J. 1978. Combined effects of body size, food concentration and season on the physiology of *Mytilus edulis*. *Journal of the Marine Biology Association of the United Kingdom* 58, 109-124.
187. Williams J.B. & Ricklefs R.E. 1984. Egg temperature and embryo metabolism in some high-latitude Procellariiform birds. *Physiological Zoology* 57(1), 118-127.
188. Wittmann C. & Pfanz H. 2007. Temperature dependency of bark photosynthesis in beech (*Fagus sylvatica* L.) and birch (*Betula pendula* Roth.) trees. *Journal of Experimental Botany* 58, 4293-4306.
189. Yamamoto A.C., Doetzer A.K. & Foerster L.A. 1998. Effect of temperature on the development of *Euplectrus ronnai* (Brethes) (Hymenoptera, Eulophidae) parasitizing *Pseudaletia sequax* Franclemont (Lepidoptera, Noctuidae) and impact of parasitism on food. *Acta Biológica Paranaense* 27, 85-95.

190. Yocom T.G. & Edsall T.A. 1974. Effect of acclimation temperature and heat shock on vulnerability of fry of lake whitefish (*Coregonus clupeaformis*) to predation. Journal of the Fisheries Research Board of Canada 31, 1503-1506.
191. Zamani A.A., Talebi A.A., Fathipour Y. & Baniamiri V. 2006. Effect of temperature on biology and population growth parameters of *Aphis gossypii* Glover (Hom., Aphididae) on greenhouse cucumber. Journal of Applied Entomology 130, 453-460.
192. Zimmerman M.C. & Wissing T.E. 1978. Effects of temperature on gut-loading and gut-clearing times of the burrowing mayfly, *Hexagenia limbata*. Freshwater Biology 8, 269-277.