

Supporting Information:

IR and UV Photodissociation as Analytical Tools for Characterizing Lipid A Structures

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Experimental Section (Supporting Information)

Materials.

Strain MST10 contains in-frame gene deletions in *lpxT* (*yeiU*) and *eptA*, and harbors two plasmids pAChp0021 and pWScj0256. To generate strain MST10 an in-frame deletion of *eptA* was introduced into strain MST01 (W3110, Δ *lpxT::cat*) by P1 *vir* phage transduction using donor strain CH030 (W3110, Δ *eptA::nptII*).¹ Plasmid pAChp0021 was generated by ligation of the coding sequence of *H. pylori* gene *hp0021* plus 50 bp upstream into the BamHI and EcoRV restriction sites of plasmid pACYC184 (Novagen). Primer A (5'-GCGCGCGGATCCTCTTATTTAAACAAAATTTTGTG-3') and Primer B (5'-GCGCGCGATATCTTAAGGCTTTTTGGGGCT-3') were used to amplify *hp0021* insert. Plasmid pWScj0256 was generated by ligation of the *C. jejuni* *cj0256* coding sequence into the SacII and XhoI restriction sites of plasmid pWSK29.² Primer C (5'-GCGCGCCCGCGGAAGAAGGAGATATACATGCTTAGATTAAGTTTTCAG-3'), containing a Shine-Dalgarno sequence, and Primer D (5'-GCGCGCCTCGAGTCATGGATTTGCCTTAAGTTTAGG-3') were used to amplify the *cj0256* insert.

Mass spectrometry, Infrared Multiphoton Dissociation, and Ultraviolet Photodissociation.

An online nanoelectrospray setup was used for direct infusion and consisted of a conductive mini microfilter junction from IDEX Health and Science (Oak Harbor, WA) coupled to a New Objectives uncoated PicoTip® nanoESI emitter (Woburn, MA). The spray voltage was applied at 2 kV. Lipid A samples were diluted to 1 μ M in 50/50 methanol/chloroform, and were

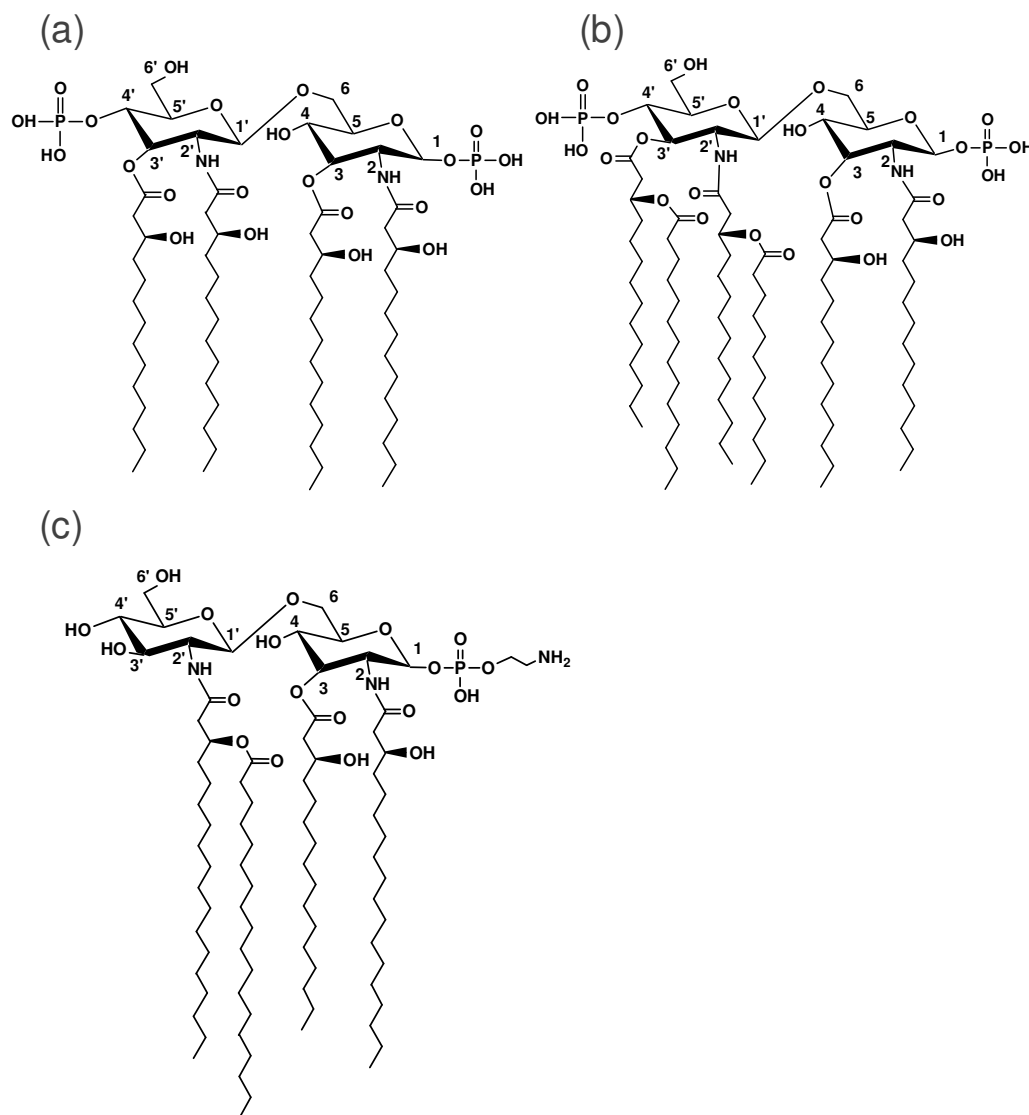
infused at 300 nL/min. Typical parameters such as a q -value of 0.25, an activation time of 30 ms, and a normalized collision energy (NCE) of 35% were used for all CID experiments (NCE = (voltage \times 30) / (0.000062 \times m/z \times 0.006325) in which 0.000062 is the TickAmpSlope and 0.006325 is the TickAmpInt in the Thermo LTQ software). IRMPD was performed using a q -value of 0.1 with the laser operated at 50 watts and an irradiation time of 30 ms. A q -value of 0.1 and a laser activation of 5 pulses (8 mJ/pulse) at a 500 Hz rep rate (over a 10 ms period) were used for all UVPD experiments. Activated – electron photodetachment dissociation (a-EPD) was performed using a q -value of 0.1 and a laser activation of 5 pulses (8 mJ/pulse) at 500 Hz (over a 10 ms period) to generate the charge reduced, radical species, which was subsequently activated using a normalized collision energy of 35%, q -value of 0.25, and an activation period of 30 ms. The maximum injection time for all MS/MS spectra was set to 100 ms.

Experimental Section (Supporting Information) References

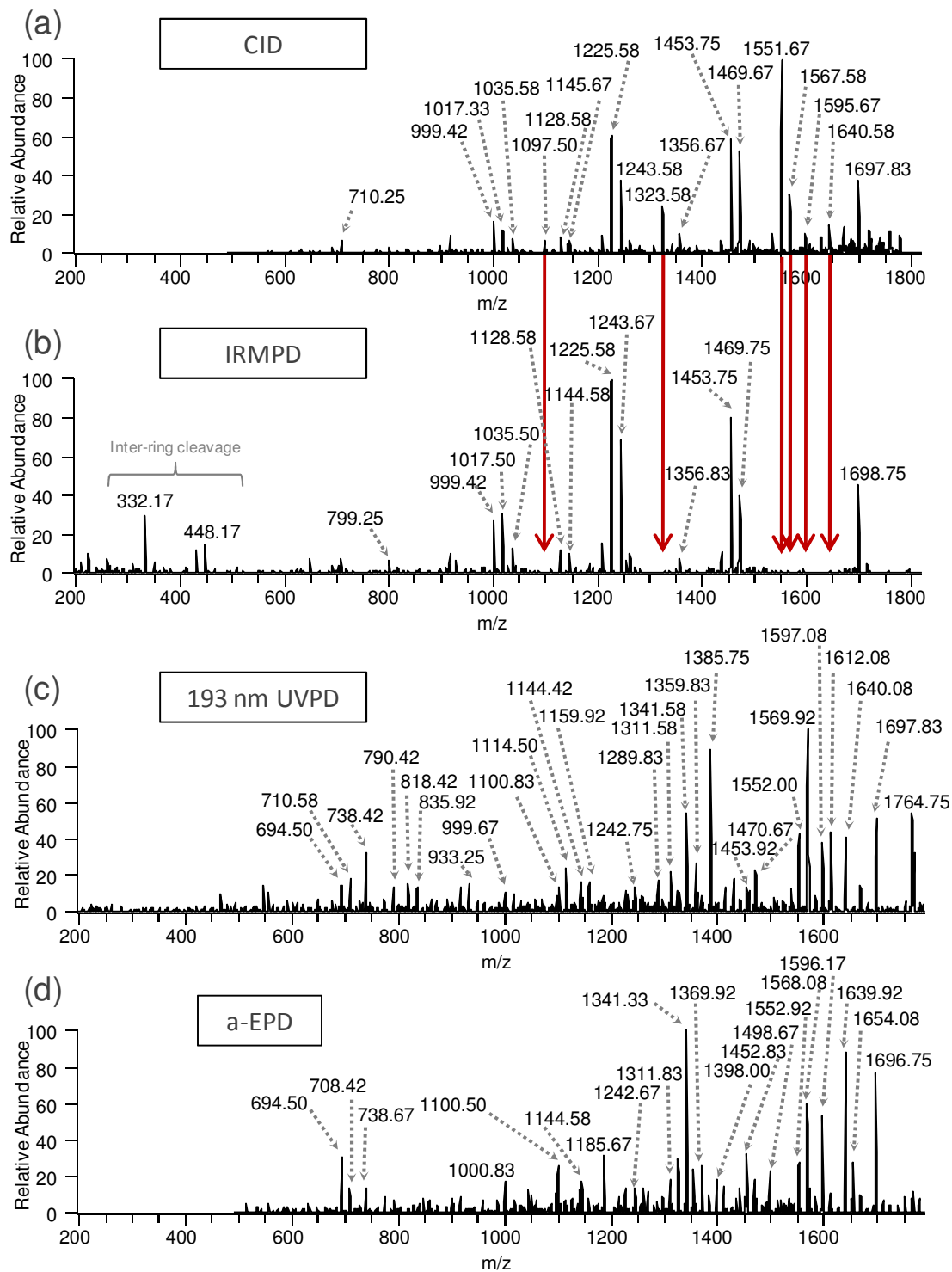
- (1) Herrera, C. M.; Hankins, J. V.; Trent, M. S. *Mol Microbiol* **2010**, *76*, 1444-1460.
- (2) Wang, R. F.; Kushner, S. R. *Gene* **1991**, *100*, 195-199.

Figures (Supporting Information)

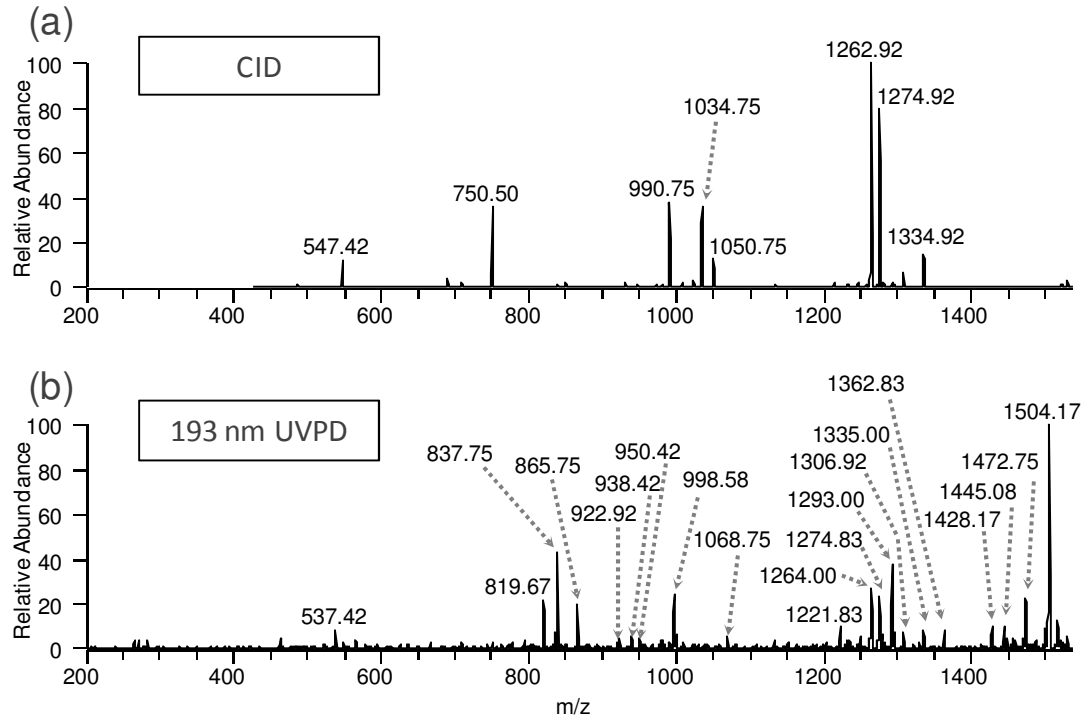
Supplementary Figure 1. Structures of lipid A analytes in this study: (a) lipid IV_A, molecular weight 1404.8, (b) *E. coli* F583 lipid A, molecular weight 1797.2, and (c) *H. pylori* lipid A, molecular weight 1548.2.



Supplementary Figure 2. MS/MS spectra from the dissociation of *E. coli* F583 lipid A by (a) CID, 1-, (b) IRMPD, 1-, (c) 193 nm UVPD, 1-, and (d) a-EPD, 1-•. Red arrows represent the secondary dissociated product ions after IRMPD. The 1- precursor was m/z 1796.0 and the 1-• precursor (for a-EPD) was m/z 1795.0.



Supplementary Figure 3. MS/MS spectra from the dissociation of *H. pylori* lipid A by (a) CID, 1-, and (b) 193 nm UVPD, 1-. The 1- precursor was m/z 1547.2.



Supplementary Figure 4. Structures of phosphorylethanolamine modified lipid A analytes in this study:

(a) *H. pylori* lipid A, molecular weight 1548.2, (b) *C. jejuni* lipid A, molecular weight 1922.4, and (c) *E. coli* strain MST10 (W3110 $\Delta lpxT$, $\Delta eptA/pAChp0021$, pWScj0256), molecular weight 1840.3. The modification is highlighted in pink.

