

Supplementary Material

Biochemical analysis of human *POLG2* variants associated with mitochondrial disease

Matthew J. Young, Matthew J. Longley, Fang-Yuan Li, Rajesh Kasiviswanathan, Lee-Jun Wong, and William C. Copeland

Supplementary Table: *POLG2* sequencing primers. Primer sequences were designed using genomic sequence from the Genebank database. All primers are M13 linked.

Exons	Primer Names	Primer Sequences (5'-3')	Length
E1	POLG2-E1F2	CTC GT TGT AAA ACG ACG GCC AGT AACGGTAGTGGTGGCTTGT	43
	POLG2-E1R2	CTG CT CAG GAA ACA GCT ATG ACC CCAGTTGCAATCCCCAAG	41
E2-3	POLG2-E2-3F	CTC GT TGT AAA ACG ACG GCC AGT CCACCAAGCTTAGCCAACAT	43
	POLG2-E2-3F3	CTC GT TGT AAA ACG ACG GCC AGT TAGGCGTGAGGCCACCAAG	41
	POLG2-E2-3R	CTG CT CAG GAA ACA GCT ATG ACC ACACATCTGAGCCCAACAAA	43
E4	POLG2-E4F	CTC GT TGT AAA ACG ACG GCC AGT TGCTGAATAAAAAGTATCGGACAA	47
	POLG2-E4R	CTG CT CAG GAA ACA GCT ATG ACC GGCTCTGTGGCAATAGTGG	43
E5	POLG2-E5F	CTC GT TGT AAA ACG ACG GCC AGT CCGTCTCAAAACAAACAAACA	45
	POLG2-E5R2	CTG CT CAG GAA ACA GCT ATG ACC AGAAACCGAGCACACTAACATT	45
E6	POLG2-E6F	CTC GT TGT AAA ACG ACG GCC AGT CTGCCAACATCTAAGTGGAGA	45
	POLG2-E6R	CTG CT CAG GAA ACA GCT ATG ACC TGAGTTCAAAGGAAATGCTCA	44
	POLG2-E6R2	AACTCACCTGTCTTAGTTCC	20
E7	POLG2-E7F	CTC GT TGT AAA ACG ACG GCC AGT TTGAAGAGGGTATTGTGG	43
	POLG2-E7R	CTG CT CAG GAA ACA GCT ATG ACC TTCTGTAAAAGGGAAACTGAGGA	46
E8	POLG2-E8F	CTC GT TGT AAA ACG ACG GCC AGT CCCTGAGTCTGTGACTCTGA	44
	POLG2-E8R	CTG CT CAG GAA ACA GCT ATG ACC AATGAAAGCAAGCACCAACAA	43

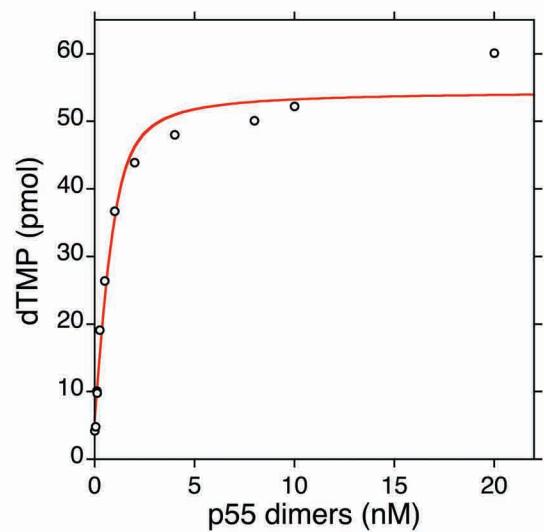
Figure Legends

Figure S1. Determination of the affinities of p55 variants for p140 ($K_{d(p140)}$). Representative p140-binding isotherms for wild-type and each of the p55 variants. Reactions (50 μ l) contained 1 nM of p140 and varying amounts (0 – 20 nM) of dimeric p55 or variant at the fixed concentration of 220 mM NaCl. Reaction times were 10 minutes at 37°C. On the Y-axis, pmol dTMP represents the amount of dTMP incorporated into the poly(rA)•oligo(dT)₁₂₋₁₈ (polymerase activity) as measured by counting TCA-insoluble radioactivity. Using the KaleidaGraph program each $K_{d(p140)}$ value was estimated from the quadratic curve fits by nonlinear regression analysis as described in Materials and Methods. Each version of p55 was measured at least twice and the average $K_{d(p140)}$ value \pm the standard deviation is reported in Table 2.

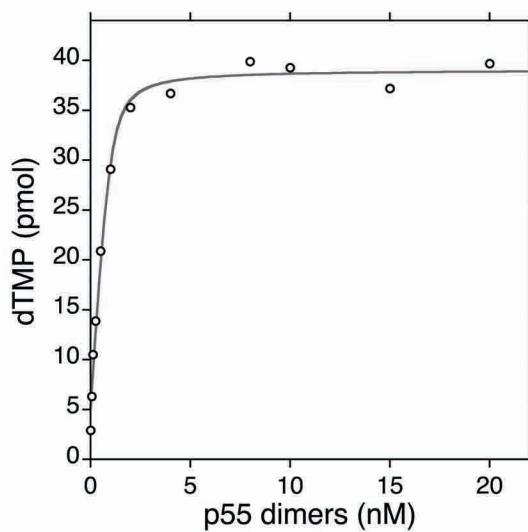
Figure S2. Determination of the dissociation constants for DNA binding ($K_{d(DNA)}$) of p55 and p55 variants. Representative DNA-binding isotherms for wild-type and each of the p55 variants. Electrophoretic mobility shift assays were performed to estimate the $K_{d(DNA)}$ values using various concentrations of p55 or p55 variant (0 – 160 nM dimers) incubated separately with 15 nM of double stranded oligonucleotide in 20 μ l binding reactions (Materials and Methods). The fraction of DNA bound is plotted against the concentration of p55 or variant dimers and binding isotherms were fit to a quadratic equation by nonlinear regression analysis using the KaleidaGraph program to estimate $K_{d(DNA)}$ values. Each version of p55 was measured at least twice and the average $K_{d(DNA)}$ value \pm the standard deviation is reported in Table 2.

Figure S1 Page 1 of 2

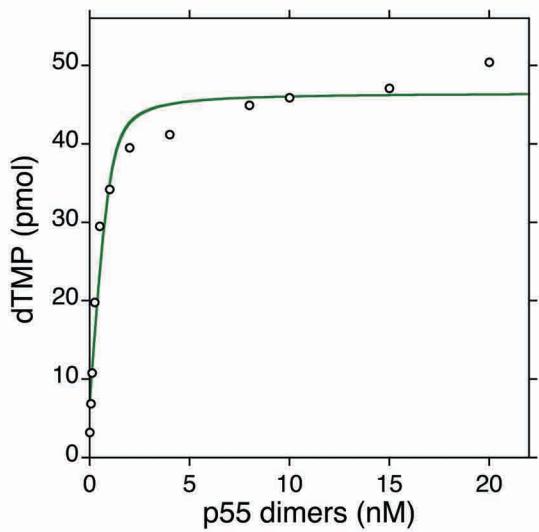
WT p55



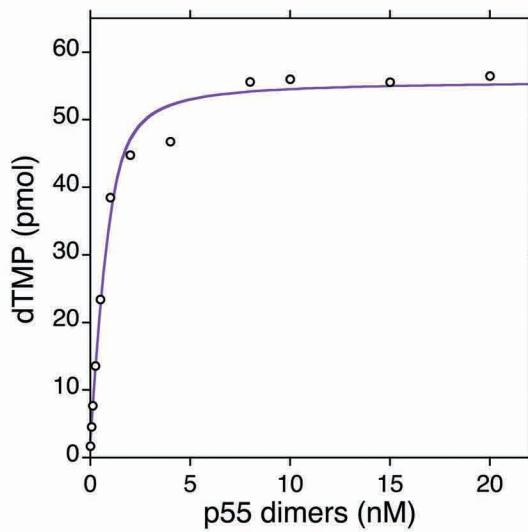
G103S p55



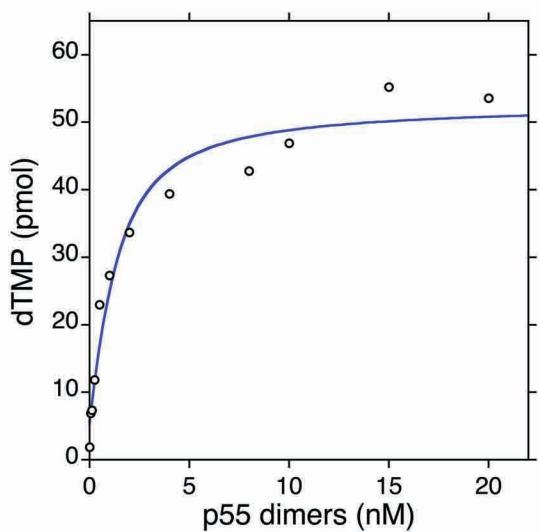
L153V p55



P205R p55



R369G p55



D386E p55

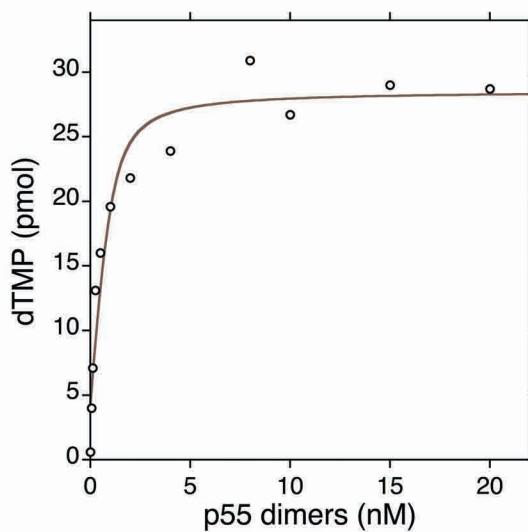


Figure S1 Page 2 of 2

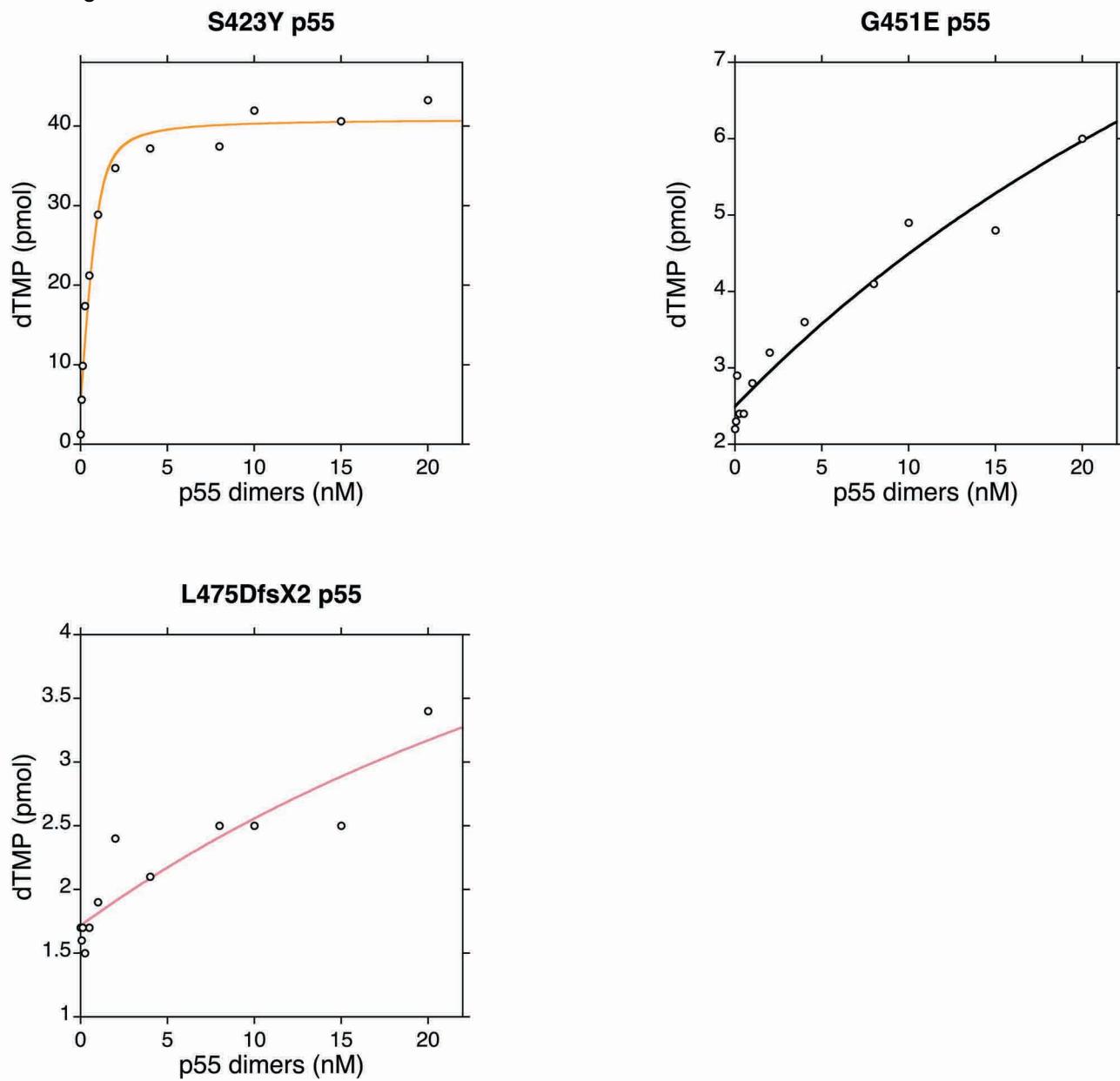


Figure S1.

Figure S2 Page 1 of 2

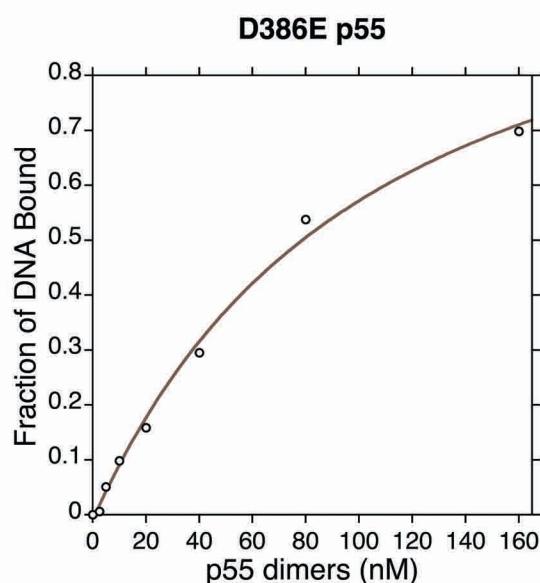
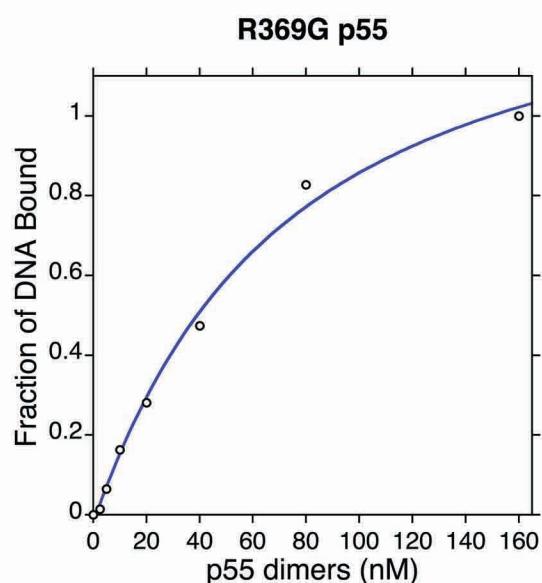
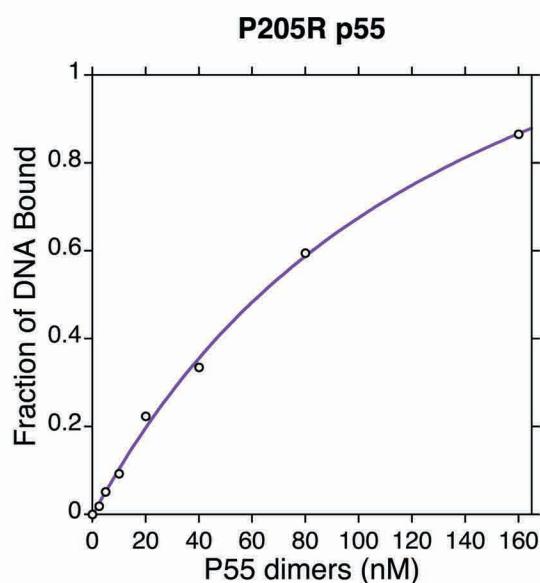
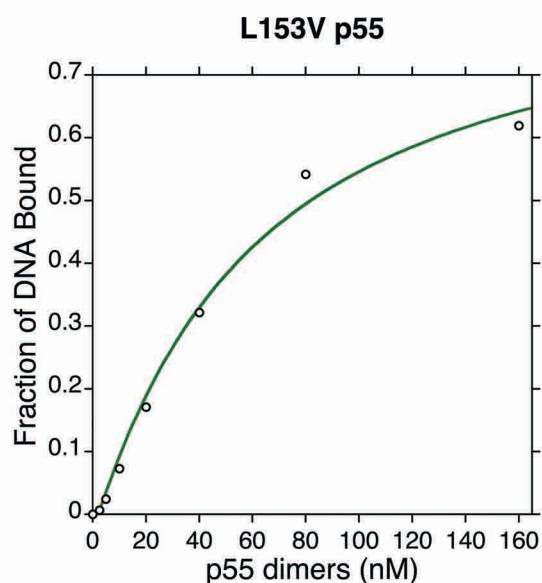
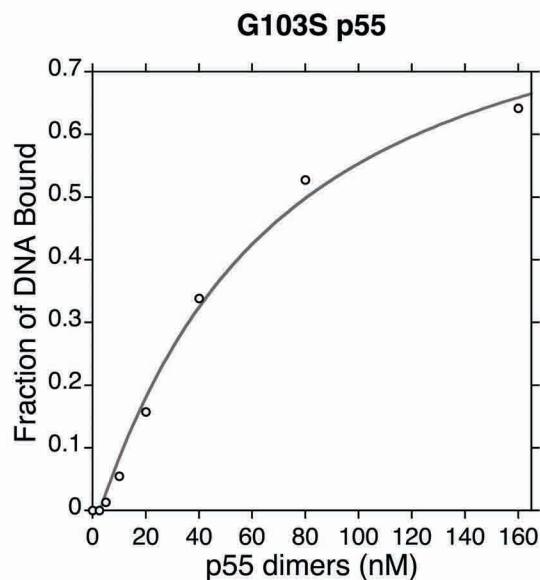
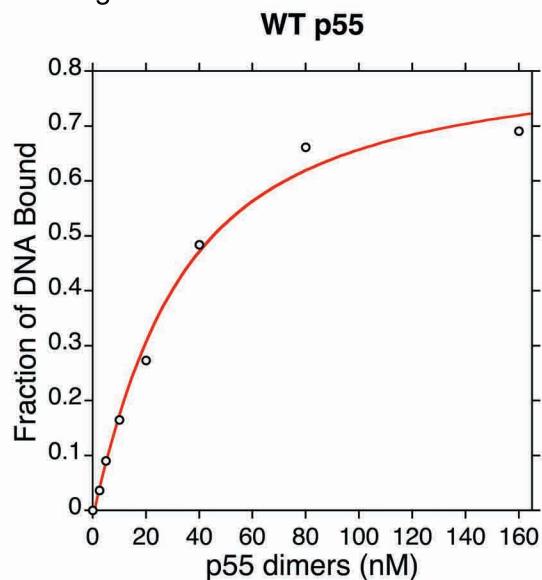


Figure S2 Page 2 of 2

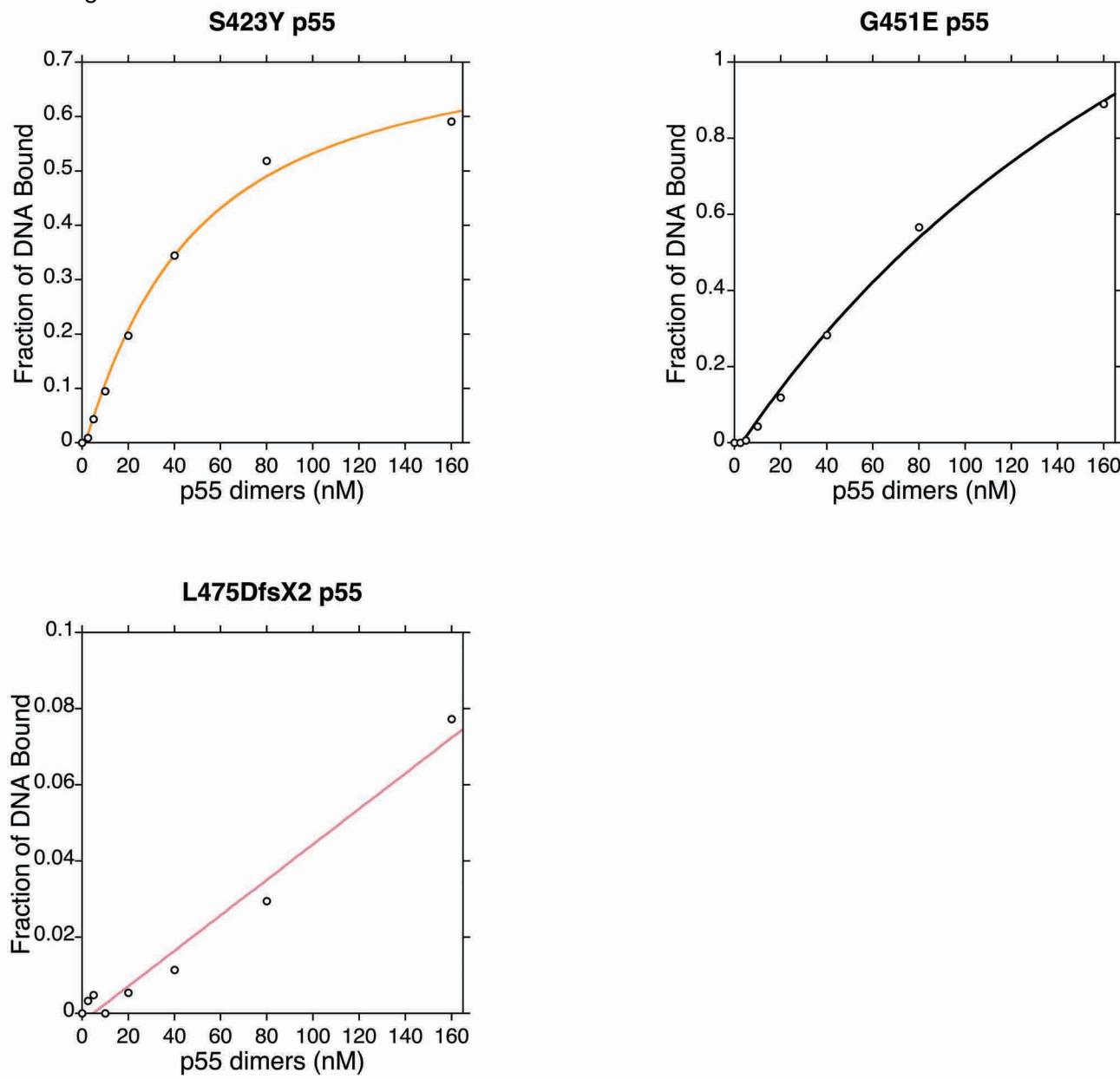


Figure S2.