

New Candidaspongiolides, Tedanolide Analogs that Selectively Inhibit Melanoma Cell Growth

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General Experimental Procedures. Optical rotations were measured on a Perkin-Elmer 241 polarimeter. UV spectra were acquired in spectroscopy grade MeOH using a Varian Cary 50 UV-Vis spectrophotometer. NMR data were collected using an Avance III 600 (^1H 600 MHz, ^{13}C 150 MHz) NMR spectrometer (Bruker Biospin) with a 3-mm PATXI probe, referenced to residual solvent. MS spectra were measured with an Agilent Technologies 6510 Q-TOF LC-MS and an Applied Biosystems, Inc. QSTAR XL hybrid triple-quad time-of-flight (QqTOF) mass spectrometer. Initial fractionation was performed on Diol SPE cartridges (Applied Separations) and Sephadex LH-20 resin (Amersham Biosciences). HPLC purification was performed on a Rainin SD-1/UV-1 system.

Biological Material. Two different collections from Papua New Guinea (OCDN1808 and OCDN5955), used in this investigation, were initially identified as *Euryspongia* sp. They were subsequently compared to Great Barrier Reef specimens of *C. flabellata* (the original source of the candidaspongiolides) and reclassified as *Candidaspongia* sp.¹ The Papua New Guinea specimens are a darker color and have somewhat sharper conules, while the *C. flabellata* specimens from Australia had thicker fibers and a thicker sand coat on the surface. Vouchers for the Papua New Guinea collections are maintained at the Smithsonian Sorting Center, Suitland, Maryland.

Extraction and Isolation. The Papua New Guinea *Candidaspongia* sp. specimens were repeatedly extracted according to the methodology outlined in McCloud² to give the aqueous crude extracts. A portion of this extract (520 mg) was subjected to size-exclusion chromatography on Sephadex LH-20 (2.5 × 90 cm) using hexanes/CH₂Cl₂/MeOH (2:5:1) to yield seven fractions (141A-141G). Fraction 141F was chromatographed on C₁₈ (2.0 × 17 cm)

¹ Meragelman, T. L.; Willis, R. H.; Woldemichael, G. M.; Heaton, A.; Murphy, P. T.; Snader, K. M.; Newman, D. J.; van Soest, R.; Boyd, M. R.; Cardellina II, J. H.; McKee, T. C. *J. Nat. Prod.* **2007**, *70*, 1133.

² McCloud, T. G. *Molecules*, **2010**, *15*, 4526.

using 50/50 50% CH₃CN/50% H₂O (+0.1% AcOH) to yield precandidaspongiolides A/B (**1/2**, 11.2 mg). Fraction 141C was chromatographed on C₁₈ (2.0 × 17 cm) using 55% CH₃CN/45% H₂O (+0.1% AcOH) to yield seven fractions (154A-154G). Fraction 154F was purified by HPLC using a Rainin Dynamax C₁₈ column (250 × 10 mm) employing a gradient of 35% CH₃CN/65% H₂O (+0.1% AcOH) to 85% CH₃CN at 4.5 mL/min over 20 min to yield candidaspongiolides A/B (**3/4**, 0.9 mg). Fraction 141D was purified by HPLC utilizing the same method to yield tedanolide (**5**, 0.4 mg).

Precandiaspongiolides A and B (1/2): $[\alpha]_D^{25} + 58.3$ (*c* 0.23, MeOH); UV (MeOH) λ_{\max} (log ϵ) 205 (3.69) nm; ¹H NMR and ¹³C NMR data, see Table 1; HRESIMS *m/z* 665.3146 [M+Na]⁺ (calcd for C₃₂H₅₀O₁₃Na, 665.3144).

Candidaspongiolides A and B (3/4): $[\alpha]_D^{25} + 20.0$ (*c* 0.11, MeOH); UV (MeOH) λ_{\max} (log ϵ) 204 (3.48) nm; ¹H NMR and ¹³C NMR data, see Table 1; HRESIMS *m/z* 707.3235 [M+Na]⁺ (calcd for C₃₄H₅₂O₁₄Na, 707.3249).

(+)-Tedanolide (5): $[\alpha]_D^{25} + 20$ (*c* 0.02, MeOH); UV (MeOH) λ_{\max} (log ϵ) 204 (3.79) nm; ¹H NMR and ¹³C NMR data in CDCl₃ and CD₃OD, see Tables S2 and S3; HRESIMS *m/z* 633.3232 [M+Na]⁺ (calcd for C₃₂H₅₀O₁₁Na, 633.3245).

Acetylation of Precandidaspongiolides A and B (1/2). A stock solution of AcCl was prepared on ice by dissolving 1 μ L of AcCl in 50 μ L of anhydrous CH₂Cl₂. 7 μ L of the AcCl solution (2 μ mol) was added to a stirring solution of **1/2** (1.1 mg, 1.7 μ mol) in 2,4,6-trimethylpyridine (excess) at -40 °C.³ The reaction was monitored by LC-MS for the production of mono-acetylated product and allowed to stir at -40 °C for 3h and then 25°C for 19h. After the mono-acetylated product formation was observed, the reaction mixture was diluted with H₂O and

³ Ishihara, K.; Kurihara, H.; Yamamoto, H. *J. Org. Chem.* 1993, 58, 3791.

dried under N₂. The reaction mixture was then purified by HPLC using a Rainin Dynamax C₁₈ column (250 × 10 mm) employing a gradient of 35% CH₃CN/65% H₂O (+0.1% AcOH) to 85% CH₃CN at 4.5 mL/min over 20 min to yield 28-acetyl-precandidaspongiolide A (**6**, 0.48 mg, 41% yield) and unreacted precandidaspongiolides A/B (**1/2**, 0.57 mg, 52% yield).

28-acetyl-precandidaspongiolide A (6): $[\alpha]_D^{25} + 28.6$ (*c* 0.12, MeOH); UV (MeOH) λ_{\max} (log ϵ) 204 (3.38) nm; ¹H NMR and ¹³C NMR data, see Table S4; HRESIMS *m/z* 707.3245 [M+Na]⁺ (calcd for C₃₄H₅₂O₁₄Na, 707.3249).

Reduction of Precandidaspongiolides A and B (1/2). NaBH₄ (~3 mg, excess) was added to a semi-pure (~75%) solution of **1/2** (13.0 mg, 20.2 μ mol) in MeOH (200 μ L). The reaction was allowed to stir at rt for 10 min. The reaction mixture was diluted with H₂O and desalted by passing through a C₁₈ SPE column. The products were then purified by HPLC using a Rainin Dynamax C₁₈ column (250 × 10 mm) employing a gradient of 25% CH₃CN/75% H₂O (+0.1% AcOH) to 75% CH₃CN at 4.5 mL/min over 20 min to yield 11*R*-dihydro-precandidaspongiolide A (**7**, 4.76 mg, 37% yield) and 11*S*-dihydro-precandidaspongiolide A (**8**, 0.49 mg, 4% yield).

11*R*-dihydro precandidaspongiolide A (7): $[\alpha]_D^{25} + 49.7$ (*c* 0.18, MeOH); UV (MeOH) λ_{\max} (log ϵ) 204 (3.60) nm; ¹H NMR and ¹³C NMR data, see Table S5; HRESIMS *m/z* 667.3304 [M+Na]⁺ (calcd for C₃₂H₅₂O₁₃Na, 667.3300).

11*S*-dihydroprecandidaspongiolide A (8): $[\alpha]_D^{25} + 43.5$ (*c* 0.02, MeOH); UV (MeOH) λ_{\max} (log ϵ) 204 (3.41) nm; ¹H NMR and ¹³C NMR data, see Table S6; HRESIMS *m/z* 667.3290 [M+Na]⁺ (calcd for C₃₂H₅₂O₁₃Na, 667.3300).

MTT Cytotoxicity Assay. Cell survival was measured by the MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) cytotoxicity assay (Sigma, St Louis, MO).⁴ Cells were seeded in 100 μ L of medium at a density of 4×10^3 cells/well and allowed to incubate at 37°C in 5% CO₂ for 24 hours. Serially diluted drugs were then added in an additional 100 μ L of medium and incubated for 72 hours. After removal of medium containing drug, MTT (0.5mg/mL) in IMDM growth medium was added to each well and incubated for four hours. The media solution was then removed from the wells, and 100 μ L acidified 80% ethanol solution was added to lyse cells and dissolve the formazan product. Cell viability was measured spectrophotometrically at 570 nm and background corrected at 690 nm. All MTT assays were performed three times in triplicate. Cytotoxicity (IC₅₀) was defined as the drug concentration that reduced cell viability to 50% of the untreated control. Both KB-3-1 and KB-V1 were also coincubated with 1/2 and 100 nM tariquidar (Dr. Susan Bates, NCI).

Cell lines. The cell lines used were: the human epithelial adenocarcinoma cell line KB-3-1 and its P-gp-expressing multidrug resistant sub-line KB-V1; the human breast cancer cell line MCF-7; the human lung carcinoma cell line H460; and the human melanoma cell lines M14, LOX IMVI, and UACC-257. All cell lines were grown at 37°C in 5% CO₂ and cultured as follows. The KB, MCF7, M14, and UACC-257 cell lines were cultured in Dulbecco's Modified Eagle's Medium (DMEM,) supplemented with 10% fetal bovine serum, 5 mM L-glutamine, 50 units/mL penicillin, and 50 μ g/mL streptomycin, all obtained from Life Technologies (Carlsbad, California, USA). LOX IMVI cells were cultured in Roswell Park Memorial Institute (RPMI) 1640 medium from Life Technologies (Carlsbad, California, USA) and supplemented as

⁴Brimacombe, K. R., Hall, M. D., Auld, D. S., Inglese, J., Austin, C. P., Gottesman, M. M., and Fung, K. L., *Assay Drug Dev. Technol.* **2009**, 7, 233.

described above. Additionally, the multidrug resistant cell line KB-V1 was cultured in 1 µg/mL vinblastine to maintain P-glycoprotein expression.⁵

NCI-60 cell line screen. Growth inhibition of 50% (GI₅₀) is defined as the concentration of a compound that causes a 50% reduction in cell growth compared to the untreated control. The GI₅₀ is comparable to an IC₅₀, but takes into account the cell count at time zero and in untreated controls at the end of the assay period; the IC₅₀ is calculated based on the number of cells in the untreated control when the assay endpoint is read. The GI₅₀ is calculated as $100 \times (T - T_0)/(C - T_0) = 50$, where T is the test optical density, T₀ is the optical density at time zero, and C is the control optical density. The total growth inhibition (TGI) signifies a cytostatic effect, and is calculated as $100 \times (T - T_0)/(C - T_0) = 0$. The lethal concentration of 50% (LC₅₀) signifies a cytotoxic effect, and is calculated as $100 \times (T - T_0)/T_0 = -50$. The control optical density is not used in the calculation of LC₅₀.⁶

⁵ Shen, D. W., Cardarelli, C., Hwang, J., Cornwell, M., Richert, N., Ishii, S., Pastan, I., and Gottesman, M. M. *J. Biol. Chem.*, **1986**, 261, 7762.

⁶ Monks, A., Scudiero, D., Skehan, P., Showmaker, R., Paull, K., Vistica, D., Hose, C., Langley, J., Cronise, P., Vaigro-Wolff, A., Gray-Goodrich, M., Campbell, H., Mayo, J., Boyd, M., *J. Natl. Cancer Inst.*, **1991**, 83, 757.

Table S1. NMR Data for Candidaspongiolide A (**3**) in Acetone-*d*₆ (600 MHz, 500MHz^a)

No.	δ_C	δ_H , mult (<i>J</i> , Hz)
1	172.6 ^a	—
2	72.3	3.79, d (1.9)
3	84.7	3.83, dd (9.5, 1.9)
4	48.5	3.19, dq (9.5, 7.0)
5	214.6 ^a	—
6	49.0	3.41, dq (10.6, 7.0)
7	80.3	5.40, d (10.6)
8	134.2 ^a	—
9	132.5	5.46, br d (9.8)
10	45.7	3.43, dq (9.8, 6.8)
11	210.9 ^a	—
12a	43.8	2.72, dd (17.7, 9.7)
12b		2.31, dd (17.7, 1.7)
13	69.2	4.50, dd (9.7, 1.7)
14	85.2 ^a	—
15	215.9 ^a	—
16	48.0	4.12, ddd (11.0, 10.8, 4.1)
17	77.9	3.22, d (10.8)
18	63.0 ^a	—
19	66.2	2.64, d (9.3)
20	31.9	2.47, ddd (10.7, 9.3, 6.6)
21	131.4	5.34 ddq (10.9, 10.7, 1.7)
22	125.3	5.49, dq (10.9, 6.8)
23	13.3	1.64, dd (6.8, 1.7)
24	14.9 ^b	1.21, d (7.0)
25	14.6	1.16, d (7.0)
26	10.6	1.67, br s
27	15.2	0.96, d (6.8)
28	65.3 ^c	3.77, ^c s
29a	64.4	4.34, dd (10.6, 4.1)
29b		3.90, dd (11.0, 10.6)
30	11.3	1.35, s
31	18.6	1.08, d (6.6)
32	60.9	3.40, s
33	170.0 ^a	—
34	20.5	2.01, s

^a Quaternary carbons obtained from original candidaspongiolide core NMR data (500 MHz).

^b Originally misassigned. Careful inspection of original HSQC data for candidaspongiolide A (candidaspongiolide core) showed no correlation between δ_C 22.9 and δ_H 1.21.

^c The multiplicity-edited HSQC used to characterize **3** clearly indicated a CH₂ multiplicity (opposite phase to CH/CH₃) for the correlation between δ_C 65.3 and δ_H 3.77, suggesting it was the C-28 primary alcohol. However, the ¹H/¹³C chemical shifts did not match those in the literature for candidaspongiolide A (candidaspongiolide core). Further evaluation of the original COSY and HMBC spectra for candidaspongiolide A (candidaspongiolide core) indicated that δ_C 65.3 and δ_H 3.77 were the correct assignments for C-28, and H-28, respectively, and a minor contaminant was responsible for the correlation observed between δ_C 70.9 and δ_H 3.61.⁷

⁷ See Supporting information for Meragelman, T. L.; Willis, R. H.; Woldemichael, G. M.; Heaton, A.; Murphy, P. T.; Snader, K. M.; Newman, D. J.; van Soest, R.; Boyd, M. R.; Cardellina II, J. H.; McKee, T. C. *J. Nat. Prod.* **2007**, *70*, 1133.

Table S2. NMR Data for Tedanolide (**5**) (600 MHz, CDCl₃)

No.	δ_C	δ_H , mult (<i>J</i> , Hz)
1	171.8	—
2	71.7	3.86, br dd (8.7, 1.7)
3	83.5	3.66, dd (8.6, 1.7)
4	48.8	3.02, ^a (8.6, 7.0)
5	215.7	—
6	50.3	3.02, ^a (10.2, 6.9)
7	80.1	4.10, br dd (10.2, 2.5)
8	136.6	—
9	129.8	5.46, ^a
10	45.7	3.40, m (6.8)
11	213.1	—
12a	45.1	2.57, dd (17.0, 9.5)
12b		2.47, dd (17.0, 2.9)
13	68.7	4.29, m (9.5, 2.9)
14	53.6	3.03, ^a (6.9)
15	214.5	—
16	52.4	3.53, ddd (11.6, 9.6, 4.0)
17	77.4	3.23, br d (9.6, 3.2)
18	63.2	—
19	67.0	2.60, d (9.3)
20	31.6	2.42, ddq (10.6, 9.3, 6.5)
21	130.6	5.22, ddq (10.8, 10.6, 1.6)
22	125.5	5.45, ^a (10.8, 6.8)
23	13.6	1.60, dd (6.8, 1.6)
24	14.5	1.21, d (7.0)
25	15.8	1.27, d (6.9)
26	10.7	1.61, s
27	16.9	1.07, d (6.8)
28	10.9	1.10, d (6.9)
29a	64.3	4.25, dd (10.5, 4.0)
29b		4.11, dd (11.6, 10.5)
30	11.8	1.37, s
31	18.8	1.11, d (6.5)
32	60.8	3.29, s
2-OH		2.75, br d (8.7)
7-OH		1.44, br d (2.5)
13-OH		3.32, br d (3.2)
17-OH		2.13, br d (3.2)

^a Signals overlapped.

Table S3. NMR Data for Tedanolide (**5**) (600 MHz, CD₃OD)

No.	δ_C	δ_H , mult (<i>J</i> , Hz)
1	171.5	—
2	72.6	3.76, d (1.7)
3	84.7	3.68, dd (9.6, 1.7)
4	51.0	3.12, ^a (9.6, 7.1)
5	216.4	—
6	49.6	3.11, ^a (10.5, 7.1)
7	80.2	4.00, d (10.5)
8	138.9	—
9	130.1	5.35, br d (10.0)
10	47.0	3.41, dq (10.0, 6.9)
11	211.2	—
12a	47.5	2.60, dd (16.9, 9.6)
12b		2.34, dd (16.9, 1.7)
13	69.3	4.22, ddd (9.6, 7.0, 1.7)
14	55.8	2.91, dq (7.0, 7.0)
15	214.1	—
16	54.5	3.43, ddd (11.2, 10.5, 3.9)
17	78.0	3.13, ^a (10.5)
18	63.9	—
19	67.7	2.58, d (9.3)
20	32.7	2.49, ddq (10.4, 9.3, 6.6)
21	132.1	5.31, ddq (10.8, 10.4, 1.6)
22	126.4	5.48, dq (10.8, 6.9)
23	13.6	1.63, dd (6.9, 1.6)
24	15.2	1.24, d (7.1)
25	16.1	1.27, d (7.1)
26	10.6	1.65, s
27	15.9	1.05, d (6.9)
28	12.0	1.15, d (7.0)
29a	65.5	4.25, dd (10.7, 3.9)
29b		4.00, dd (11.2, 10.7)
30	11.6	1.37, s
31	18.8	1.10, d (6.6)
32	61.1	3.34, s

^a Signals overlapped.

Table S4. NMR Data for 28-acetyl-precandidaspongiolide A (**6**) (600 MHz, CD₃OD)

No.	δ_C	δ_H , mult (<i>J</i> , Hz)
1	172.5	—
2	72.2	3.79, d (2.1)
3	83.7	3.74, dd (9.3, 2.1)
4	49.0	3.10, dq (9.3, 7.1)
5	216.5	—
6	50.0	3.12, dq (10.1, 7.0)
7	79.5	4.00, d (10.1)
8	137.7	—
9	129.0	5.34, br d (9.6)
10	45.9	3.42, dq (9.6, 6.9)
11	212.1	—
12a	43.3	2.75, dd (17.1, 9.3)
12b		2.36, dd (17.1, 2.1)
13	68.7	4.38, dd (9.3, 2.1)
14	82.0	—
15	212.5	—
16	48.0	4.02, ^a (10.3, 3.4)
17	77.4	3.09, d (10.3)
18	63.1	—
19	66.5	2.61, d (9.3)
20	31.6	2.49, ddq (10.5, 9.3, 6.6)
21	130.8	5.31, ddq (10.8, 10.5, 1.6)
22	125.2	5.50, dq (10.8, 6.9)
23	12.7	1.64, dd (6.9, 1.6)
24	14.0	1.22, d (7.1)
25	14.9	1.25, d (7.0)
26	9.8	1.66, br s
27	15.4	1.06, d (6.9)
28a	64.2	4.25, d (11.0)
28b		4.20, d (11.0)
29a	63.2	4.35, dd (11.2, 3.4)
29b		4.01, ^a (11.2)
30	10.6	1.38, s
31	18.0	1.11, d (6.6)
32	60.5	3.36, s
33	171.6	—
34	19.9	2.01, s

^a Signals overlapped.

Table S5. NMR Data for 11*R*-dihydro precandidaspongiolide A (**7**) (600 MHz, CD₃OD)

No.	δ_C	δ_H , mult (<i>J</i> , Hz)
1	172.2	—
2	71.6	3.64, d (2.2)
3	84.8	3.72, ^a dd (10.0, 2.2)
4	47.9	3.26, dq (10.0, 6.9)
5	217.0	—
6	51.1	3.23, dq (9.7, 7.2)
7	78.3	4.17, d (9.7)
8	135.8	—
9	130.0	5.62, br d (9.0)
10	38.1	2.19, dq (9.0, 7.1, 1.7)
11	76.3	3.74, ^a (10.5, 1.7, 1.7)
12a	36.2	1.53, ddd (14.1, 10.5, 10.5)
12b		1.06, ddd (14.1, 1.7, 1.7)
13	73.2	4.28, dd (10.5, 1.7)
14	84.9	—
15	215.6	—
16	47.1	4.03, ddd (11.1, 10.9, 4.1)
17	77.3	3.29, ^b (10.9)
18	62.8	—
19	66.4	2.64, d (9.3)
20	31.6	2.46, ddq (10.5, 9.3, 6.5)
21	130.8	5.37, ddq (10.7, 10.5, 1.6)
22	125.2	5.54, dq (10.7, 6.9)
23	12.5	1.62, dd (6.9, 1.6)
24	14.8	1.20, d (6.9)
25	13.9	1.26, d (7.2)
26	9.3	1.54, s
27	17.1	1.00, d (7.1)
28a	64.8	3.81, d (11.6)
28b		3.77, d (11.6)
29a	64.7	4.26, dd (10.6, 4.1)
29b		3.88, dd (11.1, 10.6)
30	10.6	1.30, s
31	17.7	1.11, d (6.5)
32	60.4	3.43, s

^a Signals overlapped. ^b Buried under CD₃OD signal.

Table S6. NMR Data for 11*S*-dihydro precandidaspongiolide A (**8**) (600 MHz, CD₃OD)

No.	δ_C	δ_H , mult (<i>J</i> , Hz)
1	172.2	—
2	71.5	3.59, d (1.9)
3	84.1	3.84, ^a (10.0, 1.9)
4	48.1	3.25, dq (10.0, 6.9)
5	217.8	—
6	50.8	3.18, dq (10.3, 7.0)
7	78.7	4.07, d (10.3)
8	135.0	—
9	133.2	5.18, br d (8.9)
10	39.5	2.09, ddq (9.9, 8.9, 6.8)
11	73.3	3.40, ddd (11.0, 9.9, 1.5)
12a	38.4	1.48, ddd (14.1, 10.9, 1.5)
12b		0.88, ddd (14.1, 11.0, 1.8)
13	69.5	4.27, ^a (10.9, 1.8)
14	85.2	—
15	216.7	—
16	47.3	4.05, ddd (11.3, 11.1, 4.0)
17	77.2	3.29, d (11.1)
18	62.6	—
19	66.5	2.63, d (9.5)
20	31.5	2.46, ddq (10.6, 9.5, 6.6)
21	130.9	5.36, ddq (10.8, 10.6, 1.3)
22	125.2	5.54, ddq (10.8, 6.8)
23	12.6	1.62, dd (6.8, 1.3)
24	14.6	1.22, d (6.9)
25	14.3	1.26, d (7.0)
26	9.5	1.55, br s
27	17.1	1.01, d (6.8)
28a	66.5	3.85, d (11.7)
28b		3.81, d (11.7)
29a	64.9	4.27, ^a (4.0)
29b		3.82, ^a (11.3)
30	10.4	1.30, s
31	17.6	1.11, d (6.6)
32	60.3	3.42, s

^a Signals overlapped.

Figure S1. NCI 60-cell line screen, single dose (10^{-5} M) of 1/2

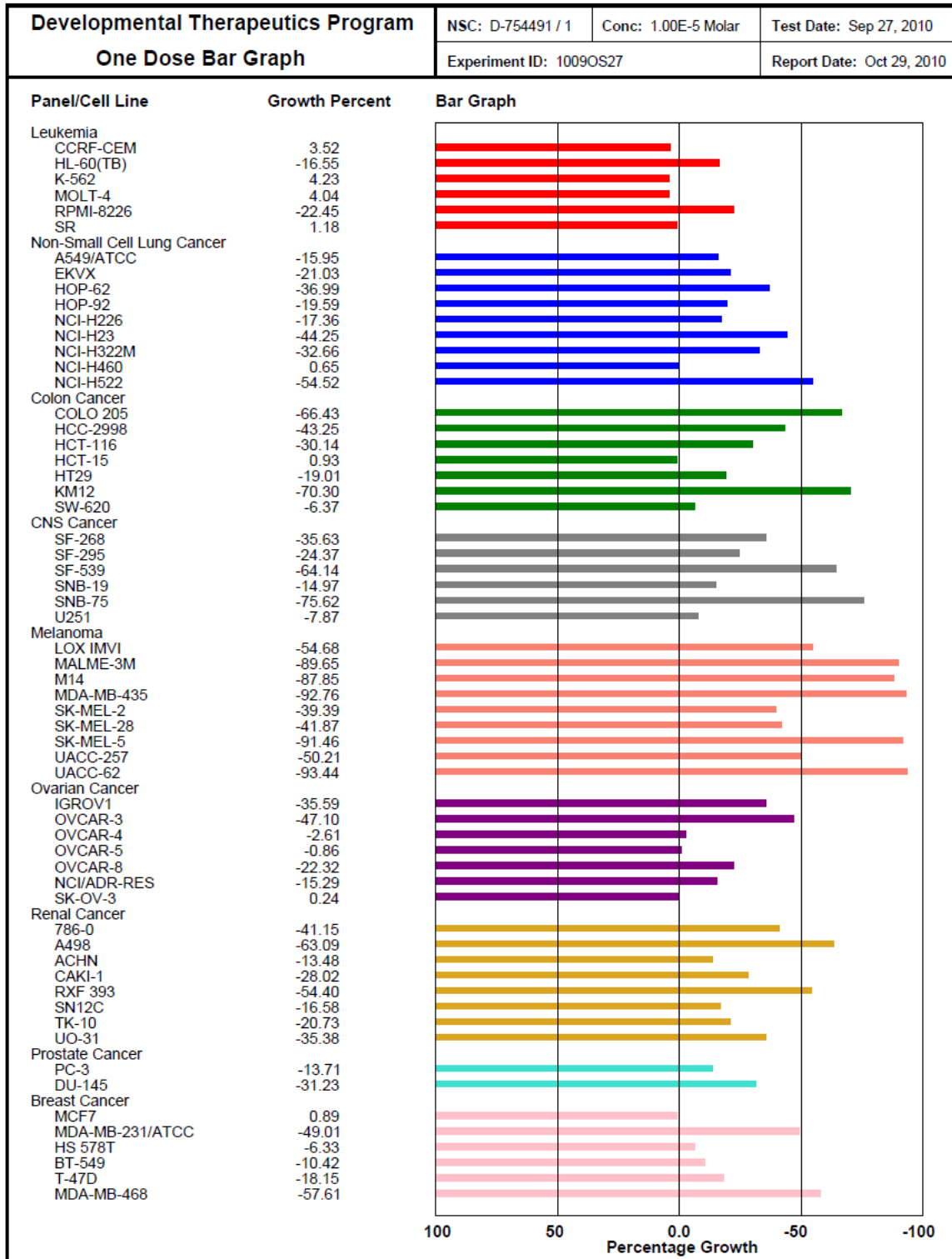


Figure S2. NCI 60-cell line screen, single dose (10^{-5} M) of 1/2

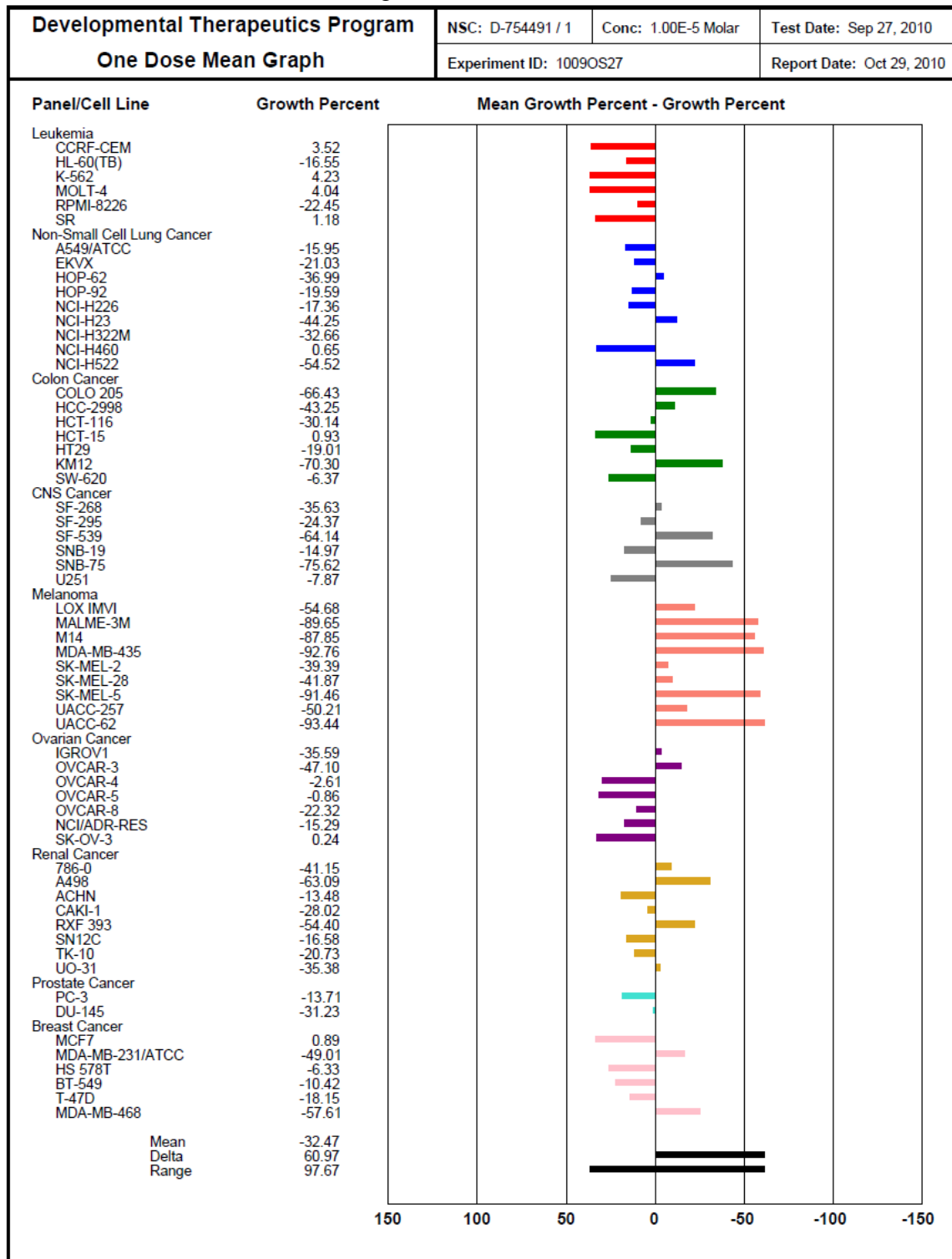


Figure S3. NCI 60-cell line screen, dose response curves for 1/2

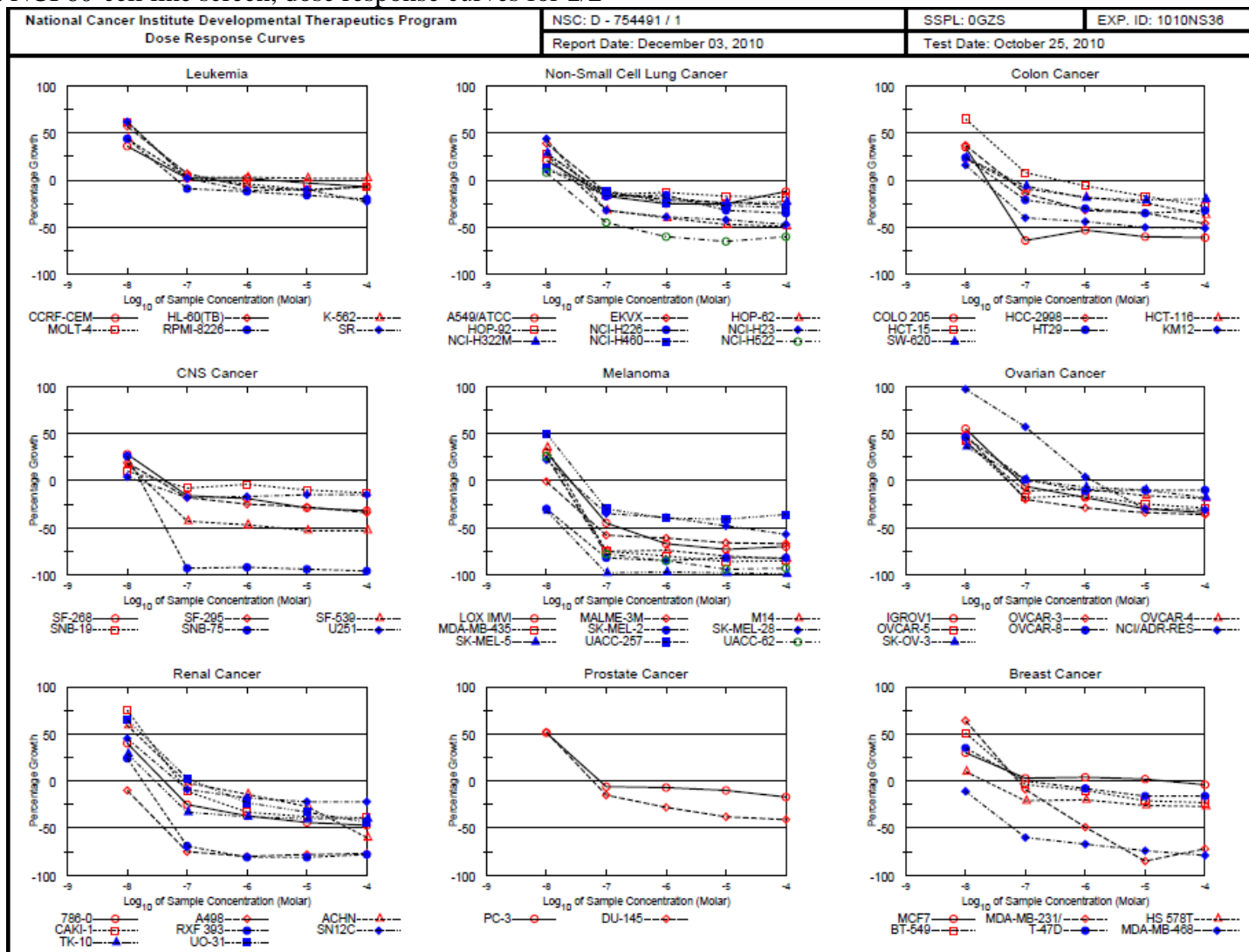


Figure S4. NCI 60-cell line screen, mean bar graphs for 1/2

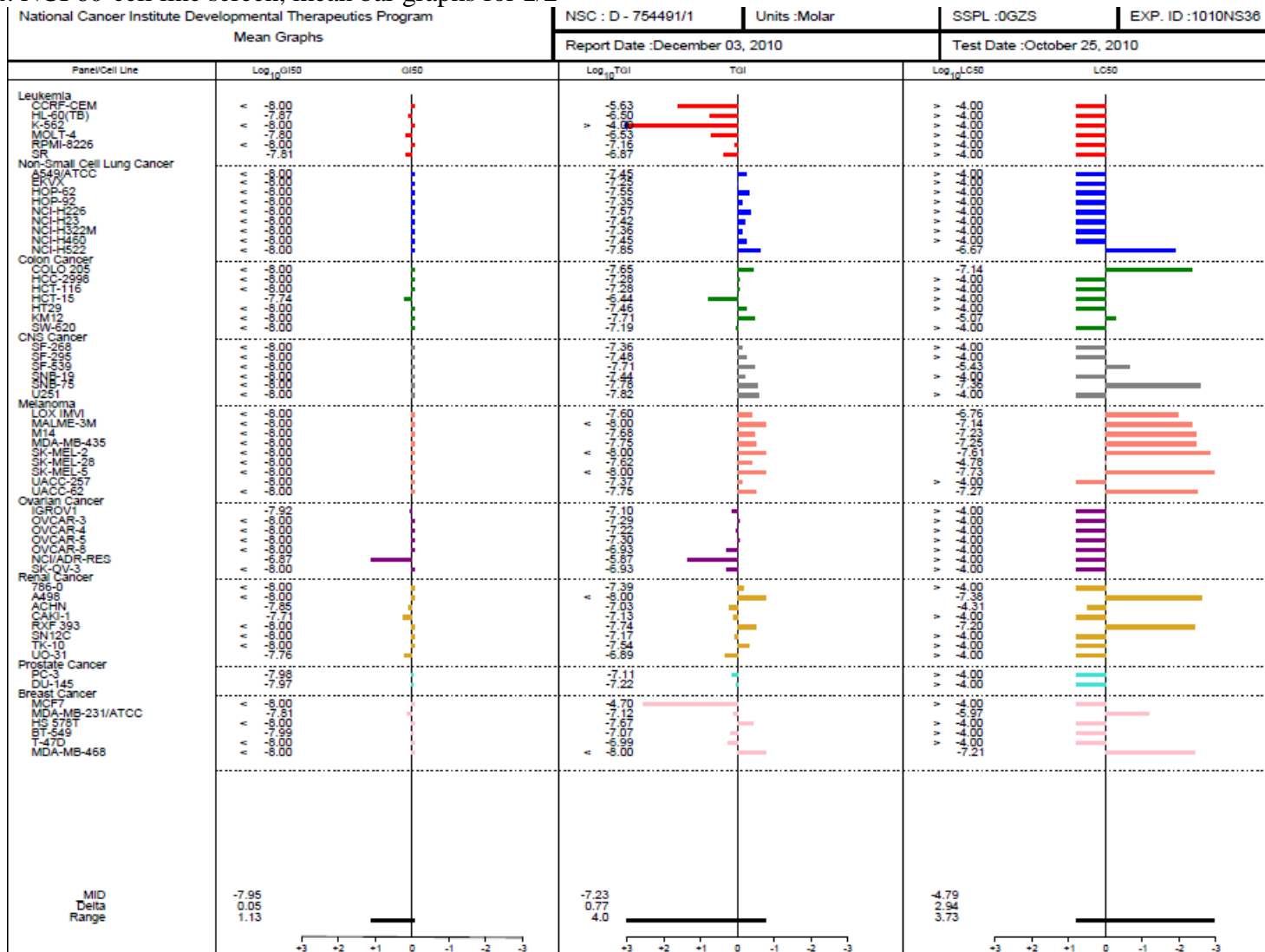


Figure S5. NCI 60-cell line screen, dose response curves for candidaspongolide acyl ester mixture

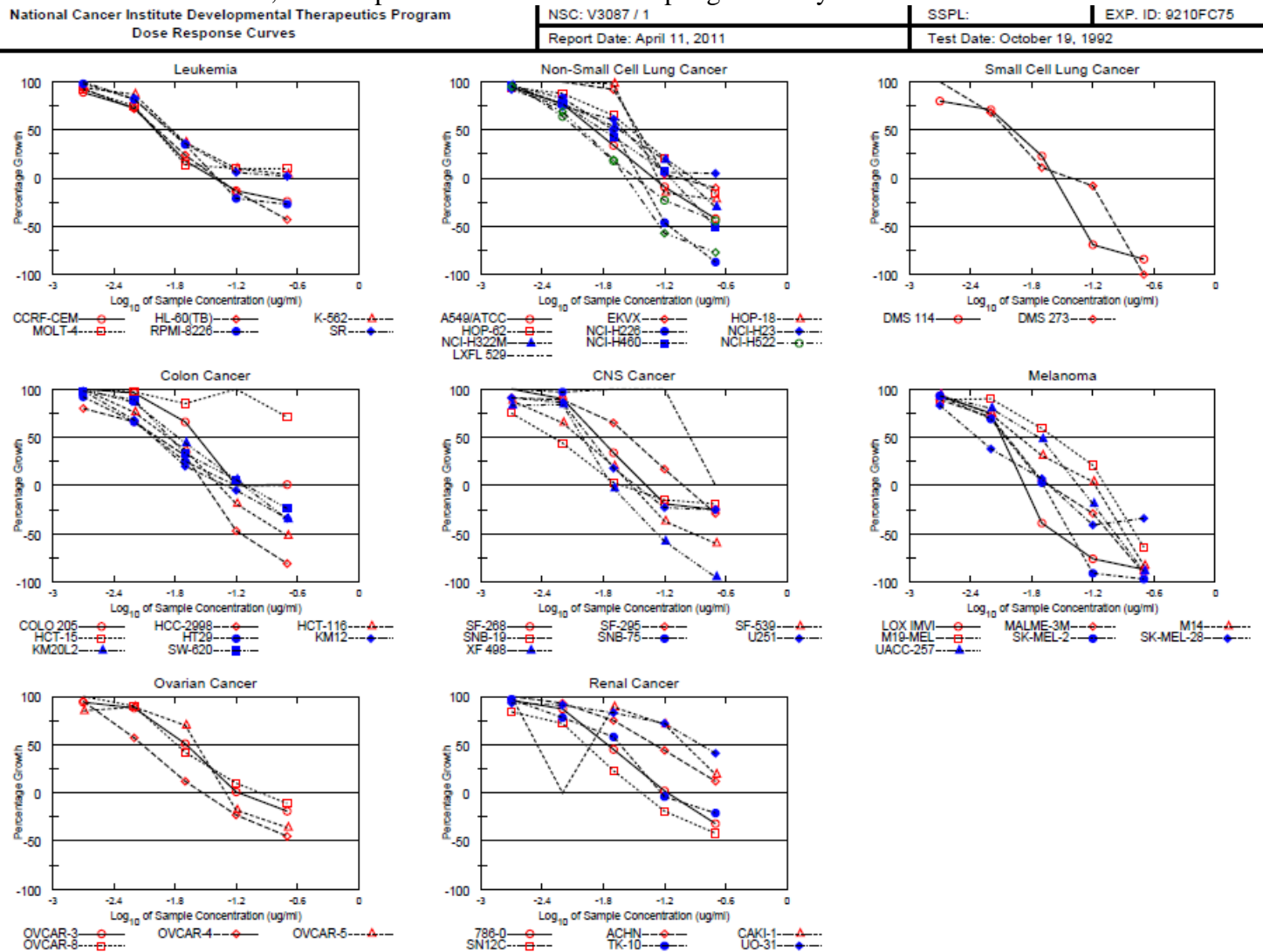


Figure S6. NCI 60-cell line screen, mean bar graphs for candidaspongolide acyl ester mixture

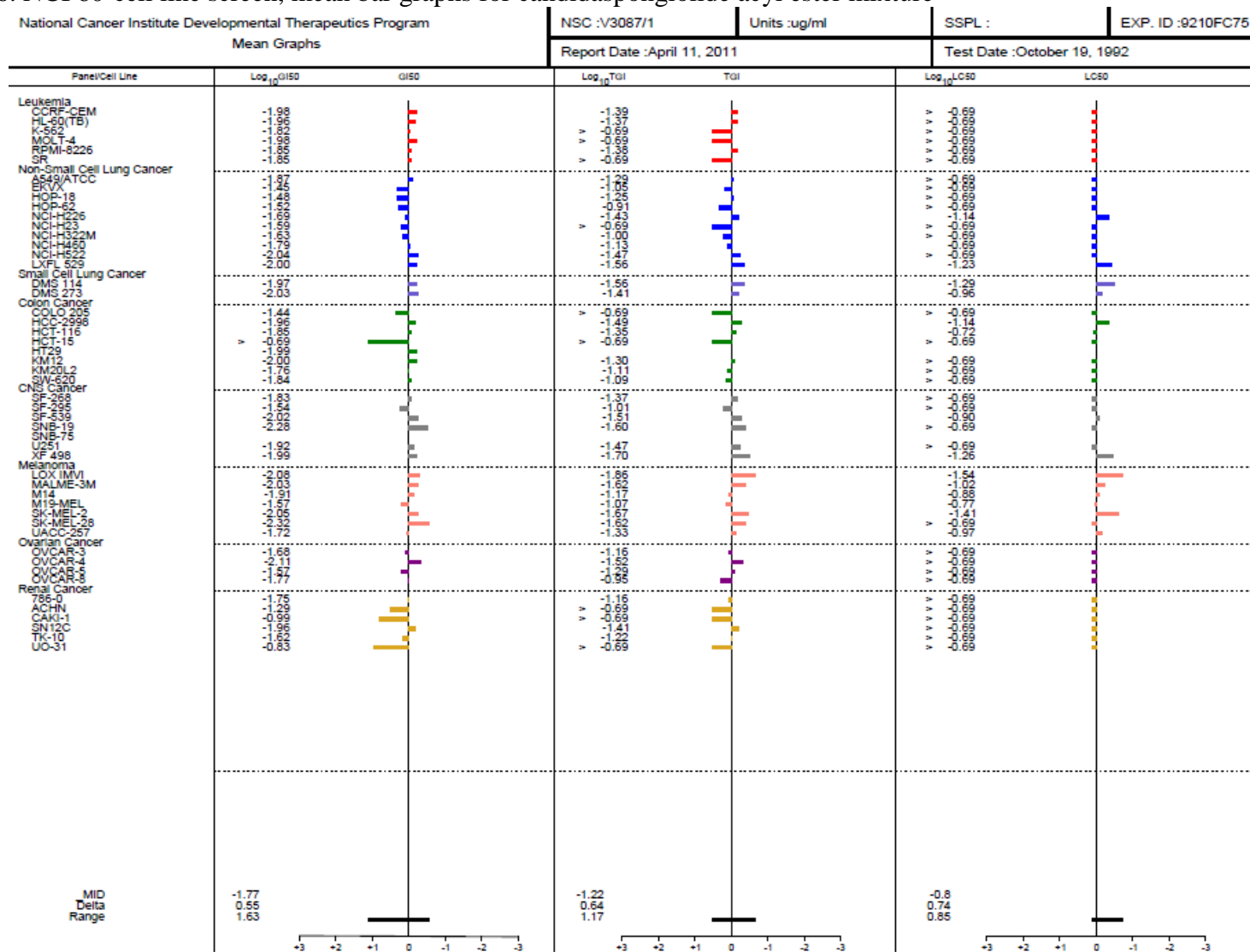


Figure S7. NCI 60-cell line screen, dose response curves for candidaspongolide A (macrolide core)

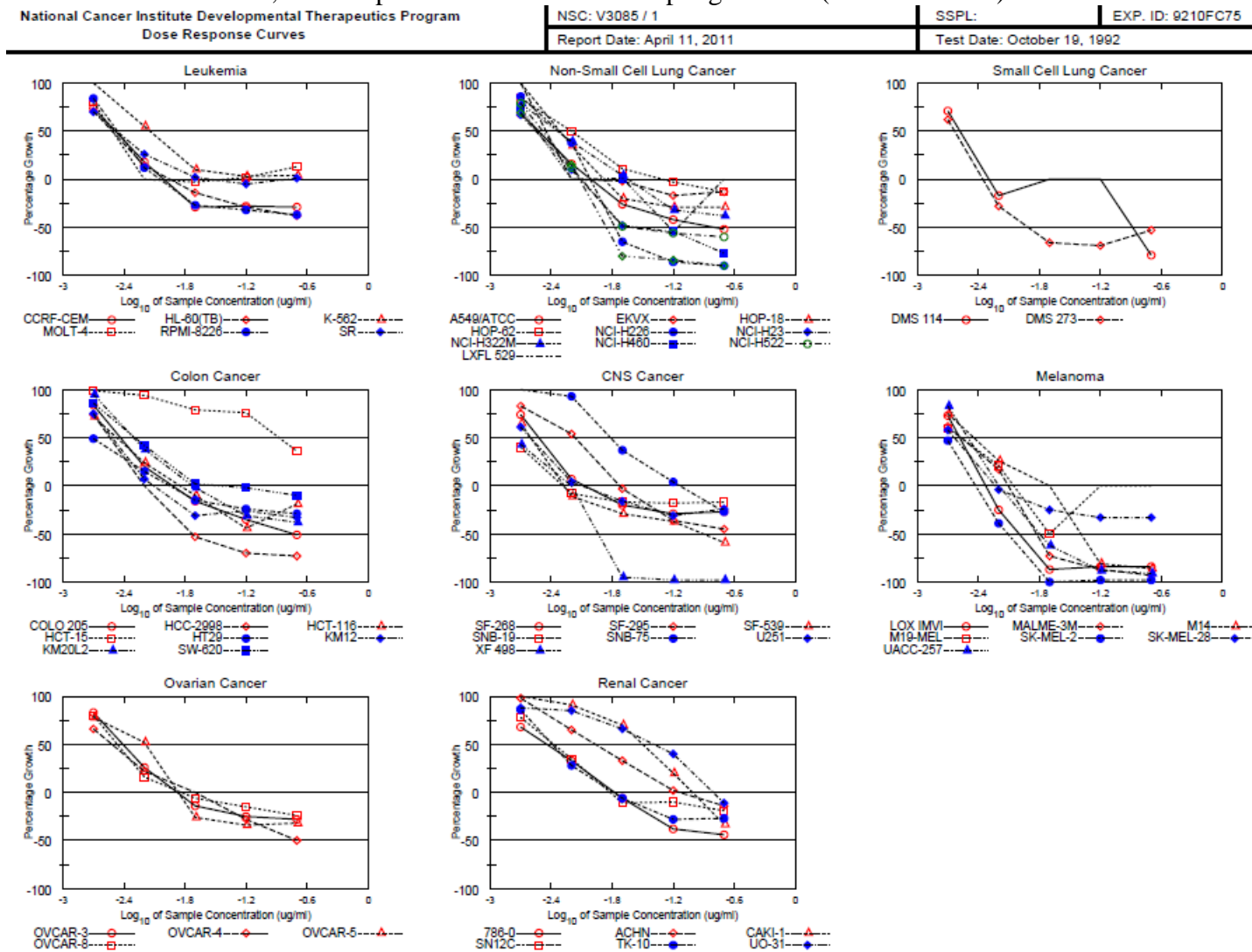


Figure S8. NCI 60-cell line screen, mean bar graphs for candidaspongolide A (macrolide core)

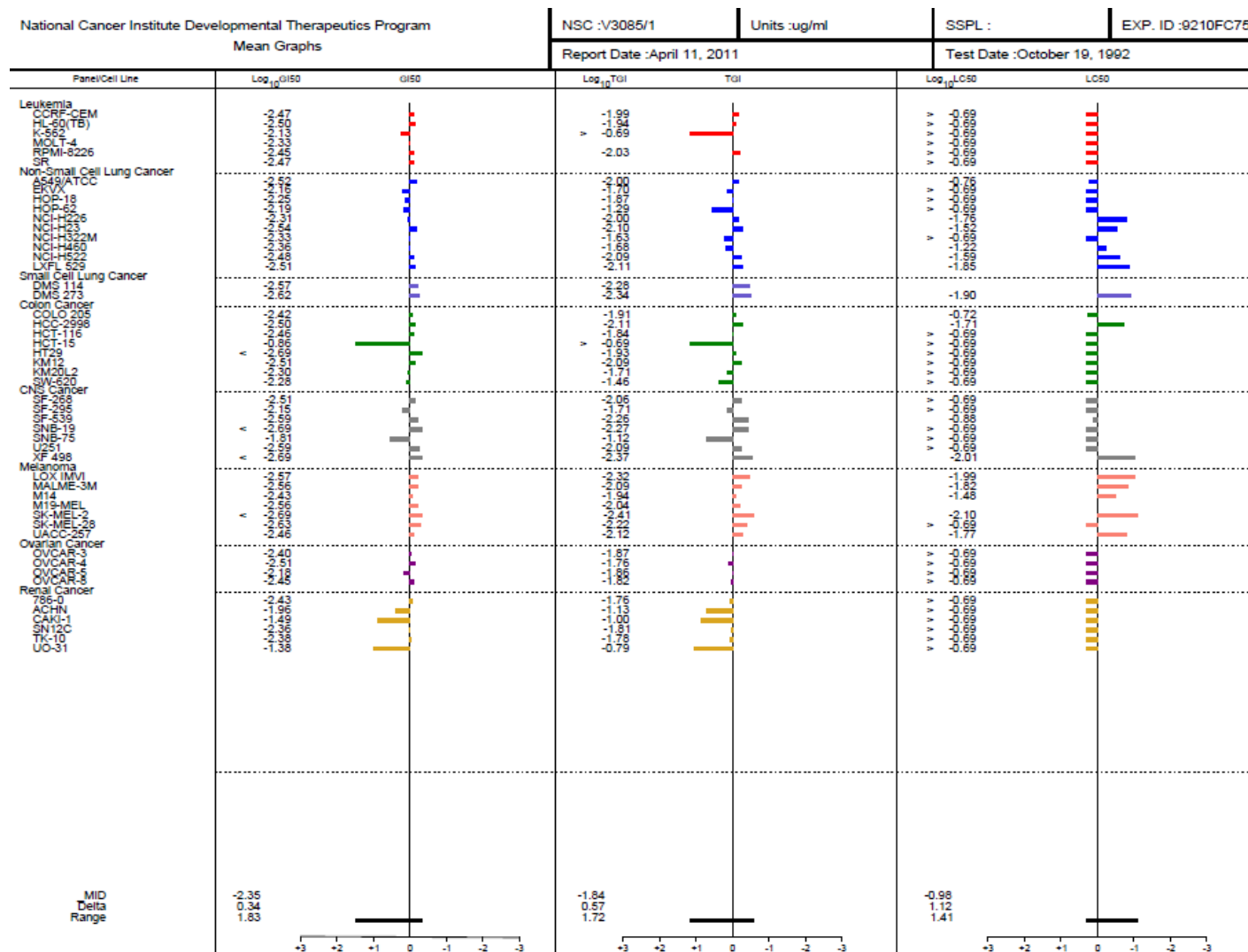


Table S7. Melanoma GI₅₀'s for candidaspongiodide acyl ester mixture

Melanoma Cell Line	GI ₅₀ (nM) ^a
LOX IMVI	9.0
MALME-3M	10.1
M14	13.3
M19-MEL	29.0
SK-MEL-2	9.6
SK-MEL-28	5.2
UACC-257	20.6
Melanoma Average	13.8
Mean GI ₅₀ All NCI-60 Cell Lines	18.3

^a Based on an estimated average mass of 927.1, calculated utilizing the reported fatty acid compositions (894.5 = 4.4%; 908.5 = 10.6%; 922.6 = 47.3%; 936.6 = 22.7%; 948.6 = 5.2%; 950.6 = 9.8%). Note: 10% of the fatty acid mixture was unidentified and percentages have been re-calculated to total 100%.⁸

Table S8. Melanoma GI₅₀'s for candidaspongiodide A (macrolide core)

Melanoma Cell Line	GI ₅₀ (nM)
LOX IMVI	3.9
MALME-3M	4.0
M14	5.4
M19-MEL	4.0
SK-MEL-2	< 3.0
SK-MEL-28	3.4
UACC-257	5.1
Melanoma Average	< 4.1
Mean GI ₅₀ All NCI-60 Cell Lines	6.5

⁸ Meragelman, T. L.; Willis, R. H.; Woldemichael, G. M.; Heaton, A.; Murphy, P. T.; Snader, K. M.; Newman, D. J.; van Soest, R.; Boyd, M. R.; Cardellina II, J. H.; McKee, T. C. *J. Nat. Prod.* **2007**, *70*, 1133.

Figure S9. Melanoma IC₅₀ curves for 1/2, 3/4, 5-8

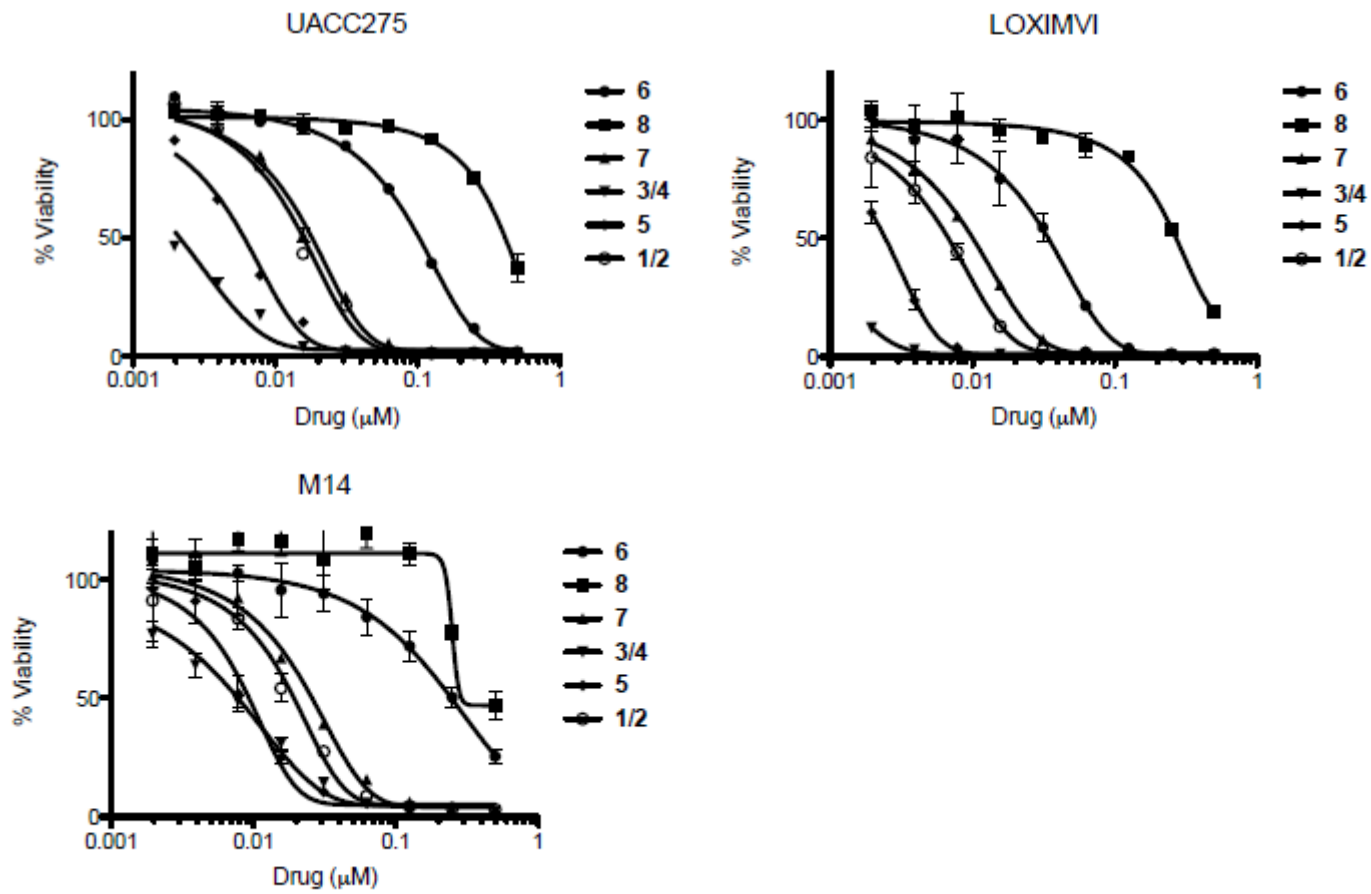


Figure S10. Breast and Lung IC₅₀ curves for 1/2, 3/4, 5-8

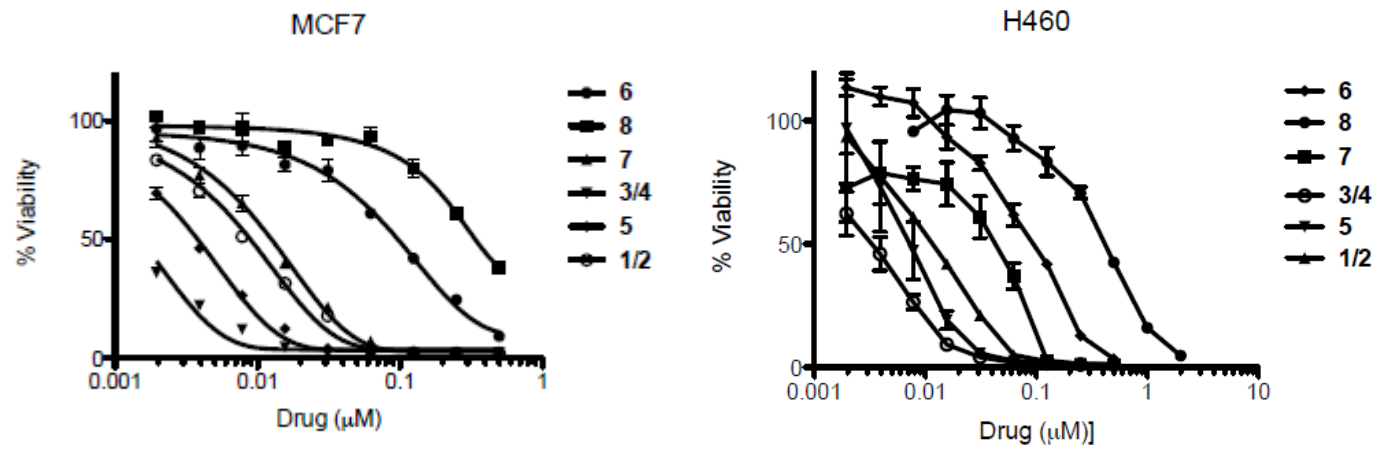
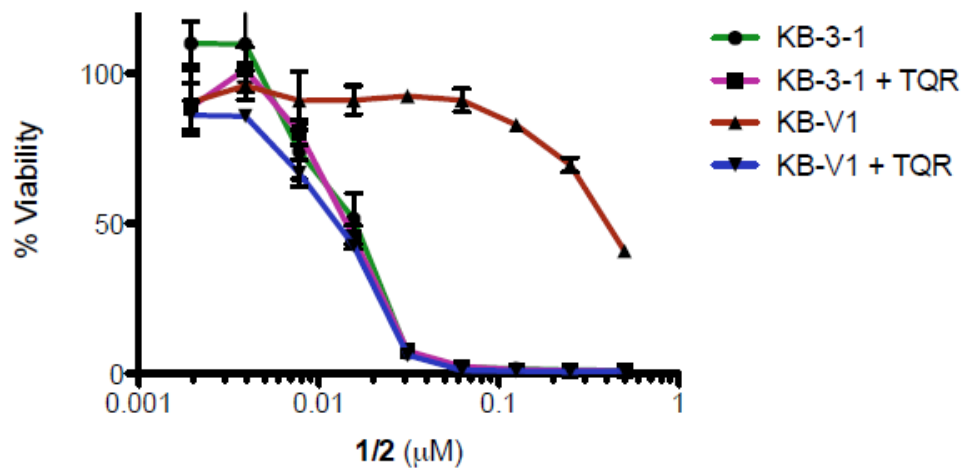


Figure S11. Precandidaspongiolides A (1) and B (2) are P-gp substrates



	1/2 (nM)	1/2 + TQR (nM)
KB-3-1	15.7 ± 4.9	14.7 ± 1.5
KB-V1	419.3 ± 15.0	13.1 ± 0.8
RR	26.7	N/A