

The sulfate-reducing bacterium *Desulfovibrio desulfuricans* ND132 as a model for understanding bacterial mercury methylation

Cynthia C. Gilmour, Dwayne A. Elias, Amy M. Kucken, Steven D. Brown, Anthony V. Palumbo, Christopher W. Schadt and Judy D. Wall

SUPPLEMENTAL MATERIAL

Table S1. Trace metal solution for SRM medium, 100X Stock

Trace metal	μM
ZnCl ₂	500
MnCl ₂ ·4H ₂ O	500
H ₂ BO ₄	800
CoCl ₂ ·6H ₂ O	1000
CuCl ₂ ·2H ₂ O	100
NiCl ₂ ·6H ₂ O	100
Na ₂ MoO ₄ ·2H ₂ O	150

Table S2. Vitamin Stock solution for SRM medium, 100X Stock

Vitamin	mg/L
Biotin	2
Nicotinic acid	20
<i>p</i> -aminobenzoic acid	10
Thiamine	20
Pantothenic acid	10
Pyridoxine·2HCl	50
Cobalamine	10

Table S3. List of *Desulfovibrio* strains used in alignment, including the accession numbers for the 16S rRNA sequences used.

Strain	Sequence source	Accession #	DSM	ATCC
<i>D. acrylicus</i>	RDP	U32578	10141	
<i>D. aerotolerans</i>	RDP	AY746987	16695	
<i>D. aespoeensis</i>	RDP	X95230	10631	
<i>D. africanus strain ADRI3</i>	RDP	AM419445.1		
<i>D. africanus strain Benghazi</i>	RDP	X99236	2603	
<i>D. alaskensis</i>	RDP	Y11984	16109	
<i>D. alcholorans</i>	RDP	AF053751	5433	
<i>D. alkalitolerans</i>	RDP	AY649785	16529	
<i>D. aminophilus</i>	RDP	AF067964	12254	
<i>D. bastinii</i>	RDP	AY359868	16055	BAA-903
<i>D. bizertensis</i>	RDP	DQ422859	18034	
<i>D. burkinensis</i>	RDP	AF053752	6830	
<i>D. caledoniensis</i>	RDP	U53465		
<i>D. carbinolicus</i>	RDP	AY626035	3852	
<i>D. carbinoliphilus</i>	RDP	DQ186200	17524	
<i>D. cuneatus</i>	RDP	X99501	11391	
<i>D. dechloroacetivorans</i>	RDP	AF230530		
<i>D. desulfuricans ND132</i>	GenBank/SERC	HQ693571.1		
<i>D. desulfuricans subs. aestuarii strain Styl 3</i>	RDP/SERC	FJ655909	17919	29578
<i>D. desulfuricans subsp. desulfuricans strain El Agheila</i>	RDP/SERC	M37316.1	1926	none
<i>D. desulfuricans subsp. desulfuricans strain Essex 6</i>	RDP	AF192153.1	642	29577
<i>D. desulfuricans subsp. desulfuricans strain MB</i>	RDP	AF192154.1	6949	27774
<i>D. desulfuricans strain G20</i>	RDP	NC_007519.1		
<i>D. desulfuricans strain LS</i>	R. Devereux			
<i>D. ferrireducens strain 61T</i>	RDP	DQ148944	16995	
<i>D. frigidus strain 18T</i>	RDP	DQ148943	17176	
<i>D. fructosivorans</i>	RDP	AF050101	3604	
<i>D. gabonensis</i>	RDP	U31080	10636	
<i>D. gigas</i>	RDP	DQ447183	1382	19364
<i>D. gracilis</i>	RDP	U53464	16080	BA-904
<i>D. halophilus</i>	RDP	U48243	5663	
<i>D. hydrothermalis</i>	RDP	AF458778	14728	
<i>D. indonesiensis</i>	RDP	Y09504	15121	
<i>D. inopinatus</i>	RDP	AF177276	10711	
<i>D. intestinalis</i>	RDP	Y12254	11275	
<i>D. littoralis</i>	RDP	X99504	11393	
<i>D. longreachensis</i>	RDP	Z24450		
<i>D. longus</i>	RDP	AY359867	6739	51456
<i>D. magneticus</i>	RDP	D43944		
<i>D. marinisediminis</i>	RDP	AB353727	17456	
<i>D. marinus</i>	RDP	DQ365924	18311	
<i>D. marrakechensis</i>	RDP	AM947130	19337	
<i>D. mexicanus</i>	RDP	AF227984	13116	
<i>D. oxamicus</i>	RDP	DQ122124	1925	33405
<i>D. oxycliniae</i>	RDP	U33316	11498	
<i>D. pasquesii</i>	RDP	AY726757	16681	
<i>D. piger</i>	RDP	AF192152		29098
<i>D. psychrotolerans</i>	RDP	AM418397	19430	
<i>D. putealis</i>	RDP	AY574979	16065	
<i>D. salexigens</i>	RDP	M34401.1	2638	14822
<i>D. senezii</i>	RDP	AF050100	8436	
<i>D. sp BerOcl</i>	RDP	EU137840		
<i>D. sp. T2</i>	SERC/GenBank	HQ693572		
<i>D. sp. X2</i>	SERC/GenBank	HQ693573		
<i>D. sulfodismutans</i>	RDP	Y17764	3696	43913
<i>D. termidus</i>	RDP	X87409	5308	49858
<i>D. tunisiensis</i>	RDP	EF577029	19275	
<i>D. vietnamensis</i>	RDP	X93994	10520	
<i>D. vulgaris strain Hildenborough</i>	RDP	AF418179	644	29579
<i>D. zosteriae</i>	RDP	Y18049	11974	
<i>Desulfocurvus vexinensis</i>	RDP	DQ841177	17965	

Table S4. Compilation of literature reports of Hg methylation assays for *Desulfovibrio* species.

Strain	Culture ID			MeHg ^a production	Reference
	DSMZ	ATCC	Other		
<i>D. desulfuricans</i> LS				+	(1)
<i>D. desulfuricans</i> ND132				+	This study
<i>Desulfovibrio</i> T2				+	This study
<i>Desulfovibrio</i> X2				+	This study
<i>D. africanus</i> Benghazi	2603			+	(2, 6), this study
<i>D. africanus</i>			ADR 13	+	(6)
<i>D. desulfuricans aestuarii</i> Sylt3		29578	NCIB 9335	-	This study
<i>D. desulfuricans subsp. desulfuricans</i>	1924	13541	NCIB 9467	+	(3, 4)
<i>D. desulfuricans subsp. desulfuricans</i> MB	6949	27774	NCIB 12833	-	(6), this study
<i>D. desulfuricans subsp. desulfuricans</i> Essex 6	642	29577	NCIB 8307	-	This study
<i>D. desulfuricans subsp. desulfuricans</i> El Agheila Z	1926		NCIB 8380	-	This study
<i>D. desulfuricans subsp. desulfuricans</i> G20				-	This study
<i>D. desulfuricans</i> M8 and M9				+	(5)
<i>D. gigas</i>	1382	19364	NCIB 8332	-	This study
<i>D. salexigens</i>	2638	14822		-	This study
<i>D. vulgaris</i> Hildenborough	644	29579		-	(6), this study
<i>D. vulgaris subsp. vulgaris</i> Marburg	2119			-	(2)
<i>D. strain BerOc1</i>			BerOc 1	+	(6)

^aMethylation is listed as positive if MeHg production was reported at levels significantly above abiotic or negative controls. Because rates were measured under different culture conditions, and over different amounts of time, a quantitative comparison of rates is not possible.

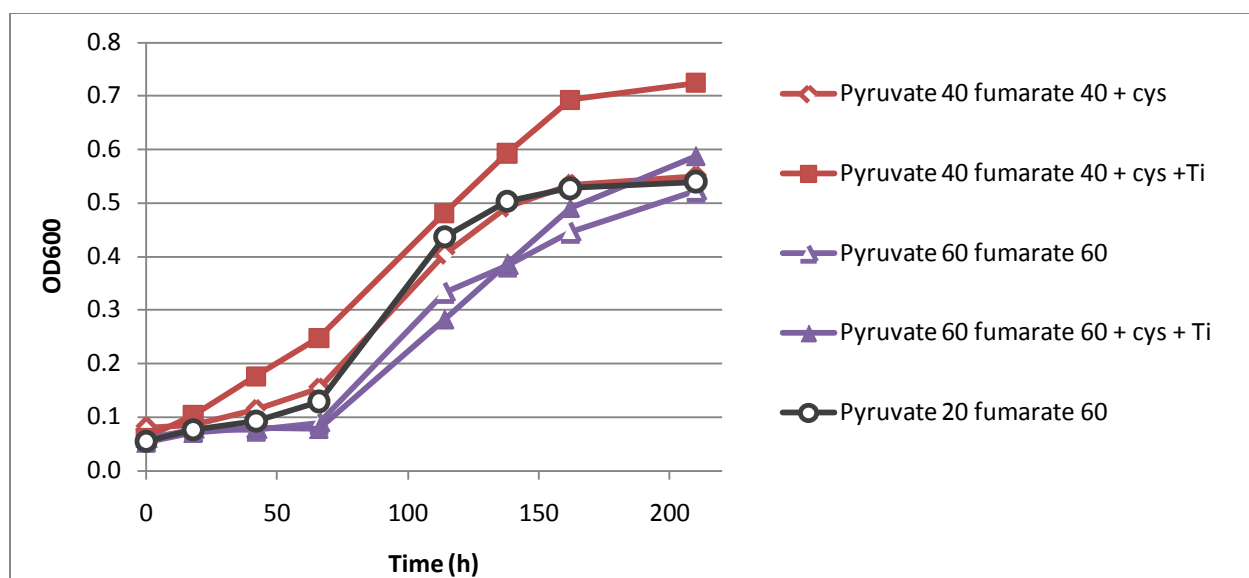


Figure S1. ND132 growth on pyruvate/fumarate medium (modified MOYPF) at 32° C. Cysteine was added to 1mM; Ti-citrate to 0.38 mM. Data points are averages for triplicate tubes.

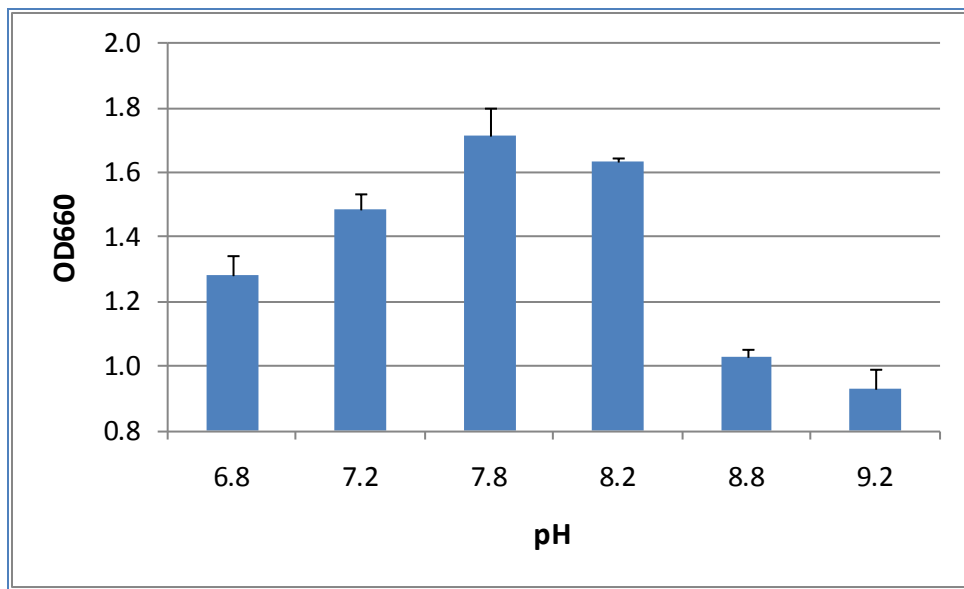
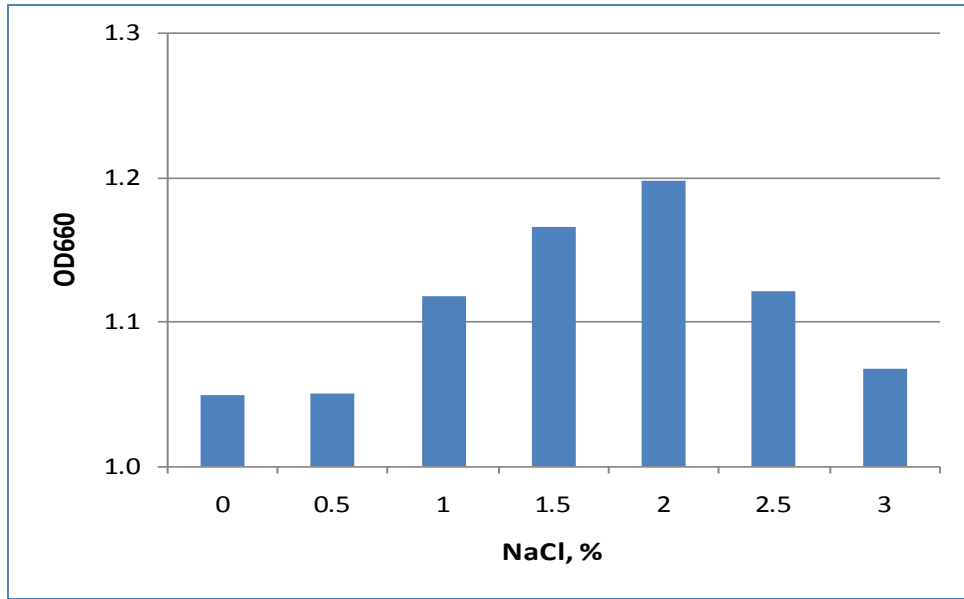


Figure S2. Optimal growth parameters for ND132 on lactate/sulfate medium (MOYLS4) at 32° C. Top, salt tolerance, bottom pH. Bars represent the average maximal optical density after growth to stationary phase (29 h) for duplicate cultures.

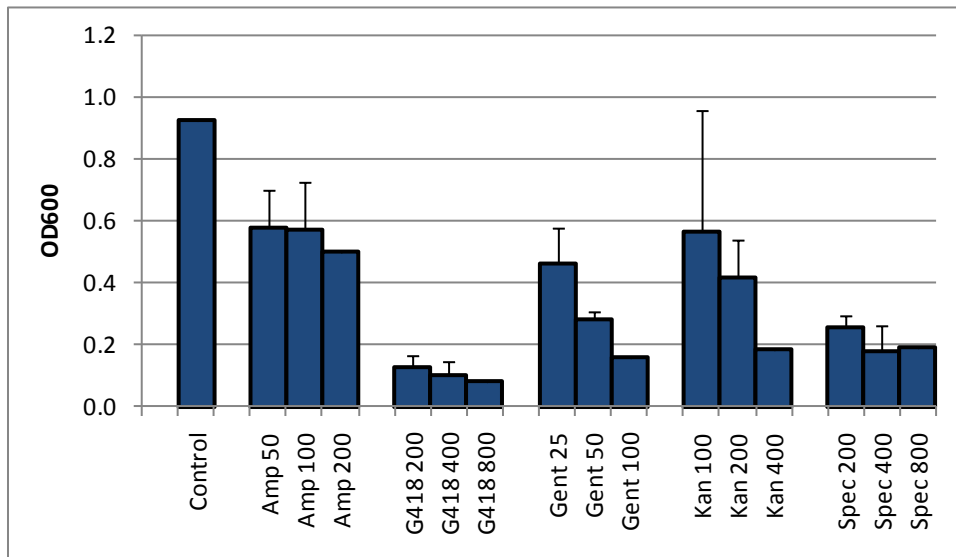


Figure S3: Antibiotic sensitivity of *D. desulfuricans* ND132, tested on lactate/sulfate (MOYLS4) medium. Growth is shown as optical density after 48 h growth at 32°C, minus the OD measured immediately after inoculation. Amp = ampicillin; Gen = gentamicin; Kan = kanamycin; Spec = spectinomycin. Antibiotic concentration values in µg/mL.

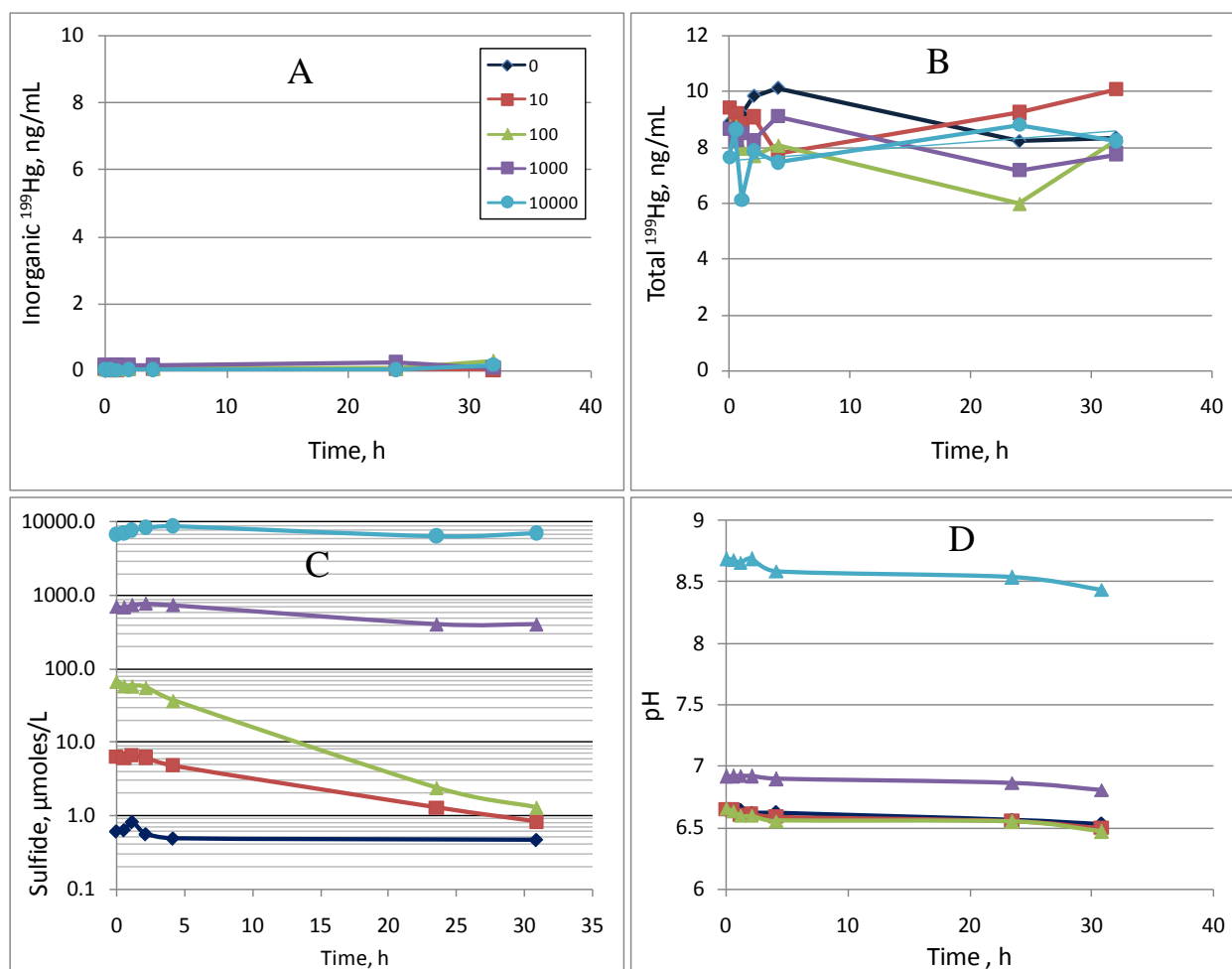


Figure S4. Lack of abiotic degradation of Me¹⁹⁹Hg by sulfide over time in un-inoculated culture medium. Me¹⁹⁹Hg was added at 10 ng/ml to un-inoculated SRM culture medium containing sulfide concentrations ranging from 10 to 1000 μM (plus a control without added sulfide), and the concentrations of inorganic ¹⁹⁹Hg (A), total ¹⁹⁹Hg (B), sulfide (C) and pH (D) were followed over 32 h. Media were reduced with 100 μM TiNTA.

Inorganic ¹⁹⁹Hg was measured directly by SnCl₂ reduction of samples; total ¹⁹⁹Hg was measured after the addition of BrCl. Bottom panels show sulfide and pH in the bottles over time. Additionally, on average, 98% of the total ¹⁹⁹Hg measured at the first time point was recovered from the bottles at the end of the experiment (after addition of BrCl to the bottles), demonstrating that MeHg was not degraded to elemental Hg.

REFERENCES

1. **Compeau, G. C., and R. Bartha.** 1985. Sulfate-reducing bacteria - principal methylators of mercury in anoxic estuarine sediment. *Applied and Environmental Microbiology* **50**:498-502.
2. **Ekstrom, E. B., F. M. M. Morel, and J. M. Benoit.** 2003. Mercury methylation independent of the acetyl-coenzyme A pathway in sulfate-reducing bacteria. *Applied and Environmental Microbiology* **69**:5414-5422.
3. **King, J. K., J. E. Kostka, M. E. Frischer, and F. M. Saunders.** 2000. Sulfate-reducing bacteria methylate mercury at variable rates in pure culture and in marine sediments. *Applied and Environmental Microbiology* **66**:2430-2437.
4. **King, J. K., J. E. Kostka, M. E. Frischer, F. M. Saunders, and R. A. Jahnke.** 2001. A quantitative relationship that demonstrates mercury methylation rates in marine sediments are based on the community composition and activity of sulfate-reducing bacteria. *Environmental Science & Technology* **35**:2491-2496.
5. **Lin, C. C., and J. A. Jay.** 2007. Mercury methylation by planktonic and biofilm cultures of *Desulfovibrio desulfuricans*. *Environmental Science & Technology* **41**:6691-6697.
6. **Ranchou-Peyruse, M., M. Monperrus, R. Bridou, R. Duran, D. Amouroux, J. C. Salvado, and R. Guyoneaud.** 2009. Overview of mercury methylation capacities among anaerobic bacteria including representatives of the sulphate-reducers: Implications for environmental studies. *Geomicrobiology Journal* **26**:1-8.