Supplementary Figure S1



Supplementary Figure S2



NOD

NOD.B6 Idd10



Α

Β

С



Supplementary Figure S4



Α

Supplementary Figure S5





Supplemental Figures:

Figure S1: Genetic map of the *Idd10* and *Idd18* regions on chromosome 3.

The NOD congenic strains used in this study and their respective introgressed regions are displayed. The NOD.B6 *Idd10/18* congenic strain (lines 1101 and 7754) has two noncontiguous, introgressed B6 regions encompassing *Idd10* and *Idd18*, whereas the NOD.B6 *Idd10* (line 3538) and NOD.B6 *Idd18* congenic strain (line 3539) have single introgressed B6 regions at *Idd10* and *Idd18*, respectively. The *Idd10* region consists of seven genes, five pseudogenes and one non-coding RNA (44, 47).

Figure S2: The severity of liver disease and the duration of bacterial persistence correlate with the level and extent of CD101 expression after infection

(A) Chronic liver disease in NOD and NOD.B6 *Idd10* mice.

Portal inflammation in 10-week-old (20x) NOD (left panel) and NOD.B6 *Idd10* (right panel) mice infected with *N. aro* at the age of 1 month.

(B) Reduction of CD101 protein expression on Gr1+ cells from heterozygous CD101^{+/-} mice.

Expression levels of CD101 on Gr1+ liver cells from 8-week-old B6 CD101^{+/+}, B6 CD101^{+/-} and B6 CD101^{-/-} mice. Respective FACS dot plots for large granular cells and the mean MFI for CD101 are displayed. Similar results were obtained for CD11c+ and TCR β + FoxP3+ cells (not shown).

(C) Prolonged bacterial persistence in B6 CD101^{-/-} compared to B6 CD101^{+/+} mice.

Copy numbers of *N. aro* in individual livers of the indicated mouse strains were analyzed by a *N. aro*-specific 16S rRNA qPCR. *N. aro* copies in the livers of B6 CD101^{+/+} and B6 CD101^{-/-} mice were compared and analyzed at the indicated weeks after infection. Statistical significance for two combined independent experiments was calculated with organs from 3 individual mice for each strain using a student's t-test. The p-value was <0.01 for the comparison at week 7. At week 8 the infection was completely cleared from the livers of both mouse strains.

Figure S3: Downregulation of CD101 expression on DCs upon infection and inverse correlation between CD101 expression and the expression of activation markers on T cells

(A+B) CD101 expression levels on regulatory T cells and DCs before and after infection.

The mean fluorescence intensity (MFI) of CD101 on TCR β + Foxp3+ (A) and CD11c+ (B) cells from the livers of uninfected and infected NOD and NOD.B6 *Idd10* mice was determined. 8-week-old, female mice of each strain were infected with 5x10⁷ *N. aro* cfus or received PBS 2 days before FACS analysis. Note the decreased CD101 expression levels in CD11c+ cells, while the MFIs remain unaffected on TCR β + cells upon infection (respective data are displayed for the FoxP3 positive T cell population, but also the FoxP3 negative population (that consisted mainly of CD44+ T cells) did not reveal any change in CD101 expression levels (41.1 in uninfected mice vs 42.3 in infected mice). Respective dot plot samples for TCR β + Foxp3+ (A) and CD11c+ (B) cells from uninfected (left column) and infected (middle column) NOD mice as well as the summary (right column) for 4 individual NOD and NOD.B6 *Idd10* mice are displayed. *In vivo* data for the suppression of CD101 expression on Gr1+ cells are shown in the bone marrow chimera studies in Fig. S6A.

(C+D) Inverse correlation between CD101 expression and the expression of activation markers on T cells

(C) CD101-negative T cells express IFN-γ and IL-17 during early stages after infection.

Ex vivo FACS analysis and intracellular cytokine staining (ICS) of liver T cells 60 hours after infection with $5 \times 10^7 N$. *aro* cfus. Dot plots are representative for three NOD.B6 *Idd10* mice tested individually.

(D) CD44 expressing T cells in NOD.B6 *Idd10* mice are CD101-negative. *Ex vivo* FACS analysis of liver T cells two weeks after the second infection. Dot plots are representative for three NOD and NOD.B6 *Idd10* mice tested individually.

Figure S4: DC subpopulations in uninfected and infected NOD and NOD.B6 Idd10 mice

(A) DCs were divided by FACS analysis into lymphoid (CD8+), myeloid (CD11b+) and plasmacytoid (B220+) subpopulations as well as CD11c positive cells expressing none of these additional markers (N.A.M.). Their recruitment was determined before and 5 days after infection in the livers of 8-week-old NOD and NOD.B6

Idd10 mice. Summaries for the absolute number (left panel) and the relative distribution (right panel) of each subpopulation are displayed for uninfected and infected NOD.B6 *Idd10* mice. In NOD mice, there was also an increase in all of these DC subpopulations upon infection without a significant change in the proportion of the subpopulations compared to NOD.B6 *Idd10* mice.

(B) Expression of CD101 on different DC subpopulations

CD11b+, B220+ and CD8+ DC populations were analyzed for their CD101 expression. Note that B220+ (plasmacytoid) DCs form the DC population that preferentially express CD101 in the liver, while the main CD101+ DC population in the spleen consists of CD11b+ CD11c+ cells.

Figure S5: Decreased CD101 expression correlates with increased numbers of N. aro.

(A-C) Long-term suppression of CD101 on DCs is liver-specific

Analysis of CD101 expression on DCs from the livers (A), pancreatic lymph nodes (B) and spleens (C) in 14 week-old NOD.B6 *Idd10* mice. Uninfected animals (0 weeks) and mice infected with *N. aro* 2 and 8 weeks before were compared. One representative histogram of three individual livers (A) and spleens (C) and an average of pancreatic lymph nodes of five individual mice (B) are displayed.

(D+E) DCs infected with *N. aro* exhibit reduced expression of CD101.

(D) CD11c+ cells were purified from the livers of three individual NOD mice three days after infection with $5x10^7$ *N. aro* cfus and separated into CD101+ and CD101- cells by cell sorting. DNA of $1x10^6$ CD101+ and CD101- DCs was extracted and tested for the presence of *N. aro* copies using a 16S rRNA specific qPCR. Data are displayed as number of *N. aro* copies of 100 CD101+ or CD101- DCs each. Statistical significance was calculated using a student's t-test and indicated as * for p<0.05.

(E) CD101-deficiency does not affect bacterial uptake

Bone marrow-derived DCs from wild-type B6 and B6 CD101^{-/-} mice were exposed to *N. aro* for 12 hours and 100 μ g/ml gentamicin for an additional hour followed by 10 μ g/ml gentamicin for the duration of the assay. Viable intracellular bacteria were quantitated by gentle lysis of the macrophages and subsequent plating on LB

agar. Experiments were performed two times.

Figure S6: Allele-intrinsic downregulation of CD101 expression on Gr1+ cells upon infection.

(A) CD101 expression on Gr1+ cells from NOD CD45.2/NOD.B6 *Idd10* bone marrow chimeras after infection *in vivo*.

CD101 expression was analyzed on Gr1+ cells from uninfected or infected - 2 days after infection - NOD CD45.2/NOD.B6 *Idd10* bone marrow chimeras. Note the enhanced CD101 expression in Gr1+ cells expressing the B6 CD101 allele from naïve chimeras and the reduced CD101 expression in Gr1+ cells expressing the B6 CD101 allele from infected chimeras.

(B) CD101 expression on Gr1+ cells after infection *in vitro*.

Liver mononuclear cells from 3 individual NOD.B6 *Idd10* mice were depleted of B and NK cells using MACS. The remaining cells were infected at a MOI of 10 with *N. aro* or left untreated and analyzed two days later for CD101 expression. Respective FACS dot plots for Gr1+ cells are displayed. While the expression of CD101 on Gr1+ cells decreased, CD11b expression remains unchanged.