Supplementary Table 1: List of genes up and down regulated in adipose tissue of mice after sCD14 treatment.

	FoldChange					
probe_id	gene_id	symbol	(absolut)	logFC	t	P.Value
1419127_at	109648	Npy	3,638	-1,863	-3,253	0,003423
1422153_a_a	68854	Asb11	3,406	-1,768	-3,832	0,000821
1450346_at	14765	Gpr50	3,125	1,644	2,970	0,006739
1452124_at	11735	Ank3	2,989	-1,580	-3,296	0,003081
1457435_x_a	17930	Myom2	2,741	-1,455	-3,120	0,004714
1434909_at	52187	Rragd	2,603	-1,380	-5,295	0,000020
1439036_a_a	11931	Atp1b1	2,563	-1,358	-3,006	0,006192
1454752_at	666794	Rbm24	2,548	-1,350	-4,275	0,000270
1449398_at	66211	Rpl3I	2,541	-1,346	-4,621	0,000112
1418095_at	66106	Smpx	2,414	-1,271	-3,200	0,003890
1418373_at	56012	Pgam2	2,310	-1,208	-2,911	0,007727
1426615_s_a	234593	Ndrg4	2,279	-1,189	-2,953	0,007002
1448182_a_a	12484	Cd24a	2,269	-1,182	-2,857	0,008777
1450826_a_a	20210	Saa3	2,262	1,177	2,971	0,006726
1422644_at	50795	Sh3bgr	2,189	-1,130	-4,200	0,000326
1451152_a_a	11931	Atp1b1	2,185	-1,128	-2,997	0,006320
1438175 x a	17930	Myom2	2,180	-1,125	-4,188	0,000336
1423890 x a	11931	Atp1b1	2,142	-1,099	-3,031	0,005827
1424616 s a	233575	Pgap2	2,115	-1,081	-2,903	0,007880
1450917 at	17930	Myom2	2,095	-1,067	-2,989	0,006445
1417889 at	11811	Apobec2	2,082	-1,058	-2,983	0,006528
1418589 [°] aa	17349	MIf1	2,078	-1,055	-3,394	0,002430
1460318 at	13009	Csrp3	2,058	-1,041	-2,847	0,008993
1417951 at	13808	Eno3	2,053	-1,038	-2,993	0,006383
1418453 [°] aa	11931	Atp1b1	2,053	-1,038	-3,073	0,005271
1438399 at	58869	Pex5I	2,040	1,028	3,173	0,004156
1457275 at	233335	Synm	2,002	-1,002	-3,889	0,000713
1417715 [°] aa	14719	Got2	1,995	-0,996	-3,023	0,005943
1419762 at	24108	Ubd	1,979	0,984	2,859	0,008744
1419606 [°] aa	21955	Tnnt1	1,967	-0,976	-3,948	0,000616
1417634 at	20657	Sod3	1,965	0,975	3,817	0,000854
1451721 [°] aa	14961	H2-Ab1	1,941	0,957	3,681	0,001197
1449308 at	12274	C6	1,928	0,947	3,228	0,003637
1449178 at	53318	Pdlim3	1,912	-0,935	-3,873	0.000742
1456180 at	666794	Rbm24	1,906	-0,930	-3,497	0,001889
1418849 x a	11832	Aqp7	1,880	0,911	3,059	0,005457
1425341 at	16527	Kcnk3	1,853	0,890	3,461	0,002061
1453355 at	75607	Wnk2	1,810	-0.856	-3.175	0,004134
1417633 at	20657	Sod3	1,777	0,830	3,732	0.001054
1459860 x a	80890	Trim2	1,763	-0.818	-3.152	0.004371
1447657 s a	68760	Synpo2l	1 755	-0.811	-3.072	0.005282

Supplementary Figure 1. A. Effect of high fat diet (HFD) on oral glucose tolerance test in mice after one month (triangles) or 3 months (crosses) versus mice fed a normal chow (NC, squares). Glucose tolerance impairs over time during HFD. **B**. Effect of one month HFD on glucose tolerance test in mice grafted with BM from WT mice (triangles) vs. mice which were not grafted (not irradiated, squares). The irradiation and grafting procedure has impaired the diabetogenic effect of the HFD. **C**. Effect of a 3 month HFD on oral glucose tolerance in mice which were grafted with BM from WT donor (triangles) or not grafted (not irradiated, squares). As for figure B, the impact of HFD on glucose tolerance is reduced. **D**. Impact of a one month HFD (squares) vs. NC (diamonds) on oral glucose tolerance. The data shows that the diabetogenic impact is lower in the grafted mice than in the non grafted mice (Figure A). It requires a longer period of time. The interpretation is that the irradiation has hampered the role of some cells in the development of glucose intolerance.



Supplementary Figure 2. **A.** Circulating sCD14 levels in the different models studied. **B.** Glucose intolerance following and oral glucose glucose challenge is significantly impaired (2 way ANOVA, Bonferoni Post hoc test, p<0.05) in wildtype mice grafted with bone marrow from the WT donor and fed a HFD when compared to the same group of mice but fed a NC. Conversely, in WT mice grafted with CD14 KO BM donor glucose intolerance was not observed. Similarly, CD14 receiver mice never became glucose intolerant in response to HFD. This suggests that both the hematopoietic cells and the cells from the body require CD14 to become glucose intolerant in response to a HFD. **C.** Fasting triglycerides and free fatty acids in the different models (* p<0.05).



Supplementary Figure 3. Confirmation by real-time PCR of the effect of CD14 treatment on gene expression in epididimal adipose tissue from control and CD14 treated *ob/ob* mice. Gene expression of defense response (A) lipid related (B) and proinflammatory (C) proteins in control (C, white bars) and CD14 treated ob/ob mice (CD14, black bars) are shown. Results are expressed as mean \pm SEM relative to the control value set as 1.0 (n=6). Statistical analyses comparing gene expression in control vs. CD14 treated ob/ob mice were performed by paired t test. Saa3, p=0.017; LBP, p=0.023; Cd74, p=0.018; Apoc1, p=0.027; Apoc2, p=0.0046; IL-1-alpha, p=0.01; IL-1-beta, p=0.05.

RNA from epididimal adipose tissue was extracted using a kit (RNeasy; Qiagen, Valencia, CA, USA). RNA (2 µg) was reverse-transcribed to cDNA using SuperscriptII enzyme (Invitrogen, USA). Real-time PCR was used to measure specific mRNAs (ABI-PRISM 7700 Sequence Detector; Perkin-Elmer Applied Biosystems, Foster City, CA, USA). All reactions were performed in 384-well optical plates (MicroAmp; Applied Biosystems, Foster City, CA, USA). Amplification mixes (10 µl) contained the diluted cDNA sample, 2X TaqMan Universal PCR Mastermix (Applied Biosystems) or Sybr green PCR Mastermix (Applied Biosystems), forward and reverse primers, and probe for the specific mRNAs. Glyceraldehyde 3-phosphate dehydrogenase (GAPDH) was used as endogenous control. Thermal cycling conditions included 10 min at 203,00°F before the onset of the PCR cycles, which consisted of 40 cycles at 95°C for 15 s and 65°C for 1 min. The oligonucleotide sequences for the primer pairs used for GAPDH was 5'-catggccttccgtgttccta -3' and 5'-gcggcacgtcagatcca -3'. The primers and probes for Saa3, Cd74, Cd24a, LBP, Apoc1, Apoc2, IL-1 alpha, IL-1 beta, IL-6 and TNF-alpha were supplied as a Mm00441203 m1. Mm00658576 m1. Mm00782538 sH. kit (TaqMan Reagents LBP. Mm00431816 m1, Mm00431816 m1, Mm00439620 m1 and Mm01336189 m1, Applied Biosystems) and used according to the manufacturer's instructions. The primers for IL-6 were: 5'-GCCCACCAAGAACGATAGTCA-3' 5'-CAAGAAGGCAACTGGATGGAA-3'; and and for TNFalpha: 5'-CACAAGATGCTGGGACAGTGA-3' and 5'-TCCTTGATGGTGGTGCATGA-3'. mRNA expression was calculated using the ΔC_1 method.



Supplementary Figures 4 and 5. Expression profiles of genes listed in Supplementary Table 1 were analyzed using The Ingenuity Pathway Analysis (IPA) methodology to compose a set of interactive networks, taking into consideration canonical pathways and the relevant biological interactions. Focus genes were defined as those with an absolute mean fold-change of at least 1.5. A number of canonical pathways came up as playing an important role, especially those related with Glycolysis and Gluconeogenesis and IL-4 signaling. Next, two significant biological networks were identified by IPA. Further analysis of the highest scored network (score 24, 14 focus genes) identified inflammatory response (p<0.028), Genetic Disorder (p<0.048) and Inflammatory Disease (p<0.0375) as the most significant biological functions linked to these networks.



