

Supporting Information

Chemoselective and Enantioselective Oxidation of Indoles Employing Aspartyl Peptide Catalysts

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Table of Contents

A.	General.....	3
B.	Preparation of 2-ArylindoleDerivatives.....	4
C.	Structure–Enantioselectivity Relationships for Peptide Catalyzed Indole Oxidation.....	17
D.	Peptide Sequence Optimization for Enantioselective Indole Oxidation.	18
E.	Preparation of Peptides 22 and <i>ent</i> -22.	19
F.	General Procedure for the Peptide Catalyzed Oxidation of 2,3-Disubstituted Indoles.....	21
G.	Analytical Data for Indole Oxidation Products.....	22
H.	X-Ray Structure of the Compound 41c and Proof of Absolute Stereochemistry.....	32
I.	Procedure for the Stereospecific Rearrangement of 3-Hydroxy-indolenines.....	33
J.	NMR Spectra.....	35
K.	X-Ray Diffraction Report for Compound 41c.....	77
L.	Chromatograms.	105
M.	References.....	123

A. General.

^1H NMR data were collected at 500 MHz or 400 MHz. ^1H NMR chemical shifts are reported in parts-per-million (δ , ppm) relative to tetramethylsilane ($\delta = 0$ ppm) with residual CHCl_3 ($\delta = 7.26$ ppm) as the internal standard. ^1H NMR spectral data are reported as follows: chemical shift (multiplicity [singlet (s), doublet (d), triplet (t), quartet (q), quintet (p), multiplet (m), broad (br)], coupling constants [Hz], integration). Proton decoupled ^{13}C NMR spectra were recorded at 126 MHz or 100 MHz. ^{13}C chemical shifts are reported in parts-per-million (δ) relative to tetramethylsilane ($\delta = 0$ ppm), with the central line of the CDCl_3 triplet (77.23 ppm) serving as the internal standard. ^{19}F spectra were collected at 376 MHz. ^{19}F NMR chemical shifts are reported in parts-per-million (δ) relative to trichlorofluoromethane ($\delta = 0$ ppm) using monofluorobenzene as the internal standard ($\delta = -113.1$ ppm). All NMR data were collected at room temperature (23 °C).

Thin film and attenuated total reflectance (ATR) infrared (IR) spectra of neat samples were recorded on a FT-IR spectrometer; IR data (ν_{max} in cm^{-1}) is reported for diagnostic bands as well as other notable frequencies.

All of the samples were characterized by ultra high performance liquid chromatography-mass spectrometry (UPLC-MS) on an instrument equipped with a reverse-phase C18 column (1.7 μm particle size, 2.1 \times 50 mm), dual atmospheric pressure chemical ionization (API)/electrospray (ESI) mass spectrometry detector, or Direct Analysis in Real Time (DART) ionization, and a photodiode array detector.

Chiral analytical normal phase HPLC was performed at a column temperature of 20 °C on a chromatograph equipped with a diode array detector (210 nm, 230 nm, 250 nm, 254 nm, or 280 nm).

Optical rotations were recorded on a polarimeter at 546, 578, and 589 nm, 23°C, with a 0.5 dm path length sample holder. Concentration are given in g/100 mL.

Analytical and preparative thin-layer chromatography (TLC) was performed using pre-coated plates (0.25 mm thickness); TLC visualization was accomplished by irradiation with a UV lamp (254 nm) and/or staining with *p*-anisaldehyde or KMnO_4 solutions.

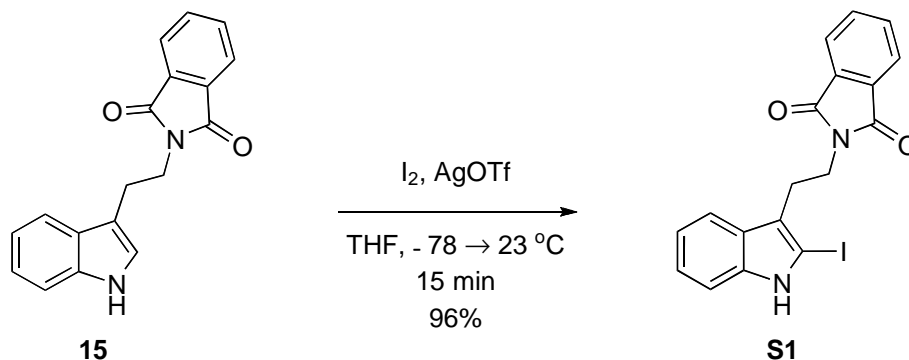
Flash column chromatography was performed using silica gel 60 Å (40-63 micron) or using an automated flash purification system. Gradient elution volumes are reported as column volumes (CV).

All yields refer to chromatographically and spectroscopically (^1H NMR) homogeneous materials unless indicated otherwise. Solvents were purified using a purification system. All commercially available materials were purchased from suppliers and used as received unless indicated otherwise.

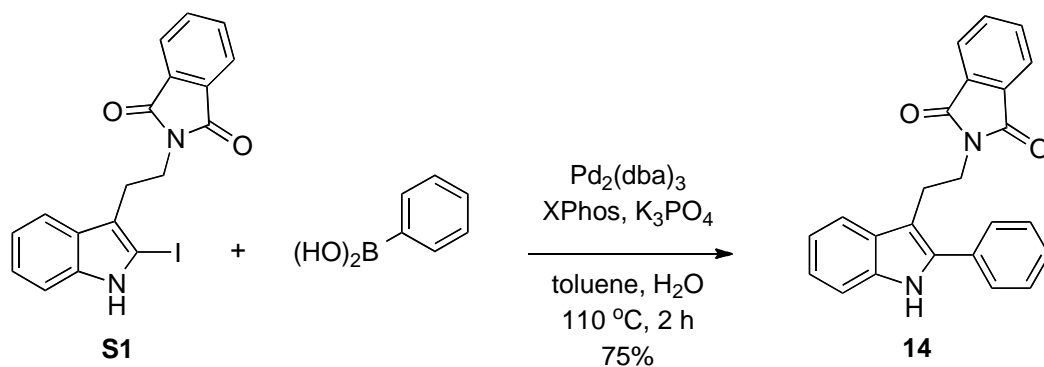
Abbreviations: AcOH = acetic acid, Boc = *tert*-butoxycarbonyl, CAM = ceric ammonium molybdate, DCM = dichloromethane, DMAP = *N,N*-dimethyl-4-aminopyridine, *N,N*-DMF = *N,N*-dimethylformamide, DIC = *N,N*-diisopropylcarbodiimide, EDC = 1-Ethyl-3-(3-dimethylaminopropyl)carbodiimide, EtOAc = ethyl acetate, Hex = Hexanes, HOBt = 1-Hydroxybenzotriazole, TEA = triethylamine, THF = tetrahydrofuran. Standard three-letter abbreviations used for proteinogenic amino acids.

B. Preparation of 2-Arylindole Derivatives.

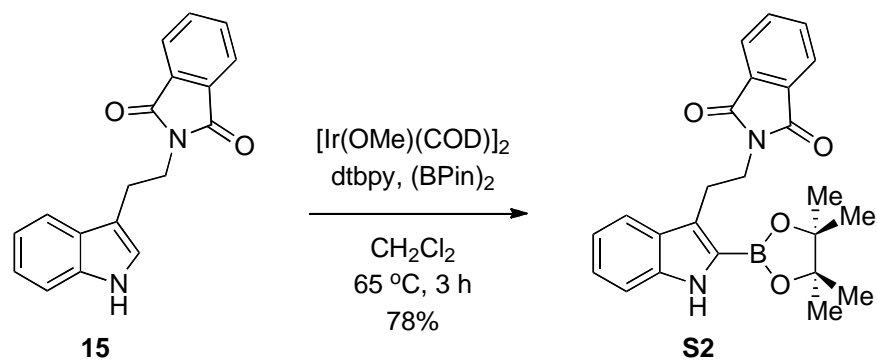
With the exception of compound **19** (*vide infra*) all substrates used in this study were prepared according to one of two routes outlined in Scheme 1. The following detailed procedures below are representative of the conditions used to prepare the desired substrates:



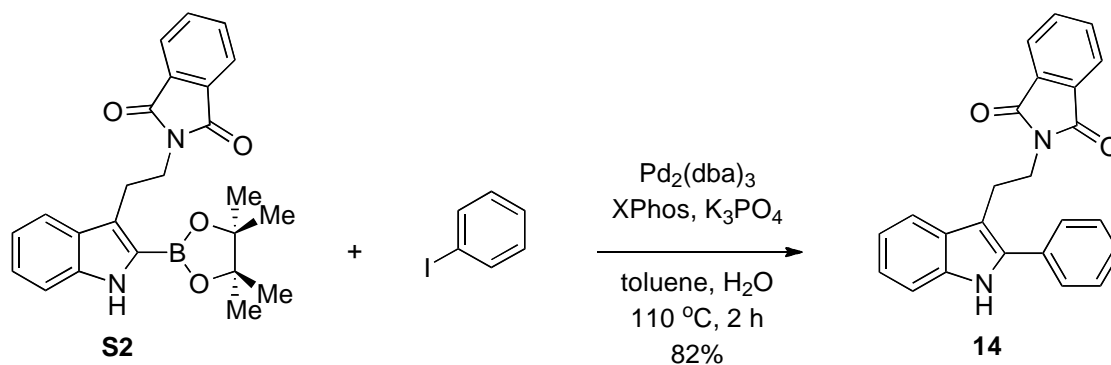
***N,N*-Phthaloyl-2-iodotryptamine (S1, Scheme 1 - Route a).** A flame dried 200-mL three-neck round bottom flask was charged with *N,N*-phthaloyltryptamine **15** (3.23 g, 11.1 mmol, 1 equiv.), iodine (3.12 g, 12.3 mmol, 1.11 equiv.), and a stirring bar and sealed under argon atmosphere. Anhydrous THF (110 mL) was added via syringe and the resulting mixture was agitated at 23 $^{\circ}C$ and then cooled to -78 $^{\circ}C$. After 15 min, AgOTf (3.12 g, 12.3 mmol, 1.11 equiv.) was added via a solid addition funnel. After 5 min, the reaction mixture became a fine yellow suspension and agitation was maintained. After 25 min, TLC analysis (30% EtOAc/hexanes) indicated completion of the reaction. Solid sodium bicarbonate (2.06 g, 24.5 mmol, 2.21 equiv.) was added and the cold bath was removed. After 30 min, the resulting yellow suspension was diluted with EtOAc (100 mL) and a mixture of saturated aqueous sodium thiosulfate-saturated aqueous sodium bicarbonate (1:1, 80 mL) at 23 $^{\circ}C$. The mixture was filtered through Celite and the filter cake was rinsed with EtOAc (300 mL) and the organic layers were combined. The combined organic layer was washed with brine (200 mL) and dried over solid anhydrous sodium sulfate. The organic layer was filtered and concentrated at reduced pressure on a rotary evaporator to afford **S1** as a yellow crystalline solid (4.45 g, 96%). The crude product was sufficiently pure to be used in subsequent reactions without further purification. A 1-gram sample was purified for characterization by flash column chromatography on silica gel (2.4 \times 14.0 cm, eluent 20% EtOAc/hexanes) to provide the product **S1** as a bright yellow solid (960 mg, 96% mass recovery). 1H NMR (500 MHz, $CDCl_3$, 20 $^{\circ}C$, J in Hz) δ 7.98 (br s, 1H, NH), 7.78 (dd, 2H, J = 3.0, 5.5 Hz, phthalimide), 7.66 (dd, 2H, J = 3.0, 5.5 Hz, phthalimide), 7.61 (d, 1H, J = 8.0 Hz, ArH), 7.25 (dt, 1H, J = 1.0, 8.0 Hz, ArH), 7.08 (dt, 1H, J = 1.0, 7.0 Hz, ArH), 7.03 (dt, 1H, J = 1.5, 7.0 Hz, ArH), 3.90 (t, 2H, J = 7.5 Hz, =C-CH₂CH₂N), 3.06 (t, 2H, J = 7.5 Hz, =C-CH₂CH₂N); ^{13}C NMR (126 MHz, $CDCl_3$, 20 $^{\circ}C$) δ 168.5, 139.0, 134.0, 132.4, 127.7, 123.4, 122.6, 120.3, 118.8, 118.1, 110.6, 78.5, 37.9, 26.2; FTIR (neat, cm^{-1}) 3352 (br m), 3050 (w), 2935(w), 1771 (m), 1701 (s), 1653 (m), 1558 (m), 1540 (m), 1396 (s), 1362 (m), 1338 (m), 1101 (w), 742 (w), 717 (m); HRMS (DART) calc'd for C₁₈H₁₂IN₂O₂ [M-H]⁻: 414.9949, found: 414.9965; TLC R_f = 0.50 (30% EtOAc/hexanes, UV, CAM).



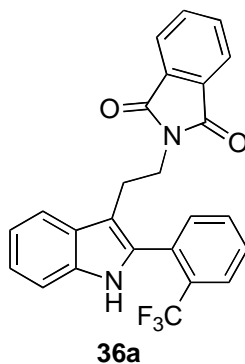
***N,N*-Phthaloyl-2-phenyltryptamine (14, Scheme 1 - Route a).** A flame dried 15-mL pressure flask was charged with 2-iodotryptamine **S1** (100 mg, 0.240 mmol, 1 equiv.), phenylboronic acid (45.3 mg, 0.360 mmol, 1.50 equiv.), $\text{Pd}_2(\text{dba})_3$ (5.5 mg, 6.0 μmol , 0.025 equiv.), XPhos (11.7 mg, 0.0240 mmol, 0.100 equiv.), anhydrous potassium phosphate tribasic (104 mg, 0.480 mmol, 2.00 equiv.), and a stirring bar and sealed with a septum under an atmosphere of argon. Toluene (2.4 mL) followed by deionized water (240 μL) were added via syringe and the mixture was agitated to give a clear red solution. The septum was replaced with the pressure flask's Teflon seal and the entire mixture was heated in an oil bath set to 110 $^\circ\text{C}$. After 5 min, the reaction mixture turned into a pale yellow clear solution and the mixture was maintained at 110 $^\circ\text{C}$. After 2 h, the oil bath was removed and the mixture was allowed to cool to 23 $^\circ\text{C}$ and TLC analysis (10% EtOAc in hexanes) indicated completion of the reaction. The volatiles were removed under reduced pressure and the crude mixture was purified by flash column chromatography on silica gel (3.0 \times 14.0 cm, eluent 15% EtOAc/hexanes) to afford the product **14** as yellow crystals (66.2 mg, 75%). $^1\text{H NMR}$ (500 MHz, CDCl_3 , 20 $^\circ\text{C}$, J in Hz) δ 8.08 (br. s, 1H, NH), 7.79 (d, 1H, $J = 7.5$ Hz, ArH), 7.75 (dd, 2H, $J = 3.5, 6.0$ Hz, phthalimide), 7.65 (dd, 2H, $J = 3.0, 5.5$ Hz, phthalimide), 7.58-7.55 (m, 2H, ArH), 7.40 (tt, 2H, $J = 1.5, 8.0$ Hz, ArH), 7.35 (dt, 1H, $J = 1.0, 8.0$ Hz, ArH), 7.29 (tt, 1H, $J = 1.0, 7.0$ Hz, ArH), 7.19 (tt, 1H, $J = 1.0, 8.0$ Hz, ArH), 7.14 (tt, 1H, $J = 1.0, 7.0$ Hz, ArH), 3.97 (t, 2H, $J = 8.0$ Hz, =C-CH₂CH₂N), 3.24 (t, 2H, $J = 8.0$ Hz, =C-CH₂CH₂N); $^{13}\text{C NMR}$ (126 MHz, CDCl_3 , 20 $^\circ\text{C}$) δ 168.4, 136.0, 135.4, 133.9, 132.8, 132.4, 129.3, 129.2, 128.0, 128.0, 123.3, 122.7, 120.2, 119.2, 111.1, 109.3, 38.6, 24.1; FTIR (neat, cm^{-1}) 3359 (br. m), 3050 (w), 3021 (w), 2942 (w), 2863 (w), 1766 (m), 1701 (s), 1446 (m), 1431 (m), 1396 (s), 1356 (m), 742 (s), 716 (s), 695 (s); HRMS (DART) calc'd for $\text{C}_{24}\text{H}_{17}\text{N}_2\text{O}_2$ $[\text{M}-\text{H}]^-$: 365.1296, found: 365.1301; TLC $R_f = 0.31$ (20% EtOAc/hexanes, UV, CAM).



***N,N*-Phthaloyl-2-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)tryptamine (S2, Scheme 1 - Route b).** A flame dried 15-mL pressure flask was charged with (1,5-cyclooctadiene)(methoxy)iridium(I) dimer (3.4 mg, 5.2 μmol , 0.015 equiv.), 4,4'-di-*tert*-butyl-2,2'-dipyridyl (2.8 mg, 1.0 μmol , 0.03 equiv.), bis(pinacolato)diboron (177 mg, 0.690 mmol, 2.00 equiv.), *N,N*-phthaloyltryptamine **15** (100 mg, 0.345 mmol, 1 equiv.), and a stirring bar and sealed with a septum under an atmosphere of argon. Anhydrous dichloromethane (2.2 mL) was added via syringe to give a colorless suspension. The septum was replaced with the pressure flask's Teflon seal and the entire mixture was heated in an oil bath set to 65 $^\circ\text{C}$. The reaction mixture gradually turned into a clear dark amber solution. After 3 h, the mixture was allowed to cool to 23 $^\circ\text{C}$ and the volatiles were removed under reduced pressure and the crude mixture was purified by flash column chromatography on silica gel (2.5 \times 11.0 cm, eluent 20% EtOAc/hexanes) to afford product **S2** as a pale yellow powder (112 mg, 78%). **^1H NMR** (500 MHz, CDCl_3 , 20 $^\circ\text{C}$, J in Hz) 8.39 (br. s, 1H, **NH**), 7.75 (dd, 2H, $J = 3.0, 5.0$ Hz, phthalimide), 7.72 (d, 1H, $J = 8.0$ Hz, **ArH**), 7.62 (dd, 2H, $J = 3.0, 5.5$ Hz, phthalimide), 7.29 (d, 1H, $J = 8.5$ Hz, **ArH**), 7.16 (t, 1H, $J = 7.5$ Hz, **ArH**), 7.03 (t, 1H, $J = 7.5$ Hz, **ArH**), 3.97 (t, 2H, $J = 7.5$ Hz, =C- $\text{CH}_2\text{CH}_2\text{N}$), 3.37 (t, 2H, $J = 7.5$ Hz, =C- $\text{CH}_2\text{CH}_2\text{N}$), 1.28 (s, 12H, -B[OC(CH₃)₂]₂); **^{13}C NMR** (126 MHz, CDCl_3 , 20 $^\circ\text{C}$) δ 168.4, 138.2, 133.8, 132.5, 128.4, 125.1, 123.9, 123.2, 119.8, 119.6, 111.5, 84.1, 39.4, 24.9, 24.7; **FTIR** (neat, cm^{-1}) 3453 (m), 3381 (br. s), 3058 (w), 2978 (s), 2935 (m), 2252 (w), 1770 (s), 1705 (s), 1619 (w), 1576 (m), 1551 (s), 1464 (m), 1436 (m), 1392 (s), 1263 (m), 1137 (s), 1105 (m), 1080 (m), 1018 (w), 961 (w), 907 (w), 857 (w), 734 (s), 713 (s); **HRMS** (DART) calc'd for $\text{C}_{24}\text{H}_{26}\text{BN}_2\text{O}_4$ [M+H]⁺: 417.1986, found: 417.1978; **TLC** $R_f = 0.48$ (30% EtOAc/hexanes, UV, CAM).

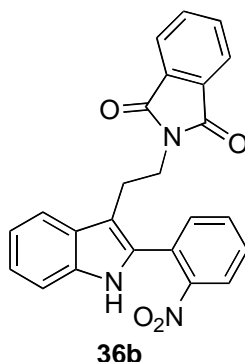


***N,N*-Phthaloyl-2-phenyltryptamine (14)**, Scheme 1 - Route b). A flame dried 15-mL pressure flask was charged with the boronic acid ester **S2** (150 mg, 0.360 mmol, 1.50 equiv.), $\text{Pd}_2(\text{dba})_3$ (5.5 mg, 6.0 μmol , 0.025 equiv.), XPhos (11.7 mg, 0.024 mmol, 0.10 equiv.), anhydrous potassium phosphate tribasic (104 mg, 0.480 mmol, 2.00 equiv.), and a stirring bar and sealed with a septum under an atmosphere of argon. Iodobenzene (27.4 μL , 0.240 mmol, 1 equiv.), toluene (2.4 mL), and deionized water (240 μL) were added via syringe and the mixture was agitated to give a clear red solution. The septum was replaced with the pressure flask's Teflon seal and the entire mixture was heated in an oil bath set to 110 $^\circ\text{C}$. After 5 min, the reaction mixture turned into a pale yellow clear solution and the mixture was maintained at 110 $^\circ\text{C}$. After 2 h, the oil bath was removed and the mixture was allowed to cool to 23 $^\circ\text{C}$ and TLC analysis (10% EtOAc/hexanes) indicated the completion of the reaction. The volatiles were removed under reduced pressure and the crude mixture was purified by flash column chromatography on silica gel (3.0 \times 14.0 cm, eluent 15% EtOAc/hexanes) to afford the product **14** as yellow crystals (72.4 mg, 82%). (For characterization data, please see the procedure for synthesis of **14** from **S1**.)

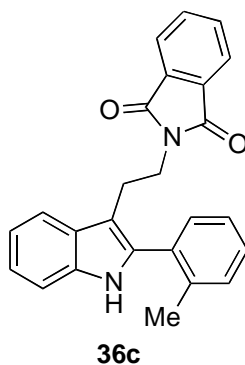


2-(2-(2-(2-(Trifluoromethyl)phenyl)-1*H*-indol-3-yl)ethyl)isoindoline-1,3-dione (36a). Purified by flash column chromatography on silica gel (20% EtOAc/Hex). Yellow foam, 188.9 mg, 90.70% yield. ^1H NMR (500 MHz, CDCl_3 , 20 $^\circ\text{C}$, J in Hz) δ 8.11 (br. s, 1H, NH), 7.81 (d, 1H, $J = 8.0$ Hz, ArH), 7.78-7.72 (complex m, 3H, ArH/phthalimide), 7.64 (m, 2H, phthalimide), 7.62 (d, 1H, $J = 7.5$ Hz, ArH), 7.57 (d, 1H, $J = 7.5$ Hz, ArH), 7.54 (t, 1H, $J = 8.0$ Hz, ArH), 7.35 (d, 1H, $J = 8.0$ Hz, ArH), 7.20 (t, 1H, $J = 8.0$ Hz, ArH), 7.14 (t, 1H, $J = 7.5$ Hz, ArH), 3.87 (t, 2H, $J = 8.0$ Hz, =C- $\text{CH}_2\text{CH}_2\text{N}$), 3.00 (t, 2H, $J = 8.0$ Hz, =C- $\text{CH}_2\text{CH}_2\text{N}$); ^{13}C NMR (126 MHz, CDCl_3 , 20 $^\circ\text{C}$) 168.4, 135.9, 134.0, 133.7, 132.4, 132.3, 132.0, 131.1, 129.9 (q, $J = 30.0$ Hz), 129.1, 128.0, 126.6 (q, $J = 5.2$ Hz), 124.1 (q, $J = 274.6$ Hz), 123.3, 122.8, 120.1, 119.4, 111.4, 111.1, 38.4, 23.9; FTIR (neat, cm^{-1}) 3460 (m), 3367 (s), 3065 (m), 3022 (m), 2942 (m), 2856 (w), 1766 (m), 1709 (s), 1608 (m), 1579 (w), 1489 (m), 1450 (m), 1436 (m), 1400 (s), 1360

(m), 1310 (s), 1267 (m), 1231 (m), 1216 (m), 1173 (s), 1126 (s), 1108 (m), 1076 (m), 1036 (m), 997 (m), 961 (w), 936 (w), 867 (w), 767 (s), 749 (s), 716 (s); **HRMS** (DART) calc'd for $C_{25}H_{18}F_3N_2O_2$ $[M+H]^+$: 435.1315, found:435.1328; **TLC** R_f = 0.45 (30% EtOAc/Hex).

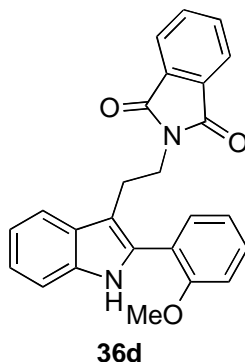


2-(2-(2-(2-Nitrophenyl)-1H-indol-3-yl)ethyl)isoindoline-1,3-dione (36b). Purified by flash column chromatography on silica gel (20% acetone/Hex). Orange foam, 171.8 mg, 87.00% yield. 1H NMR (500 MHz, $CDCl_3$, 20 °C, J in Hz) δ 8.13 (br. s, 1H, NH), 7.93 (dd, 1H, J = 1.0, 8.0 Hz, ArH), 7.78 (d, 1H, J = 8.0 Hz, ArH), 7.74 (dd, 2H, J = 3.0, 5.5 Hz, phthalimide), 7.67 (dt, 1H, J = 1.5, 7.5 Hz, ArH), 7.65 (dd, 2H, J = 2.5, 5.5 Hz, phthalimide), 7.61 (dd, 1H, J = 1.5, 7.5 Hz, ArH), 7.55 (dt, 1H, J = 1.5, 8.0 Hz, ArH), 7.33 (d, 1H, J = 8.0 Hz, ArH), 7.20 (dd, 1H, J = 1.5, 8.0 Hz, ArH), 7.13 (dt, 1H, J = 1.0, 8.0 Hz, ArH), 3.86 (t, 2H, J = 8.0 Hz, =C-CH₂CH₂N), 3.00 (t, 2H, J = 8.0 Hz, =C-CH₂CH₂N); ^{13}C NMR (126 MHz, $CDCl_3$, 20 °C) δ 168.3, 149.6, 136.3, 134.0, 133.6, 132.9, 132.2, 130.3, 129.7, 128.1, 127.0, 124.6, 123.3, 123.1, 120.2, 119.4, 111.7, 111.4, 38.3, 23.8; **FTIR** (neat, cm^{-1}) 3374 (s), 3058 (m), 3022 (m), 2942 (m), 2863 (m), 1766 (m), 1705 (s), 1612 (m), 1522 (s), 1453 (m), 1439 (m), 1400 (s), 1356 (s), 1346 (s), 1184 (m), 1126 (m), 1101 (m), 993 (m), 849 (m), 749 (s), 716 (s); **HRMS** (DART) calc'd for $C_{24}H_{18}N_3O_4$ $[M+H]^+$: 412.1292, found: 412.1250; **TLC** R_f = 0.21 (30% EtOAc/Hex).

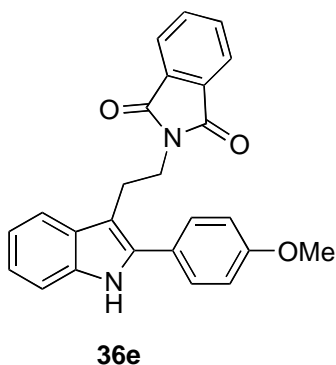


2-(2-(2-(o-Tolyl)-1H-indol-3-yl)ethyl)isoindoline-1,3-dione (36c). Purified by flash column chromatography on silica gel (20% EtOAc/Hex). Yellow foam, 180 mg, 99.0% yield. 1H NMR (500 MHz, $CDCl_3$, 20 °C, J in Hz) δ 7.95 (br. s, 1H, NH), 7.76 (d, 1H, J = 7.5 Hz, ArH), 7.71 (dd, 2H, J = 3.0, 5.0 Hz, phthalimide), 7.63 (dd, 2H, J = 3.0, 5.5 Hz), 7.34-7.29 (complex m, 2H, ArH), 7.26 (dt, 1H, J = 1.5, 7.5 Hz, ArH), 7.22 (dd, 1H, J = 0.5, 7.5 Hz, ArH), 7.19-7.14 (complex m, 2H, ArH), 7.10 (dt, 1H, J = 1.0, 7.5 Hz, ArH), 3.86 (t, 2H, J = 7.5 Hz, =C-CH₂CH₂N), 3.01 (t, 2H, J = 7.5 Hz, =C-CH₂CH₂N), 2.22 (s, 3H, -CH₃); ^{13}C NMR (126 MHz, $CDCl_3$, 20 °C) δ 168.4, 137.6, 135.8, 135.5, 133.9, 132.4, 132.2,

131.0, 130.6, 128.8, 128.5, 125.9, 123.2, 122.2, 119.9, 119.1, 111.0, 110.1, 38.5, 23.9, 20.2; **FTIR** (neat, cm^{-1}) 3453 (m), 3374 (s), 3050 (m), 3022 (m), 2928 (m), 2849 (m), 2245 (m), 1770 (s), 1705 (s), 1615 (m), 1486 (m), 1453 (s), 1436 (s), 1396 (s), 1356 (s), 1338 (s), 1302 (s), 1234 (w), 1187 (w), 1169 (m), 1123 (m), 1101 (s), 1087 (m), 1040 (m), 1008 (s), 936 (m), 871 (m), 763 (s), 734 (s), 720 (s); **HRMS** (DART) calc'd for $\text{C}_{25}\text{H}_{21}\text{N}_2\text{O}_2$ $[\text{M}+\text{H}]^+$: 381.1598, found: 381.1608; **TLC** R_f = 0.24 (20% acetone/Hex).

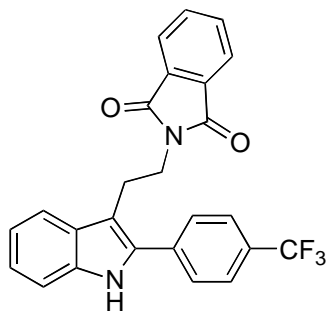


2-(2-(2-(2-methoxyphenyl)-1H-indol-3-yl)ethyl)isoindoline-1,3-dione (36d). Purified by flash column chromatography on silica gel (20% EtOAc/Hex). Yellow foam, 135.9 mg, 23.00% yield (unoptimized). **^1H NMR** (500 MHz, CDCl_3 , 20 °C, J in Hz) δ 7.83 (dd, 2H, J = 3.0, 5.0 Hz, phthalimide), 7.77 (m, 1H, ArH), 7.69 (dd, 2H, J = 3.0, 5.5 Hz, phthalimide), 7.34 (dt, 1H, J = 2.0, 7.5 Hz, ArH), 7.33 (d, 1H, J = 8.0 Hz, ArH), 7.18-7.12 (complex m, 4H, phthalimide/ ArH), 7.09-7.02 (complex m, 2H, ArH), 4.04 (t, 2H, J = 8.0 Hz, =C- $\text{CH}_2\text{CH}_2\text{N}$), 3.73 (s, 3H, -OMe), 3.18 (t, 2H, J = 8.0 Hz, =C- $\text{CH}_2\text{CH}_2\text{N}$); **^{13}C NMR** (126 MHz, CDCl_3 , 20 °C) δ 168.6, 154.5, 137.2, 134.1, 132.5, 128.5, 128.3, 128.2, 128.2, 127.4, 123.4, 122.3, 121.1, 119.9, 119.2, 112.6, 112.5, 111.2, 55.9, 38.7, 24.8; **FTIR** (neat, cm^{-1}) 3050 (w), 2928 (w), 2849 (w), 2835 (w), 1770 (m), 1709 (s), 1608 (w), 1594 (m), 1507 (s), 1461 (s), 1436 (m), 1396 (s), 1371 (m), 1356 (m), 1249 (m), 1227 (m), 1094 (m), 1018 (m), 993 (m), 867 (w), 745 (s), 716 (s); **HRMS** (DART) calc'd for $\text{C}_{25}\text{H}_{21}\text{N}_2\text{O}_3$ $[\text{M}+\text{H}]^+$: 397.1547, found: 397.1535; **TLC** R_f = 0.38 (30% EtOAc/Hex).



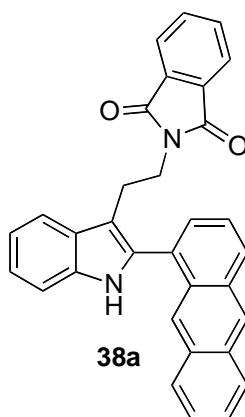
2-(2-(2-(4-methoxyphenyl)-1H-indol-3-yl)ethyl)isoindoline-1,3-dione (36e). Purified by flash column chromatography on silica gel (20% EtOAc/Hex). Yellow foam, 253 mg, 99.0% yield. **^1H NMR** (500 MHz, CDCl_3 , 20 °C, J in Hz) δ 7.98 (br. s, 1H, NH), 7.76-7.72 (complex m, 3H, phthalimide/ArH), 7.66 (dd, 2H, J = 3.0, 5.5 Hz, phthalimide), 7.49 (d, 2H, J = 8.0 Hz, ArH), 7.33 (d, 1H, J = 7.5 Hz, ArH), 7.17 (t, 1H, J = 7.0 Hz, ArH), 7.13 (t, 1H, J = 7.0 Hz, ArH), 6.93 (d, 2H, J = 8.0 Hz, ArH), 3.96 (t, 2H, J = 7.5 Hz, =C- $\text{CH}_2\text{CH}_2\text{N}$), 3.81 (s, 3H, -OMe), 3.21 (t, 2H, J = 8.0 Hz, =C- $\text{CH}_2\text{CH}_2\text{N}$); **^{13}C NMR** (126 MHz,

CDCl₃, 20 °C) δ 168.5, 159.5, 135.9, 135.4, 133.9, 132.4, 129.4, 129.3, 125.4, 123.3, 122.4, 120.1, 119.0, 114.6, 110.9, 108.6, 55.5, 38.6, 24.1; **FTIR** (neat, cm⁻¹) 3367 (s), 3058 (w), 2921 (m), 2849 (m), 2835 (m), 1770 (m), 1705 (s), 1608 (m), 1507 (m), 1461 (s), 1443 (s), 1396 (s), 1356 (m), 1281 (m), 1249 (s), 1177 (m), 1123 (m), 1101 (m), 1029 (m), 997 (m), 907 (m), 835 (m), 734 (s), 716 (s); **HRMS** (DART) calc'd for C₂₅H₂₁N₂O₃ [M+H]⁺: 397.1547, found: 397.1532; **TLC** R_f = 0.30 (30% EtOAc/Hex).



36f

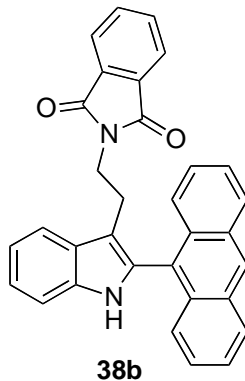
2-(2-(2-(4-(Trifluoromethyl)phenyl)-1H-indol-3-yl)ethyl)isoindoline-1,3-dione (36f). Purified by flash column chromatography on silica gel (20% EtOAc/Hex). Yellow foam, 268 mg, 96.4% yield. **¹H NMR** (500 MHz, CDCl₃, 20 °C, *J* in Hz) 8.09 (br. s, 1H, NH), 7.78 (d, 1H, *J* = 8.0 Hz, ArH), 7.72 (dd, 2H, *J* = 3.0, 5.5 Hz, phthalimide), 7.68-7.63 (complex m, 4H), 7.60 (d, 2H, *J* = 8.5 Hz, ArH), 7.37 (d, 1H, *J* = 8.0 Hz, ArH), 7.23 (dt, 1H, *J* = 1.0, 8.0 Hz, ArH), 7.17 (dt, 1H, *J* = 1.0, 8.0 Hz, ArH), 3.95 (t, 2H, *J* = 7.5 Hz, =C-CH₂CH₂N), 3.27 (t, 2H, *J* = 8.0 Hz, =C-CH₂CH₂N); **¹³C NMR** (126 MHz, CDCl₃, 20 °C) 168.4, 136.3, 136.3, 134.1, 133.7, 132.2, 129.6 (q, *J* = 32.9 Hz), 129.2, 128.0, 126.1 (q, *J* = 4.0 Hz), 124.2 (q, *J* = 27.2 Hz), 123.5, 123.3, 120.5, 119.4, 111.3, 111.0, 38.5, 24.0; **FTIR** (neat, cm⁻¹) 3460 (w), 3374 (m), 3065 (w), 3022 (w), 2935 (w), 2863 (w), 1770 (m), 1705 (s), 1615 (m), 1457 (m), 1439 (m), 1396 (s), 1356 (m), 1320 (s), 1256 (w), 1166 (s), 1119 (s), 1069 (s), 1015 (m), 997 (w), 936 (w), 846 (m), 749 (s), 716 (s); **HRMS** (DART) calc'd for C₂₅H₁₈F₃N₂O₂ [M+H]⁺: 435.1315, found: 435.1320; **TLC** R_f = 0.38 (20% EtOAc/Hex).



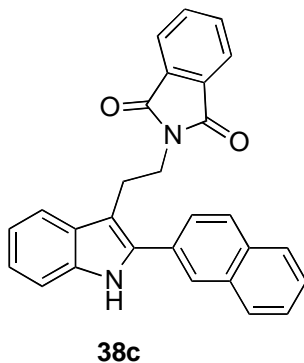
38a

2-(2-(2-(Anthracen-1-yl)-1H-indol-3-yl)ethyl)isoindoline-1,3-dione (38a). Purified by flash column chromatography on silica gel (20% EtOAc/Hex). Yellow foam, 206.2 mg, 92.14% yield. **¹H NMR** (500 MHz, CDCl₃, 20 °C, *J* in Hz) 8.32 (s, 1H, ArH), 8.24 (s, 1H, ArH), 8.21 (br. s, 1H, NH), 7.96 (d, 1H, *J* = 8.5 Hz, ArH), 7.92 (d, 1H, *J* = 8.5 Hz, ArH), 7.82 (d, 1H, *J* = 8.0 Hz, ArH), 7.77 (d, 1H, *J* = 8.5 Hz,

ArH), 7.52 (d, 1H, $J = 6.5$ Hz, ArH), 7.47-7.38 (m, 3H, ArH), 7.38-7.30 (m, 5H, phthalimide/ArH), 7.25 (d, 1H, $J = 7.5$ Hz, ArH), 7.18 (t, 1H, $J = 8.0$ Hz, ArH), 3.85 (br. s, 2H, =C-CH₂CH₂N), 3.19 (br. s, 2H, =C-CH₂CH₂N); ¹³C NMR (126 MHz, CDCl₃, 20 °C) δ 168.2, 136.1, 134.6, 133.3, 132.0, 131.8, 131.8, 131.7, 130.7, 130.1, 129.3, 128.8, 128.6, 128.5, 128.0, 127.0, 125.9, 125.7, 124.9, 124.8, 122.6, 122.5, 120.0, 119.1, 111.5, 111.2, 38.7, 23.5; FTIR (neat, cm⁻¹) 3356 (s), 3048 (m), 3014 (w), 2924 (m), 2852 (w), 1768 (m), 1709 (s), 1614 (m), 1460 (m), 1438 (m), 1393 (s), 1362 (m), 1337 (m), 1309 (m), 1102 (m), 1018 (m), 878 (m), 747 (s), 738 (s), 713 (s), 666 (m); HRMS (DART) calc'd for C₃₂H₂₃N₂O₂ [M+H]⁺: 467.1754, found: 467.1738; TLC R_f = 0.47 (30% EtOAc/Hex).

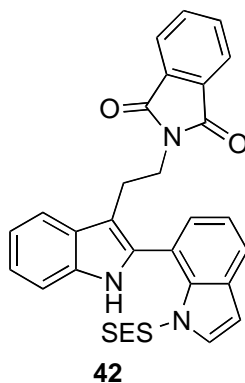


2-(2-(2-(Anthracen-9-yl)-1H-indol-3-yl)ethyl)isoindoline-1,3-dione (38b). Purified by flash column chromatography on silica gel (20% EtOAc/Hex). Yellow foam, 211 mg, 94.2% yield. ¹H NMR (500 MHz, CDCl₃, 20 °C, J in Hz) δ 8.51 (s, 1H, ArH), 8.15 (br. s, 1H, NH), 8.00 (d, 2H, $J = 8.5$ Hz, ArH), 7.83 (d, 1H, $J = 7.5$ Hz, ArH), 7.69 (d, 2H, $J = 8.5$ Hz, ArH), 7.54 (app. br. s, 4H, phthalimide), 7.41-7.37 (complex m, 3H, ArH), 7.28-7.22 (complex m, 3H, ArH), 7.16 (dt, 1H, $J = 1.0, 8.0$ Hz, ArH), 3.75 (t, 2H, $J = 7.0$ Hz, =C-CH₂CH₂N), 2.92 (t, 2H, $J = 7.5$ Hz, =C-CH₂CH₂N); ¹³C NMR (126 MHz, CDCl₃, 20 °C) δ 168.2, 136.5, 133.7, 132.2, 131.9, 131.8, 131.4, 128.7, 128.6, 128.5, 126.5, 126.5, 126.4, 125.5, 123.0, 122.4, 120.0, 119.2, 113.4, 111.1, 38.4, 24.1; FTIR (neat, cm⁻¹) 3356 (s), 3053 (m), 3025 (m), 2936 (w), 2852 (w), 1768 (m), 1706 (s), 1614 (w), 1488 (w), 1460 (m), 1438 (m), 1399 (s), 1359 (m), 1334 (m), 1217 (m), 1119 (m), 1024 (m), 892 (m), 738 (s), 710 (s); HRMS (DART) calc'd for C₃₂H₂₃N₂O₂ [M+H]⁺: 467.1754, found: 467.1763; TLC R_f = 0.42 (30% EtOAc/Hex).

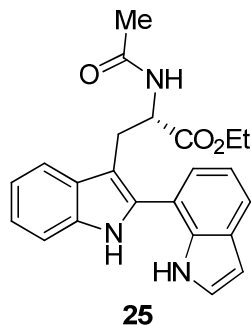


2-(2-(2-(Naphthalen-2-yl)-1H-indol-3-yl)ethyl)isoindoline-1,3-dione (38c). Purified by flash column chromatography on silica gel (20% acetone/Hex). Yellow foam, 266 mg, 99.0% yield. ¹H NMR (500 MHz, CDCl₃, 20 °C, J in Hz) δ : 8.18 (br. s, 1H, NH), 8.01 (s, 1H, ArH), 7.90 (d, 1H, $J = 8.0$ Hz, ArH),

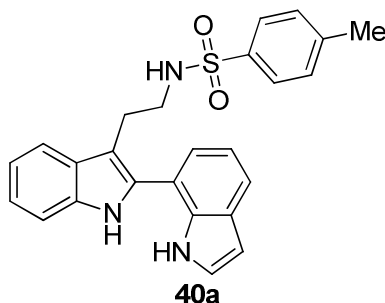
7.84 (d, 1H, $J = 8.0$ Hz, ArH), 7.80 (dd, 1H, $J = 0.5, 8.0$ Hz, ArH), 7.77 (d, 1H, $J = 7.5$ Hz, ArH), 7.68 (dd, 1H, $J = 2.0, 8.5$ Hz, ArH), 7.64 (dd, 2H, $J = 3.0, 5.5$ Hz, phthalimide), 7.52 (dd, 2H, $J = 3.0, 5.5$ Hz, phthalimide), 7.49 (dt, 1H, $J = 1.0, 7.0$ Hz, ArH), 7.45 (dt, 1H, $J = 1.5, 7.5$ Hz, ArH), 7.39 (dd, 1H, $J = 0.5, 8.0$ Hz, ArH), 7.21 (dd, 1H, $J = 1.5, 8.0$ Hz, ArH), 7.17 (dt, 1H, $J = 1.0, 7.0$ Hz, ArH), 4.02 (t, 2H, $J = 7.5$ Hz, =C-CH₂CH₂N), 3.34 (t, 2H, $J = 7.5$ Hz, =C-CH₂CH₂N); ¹³C NMR (126 MHz, CDCl₃, 20 °C) 168.4, 136.2, 135.3, 133.8, 133.7, 132.7, 132.2, 130.2, 129.5, 128.9, 128.5, 127.9, 126.9, 126.7, 126.4, 125.6, 123.1, 122.9, 120.2, 119.2, 111.1, 109.9, 38.6, 24.2; FTIR (neat, cm⁻¹) 3453 (m), 3374 (s), 3058 (m), 3022 (m), 2935 (m), 2863 (w), 1766 (s), 1709 (s), 1612 (m), 1601 (m), 1551 (m), 1453 (m), 1436 (s), 1396 (s), 1360 (s), 1335 (m), 1306 (m), 1259 (m), 1231 (m), 1213 (m), 1144 (m), 1011 (m), 1000 (m), 860 (m), 817 (m), 749 (s), 716 (s); HRMS (DART) calc'd for C₂₈H₂₁N₂O₂ [M+H]⁺: 417.1598, found: 417.1593; TLC R_f = 0.47 (30% EtOAc/Hex).



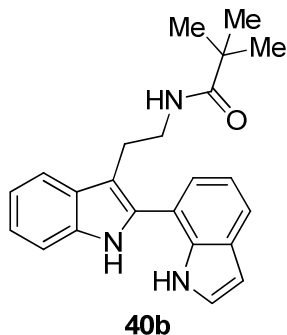
2-(2-(1'-((2-(trimethylsilyl)ethyl)sulfonyl)-1H,1'-[2,7'-biindol]-3-yl)ethyl)isoindoline-1,3-dione (42). Purified by flash column chromatography on silica gel (1:1:8 CH₂Cl₂/EtOAc/Hex). Yellow solid, 74.8 mg, 96.0% yield. ¹H NMR (500 MHz, CDCl₃, 20 °C, J in Hz) 8.74 (br. s, 1H, NH), 7.71 (dd, 2H, $J = 3.0, 5.5$ Hz, phthalimide), 7.70-7.65 (complex m, 2H, ArH), 7.63 (dd, 2H, $J = 3.0, 5.5$ Hz, phthalimide), 7.51 (d, 1H, $J = 3.5$ Hz, ArH), 7.39 (d, 1H, $J = 8.0$ Hz, ArH), 7.36 (dd, 1H, $J = 1.5, 7.5$ Hz, ArH), 7.31 (t, 1H, $J = 7.5$ Hz, ArH), 7.19 (dt, 1H, $J = 1.0, 8.0$ Hz, ArH), 7.07 (t, 1H, $J = 7.5$ Hz, ArH), 7.66 (d, 1H, $J = 3.5$ Hz, ArH), 3.81 (t, 2H, $J = 7.5$ Hz, =C-CH₂CH₂N), 2.83 (app. br. s, 2H, =C-CH₂CH₂N), 2.61 (m, 2H, -SO₂CH₂CH₂SiMe₃), 0.66 (m, 2H, -SO₂CH₂CH₂SiMe₃), -0.28 (s, 9H, -SiMe₃); ¹³C NMR (126 MHz, CDCl₃, 20 °C) δ 168.3, 135.7, 134.6, 134.0, 133.7, 133.3, 132.3, 130.1, 130.0, 127.4, 123.5, 123.3, 123.0, 122.9, 120.2, 119.3, 118.9, 112.1, 111.5, 108.0, 51.4, 38.5, 24.3, 10.2, -2.0; FTIR (neat, cm⁻¹) 3381 (m), 3108 (w), 3050 (w), 3022 (w), 2950 (m), 2892 (w), 1766 (m), 1705 (s), 1612 (w), 1457 (m), 1436 (m), 1400 (s), 1360 (s), 1306 (m), 1249 (s), 1231 (m), 1169 (s), 1155 (s), 1126 (s), 1101 (m), 1069 (m), 1018 (m), 979 (m), 889 (w), 857 (m), 839 (m), 799 (m), 745 (s), 716 (s), 698 (m), 662 (m); HRMS (DART) calc'd for C₃₁H₃₀N₃O₄SSi [M-H]⁻: 568.1732, found: 568.1732; TLC R_f = 0.16 (1:1:8 CH₂Cl₂/EtOAc/Hex).



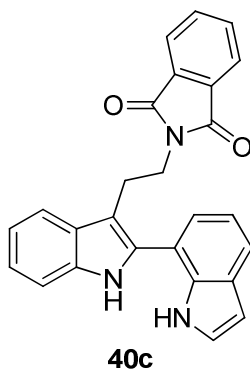
(2'S)-3-(2-N-Acetylamino-2-ethoxycarbonylethyl)-2-(7-indolyl)-1H-indole (25). Purified by column chromatography on silica gel (50% EtOAc/Hex). Yellow foam, 514.91 mg, 73% yield. **¹H NMR** (500 MHz, CDCl₃, *J* in Hz) δ 9.02 (s, 1H), 8.19 (s, 1H), 7.73 (dd, *J* = 7.5, 0.8, 1H), 7.63 (d, *J* = 7.9, 1H), 7.37 (d, *J* = 8.0, 1H), 7.27 – 7.17 (m, 5H), 6.64 (dd, *J* = 3.1, 2.0, 1H), 5.77 (d, *J* = 8.3, 1H), 4.78 (ddd, *J* = 8.3, 6.8, 5.2, 1H), 3.83 (dq, *J* = 10.7, 7.1, 1H), 3.46 (dq, *J* = 10.7, 7.1, 1H), 3.40 (dd, *J* = 14.6, 5.1, 1H), 3.30 (dd, *J* = 14.6, 6.8, 1H), 1.48 (s, 3H), 1.01 (t, *J* = 7.1, 3H); **¹³C NMR** (126 MHz, CDCl₃) δ 172.0, 170.0, 136.2, 135.0, 133.6, 129.5, 128.8, 125.3, 123.1, 122.7, 121.6, 120.3, 120.2, 119.1, 116.4, 111.3, 108.7, 103.2, 61.5, 53.1, 27.5, 22.8, 14.0; **FTIR** (cm⁻¹) 3395, 2929, 1727, 1653, 1514, 1435, 1335, 1211, 1025; **UPLC-MS** (ESI) *m/z* 376.17 (calculated for C₂₂H₂₁N₃O₃[M + H⁺] = 376.16); **TLC** R_f = 0.31 (50% EtOAc/Hex); [α]_{546 nm}^{20 °C} = +177.7 (*c* = 0.115, CHCl₃).



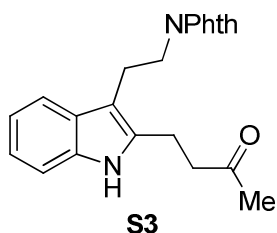
3-(2-N-(4-Toluenesulfonyl)aminoethyl)-2-(7-indolyl)-1H-indole (40a). Purified by column chromatography on silica gel (30% EtOAc/Hex). Yellow foam, 1.30 g, 43% yield. **¹H NMR** (500 MHz, CDCl₃, *J* in Hz) δ 9.19 (brs, 1H), 8.56 (brs, 1H), 7.80 – 7.74 (m, 1H), 7.72 (t, *J* = 6.4, 1H), 7.63 (t, *J* = 7.0, 1H), 7.48 (t, *J* = 7.3, 2H), 7.30 – 7.23 (m, 4H), 7.19 – 7.11 (m, 3H), 6.71 – 6.65 (m, 1H), 3.28 – 3.19 (m, 2H), 2.97 (q, *J* = 6.6, 2H), 2.39 (s, 3H); **¹³C NMR** (126 MHz, CDCl₃) δ 143.4, 141.7, 136.7, 134.5, 133.3, 130.1, 129.8, 128.7, 127.4, 127.2, 125.2, 123.1, 122.3, 121.5, 120.3, 119.9, 116.4, 109.1, 103.3, 84.2, 43.2, 25.2, 21.7; **FTIR** (cm⁻¹) 3389, 3057, 2926, 2852, 1597, 1428, 1153, 1092; **UPLC-MS** (ESI)*m/z*430.16 (calculated for C₂₅H₂₃N₃O₂S [M + H⁺] = 430.15); **TLC** R_f = 0.59 (6% EtOAc/DCM).



3-(2-*N*-Pivaloylaminoethyl)-2-(7-indolyl)-1*H*-indole(40b). Purified by column chromatography on silica gel (25% EtOAc/Hex). Yellow foam, 1.46 g, 55% yield. ¹H NMR (500 MHz, CDCl₃, *J* in Hz) δ 9.17 (s, 1H), 8.13 (s, 1H), 7.72 (dd, *J* = 7.6, 4.3, 2H), 7.40 (d, *J* = 7.9, 1H), 7.30 – 7.28 (m, 1H), 7.26 – 7.18 (m, 3H), 6.64 (dd, *J* = 3.1, 2.0, 1H), 5.71 – 5.63 (m, 1H), 3.52 (dd, *J* = 12.9, 6.7, 2H), 2.99 (t, *J* = 6.8, 2H), 0.91 (s, 9H); ¹³C NMR (126 MHz, CDCl₃) δ 178.8, 136.4, 134.8, 132.9, 129.3, 128.7, 125.3, 123.1, 122.6, 121.4, 120.1, 120.0, 119.4, 116.5, 111.4, 111.3, 103.0, 40.1, 38.6, 27.4, 25.0; FTIR (cm⁻¹) 3401, 2964, 1638, 1514, 1431, 1334, 1198; UPLC-MS (ESI) *m/z* 360.22 (calculated for C₂₃H₂₅N₃O [M + H⁺] = 360.20); TLC R_f = 0.47 (35% EtOAc/Hex).



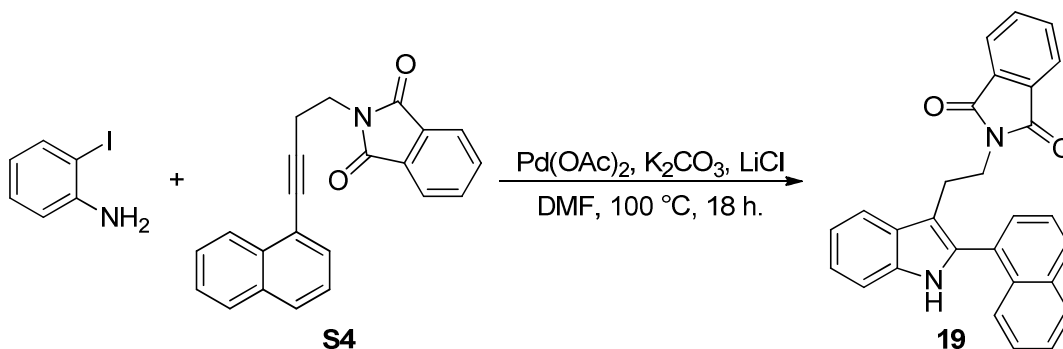
3-(2-*N,N*-Phthaloylaminoethyl)-2-(7-indolyl)-1*H*-indole (40c). Purified by column chromatography on silica gel (25% EtOAc/Hex). Yellow solid, 138 mg, 69% yield. ¹H NMR (500 MHz, CDCl₃, *J* in Hz) δ 9.07 (s, 1H), 8.07 (s, 1H), 7.67 – 7.59 (m, 5H), 7.55 (d, *J* = 8.0 Hz, 1H), 7.33 (d, *J* = 8.1 Hz, 1H), 7.27 – 7.23 (m, 2H), 7.18 – 7.14 (m, 1H), 7.14 – 7.10 (m, 1H), 6.97 – 6.93 (m, 1H), 6.58 (dd, *J* = 3.1, 2.1 Hz, 1H), 3.88 (t, *J* = 6.7 Hz, 2H), 3.19 (t, *J* = 6.7 Hz, 2H); ¹³C NMR (126 MHz, CDCl₃) δ 168.73, 136.17, 135.10, 133.82, 133.10, 132.03, 129.18, 128.52, 125.08, 123.25, 123.06, 122.43, 121.34, 120.09, 120.03, 118.72, 116.31, 111.11, 111.08, 103.16, 39.04, 23.62; FTIR (cm⁻¹) 3339, 3289, 3066, 2947, 2911, 1761, 1694, 1614, 1337, 1277; UPLC-MS (ESI) *m/z* 406.15 (calculated for C₂₆H₁₉N₃O₂[M + H⁺] = 406.15); TLC R_f = 0.31 (25% EtOAc/Hex).



3-(2-*N,N*-Phthaloylaminoethyl)-2-(3-oxobutyl)-1*H*-indole (S3). Prepared by the reported procedure.⁵ Purified by preparative TLC (50% EtOAc/Hex). Yellow solid, 87.8 mg, 25% yield. ¹H NMR (500 MHz, CDCl₃, *J* in Hz) δ 8.51 (s, 1H), 7.85 (dd, *J* = 5.4, 3.0, 2H), 7.72 (dd, *J* = 5.5, 3.0, 2H), 7.66 (d, *J* = 7.7, 1H), 7.27 (d, *J* = 7.9, 2H), 7.13 – 7.09 (m, 1H), 7.09 – 7.04 (m, 1H), 3.91 – 3.85 (m, 2H), 3.09 – 3.01 (m, 4H), 2.93 – 2.87 (m, 2H), 2.19 (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 209.9, 168.5, 135.9, 135.3, 134.1, 132.4, 128.2, 123.4, 121.6, 119.5, 118.4, 110.8, 107.7, 44.1, 38.9, 30.3, 23.7, 19.4; FTIR (cm⁻¹) 1770, 1703, 1615, 1436, 1395, 1359, 1265, 1162, 1020; UPLC-MS (ESI) *m/z* 361.15 (calculated for C₂₂H₂₀N₂O₃[M + H⁺] = 361.15); TLC R_f = 0.56 (50% EtOAc/Hex).

3-(2-*N,N*-Phthaloylaminoethyl)-2-(1-naphthyl)-1*H*-indole (19)

Compound was prepared by a modification of a reported procedure.⁶



Notes:

- 2-Iodoaniline was recrystallized from boiling hexanes.
- Lithium chloride was kept under high vacuum and flame-dried for 60 seconds prior to use.
- Dry *N,N*-dimethylformamide was degassed by freeze-pump-thaw procedure.

A flame dried 500-mL round bottom flask was charged with 2-iodoaniline (2.18 g, 9.95 mmol, 1.3 equiv.), Pd(OAc)₂ (340 mg, 1.51 mmol, 0.2 equiv.), K₂CO₃ (3.17 g, 22.94 mmol, 3.0 equiv.) and LiCl (200 mg, 4.72 mmol, 0.62 equiv.). The atmosphere of the flask was evacuated and refilled with nitrogen. To this mixture was added *N,N*-DMF (103 mL), followed by the solution of alkyne **S4** (2.49 g, 7.65 mmol, 1.0 equiv.) in *N,N*-DMF (50 mL). The flask was sealed, flushed with nitrogen and heated at 100 °C in an oil bath for 18 hours. After this time, alkyne **S4** could not be detected by TLC. The reaction mixture was cooled to room temperature, diluted with 500 mL of EtOAc, transferred to a separatory funnel and extracted sequentially with 500 mL of distilled water, 500 mL of 1 M HCl, 500 mL of saturated NaHCO₃,

and 500 mL of brine. Organic phase was dried over MgSO_4 , filtered, and the solvent was removed *in vacuo*. The black residue was purified by column chromatography (20% EtOAc/Hex), and thus obtained material was further purified by trituration in boiling DCM and dried under high vacuum. Compound **19** was obtained as a pale yellow solid, 1.27 g (39% yield).

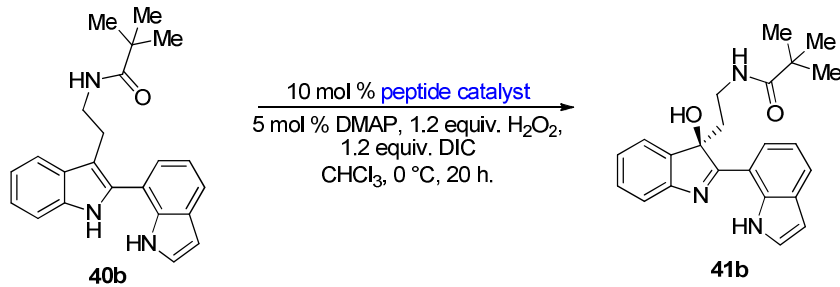
^1H NMR (500 MHz, CDCl_3 , J in Hz) δ 8.15 (s, 1H), 7.88 – 7.84 (m, 2H), 7.83 (d, $J = 7.9$, 1H), 7.78 (d, $J = 8.4$, 1H), 7.64 – 7.57 (m, 5H), 7.50 (dd, $J = 8.2$, 7.1, 1H), 7.45 (ddd, $J = 8.1$, 6.8, 1.1, 1H), 7.39 (d, $J = 8.0$, 1H), 7.36 (ddd, $J = 8.2$, 6.8, 1.2, 1H), 7.24 (ddd, $J = 8.3$, 7.2, 1.1, 1H), 7.17 (ddd, $J = 8.5$, 7.2, 0.9, 1H), 3.88 (t, $J = 7.3$, 2H), 3.18 – 3.07 (m, 2H); **^{13}C NMR** (126 MHz, CDCl_3) δ 168.3, 136.0, 134.3, 133.9, 133.7, 132.5, 132.2, 130.2, 129.1, 128.9, 128.6, 128.5, 126.8, 126.2, 125.9, 125.5, 123.0, 122.5, 120.0, 119.2, 111.4, 111.0, 38.7, 23.8; **FTIR** (cm^{-1}) 3447, 3354, 3054, 2952, 2915, 1765, 1692, 1614, 1431, 1397, 1360, 1017, 971; **UPLC-MS** (ESI) m/z 417.16 (calculated for $\text{C}_{28}\text{H}_{20}\text{N}_2\text{O}_2[\text{M} + \text{H}^+] = 417.15$); **TLC** $R_f = 0.44$ (30% EtOAc/Hex).

C. Structure–Enantioselectivity Relationships for Peptide Catalyzed Indole Oxidation.

Entry	Residue Position							e.r.
	<i>i</i> -1	<i>i</i> = catalytic	<i>i</i> +1	<i>i</i> +2	<i>i</i> +3	<i>i</i> +4	<i>i</i> +5	
1		Asp	Pro	D-Val	Leu	Gly		67:33
2		Asp	Pro	D-Val	Dhl	Gly		67:33
3		Asp	Pro	D-Val	Tba	Gly		64:36
4		Asp	Pro	D-Val	Cha	Gly		61:39
5		Asp	Pro	D-Val	Cpa	Gly		59:41
6		Asp	Pro	D-Val	D-Leu	Gly		53:47
7		Asp	Pro	D-Val	Ile	Gly		62:38
8		Asp	Pro	D-Val	Val	Gly		61:39
9		Asp	Pro	D-Val	Val	Val	Gly	59:41
10		Asp	Pro	D-Val	Chg	Gly		60:40
11		Asp	Pro	D-Val	Phe			59:41
12		Asp	Pro	D-Val	MEA			57:43
13		Asp	Pro	D-Val	<i>tert</i> -Gln	Gly		56:44
14		Asp	Pro	D-Val	Boc ₂ Arg	Gly		55:45
15		Asp	Pro	D-Val	<i>tert</i> -Leu	Gly		52:48
16		Asp	Pro	D-Val	(<i>R</i>)-PEA			60:40
17		Asp	Pro	D-Val	(<i>R</i>)-2-NEA			62:38
18		Asp	Pro	D-Val	(<i>R</i>)-1-NEA			58:42
19		Asp	Pro	D-Val	(<i>S</i>)-1-NEA			52:48
20		Asp	Pip	D-Val	Leu	Gly		67:33
21		Asp	Bn-Hyp	D-Val	Leu	Gly		67:33
22		Asp	Ala	D-Val	Leu	Gly		62:38
23		Asp	Pro	D- <i>t</i> Bu-Thr	Leu	Gly		55:45
24		Asp	D-Pro	Val	Phe			47:53
25		Asp	D-Pro	Aib	Phe			49:51
26		Asp	Val	Pro	D-Val	Val	DPMA	52:48
27		D-Asp	D-Pro	Val	D-Val	Abi		41:59
28		Asp	Pro	D-Val	Cha	Gly	NBA	59:41
29		Asp	Pro	D-Val	Leu	NPA		62:38
30	Val-Val-Val-D-Pro-D-Val	Asp	Pro	D-Val	Val	Gly		60:40
31	Val-Val-Val-Pro-D-Val	Asp	Pro	D-Val	Val	Gly		55:45

Table S1. Peptide primary structure/enantioselectivity studies for a model indole oxidation reaction. Abbreviations: Dhl = *L*-4,5-di-dehydroleucine, Tba = *L*-β-*tert*-butylalanine, Cha = *L*-β-cyclohexylalanine, Cpa = *L*-β-cyclopropylalanine, Chg = *L*-cyclohexylglycine, MEA = methylamine, PEA = 1-phenylethylamine, 2-NEA = 1-(2-naphthyl)ethylamine, 1-NEA = 1-(1-naphthyl)ethylamine, Pip = *L*-pipercolic acid, Hyp = (2*S*,4*R*)-4-hydroxyproline, Aib = 2-aminoisobutyric acid, DPMA = 1,1-diphenylmethylamine, Abi = abietic acid imide, NBA = *n*-butylamine, NPA = *n*-propylamine.

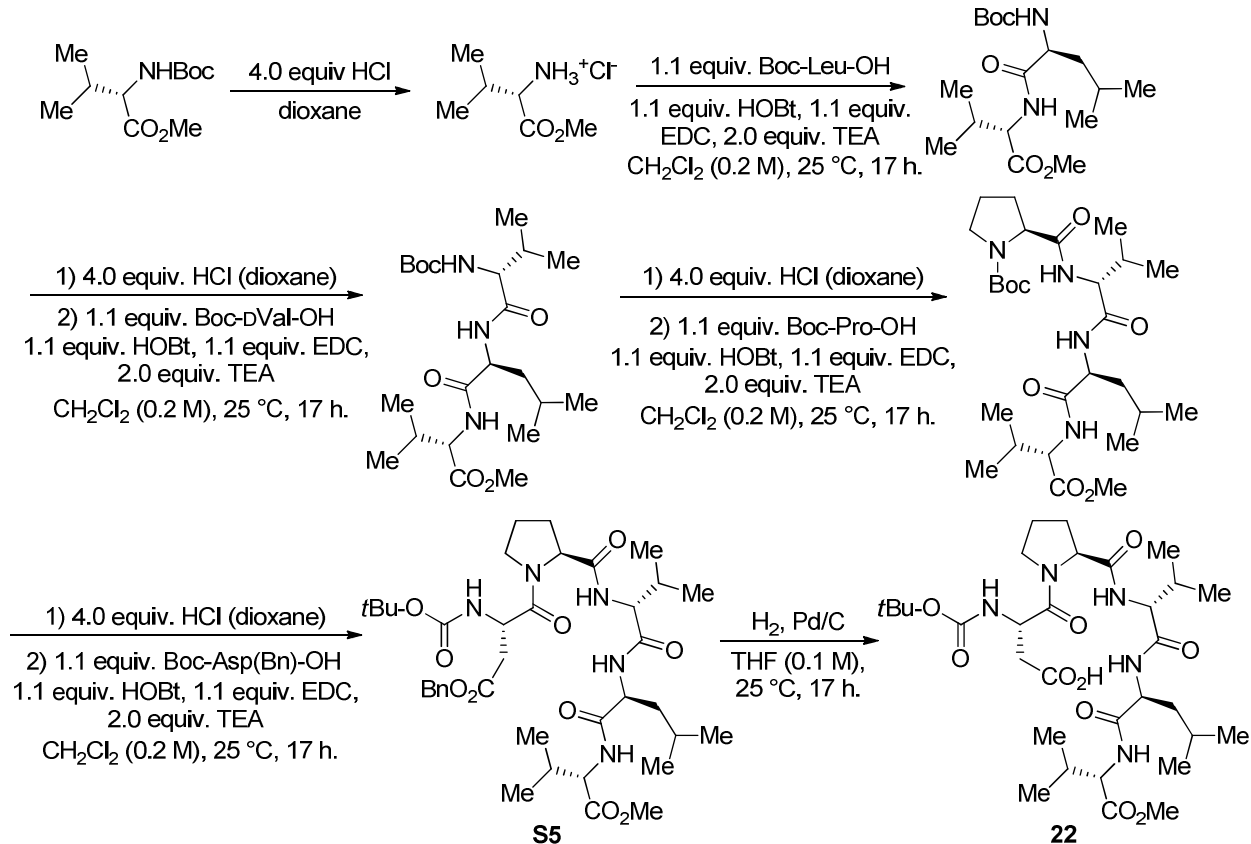
D. Peptide Sequence Optimization for Enantioselective Indole Oxidation.



Entry	Residue Position					
	<i>i</i> = catalytic	<i>i</i> +1	<i>i</i> +2	<i>i</i> +3	<i>i</i> +4	<i>e.r.</i>
1	Asp	Pro	D-Val	Leu	Gly	79:21
2	Glu	Pro	D-Val	Leu	Gly	60:40
3	Asp	<i>t</i> Bu-Hyp	D-Val	Leu	Gly	77:23
4	Asp	Pip	D-Val	Leu	Gly	75:25
5	Asp	Leu	D-Val	Leu	Gly	74:26
6	Asp	Pro	D- <i>tert</i> -Leu	Leu	Gly	79:21
7	Asp	Pro	D-Val	Cpa	Gly	75:25
8	Asp	Pro	D-Val	Tba	Gly	78:22
9	Asp	Pro	D-Val	Leu	Val	84:16
10	Asp	D-Pro	Aib	Phe	Gly	50:50
11	Asp	Pro	D-Val	Leu	Ala	82:18
12	Asp	Pro	D-Val	Leu	Phe	82:18
13	Asp	Pro	D-Val	Leu	Leu	82:18
14	Asp	Pro	D-Val	Leu	Ile	81:19
15	Asp	Pro	D-Val	Leu	Pro	76:24
16	Asp	Pro	D-Val	Leu	D-Val	64:36

Table S2. Optimization of the peptide primary structure for enantioselectivity in the oxidation of the model bis(indole) **40b**. Currently optimized sequence (peptide catalyst **22**) depicted in entry 9. Abbreviations: Hyp = (2*S*,4*R*)-4-hydroxyproline, Pip = L-pipecolic acid, Cpa = L- β -cyclopropylalanine, Tba = L- β -*tert*-butylalanine, Aib = 2-aminoisobutyric acid.

E. Preparation of Peptides 22 and *ent*-22.



Boc-Val-OMe (1.38 g, 5.96 mmol, 1.0 equiv.) was placed in a flask equipped with magnetic stir bar, flushed with nitrogen, and treated with 4 M solution of HCl in dioxane (6.0 mL). The dissolution of the material was accompanied by intense gas evolution, so in order to avoid pressure build-up inside the flask, the septum was pierced with a needle. After 0.5 h starting material could not be detected by TLC, and the reaction mixture was concentrated under reduced pressure in a rotary evaporator (caution: HCl gas released). Solvent and any residual HCl were removed by exposing the product to high vacuum for a minimum of 1 h. To the resulting white powder were added HOBt·H₂O (1.01 g, 6.56 mmol, 1.1 equiv.), EDC·HCl (1.26 g, 6.56 mmol, 1.1 equiv.), and Boc-Leu-OH·H₂O (1.64 g, 6.56 mmol, 1.1 equiv.), the mixture was suspended in dry DCM (30 mL, 0.2 M), and triethylamine (1.66 mL, 11.93 mmol, 2.0 equiv.) was added in one portion. The resulting clear, colorless solution was stirred at room temperature for 17 h. The reaction mixture was then diluted with 350 mL of EtOAc, transferred to a separatory funnel and sequentially washed with 150 mL of 0.5 M aqueous solution of citric acid and 150 mL of saturated aqueous solution of NaHCO₃. The organic phase was dried over MgSO₄, filtered, and concentrated under reduced pressure. This product was exposed to high vacuum for 1 hour, and then subjected to the removal of the Boc group in a manner described above. To the white powder obtained after extensive drying of the crude deprotection product at high vacuum were added HOBt·H₂O (1.01 g, 6.56 mmol, 1.1 equiv.), EDC·HCl (1.26 g, 6.56 mmol, 1.1 equiv.), and Boc-D-Val-OH (1.43 g, 6.56 mmol, 1.1 equiv.), the mixture was suspended in dry DCM (30 mL, 0.2 M), and triethylamine (1.66 mL, 11.93 mmol, 2.0 equiv.) was added in one portion. Subsequent coupling, work-up, and Boc removal were carried out in a manner analogous to the one described above. To the dry deprotection product thus obtained were added

HOBt·H₂O (1.01 g, 6.56 mmol, 1.1 equiv.), EDC·HCl (1.26 g, 6.56 mmol, 1.1 equiv.), and Boc-Pro-OH (1.41 g, 6.56 mmol, 1.1 equiv.), the mixture was suspended in dry DCM (30 mL, 0.2 M), and triethylamine (1.66 mL, 11.93 mmol, 2.0 equiv.) was added in one portion. After carrying out the peptide coupling, work-up, and Boc removal procedures as described above, final peptide coupling step was performed using following amount of materials: HOBt·H₂O (1.01 g, 6.56 mmol, 1.1 equiv.), EDC·HCl (1.26 g, 6.56 mmol, 1.1 equiv.), and Boc-Asp(Bn)-OH (2.12 g, 6.56 mmol, 1.1 equiv.), dry DCM (30 mL, 0.2 M), and triethylamine (1.66 mL, 11.93 mmol, 2.0 equiv.). Crude product obtained after the final peptide coupling was purified by flash column chromatography on normal phase silica gel (30% → 50% acetone/hexanes) to yield 2.30 g (52% yield) of benzyl ester **S5** as a white solid. This compound was dissolved in THF (30 mL, 0.1 M), and the atmosphere of the flask was flushed with dry N₂ for 5 minutes. Then Pd on charcoal (10wt%, 500 mg, 0.470 mmol, 0.15 equiv.) was added, and a balloon filled with H₂ was placed on the reaction flask. The resulting suspension was stirred at room temperature for 17 hours. At this point, no starting material could be detected by thin layer chromatography. The suspension was diluted with 250 mL of EtOAc and filtered through Celite. The filtrate was concentrated under reduced pressure to afford 1.82 g of the free acid **22** as a white foam (47% yield). This material was purified via the following sequence: 1) Flash purification on an automated system using a reverse phase (C18) column, eluting with the gradient 0% → 99% MeOH/H₂O over 25 CV, then 100% MeOH for 2 CV; 30 mL/min, detection at 210 nm, 262 nm. 2) Flash chromatography on normal phase silica gel eluting with the gradient 95:5:1 → 85:15:1 DCM:MeOH:AcOH, followed by several cycles of azeotropic removal of acetic acid with toluene under reduced pressure on a rotary evaporator, and the final drying of the product under high vacuum for a minimum of 72 h. White powder (1.61 g, 41% yield).

¹H NMR (500 MHz, CDCl₃, *J* in Hz) δ 7.61 (d, *J* = 8.5, 1H), 7.56 (d, *J* = 7.9, 1H), 7.50 (d, *J* = 7.4, 1H), 5.66 (d, *J* = 9.5, 1H), 4.86 (td, *J* = 11.1, 4.1, 1H), 4.77 (d, *J* = 7.2, 1H), 4.60 – 4.54 (m, 1H), 4.41 (dd, *J* = 8.5, 5.4, 1H), 4.27 (dd, *J* = 7.4, 3.5, 1H), 3.96 (t, *J* = 8.9, 1H), 3.88 (dd, *J* = 17.3, 9.1, 1H), 3.69 (s, 3H), 3.08 (dd, *J* = 17.5, 11.5, 1H), 2.77 (dd, *J* = 17.5, 4.3, 1H), 2.60 (dd, *J* = 11.2, 5.1, 1H), 2.34 – 2.23 (m, 1H), 2.10 (dq, *J* = 13.4, 6.8, 1H), 2.05 – 1.96 (m, 1H), 1.93 – 1.82 (m, 1H), 1.82 – 1.71 (m, 2H), 1.58 – 1.42 (m, 3H), 1.39 (s, 9H), 0.93 (d, *J* = 6.9, 3H), 0.89 – 0.80 (m, 12H), 0.69 (d, *J* = 6.9, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 175.0, 174.8, 173.1, 172.2, 171.8, 171.5, 155.1, 80.4, 60.2, 59.7, 57.9, 52.3, 52.1, 47.8, 39.5, 37.6, 31.0, 30.7, 28.4, 26.8, 24.8, 24.2, 22.9, 21.9, 19.8, 19.0, 17.9, 16.4; FTIR (cm⁻¹) 3306, 2962, 1711, 1690, 1634, 1516, 1436, 1294, 1212, 1160, 1004; UPLC-MS (ESI) *m/z* 656.44 (calculated for C₃₁H₅₃N₅O₁₀ [M + H⁺] = 656.38); TLC R_f = 0.64 (85:15:1 DCM:MeOH:AcOH). [α]_D²⁰ = -159.7 (c = 5.225, CHCl₃).

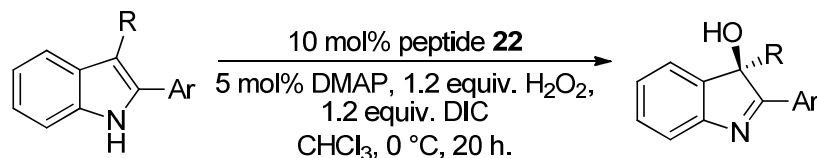
The enantiomer of the catalyst, *ent*-**22**, was prepared in an analogous fashion, using following monomers and reagents:

Boc-D-Val-OMe (1.62 g, 7.00 mmol, 1.0 equiv.), Boc-D-Leu-OH·H₂O (1.92 g, 7.7 mmol, 1.1 equiv.), Boc-Val-OH (1.67g, 7.7 mmol, 1.1 equiv.), Boc-D-Pro-OH (1.66 g, 7.7 mmol, 1.1 equiv.), Boc-D-Asp(Bn)-OH (2.49 g, 7.7 mmol, 1.1 equiv.), HOBt·H₂O (1.18 g, 7.7 mmol, 1.1 equiv.), EDC·HCl (1.48 g, 7.7 mmol, 1.1 equiv.), TEA (1.95 mL, 14.0 mmol, 2.0 equiv.). White powder (2.11 g, 46%).

¹H NMR (500 MHz, CDCl₃, *J* in Hz) δ 7.61 (d, *J* = 8.5, 1H), 7.56 (d, *J* = 7.9, 1H), 7.50 (d, *J* = 7.4, 1H), 5.66 (d, *J* = 9.5, 1H), 4.86 (td, *J* = 11.1, 4.1, 1H), 4.77 (d, *J* = 7.2, 1H), 4.60 – 4.54 (m, 1H), 4.41 (dd, *J* = 8.5, 5.4, 1H), 4.27 (dd, *J* = 7.4, 3.5, 1H), 3.96 (t, *J* = 8.9, 1H), 3.88 (dd, *J* = 17.3, 9.1, 1H), 3.69 (s, 3H),

3.08 (dd, $J = 17.5, 11.5$, 1H), 2.77 (dd, $J = 17.5, 4.3$, 1H), 2.60 (dd, $J = 11.2, 5.1$, 1H), 2.34 – 2.23 (m, 1H), 2.10 (dq, $J = 13.4, 6.8$, 1H), 2.05 – 1.96 (m, 1H), 1.93 – 1.82 (m, 1H), 1.82 – 1.71 (m, 2H), 1.58 – 1.42 (m, 3H), 1.39 (s, 9H), 0.93 (d, $J = 6.9$, 3H), 0.89 – 0.80 (m, 12H), 0.69 (d, $J = 6.9$, 3H); ^{13}C NMR (126 MHz, CDCl_3) δ 175.0, 174.7, 173.1, 172.2, 171.8, 171.5, 155.1, 80.3, 60.2, 59.7, 57.9, 52.3, 52.1, 47.8, 39.5, 37.5, 31.0, 30.7, 28.4, 26.8, 24.8, 24.2, 22.9, 21.9, 19.7, 18.9, 17.9, 16.5; FTIR (cm^{-1}) 3306, 2963, 1634, 1516, 1437, 1367, 1293, 1249, 1211, 1159; UPLC-MS (ESI) m/z 656.47 (calculated for $\text{C}_{31}\text{H}_{53}\text{N}_5\text{O}_{10}$ $[\text{M} + \text{H}^+] = 656.38$); TLC $R_f = 0.64$ (85:15:1DCM:MeOH:AcOH). $[\alpha]_{\text{D}}^{20} = +147.5$ ($c = 4.407$, CHCl_3).

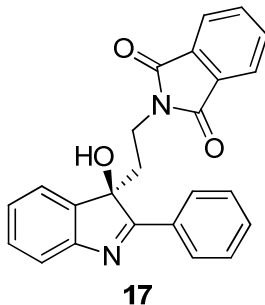
F. General Procedure for the Peptide Catalyzed Oxidation of 2,3-Disubstituted Indoles.



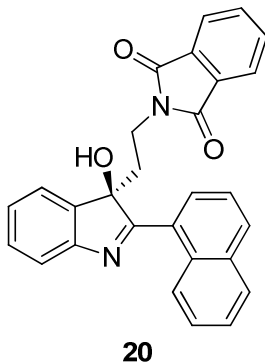
The oxidation of indoles was carried out in ¼-dram or 2-dram glass vials mounted to a cryogenic cooler with temperature control of ± 0.1 °C. Reactions were set up on the benchtop without exclusion of air or moisture. In a typical experiment, indole (0.1 mmol) was dissolved in 890 μL of chloroform (for deviations from the standard concentration see section G) and peptide **22** was added as a chloroform solution (typically 100 μL of 0.1 M solution, 0.01 mmol). To this solution was added a chloroform solution of *N,N*-dimethyl-4-aminopyridine (typically 10 μL of 0.5 M solution, 5 μmol), followed by 12.2 μL of aqueous 30% hydrogen peroxide (0.12 mmol). The biphasic mixture was stirred at 0 °C for 5 minutes, and the reaction was initiated by the addition 18.6 μL of diisopropylcarbodiimide (0.12 mmol). After 20 hours, the reaction mixture was diluted with 2.0 mL of chloroform and directly purified by flash chromatography or preparative TLC.

Racemic standards for the indole oxidation products were prepared by reacting the indole with 4.0 equiv. of peracetic acid ($\text{CH}_3\text{CO}_3\text{H}$) in chloroform (typically 0.1 M) at room temperature for 6-10 h. Standards were purified without previous workup in a manner analogous to that for the peptide catalyzed reactions.

G. Analytical Data for Indole Oxidation Products.

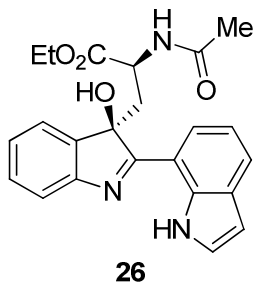


(R)-3-Hydroxy-3-(2-*N,N*-phthaloylaminoethyl)-2-phenyl-3*H*-indole (17). Purified by preparative TLC (50% EtOAc/Hex). Compound isolated as a mixture of conformers. White foam, 9.6 mg, 59% yield. $^1\text{H NMR}$ (500 MHz, CDCl_3 , J in Hz) δ 8.27 – 8.15 (m, 2H), 7.72 – 7.65 (m, 2H), 7.65 – 7.58 (m, 2H), 7.52 (d, $J = 6.6$, 1H), 7.48 (d, $J = 7.6$, 1H), 7.41 (dd, $J = 13.2$, 5.9, 1H), 7.36 (dd, $J = 13.8$, 6.2, 2H), 7.32 – 7.27 (m, 1H), 7.19 (t, $J = 7.4$, 1H), 3.58 – 3.48 (m, 1H), 3.44 – 3.35 (m, 1H), 2.82 – 2.62 (m, 2H), 2.52 – 2.43 (m, 1H); $^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 179.0, 167.9, 153.05, 140.2, 133.9, 132.1, 131.8, 131.6, 130.4, 128.7, 128.6, 126.8, 123.2, 122.6, 121.6, 86.0, 36.1, 33.3; **FTIR** (cm^{-1}) 3469, 1772, 1704, 1536, 1444, 1396, 1369, 1265, 1026; **UPLC-MS** (ESI) m/z 383.19 (calculated for $\text{C}_{24}\text{H}_{18}\text{N}_2\text{O}_3$ [$\text{M} + \text{H}^+$] = 383.13); **TLC** $R_f = 0.57$ (50% EtOAc/Hex); **Chiral HPLC analysis**: 77:23 er, performed on Chiralcel OD-H (Daicel, 4.6 mm \times 250 mm, 5 μm , 20 $^\circ\text{C}$), eluting at 0.20 mL/min with 80:20 hexanes/ethanol. Retention times: $t_{\text{minor}} = 35.0$ min, $t_{\text{major}} = 37.4$ min; $[\alpha]_{546\text{ nm}}^{20\text{ }^\circ\text{C}} = -17.1$ ($c = 0.164$, CHCl_3).



(R)-3-Hydroxy-3-(2-*N,N*-phthaloylaminoethyl)-2-(1-naphthyl)-3*H*-indole (20). Purified by preparative TLC (40% EtOAc/Hex). White solid, 14.9 mg, 57% yield. Product was recrystallized by slow evaporation from 95:5 2-propanol/ethanol mixture; white crystals, 11.9 mg, 45% yield. $^1\text{H NMR}$ (500 MHz, CDCl_3 , J in Hz) δ 9.08 (d, $J = 8.6$, 1H), 8.53 (dd, $J = 7.3$, 1.1, 1H), 7.93 (d, $J = 8.2$, 1H), 7.85 (d, $J = 7.8$, 1H), 7.66 – 7.62 (m, 3H), 7.61 – 7.56 (m, 3H), 7.56 – 7.48 (m, 3H), 7.33 (td, $J = 7.6$, 1.1, 1H), 7.18 (td, $J = 7.5$, 0.9, 1H), 3.71 – 3.63 (m, 1H), 3.56 – 3.46 (m, 1H), 2.77 (s, 1H), 2.67 – 2.58 (m, 1H), 2.34 (ddd, $J = 13.8$, 7.8, 5.9, 1H); $^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 179.6, 168.0, 153.6, 139.1, 134.4, 133.9, 132.0, 131.7, 131.7, 130.4, 129.3, 128.7, 128.3, 127.8, 126.9, 126.8, 126.3, 124.9, 123.2, 122.7, 122.0, 87.5, 35.6, 33.8; **FTIR** (cm^{-1}) 3352, 2943, 1764, 1698, 1567, 1507, 1401, 1339, 1273, 1033; **UPLC-MS** (ESI) m/z 433.20 (calculated for $\text{C}_{28}\text{H}_{20}\text{N}_2\text{O}_3$ [$\text{M} + \text{H}^+$] = 433.15); **TLC** $R_f = 0.45$ (40% EtOAc/Hex); **Chiral HPLC analysis**: 94:6er (>98:2 er after recrystallization), performed on Chiralcel OD (Daicel, 4.6 mm \times 250 mm,

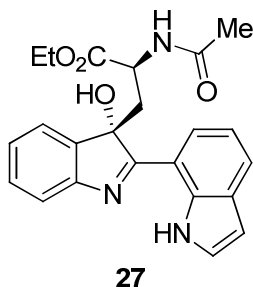
10 μm , 20 $^{\circ}\text{C}$), eluting at 0.50 mL/min with 90:10 hexanes/ethanol. Retention times: $t_{\text{major}} = 148.0$ min, $t_{\text{minor}} = 222.2$ min; $[\alpha]_{546\text{ nm}}^{20\text{ }^{\circ}\text{C}} = -174.8$ ($c = 0.078$, CHCl_3).



(2'S,3R)-3-Hydroxy-3-(2-N-acetylamino-2-ethoxycarbonyl-ethyl)-2-(7-indolyl)-3H-indole (26).

Purified by column chromatography on silica gel (60% EtOAc/Hex). Yellow foam, 586.0 mg, 73% yield.

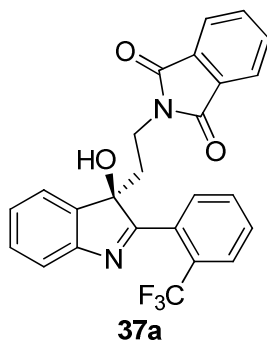
^1H NMR (400 MHz, CDCl_3 , J in Hz) δ 11.12 (s, 1H), 8.16 (d, $J = 7.5$, 1H), 7.85 (d, $J = 7.8$, 1H), 7.63 (d, $J = 7.6$, 1H), 7.53 (d, $J = 7.3$, 1H), 7.46 (td, $J = 7.6$, 1.1 Hz, 1H), 7.41 – 7.37 (m, 1H), 7.31 (t, $J = 7.4$, 1H), 7.24 (t, $J = 7.7$, 1H), 6.65 – 6.61 (m, 1H), 5.02 (d, $J = 8.5$, 1H), 4.07 (td, $J = 9.0$, 3.5, 1H), 3.96 (dq, $J = 10.8$, 7.1, 1H), 3.83 (dq, $J = 10.7$, 7.1, 1H), 3.03 (dd, $J = 14.2$, 3.5, 1H), 2.72 – 2.64 (m, 2H), 1.17 – 1.08 (m, 6H); **^{13}C NMR** (101 MHz, CDCl_3) δ 179.1, 171.6, 169.6, 153.6, 139.1, 134.7, 130.7, 129.1, 126.6, 125.4, 125.3, 123.5, 122.6, 121.2, 119.5, 115.4, 102.9, 86.1, 61.7, 48.8, 41.7, 22.0, 14.0; **FTIR** (cm^{-1}) 3380, 2984, 1731, 1657, 1522, 1433, 1372, 1335, 1266, 1212, 1059; **UPLC-MS** (ESI) m/z 406.23 (calculated for $\text{C}_{23}\text{H}_{23}\text{N}_3\text{O}_4$ [$\text{M} + \text{H}^+$] = 406.17); **TLC** $R_f = 0.35$ (70% EtOAc/Hex). $[\alpha]_{546\text{ nm}}^{20\text{ }^{\circ}\text{C}} = +89.5$ ($c = 0.140$, CHCl_3).



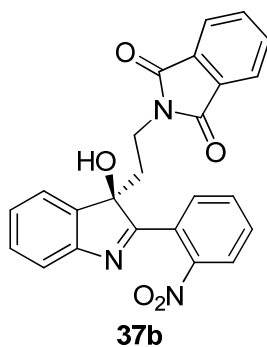
(2'S,3S)-3-Hydroxy-3-(2-N-acetylamino-2-ethoxycarbonyl-ethyl)-2-(7-indolyl)-3H-indole (27).

Purified by column chromatography on silica gel (60% EtOAc/Hex). White foam, 80.3 mg, 10% yield.

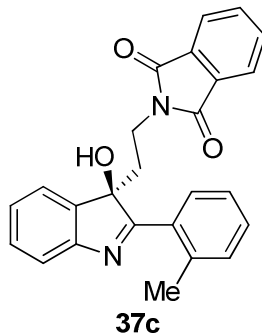
^1H NMR (500 MHz, CDCl_3 , J in Hz) δ 11.17 (s, 1H), 8.19 (d, $J = 7.6$, 1H), 7.85 (d, $J = 7.8$, 1H), 7.63 (d, $J = 7.6$, 1H), 7.57 (d, $J = 7.3$, 1H), 7.45 (t, $J = 8.0$, 1H), 7.42 – 7.39 (m, 1H), 7.28 (t, $J = 7.4$, 1H), 7.24 (t, $J = 7.7$, 1H), 6.67 – 6.62 (m, 1H), 6.33 (d, $J = 6.3$, 1H), 4.63 – 4.52 (m, 1H), 3.97 – 3.90 (m, 1H), 3.86 – 3.77 (m, 1H), 3.37 (s, 1H), 2.86 (dd, $J = 14.8$, 8.9, 1H), 2.46 (dd, $J = 14.7$, 3.5, 1H), 1.69 (s, 3H), 1.09 (t, $J = 7.1$, 3H); **^{13}C NMR** (126 MHz, CDCl_3) δ 179.9, 171.4, 170.3, 153.6, 139.2, 135.1, 130.6, 129.1, 126.3, 125.5, 125.3, 124.3, 123.4, 121.3, 119.5, 114.8, 102.9, 86.7, 61.8, 50.4, 41.1, 22.8, 14.0; **FTIR** (cm^{-1}) 3383, 3351, 2974, 2924, 2851, 1745, 1641, 1524, 1451, 1267, 1208, 1061; **UPLC-MS** (ESI) m/z 406.22 (calculated for $\text{C}_{23}\text{H}_{23}\text{N}_3\text{O}_4$ [$\text{M} + \text{H}^+$] = 406.17); **TLC** $R_f = 0.25$ (70% EtOAc/Hex); $[\alpha]_{546\text{ nm}}^{20\text{ }^{\circ}\text{C}} = -51.4$ ($c = 0.006$, CHCl_3).



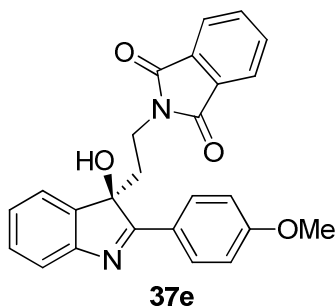
(R)-3-Hydroxy-3-(2-*N,N*-phthaloylaminoethyl)-2-(2-trifluoromethylphenyl)-3*H*-indole (37a). The concentration of the indole in this reaction was set to 0.5 M. Purified by column chromatography on silica gel (35% EtOAc/Hex). White foam, 83.4 mg, 93% yield. $^1\text{H NMR}$ (500 MHz, CDCl_3 , J in Hz) δ 8.06 (d, $J = 7.6$, 1H), 7.83 (d, $J = 7.8$, 1H), 7.81 – 7.76 (m, 2H), 7.72 – 7.67 (m, 2H), 7.66 – 7.61 (m, 2H), 7.59 (d, $J = 7.6$, 2H), 7.42 (td, $J = 7.6$, 1.2, 1H), 7.32 (td, $J = 7.5$, 1.0, 1H), 3.82 – 3.69 (m, 2H), 2.90 (s, 1H), 2.29 (ddd, $J = 13.9$, 9.5, 6.6, 1H), 2.16 – 2.06 (m, 1H); $^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 179.4, 168.3, 153.0, 139.2, 134.2, 132.3 (q, $J = 1.7$), 132.2, 131.8, 130.5, 130.0, 130.0, 129.4 (q, $J = 30.8$), 127.5, 124.1 (q, $J = 273.7$), 123.5, 123.2, 122.3, 87.0, 34.5, 33.2; $^{19}\text{F NMR}$ (376 MHz, CDCl_3) δ -57.0; **FTIR** (cm^{-1}) 3470, 2924, 1772, 1704, 1579, 1522, 1446, 1398, 1369, 1309, 1130, 1113, 1034; **UPLC-MS** (ESI) m/z 451.14 (calculated for $\text{C}_{25}\text{H}_{17}\text{F}_3\text{N}_2\text{O}_3[\text{M} + \text{H}^+] = 451.12$); **TLC** $R_f = 0.45$ (40% EtOAc/Hex); **Chiral HPLC analysis**: 94:6 er, performed on Chiralpak IA (Daicel, 4.6 mm \times 250 mm, 5 μm , 20 $^\circ\text{C}$), eluting at 0.50 mL/min with 80:20 hexanes/isopropanol. Retention times: $t_{\text{minor}} = 22.6$ min, $t_{\text{major}} = 33.3$ min; $[\alpha]_{546\text{ nm}}^{20\text{ }^\circ\text{C}} = -14.5$ ($c = 3.950$, CHCl_3).



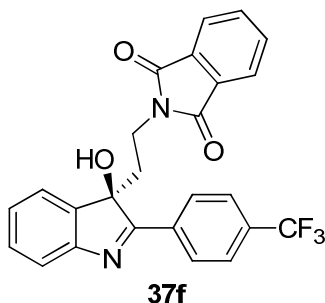
(R)-3-Hydroxy-3-(2-*N,N*-phthaloylaminoethyl)-2-(2-nitrophenyl)-3*H*-indole (37b). The concentration of the indole in this reaction was set to 0.5 M. Purified by preparative TLC (15% EtOAc/DCM). Yellow foam, 29.2 mg, 78% yield. $^1\text{H NMR}$ (500 MHz, CDCl_3 , J in Hz) δ 8.07 (d, $J = 8.2$, 1H), 8.01 (d, $J = 7.5$, 1H), 7.83 – 7.75 (m, 3H), 7.72 – 7.68 (m, 2H), 7.67 – 7.62 (m, 2H), 7.55 (d, $J = 7.6$, 1H), 7.41 (t, $J = 7.6$, 1H), 7.33 (t, $J = 7.4$, 1H), 3.69 – 3.58 (m, 2H), 2.93 (s, 1H), 2.45 – 2.36 (m, 1H), 2.18 – 2.08 (m, 1H); $^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 179.3, 168.2, 152.8, 149.1, 139.2, 134.2, 133.6, 132.2, 130.9, 130.5, 130.5, 129.0, 127.7, 125.1, 123.4, 123.2, 122.2, 87.6, 34.7, 33.1; **FTIR** (cm^{-1}) 3453, 3058, 1772, 1704, 1614, 1529, 1445, 1398, 1368, 1265, 1188; **UPLC-MS** (ESI) m/z 428.13 (calculated for $\text{C}_{24}\text{H}_{17}\text{N}_3\text{O}_5[\text{M} + \text{H}^+] = 428.12$); **TLC** $R_f = 0.45$ (15% EtOAc/DCM); **Chiral HPLC analysis**: 89:11 er, performed on Chiralcel OD-H (Daicel, 4.6 mm \times 250 mm, 5 μm , 20 $^\circ\text{C}$), eluting at 0.20 mL/min with 80:20 hexanes/ethanol. Retention times: $t_{\text{minor}} = 54.9$ min, $t_{\text{major}} = 65.0$ min; $[\alpha]_D^{20\text{ }^\circ\text{C}} = +149.0$ ($c = 0.603$, CHCl_3).



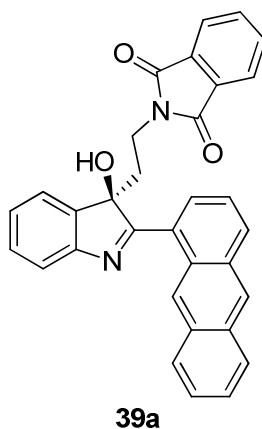
(R)-3-Hydroxy-3-(2-*N,N*-phthaloylaminoethyl)-2-(2-methylphenyl)-3*H*-indole (37c). Purified by preparative TLC (10% EtOAc/DCM). Compound isolated as ~5:1 mixture of conformers. White foam, 11.7 mg, 76% yield. $^1\text{H NMR}$ (500 MHz, CDCl_3 , J in Hz) δ 8.20 (d, $J = 7.9$, 0.15H), 8.17 (d, $J = 7.7$, 0.83H), 7.74 – 7.69 (m, 2H), 7.67 – 7.62 (m, 2H), 7.56 – 7.46 (m, 2H), 7.34 – 7.28 (m, 3H), 7.26 – 7.21 (m, 1H), 7.19 – 7.12 (m, 1H), 3.62 – 3.45 (m, 2H), 2.82 (s, 1H), 2.64 (s, 3H), 2.52 (ddd, $J = 13.9$, 8.7, 7.0, 1H), 2.28 – 2.16 (m, 1H); $^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 180.5, 168.0, 153.4, 139.1, 139.1, 134.0, 132.1, 131.9, 131.9, 130.2, 130.2, 129.2, 126.8, 125.7, 123.3, 122.7, 121.7, 87.0, 35.2, 33.2, 22.2; **FTIR** (cm^{-1}) 3450, 3057, 2925, 1772, 1706, 1600, 1534, 1447, 1397, 1368, 1265, 1188; **UPLC-MS** (ESI) m/z 397.17 (calculated for $\text{C}_{25}\text{H}_{20}\text{N}_2\text{O}_3[\text{M} + \text{H}^+] = 397.15$); **TLC** $R_f = 0.30$ (10% EtOAc/DCM); **Chiral HPLC analysis**: 88:12 er, performed on Chiralcel OJ-H (Daicel, 4.6 mm \times 250 mm, 5 μm , 20 $^\circ\text{C}$), eluting at 0.50 mL/min with 75:25 hexanes/ethanol. Retention times: $t_{\text{minor}} = 23.3$ min, $t_{\text{major}} = 68.7$ min; $[\alpha]_{546\text{ nm}}^{20\text{ }^\circ\text{C}} = -14.7$ ($c = 0.244$, CHCl_3).



(R)-3-Hydroxy-3-(2-*N,N*-phthaloylaminoethyl)-2-(4-methoxyphenyl)-3*H*-indole (37e). Purified by preparative TLC (50% EtOAc/Hex). Compound isolated as ~2:1 mixture of conformers. Yellow foam, 9.0 mg, 64% yield. $^1\text{H NMR}$ (500 MHz, CDCl_3 , J in Hz) δ 8.20 (d, $J = 8.8$, 0.67 H), 8.17 (d, $J = 8.8$, 1.29H), 7.72 – 7.66 (m, 2H), 7.66 – 7.59 (m, 2H), 7.51 (dd, $J = 7.1$, 3.4, 1H), 7.47 (d, $J = 7.4$, 0.31H), 7.42 (d, $J = 7.6$, 0.64H), 7.30 – 7.25 (m, 1H), 7.20 – 7.15 (m, 1H), 6.87 (d, $J = 9.0$, 0.58H), 6.84 (d, $J = 9.0$, 1.20H), 3.81 (s, 3H), 3.56 – 3.47 (m, 1H), 3.43 – 3.35 (m, 1H), 2.82 – 2.65 (m, 2H), 2.53 – 2.44 (m, 1H); $^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 178.6, 167.8, 162.1, 152.8, 139.9, 133.8, 132.0, 130.5, 130.1, 126.0, 124.4, 123.1, 122.5, 120.8, 113.9, 85.4, 55.4, 36.3, 33.2; **FTIR** (cm^{-1}) 3464, 2961, 1772, 1705, 1603, 1508, 1397, 1253, 1173, 1024; **UPLC-MS** (ESI) m/z 413.20 (calculated for $\text{C}_{25}\text{H}_{20}\text{N}_2\text{O}_4[\text{M} + \text{H}^+] = 413.14$); **TLC** $R_f = 0.34$ (50% EtOAc/Hex); **Chiral HPLC analysis**: 75:25 er, performed on Chiralcel OJ-H (Daicel, 4.6 mm \times 250 mm, 5 μm , 20 $^\circ\text{C}$), eluting at 0.50 mL/min with 75:25 hexanes/ethanol. Retention times: $t_{\text{major}} = 53.1$ min, $t_{\text{minor}} = 78.0$ min; $[\alpha]_{546\text{ nm}}^{20\text{ }^\circ\text{C}} = +104.4$ ($c = 0.440$, N,N -DMF).

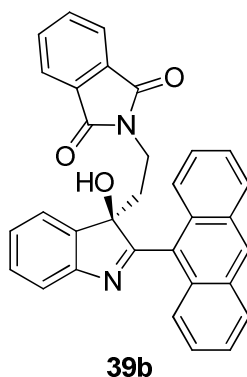


(R)-3-Hydroxy-3-(2-*N,N*-phthaloylaminoethyl)-2-(4-trifluoromethylphenyl)-3*H*-indole (37f). The concentration of the indole in this reaction was set to 0.5 M. Purified by preparative TLC (40% EtOAc/Hex). Compound isolated as ~2.5:1 mixture of conformers. White foam, 24.3 mg, 69% yield. ¹H NMR (500 MHz, CDCl₃, *J* in Hz) δ 8.20 (d, *J* = 8.1, 0.57 H), 8.13 (d, *J* = 8.1, 1.36 H), 7.76 – 7.54 (m, 4H), 7.51 – 7.38 (m, 3.28H), 7.32 (d, *J* = 7.3, 0.69H), 7.26 – 7.11 (m, 2H), 3.53 – 3.38 (m, 1H), 3.35 – 2.92 (m, 2H), 2.78 – 2.62 (m, 1H), 2.58 – 2.39 (m, 1H); ¹³C NMR (126 MHz, CDCl₃, *J* in Hz) δ 177.6, 177.5, 167.9, 167.8, 152.6, 152.5, 140.0, 140.0, 134.8, 134.7, 134.0, 132.6 (q, *J* = 32.6), 132.6 (q, *J* = 32.6), 131.9, 131.8, 130.6, 130.6, 128.7, 128.6, 127.4, 127.3, 124.0 (q, *J* = 272.0), 123.9 (q, *J* = 272.3), 123.2, 122.6, 122.0, 121.8, 85.8, 85.7, 36.0, 35.9, 33.1, 33.1; ¹⁹F NMR (376 MHz, CDCl₃) δ -63.1; FTIR (cm⁻¹) 3463, 1773, 1707, 1617, 1539, 1396, 1322, 1123, 1067, 1017; UPLC-MS (ESI) *m/z* 451.15 (calculated for C₂₅H₁₇F₃N₂O₃[M + H⁺] = 451.12); TLC *R_f* = 0.42 (40% EtOAc/Hex); Chiral HPLC analysis: 72:28 er, performed on Chiralpak IA (Daicel, 4.6 mm × 250 mm, 5 μm, 20 °C), eluting at 0.50 mL/min with 80:20 hexanes/isopropanol. Retention times: *t*_{major} = 20.2 min, *t*_{minor} = 25.6 min; [α]_{546 nm}^{20 °C} = -18.0 (*c* = 0.050, CHCl₃).

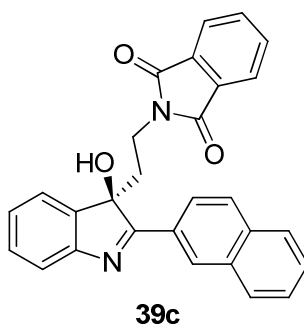


(R)-2-(1-Anthracenyl)-3-hydroxy-3-(2-*N,N*-phthaloylaminoethyl)-3*H*-indole (39a). Purified by column chromatography on silica gel (35% EtOAc/Hex). Yellow solid, 89.9 mg, 88% yield. Product was recrystallized by slow evaporation from DCM; yellow crystals, 72.7 mg, 71% yield. ¹H NMR (500 MHz, CDCl₃, *J* in Hz) δ 9.82 (s, 1H), 8.56 (d, *J* = 7.1, 1H), 8.34 (s, 1H), 8.13 – 8.07 (m, 1H), 8.03 (d, *J* = 8.6, 1H), 7.98 – 7.93 (m, 1H), 7.70 (d, *J* = 7.6, 1H), 7.54 (d, *J* = 7.2, 1H), 7.51 – 7.45 (m, 4H), 7.43 (dd, *J* = 8.4, 7.2, 1H), 7.39 (dd, *J* = 5.5, 3.0, 2H), 7.36 (dd, *J* = 7.6, 1.2, 1H), 7.21 (t, *J* = 7.0, 1H), 3.62 (dt, *J* = 14.0, 6.9, 1H), 3.47 (dt, *J* = 14.0, 7.0, 1H), 2.90 (s, 1H), 2.66 (dt, *J* = 14.0, 6.9, 1H), 2.53 (dt, *J* = 14.0, 7.0, 1H); ¹³C NMR (126 MHz, CDCl₃) δ 179.3, 167.9, 153.7, 139.1, 133.6, 132.9, 132.4, 132.4, 131.8,

131.6, 130.4, 129.8, 129.2, 129.2, 128.8, 127.8, 127.1, 126.9, 126.7, 126.1, 125.7, 124.2, 122.8, 122.6, 122.1, 87.7, 35.9, 33.5; **FTIR** (cm^{-1}) 3424, 3046, 2944, 1762, 1696, 1616, 1517, 1395, 1348, 1198, 1019; **UPLC-MS** (ESI) m/z 483.16 (calculated for $\text{C}_{32}\text{H}_{22}\text{N}_2\text{O}_3[\text{M} + \text{H}^+] = 483.16$); **TLC** $R_f = 0.43$ (40% EtOAc/Hex); **Chiral HPLC analysis**: 95:5 er (>99:1 after recrystallization), performed on Chiralcel OD-H (Daicel, 4.6 mm \times 250 mm, 5 μm , 20 $^\circ\text{C}$), eluting at 0.50 mL/min with 85:15 hexanes/ethanol. Retention times: $t_{\text{minor}} = 27.8$ min, $t_{\text{major}} = 46.7$ min; $[\alpha]_{546 \text{ nm}}^{20 \text{ }^\circ\text{C}} = -18.4$ ($c = 3.380$, N,N -DMF) (recrystallized material).

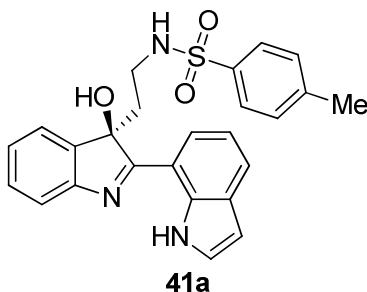


(R)-2-(9-Anthracenyl)-3-hydroxy-3-(2-*N,N*-phthaloylaminoethyl)-3H-indole (39b). Purified by preparative TLC (40% EtOAc/Hex). White solid, 1.2 mg, 7% yield. $^1\text{H NMR}$ (500 MHz, CDCl_3 , J in Hz) δ 8.50 (s, 1H), 8.07 – 8.98 (m, 3H), 7.97 – 7.85 (m, 1H), 7.75 (dd, $J = 11.2, 7.5$, 2H), 7.68 – 7.59 (m, 4H), 7.54 (td, $J = 7.7, 1.1$, 1H), 7.51 – 7.37 (m, 5H), 4.00 – 3.91 (m, 1H), 3.83 – 3.76 (m, 1H), 2.22 – 2.13 (m, 1H), 2.03 – 1.94 (m, 1H); $^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 183.7, 168.3, 153.2, 139.7, 134.1, 133.9, 132.0, 131.5, 130.4, 129.1, 128.5, 127.8, 127.3, 123.7, 123.5, 122.2, 88.7, 35.8, 33.1; **FTIR** (cm^{-1}) 3451, 3051, 2923, 2852, 1770, 1700, 1574, 1520, 1444, 1397, 1366, 1264, 1088; **UPLC-MS** (ESI) m/z 483.17 (calculated for $\text{C}_{32}\text{H}_{22}\text{N}_2\text{O}_3[\text{M} + \text{H}^+] = 483.16$); **TLC** $R_f = 0.40$ (7% EtOAc/DCM); **Chiral HPLC analysis**: 84:16 er, performed on Chiralcel OD-H (Daicel, 4.6 mm \times 250 mm, 5 μm , 20 $^\circ\text{C}$), eluting at 0.50 mL/min with 85:15 hexanes/ethanol. Retention times: $t_{\text{minor}} = 30.7$ min, $t_{\text{major}} = 50.2$ min; $[\alpha]_{546 \text{ nm}}^{20 \text{ }^\circ\text{C}} = +830.8$ ($c = 0.035$, CHCl_3).

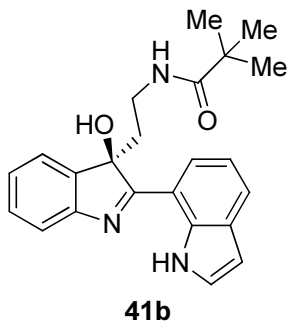


(R)-3-Hydroxy-2-(2-naphthyl)-3-(2-*N,N*-phthaloylaminoethyl)-3H-indole (39c). Purified by preparative TLC (40% EtOAc/Hex). Compound isolated as ~2:1 mixture of conformers. White solid, 14.1 mg, 74% yield. $^1\text{H NMR}$ (500 MHz, CDCl_3 , J in Hz) δ 8.60 (s, 0.31 H), 8.53 (s, 0.65 H), 8.14 (d, $J = 8.6$, 0.32 H), 8.08 (d, $J = 8.6$, 0.67 H), 7.78 (d, $J = 8.0$, 0.33 H), 7.70 (t, $J = 7.0$, 1H), 7.63 (d, $J = 7.0$, 1H),

7.54 (d, $J = 8.7$, 1H), 7.52 – 7.41 (m, 6H), 7.41 – 7.35 (m, 1H), 7.31 (d, $J = 7.5$, 1H), 7.21 – 7.13 (m, 1H), 7.11 (t, $J = 7.3$, 1H), 3.51 – 3.38 (m, 1H), 3.36 – 3.25 (m, 1H), 2.82 – 2.71 (m, 1H), 2.66 – 2.52 (m, 1H), 1.67 (brs, 1H); $^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 179.0, 178.9, 167.9, 167.8, 153.0, 152.9, 140.2, 134.7, 134.7, 133.7, 133.7, 132.9, 132.8, 131.8, 130.3, 129.5, 129.5, 129.1, 129.0, 128.3, 128.2, 127.8, 127.7, 127.6, 126.7, 126.6, 126.5, 126.4, 124.9, 124.8, 123.0, 123.0, 122.5, 121.5, 121.4, 85.9, 85.8, 36.3, 36.3, 33.3; **FTIR** (cm^{-1}) 3457, 2924, 1771, 1705, 1532, 1396, 1367, 1190, 1021; **UPLC-MS** (ESI) m/z 433.17 (calculated for $\text{C}_{28}\text{H}_{20}\text{N}_2\text{O}_3[\text{M} + \text{H}^+] = 433.15$); **TLC** $R_f = 0.31$ (10% EtOAc/DCM); **Chiral HPLC analysis**: 67:33 er, performed on Chiralcel OD-H (Daicel, 4.6 mm \times 250 mm, 5 μm , 20 $^\circ\text{C}$), eluting at 0.50 mL/min with 85:15 hexanes/ethanol. Retention times: $t_{\text{minor}} = 18.7$ min, $t_{\text{major}} = 22.9$ min; $[\alpha]_{546\text{ nm}}^{20\text{ }^\circ\text{C}} = -24.4$ ($c = 1.000$, CHCl_3).

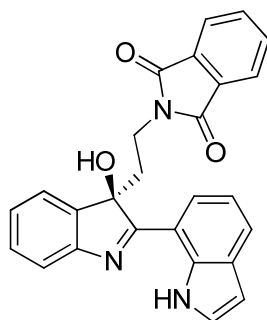


(R)-3-Hydroxy-3-(2-N-(4-toluenesulfonyl)aminoethyl)-2-(7-indolyl)-3H-indole (41a). Purified by preparative TLC (15% EtOAc/DCM). Yellow foam, 11.1 mg, 77% yield. $^1\text{H NMR}$ (500 MHz, CDCl_3 , J in Hz) δ 10.99 (s, 1H), 8.05 (dd, $J = 7.5$, 0.6, 1H), 7.83 (d, $J = 7.8$, 1H), 7.54 (t, $J = 7.9$, 3H), 7.39 (ddd, $J = 7.7$, 6.6, 2.2, 1H), 7.36 (dd, $J = 3.0$, 2.4, 1H), 7.21 (d, $J = 7.9$, 2H), 7.19 – 7.13 (m, 3H), 6.62 (dd, $J = 3.1$, 2.3, 1H), 4.91 – 4.84 (m, 1H), 2.93 – 2.83 (m, 2H), 2.68 (brs, 1H), 2.54 (ddd, $J = 14.4$, 8.2, 7.2, 1H), 2.43 (s, 3H), 1.89 (dt, $J = 14.1$, 5.8, 1H); $^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 179.6, 152.9, 143.5, 139.1, 136.7, 134.9, 130.5, 129.9, 129.1, 127.3, 126.3, 125.6, 125.3, 124.2, 122.8, 121.2, 119.4, 114.3, 102.9, 87.5, 39.2, 38.2, 21.8; **FTIR** (cm^{-1}) 3453, 3375, 3297, 3068, 2924, 1700, 1657, 1598, 1522, 1433, 1334, 1266, 1157, 1091; **UPLC-MS** (ESI) m/z 446.16 (calculated for $[\text{M} + \text{H}^+] = 446.15$); **TLC** $R_f = 0.30$ (35% EtOAc/Hex); **Chiral HPLC analysis**: 86:14 er, performed on Chiralcel OD (Daicel, 4.6 mm \times 250 mm, 10 μm , 20 $^\circ\text{C}$), eluting at 0.10 mL/min with 80:20 hexanes/ethanol. Retention times: $t_{\text{minor}} = 99.0$ min, $t_{\text{major}} = 107.8$ min.



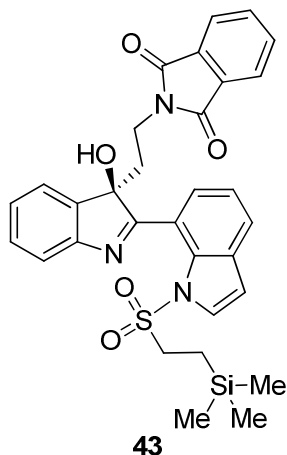
(R)-3-Hydroxy-2-(7-indolyl)-3-(2-N-pivaloylaminoethyl)-3H-indole (41b). Purified by column chromatography (35% EtOAc/Hex). White foam, 10.2 mg, 87% yield. $^1\text{H NMR}$ (500 MHz, CDCl_3 , J in Hz) δ 11.16 (s, 1H), 8.21 (d, $J = 7.6$, 1H), 7.85 (d, $J = 7.8$, 1H), 7.61 (d, $J = 7.6$, 1H), 7.51 (d, $J = 7.3$,

1H), 7.43 (td, $J = 7.6, 1.2$, 1H), 7.40 – 7.38 (m, 1H), 7.28 (t, $J = 7.4$, 1H), 7.23 (t, $J = 7.7$, 1H), 6.65 – 6.62 (m, 1H), 5.76 – 5.69 (m, 1H), 3.24 – 3.12 (m, 1H), 3.00 – 2.89 (m, 2H), 2.68 – 2.57 (m, 1H), 2.30 – 2.19 (m, 1H), 0.92 (s, 9H); $^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 180.4, 178.6, 153.2, 140.1, 135.1, 130.3, 129.2, 126.6, 125.5, 125.3, 124.3, 122.9, 121.1, 119.4, 114.8, 102.9, 87.5, 39.3, 38.5, 35.5, 27.3; **FTIR** (cm^{-1}) 3377, 2964, 1634, 1520, 1335, 1265, 1201, 1051; **UPLC-MS** (ESI) m/z 376.21 (calculated for $\text{C}_{23}\text{H}_{25}\text{N}_3\text{O}_2[\text{M} + \text{H}^+] = 376.19$); **TLC** $R_f = 0.39$ (45% EtOAc/Hex); **Chiral HPLC analysis**: 83:17 er, performed on Chiralcel OJ-H (Daicel, 4.6 mm \times 250 mm, 5 μm , 20 $^\circ\text{C}$), eluting at 0.20 mL/min with 90:10 hexanes/isopropanol. Retention times: $t_{\text{major}} = 128.6$ min, $t_{\text{minor}} = 164.5$ min; $[\alpha]_{546\text{ nm}}^{20\text{ }^\circ\text{C}} = +49.4$ ($c = 2.110$, CHCl_3).

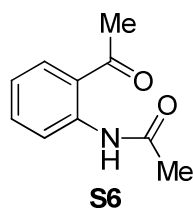


41c

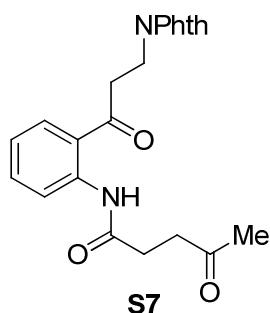
(R)-3-Hydroxy-2-(7-indolyl)-3-(2-N,N-phthaloylaminoethyl)-3H-indole (41c). Purified by preparative TLC (40% EtOAc/Hex). Yellow solid, 5.5 mg, 89% yield. Product was recrystallized by slow evaporation from 95:5 2-propanol/ethanol mixture; yellow crystals, 2.90 mg, 47% yield. $^1\text{H NMR}$ (500 MHz, CDCl_3 , J in Hz) δ 11.03 (s, 1H), 8.10 (d, $J = 7.5$, 1H), 7.64 (d, $J = 7.8$, 1H), 7.59 (d, $J = 7.6$, 1H), 7.54 (d, $J = 7.3$, 1H), 7.51 – 7.46 (m, 4H), 7.36 (td, $J = 7.7, 1.1$, 1H), 7.26 – 7.24 (m, 1H), 7.21 (t, $J = 7.4$, 1H), 7.14 (td, $J = 7.7, 1.8$, 1H), 6.45 – 6.41 (m, 1H), 3.55 – 3.44 (m, 1H), 3.40 – 3.29 (m, 1H), 2.85 (t, $J = 6.4$, 2H), 2.55 (s, 1H); $^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 179.1, 168.0, 153.6, 139.5, 134.8, 133.6, 131.5, 130.4, 128.9, 126.5, 125.2, 125.0, 124.1, 122.8, 122.5, 121.2, 119.4, 114.8, 102.7, 86.5, 37.5, 33.4; **FTIR** (cm^{-1}) 3441, 3377, 3059, 2941, 1767, 1697, 1521, 1396, 1335, 1266, 1196, 1017; **UPLC-MS** (ESI) m/z 422.21 (calculated for $\text{C}_{26}\text{H}_{19}\text{N}_3\text{O}_3[\text{M} + \text{H}^+] = 422.14$); **TLC** $R_f = 0.47$ (40% EtOAc/Hex); **Chiral HPLC analysis**: 82:18er (95:5 er after recrystallization), performed on Chiralcel OD-H (Daicel, 4.6 mm \times 250 mm, 5 μm , 20 $^\circ\text{C}$), eluting at 0.50 mL/min with 80:20 hexanes/ethanol. Retention times: $t_{\text{minor}} = 19.2$ min, $t_{\text{major}} = 20.8$ min; $[\alpha]_{578\text{ nm}}^{20\text{ }^\circ\text{C}} = +136.8$ ($c = 0.100$, CHCl_3).



(R)-3-Hydroxy-2-(N-(2-trimethylsilylethanesulfonyl)indol-7-yl)-3-(2-N,N-phthaloylaminoethyl)-3H-indole (43). Purified by preparative TLC (40% EtOAc/Hex). White foam, 8.2 mg, 70% yield. $^1\text{H NMR}$ (500 MHz, CDCl_3 , J in Hz) δ 7.81 (dd, $J = 5.4, 3.1$, 2H), 7.75 (dd, $J = 7.8, 1.1$, 1H), 7.71 – 7.67 (m, 3H), 7.61 (d, $J = 7.3$, 1H), 7.57 (d, $J = 3.7$, 1H), 7.51 (d, $J = 7.1$, 1H), 7.48 (t, $J = 7.7$, 1H), 7.43 (td, $J = 7.5, 1.4$, 1H), 7.38 (td, $J = 7.5, 1.1$, 1H), 6.77 (d, $J = 3.7$, 1H), 4.07 (s, 1H), 3.70 – 3.54 (m, 3H), 3.49 (td, $J = 14.0, 4.0$, 1H), 2.50 (ddd, $J = 13.0, 11.4, 5.8$, 1H), 2.21 (ddd, $J = 12.7, 10.8, 4.6$, 1H), 0.87 (td, $J = 13.8, 4.2$, 1H), 0.55 (td, $J = 13.8, 3.7$, 1H), 0.02 (s, 9H); $^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 184.06, 168.2, 152.5, 140.6, 134.1, 133.2, 132.9, 132.3, 130.0, 129.7, 127.5, 125.3, 123.6, 123.5, 123.4, 123.3, 121.2, 120.4, 108.3, 88.0, 50.6, 34.7, 33.5, 9.6, -1.7; **FTIR** (cm^{-1}) 3510, 2956, 1773, 1709, 1570, 1398, 1367, 1250, 1157, 1130, 833; **UPLC-MS** (ESI) m/z 586.18 (calculated for $[\text{M} + \text{H}^+] = 586.18$); **TLC** R_f = 0.40 (35% EtOAc/Hex); **Chiral HPLC analysis**: 93:7er, performed on Chiralcel OD (Daicel, 4.6 mm \times 250 mm, 10 μm , 20 $^\circ\text{C}$), eluting at 0.1 mL/min with 96:4 hexanes/ethanol. Retention times: $t_{\text{major}} = 180.6$ min, $t_{\text{minor}} = 200.3$ min; $[\alpha]_{\text{D}}^{20} = +271.1$ ($c = 0.100$, CHCl_3).



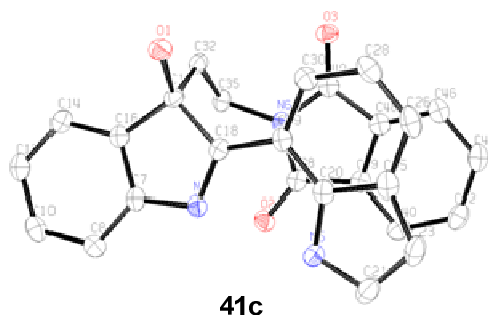
S6. Purified by preparative TLC (30% EtOAc/Hex). White solid, 25.0 mg, 45% yield. $^1\text{H NMR}$ (500 MHz, CDCl_3 , J in Hz) δ 11.70 (s, 1H), 8.75 (dd, $J = 8.5, 0.9$, 1H), 7.90 (dd, $J = 8.0, 1.5$, 1H), 7.62 – 7.50 (m, 1H), 7.18 – 7.06 (m, 1H), 2.68 (s, 3H), 2.24 (s, 3H); $^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 203.1, 169.7, 141.3, 135.4, 131.8, 122.5, 121.9, 120.9, 28.8, 25.8; **FTIR** (cm^{-1}) 3242, 2973, 2926, 1685, 1650, 1581, 1522, 1452, 1360, 1239, 1168; **UPLC-MS** (ESI) m/z 178.40 (calculated for $[\text{M} + \text{H}^+] = 178.08$); **TLC** R_f = 0.44 (30% EtOAc/Hex).



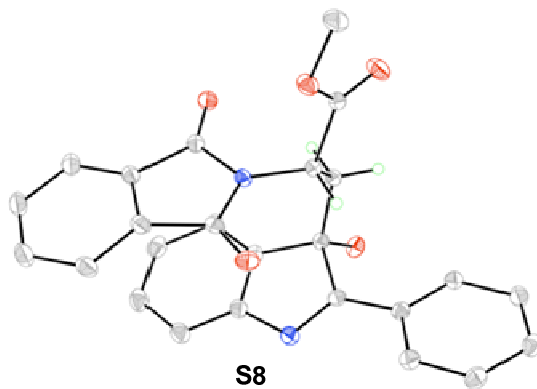
S7. Purified by preparative TLC (65% EtOAc/Hex). Yellow solid, 10.0 mg, 13% yield. $^1\text{H NMR}$ (500 MHz, CDCl_3 , J in Hz) δ 11.64 (s, 1H), 8.70 (dd, $J = 8.5, 1.0$, 1H), 7.89 – 7.83 (m, 3H), 7.74 (dd, $J = 5.5, 3.0$, 2H), 7.56 – 7.50 (m, 1H), 7.12 – 7.03 (m, 1H), 4.20 – 4.09 (m, 2H), 3.54 – 3.44 (m, 2H), 2.87 (t, $J = 6.5$, 2H), 2.75 (t, $J = 6.5$, 2H), 2.24 (s, 3H); $^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 207.1, 201.9, 171.4, 168.3, 141.2, 135.5, 134.3, 134.1, 132.3, 130.8, 123.6, 122.6, 121.1, 38.3, 33.8, 32.0, 30.2, 25.0; **FTIR** (cm^{-1}) 3473, 2951, 1771, 1706, 1650, 1583, 1522, 1450, 1397, 1365, 1200, 910; **UPLC-MS** (ESI) m/z 393.55 (calculated for $[\text{M} + \text{H}^+] = 393.14$); **TLC** $R_f = 0.62$ (65% EtOAc/Hex).

H. X-Ray Structure of the Compound **41c** and Proof of Absolute Stereochemistry.

The absolute stereochemistry of the compound **41c** was established as *R* by X-ray crystallographic analysis. The details of the analysis are in section K, following the NMR spectra.

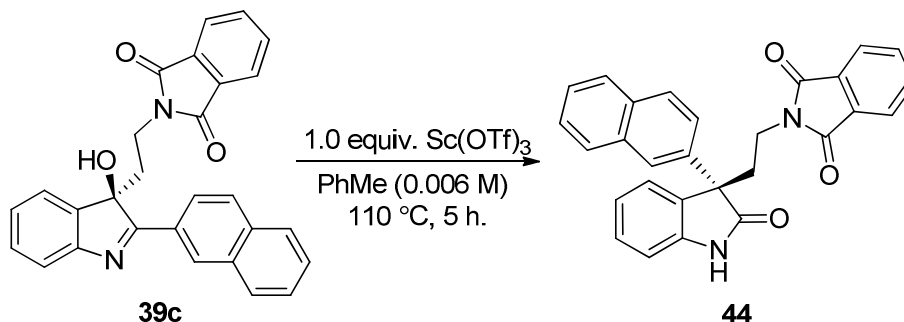


Support for the stereochemical assignment derives from the work of Movassaghi *et al.*⁴, who reported a crystal structure of a related indole oxidation product **S8**. This compound is identical in spectroscopic and chromatographic properties to the *minor* diastereomer **24** obtained in the matched case of the oxidation of indole **21** using peptide catalyst **22**.



I. Procedure for the Stereospecific Rearrangement of 3-Hydroxy-indolenines.

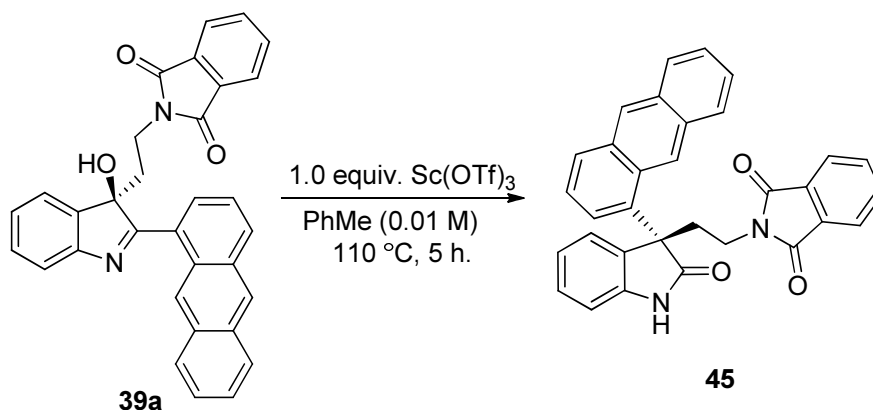
(*S*)-3-(2-*N,N*-phthaloylaminoethyl)-3-(2-naphthyl)oxindole (**44**)



A 5-mL round-bottom flask was charged with 3-hydroxy-indolenine **39c** (7.0 mg, 0.01619 mmol, 1.0 equiv.), scandium(III) triflate (8.0 mg, 0.01619 mmol, 1.0 equiv.) and degassed toluene (2.5 mL). The flask was flushed with nitrogen, sealed, and heated in an oil bath at 110 °C for 5 hours. The resulting yellow solution was directly purified by preparative TLC eluting with 10% EtOAc/DCM. After removal of the desorption solvent (EtOAc), the 2-oxindole **44** was isolated as white solid (5.0 mg, 71% yield).

¹H NMR (500 MHz, CDCl₃, *J* in Hz) δ 7.92 (s, 1H), 7.82 (d, *J* = 1.7, 1H), 7.79 – 7.66 (m, 5H), 7.61 (dd, *J* = 5.5, 3.1, 2H), 7.54 (dd, *J* = 8.7, 2.0, 1H), 7.45 – 7.40 (m, 2H), 7.38 (d, *J* = 7.4, 1H), 7.17 (td, *J* = 7.7, 1.2, 1H), 6.99 (td, *J* = 7.6, 1.0, 1H), 6.95 (d, *J* = 7.8, 1H), 3.88 – 3.66 (m, 2H), 2.95 (dt, *J* = 15.2, 7.6, 1H), 2.87 – 2.75 (m, 1H); **¹³C NMR** (126 MHz, CDCl₃) δ 179.6, 168.2, 140.9, 136.6, 133.9, 133.4, 132.7, 132.1, 131.9, 128.7, 128.6, 128.4, 127.6, 126.4, 126.3, 125.9, 125.6, 124.9, 123.2, 123.0, 110.5, 55.7, 34.9, 34.7; **FTIR** (cm⁻¹) 3236, 3051, 2917, 1765, 1699, 1615, 1469, 1349, 1234; **UPLC-MS** (ESI) *m/z* 433.57 (calculated for C₂₈H₂₀N₂O₃ [M + H⁺] = 433.15); **TLC** R_f = 0.34 (10% EtOAc/DCM); **Chiral HPLC analysis**: 67:33 er, performed on Chiralcel OJ-H (Daicel, 4.6 mm × 250 mm, 5 μm, 20 °C), eluting at 0.50 mL/min with 80:20 hexanes/ethanol. Retention times: *t*_{minor} = 64.3 min, *t*_{major} = 84.8 min; **[α]_{546 nm}^{20 °C}** = -24.4 (*c* = 1.0, CHCl₃).

(S)-3-(2-*N,N*-phthaloylaminoethyl)-3-(1-anthracenyl)oxindole (45)

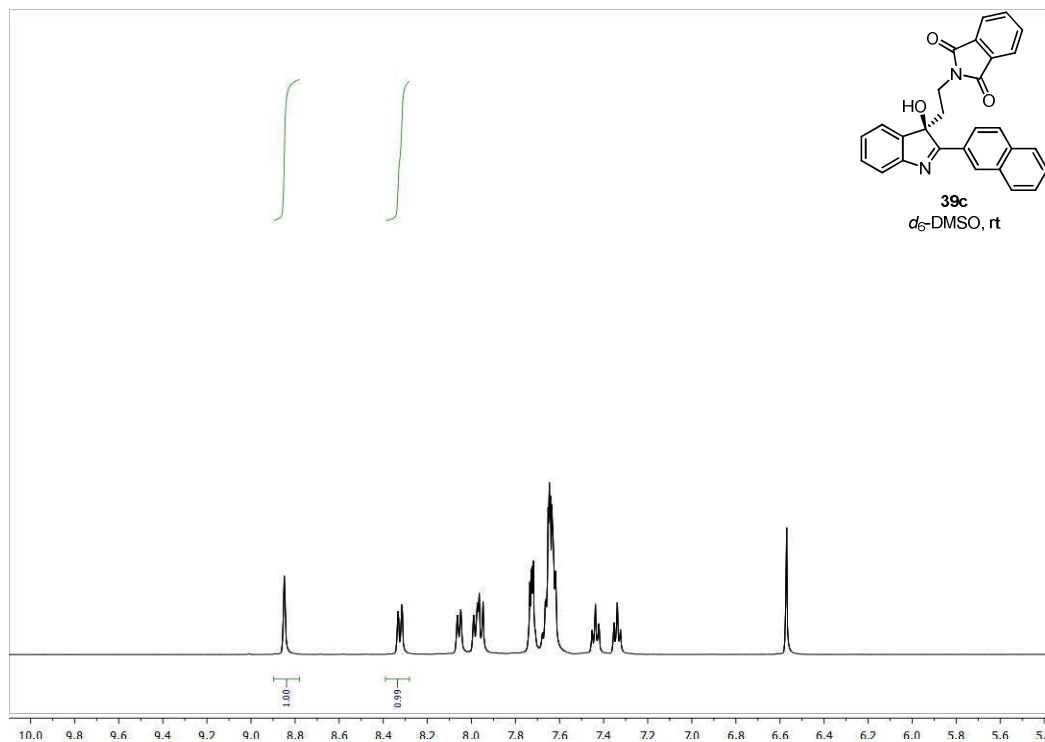
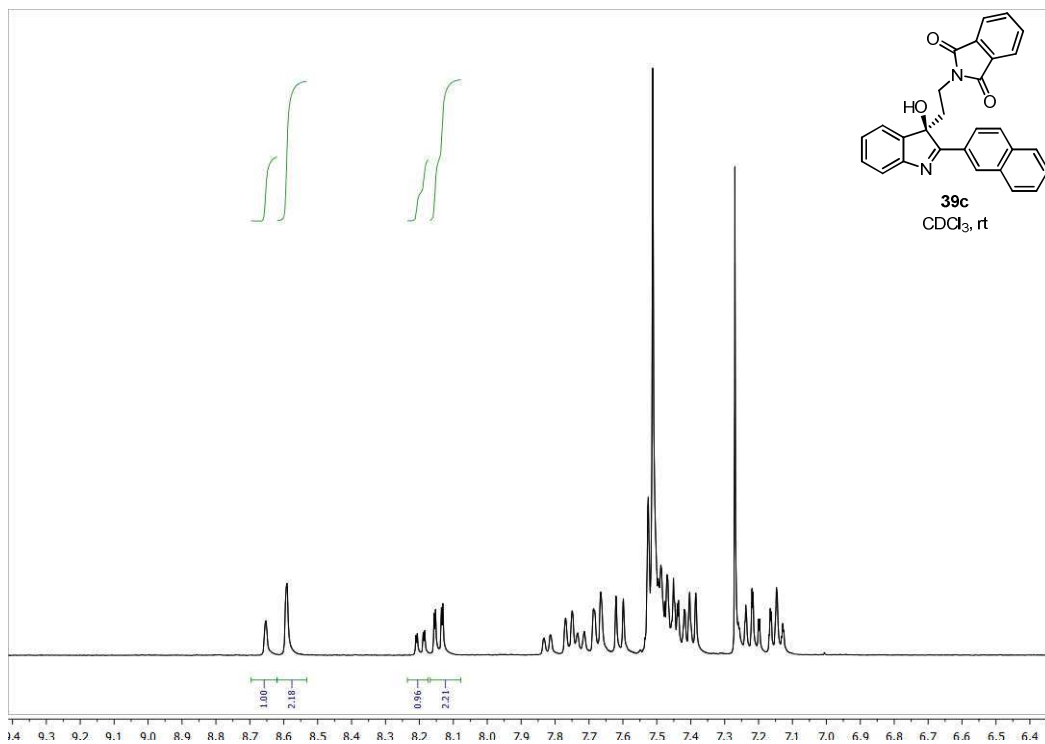


A 5-mL round-bottom flask was charged with 3-hydroxy-indolenine **39a** (19.8 mg, 0.04103 mmol, 1.0 equiv.), scandium(III) triflate (20.2 mg, 0.04103 mmol, 1.0 equiv.) and degassed toluene (4.1 mL). The flask was flushed with nitrogen, sealed, and heated in an oil bath at 110 °C for 5 hours. The resulting yellow solution was directly purified by preparative TLC eluting with 10% EtOAc/DCM. After removal of the desorption solvent (EtOAc), the 2-oxindole **45** was isolated as white solid (7.0 mg, 35% yield).

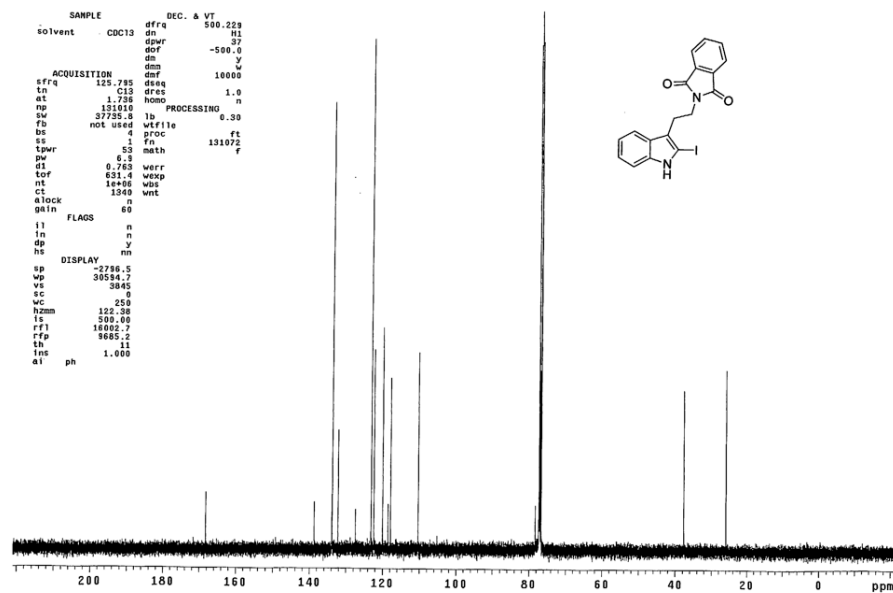
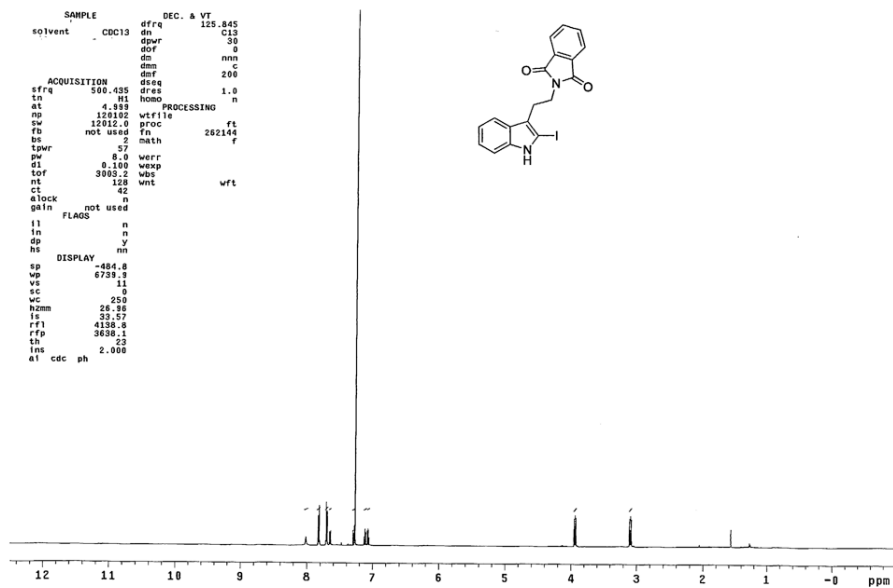
¹H NMR (500 MHz, CDCl₃, *J* in Hz) δ 8.51 (s, 1H), 8.38 (s, 1H), 8.05 (d, *J* = 7.0, 1H), 8.01 (d, *J* = 8.7, 1H), 7.91 (s, 1H), 7.88 (d, *J* = 8.3, 1H), 7.79 (dd, *J* = 5.5, 3.0, 2H), 7.68 (dd, *J* = 5.5, 3.0, 2H), 7.57 (dd, *J* = 8.4, 7.2, 1H), 7.53 (d, *J* = 8.4, 1H), 7.39 – 7.33 (m, 1H), 7.26 – 7.17 (m, 2H), 7.14 (d, *J* = 7.7, 1H), 6.95 – 6.90 (m, 1H), 6.87 (td, *J* = 7.4, 1.0, 1H), 4.02 – 3.94 (m, 1H), 3.91 – 3.83 (m, 1H), 2.96 – 2.83 (m, 2H); **¹³C NMR** (126 MHz, CDCl₃) δ 180.6, 168.3, 140.1, 134.8, 134.1, 133.7, 133.0, 132.3, 131.7, 131.2, 130.0, 129.7, 128.9, 128.8, 127.6, 127.6, 126.6, 125.8, 125.5, 124.7, 124.2, 123.6, 123.3, 123.2, 110.5, 55.9, 36.4, 34.2; **FTIR** (cm⁻¹) 3245, 2926, 1770, 1708, 1617, 1399, 1369; **UPLC-MS** (ESI) *m/z* 483.62 (calculated for C₃₂H₂₂N₂O₃ [M + H]⁺ = 483.16); **TLC** R_f = 0.45 (10% EtOAc/DCM); **Chiral HPLC analysis**: 99:1 er, performed on Chiralcel OD-H (Daicel, 4.6 mm × 250 mm, 5 μm, 20 °C), eluting at 0.7 mL/min with 55:45 hexanes/ethanol. Retention times: *t*_{major} = 13.8 min, *t*_{minor} = 35.8 min; [α]_{546 nm}^{20 °C} = -1.1 (*c* = 0.82, CHCl₃).

J. NMR Spectra.

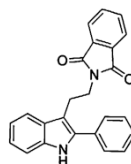
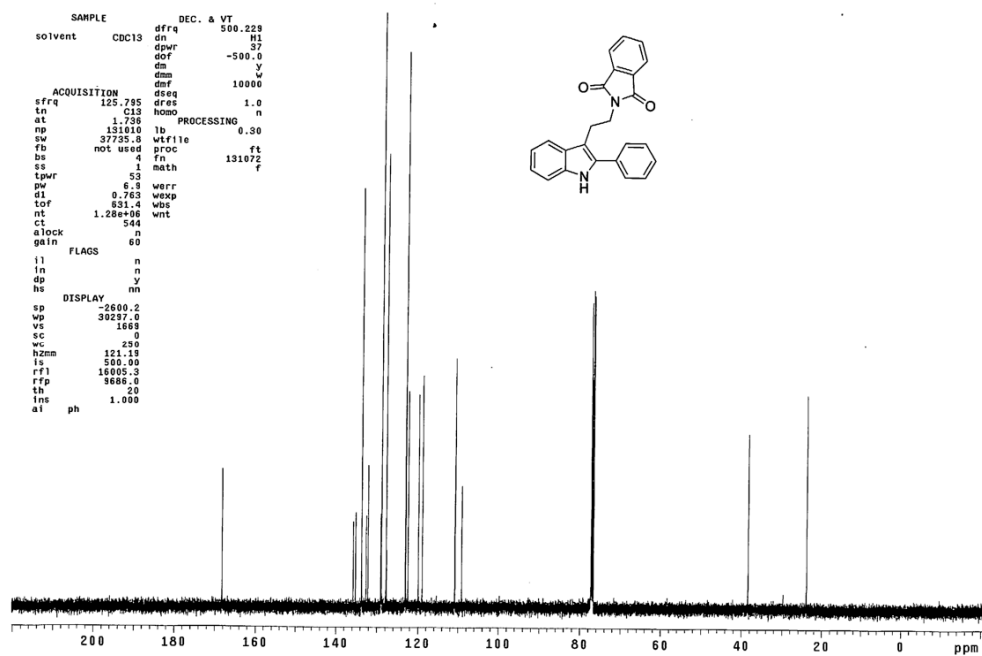
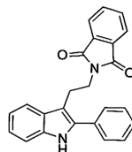
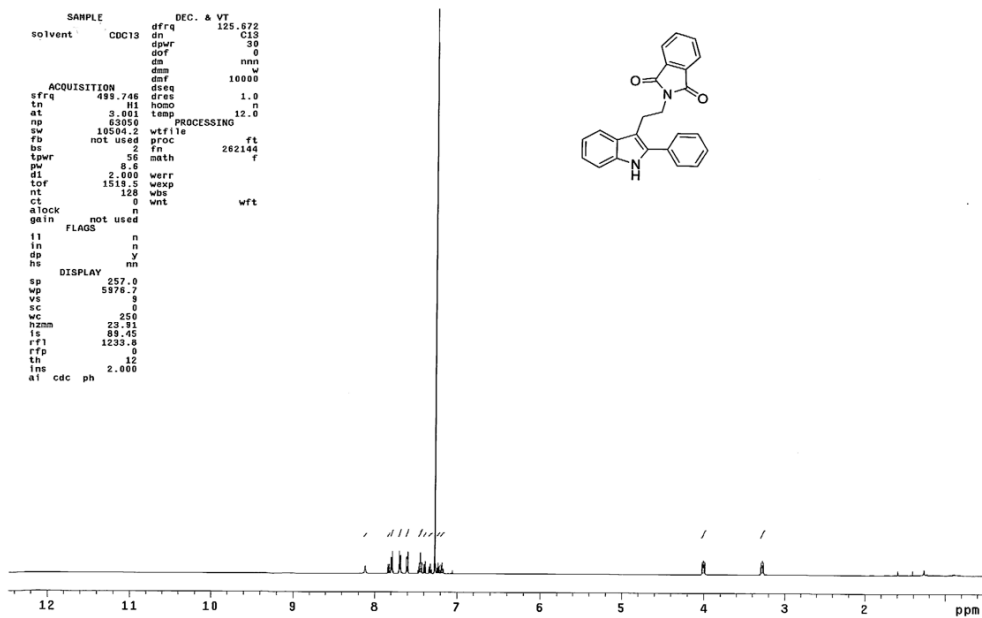
As shown below, some of the oxidation products exhibit conformational equilibria, resulting in doubling of signals at room temperature. As shown for compound **39c**, solvent exchange in the NMR experiments reveals reversible coalescence of signals with the introduction of d_6 -DMSO.



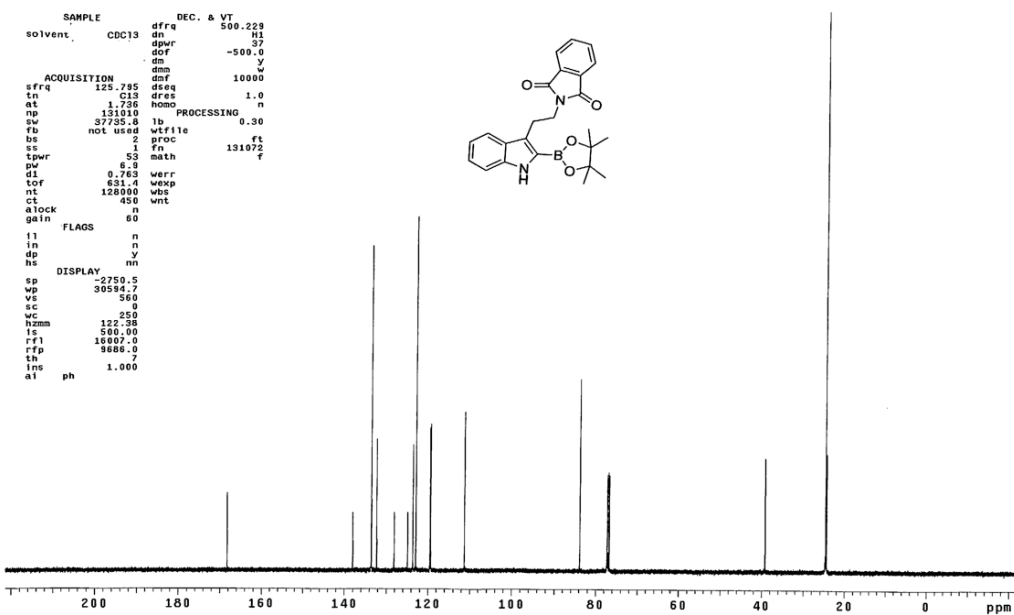
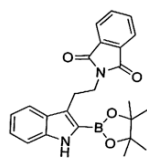
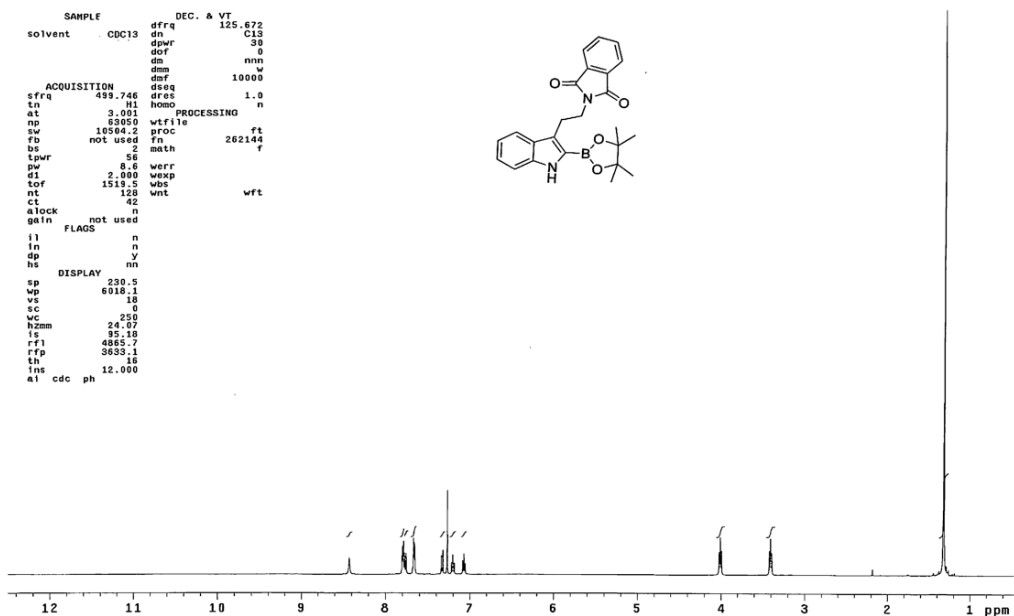
N,N-Phthaloyl-2-iodotryptamine (S1):



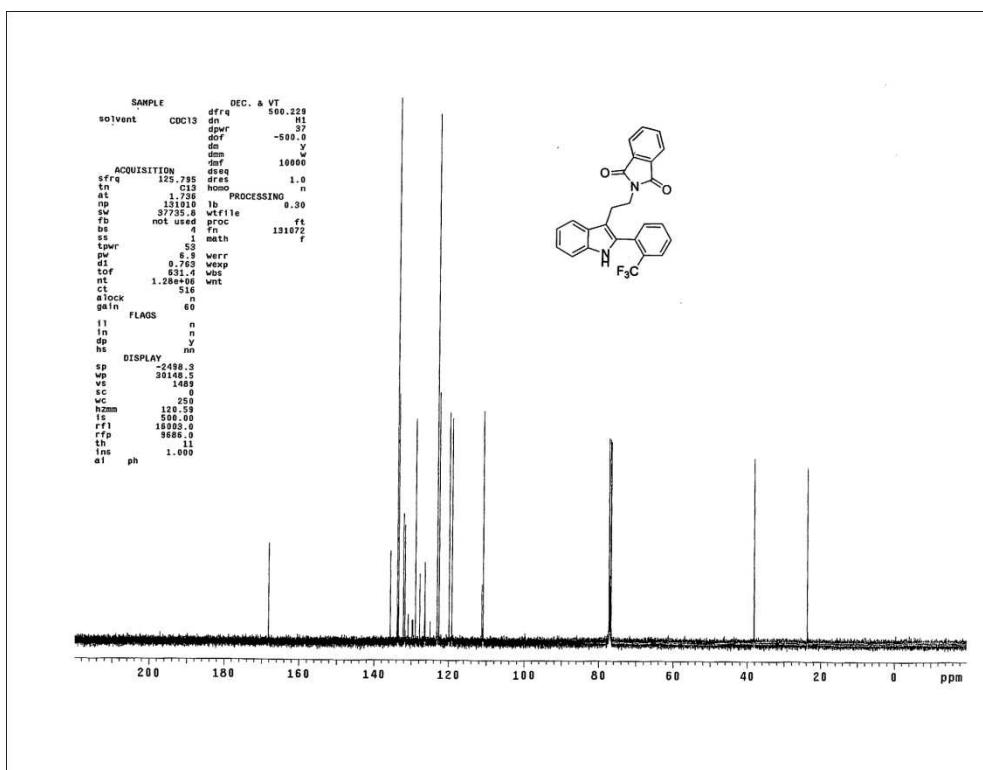
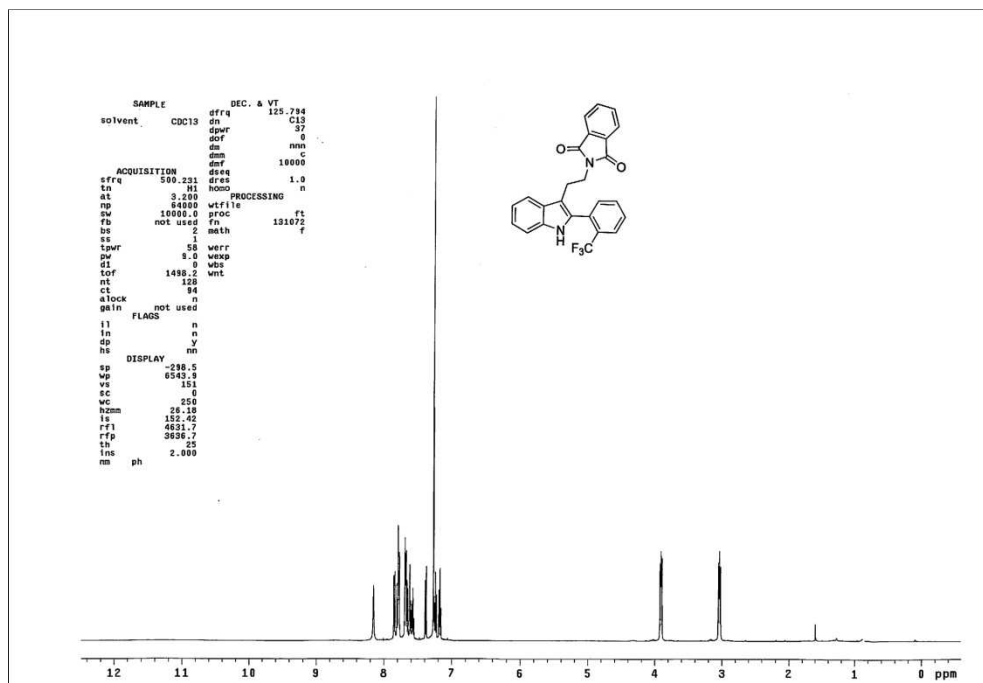
N,N-Phthaloyl-2-phenyltryptamine (14):



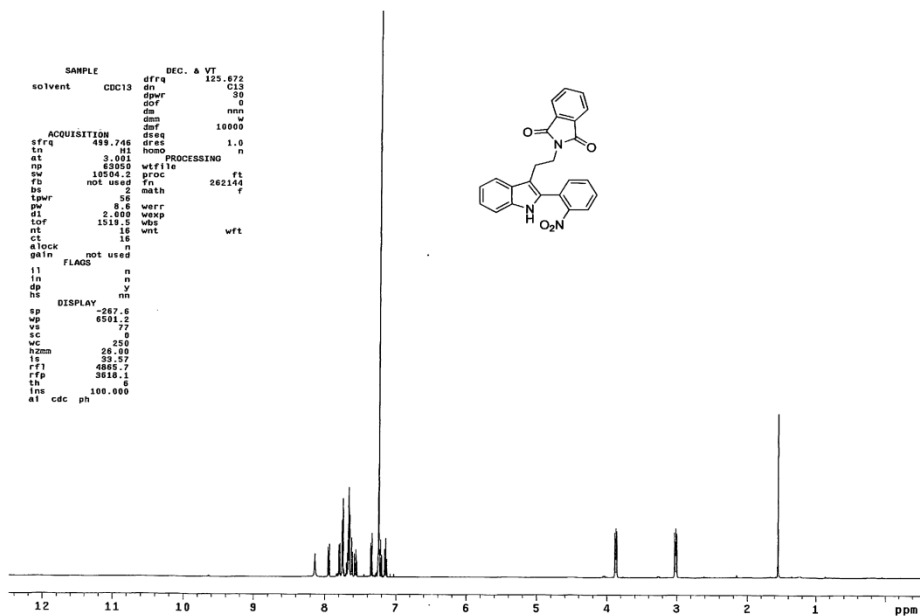
***N,N*-Phthaloyl-2-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)tryptamine (S2):**



2-(2-(2-(2-(Trifluoromethyl)phenyl)-1H-indol-3-yl)ethyl)isoindoline-1,3-dione (Table 1, Entry 36a):

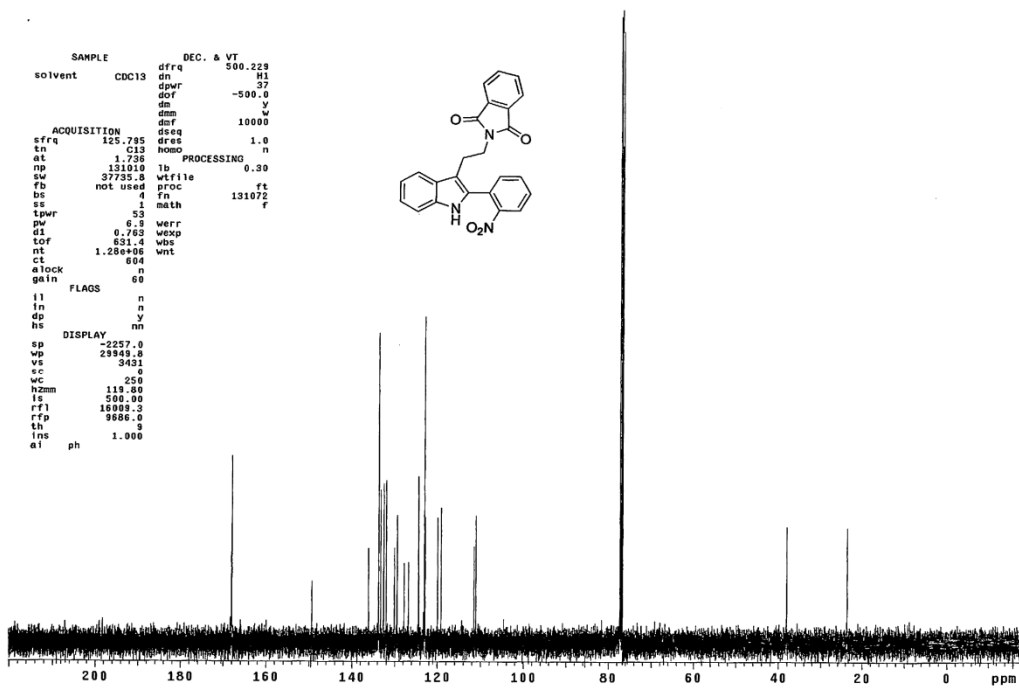
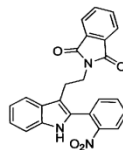


2-(2-(2-(2-Nitrophenyl)-1H-indol-3-yl)ethyl)isoindoline-1,3-dione :(Table 1, Entry 36b):



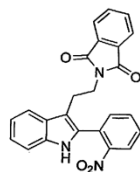
```

SAMPLE          DEC. & VT
solvent         CDC13  dffrq 125.872
                dn      C13   30
                dpwr   0
                ds     nnn
                dsm    w
                dsf    10000
ACQUISITION
sfrq           499.746  dres  1.0
tn             H1
at             3.001   homo
ns             63050
np             10504.2  proc  ft
sv             not used  fn   262144
bs             2       math  f
ss             58
tpwr           8.6    werr
d1             2.000  wexp
tof           1519.5  wbt
nt             16    wnt
ct             16
elock          n
gain           not used
FLAGS
il             n
in             n
dp             y
hs             nn
DISPLAY
sp            -287.6
vp            6501.2
vs             77
sc             0
wc            250
hzmm          26.00
ls            35.27
rf1           4885.7
rfp           2619.1
th            0
ins           100.000
at cdc ph
    
```

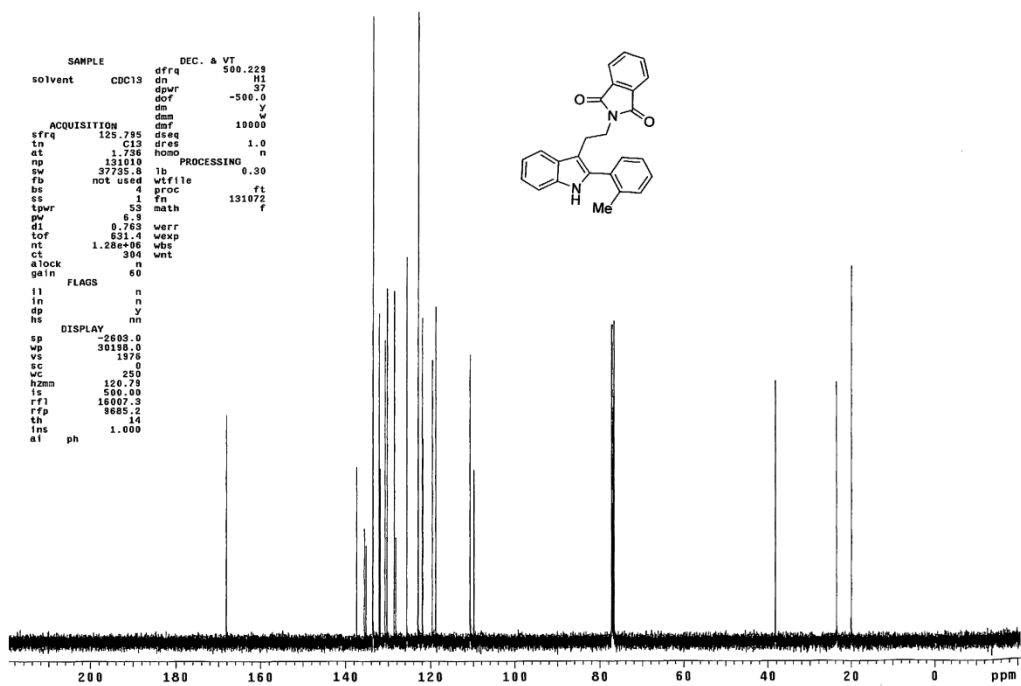
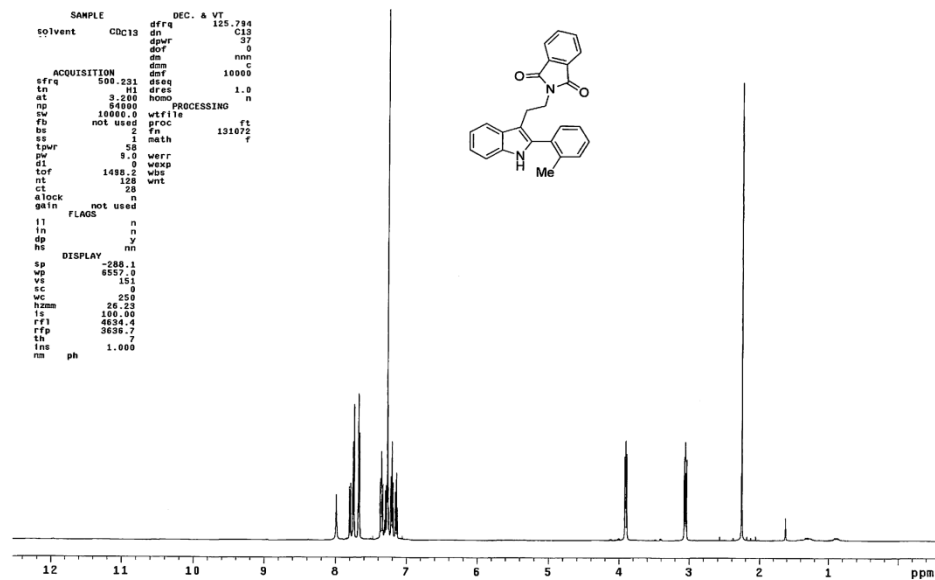


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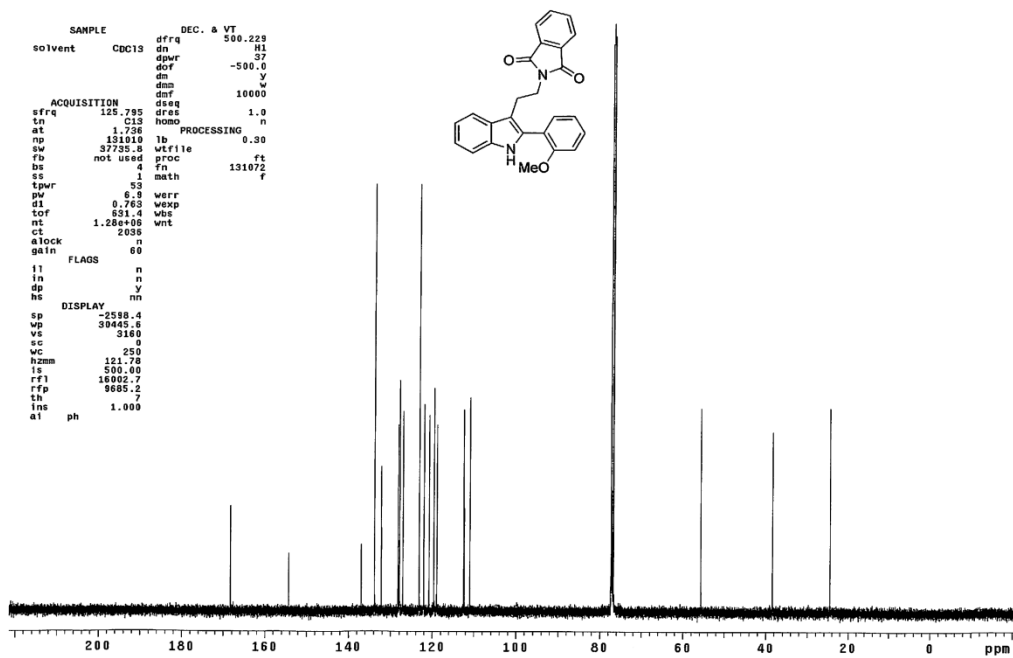
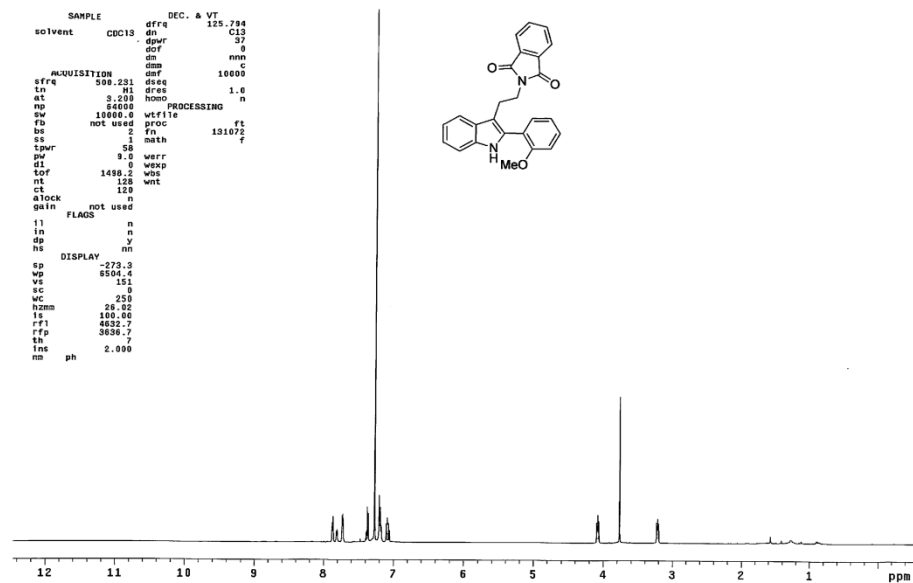
SAMPLE          DEC. & VT
solvent         CDC13  dffrq 500.229
                dn      H1
                dpwr   37
                ds     -500.0
                dsm    y
                dsf    10000
ACQUISITION
sfrq           125.795  dres  1.0
tn             C13
at             1.736   homo
ns             131010  lb   0.30
sv             37735.8  wfile
fb             not used  proc  ft
bs             4       fn   131072
ss             1       math  f
tpwr           53
d1             0.8    werr
d2             0.763  wexp
tof           831.4   wbt
nt             1.28e+06 wnt
ct             604
elock          n
gain           60
FLAGS
il             n
in             n
dp             y
hs             nn
DISPLAY
sp            -2257.0
vp            29949.8
vs             3431
sc             0
wc            250
hzmm          119.80
ls            500.00
rf1           16008.3
rfp           9886.0
th            0
ins           1.000
at ph
    
```



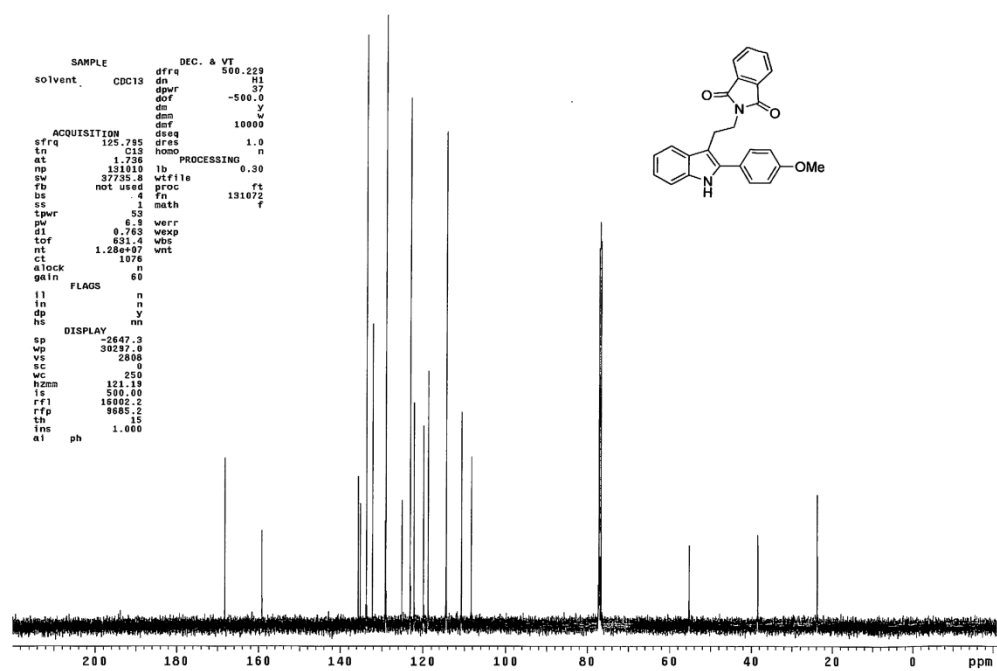
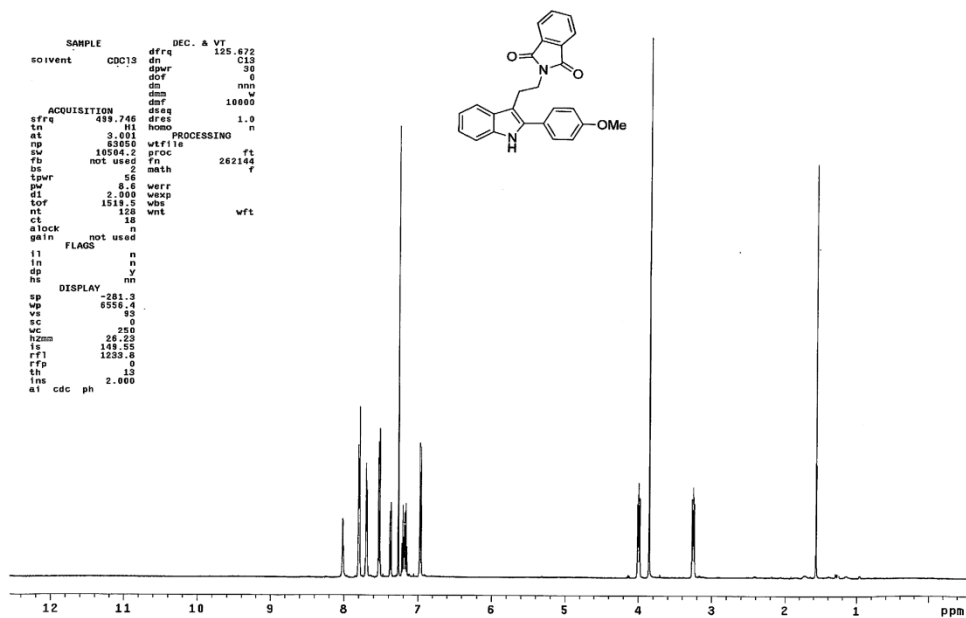
2-(2-(2-(*o*-Tolyl)-1*H*-indol-3-yl)ethyl)isoindoline-1,3-dione: (Table 1, Entry 36c):



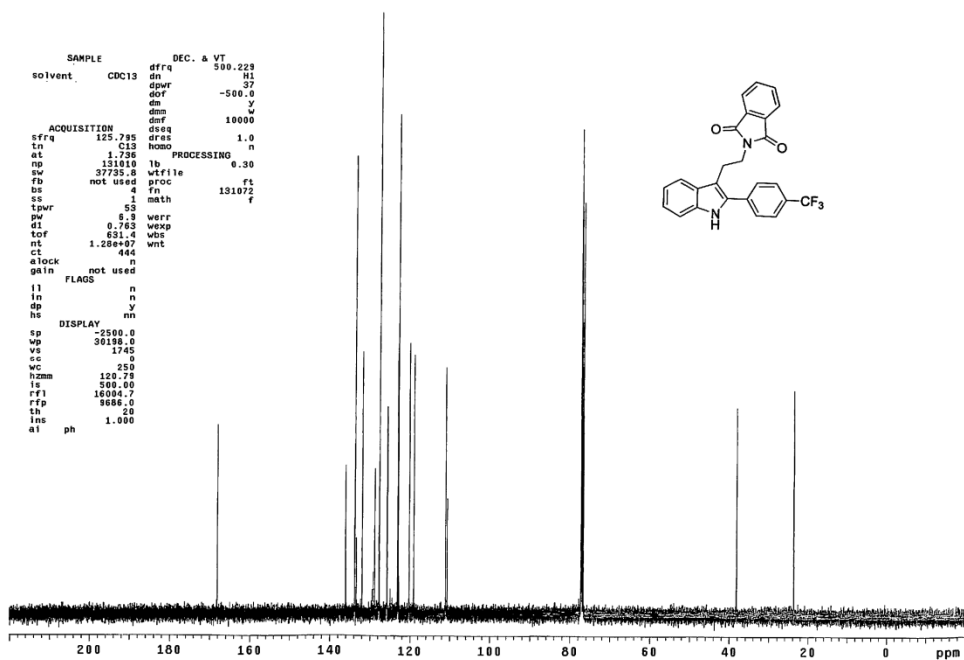
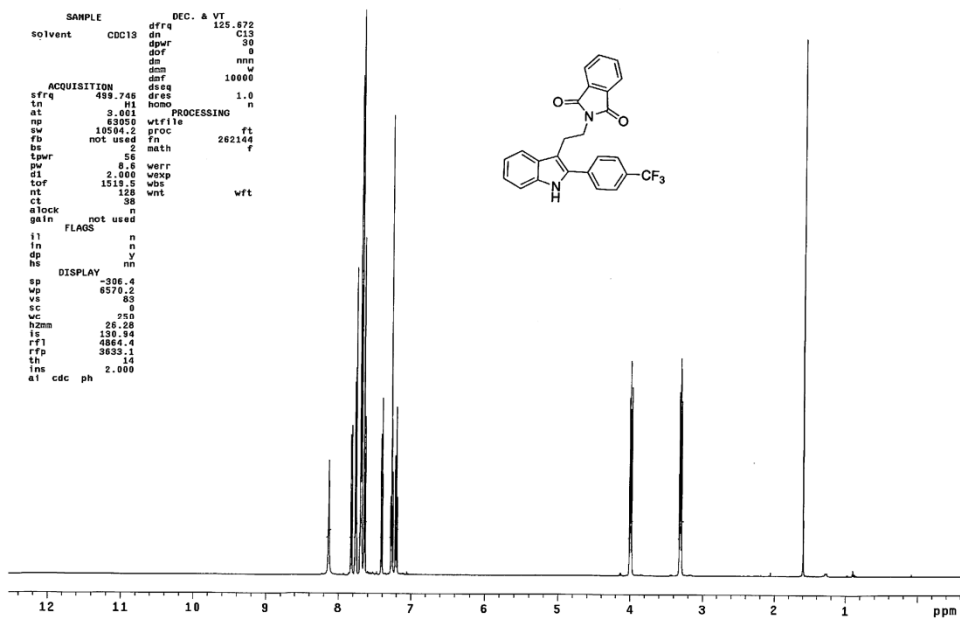
2-(2-(2-(2-Methoxyphenyl)-1H-indol-3-yl)ethyl)isoindoline-1,3-dione : (Table 1, Entry 36d):



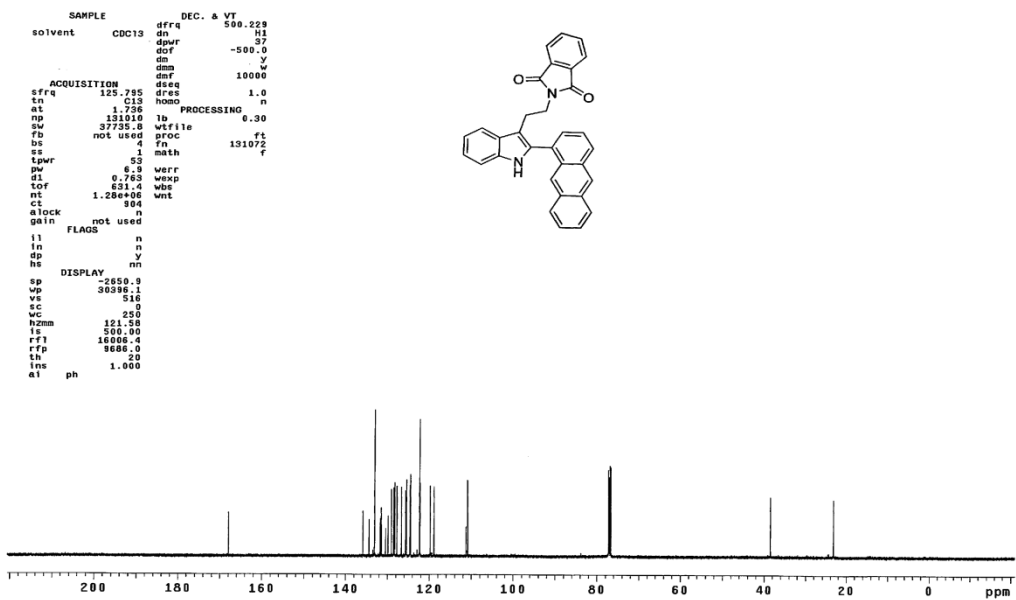
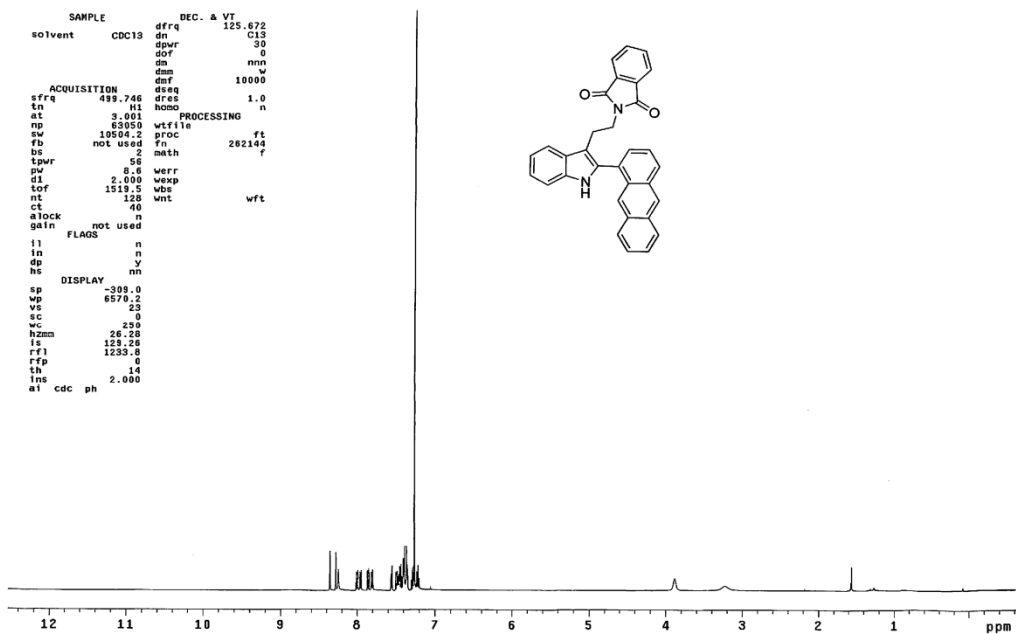
2-(2-(2-(4-Methoxyphenyl)-1H-indol-3-yl)ethyl)isoindoline-1,3-dione: (Table 1, Entry 36e):



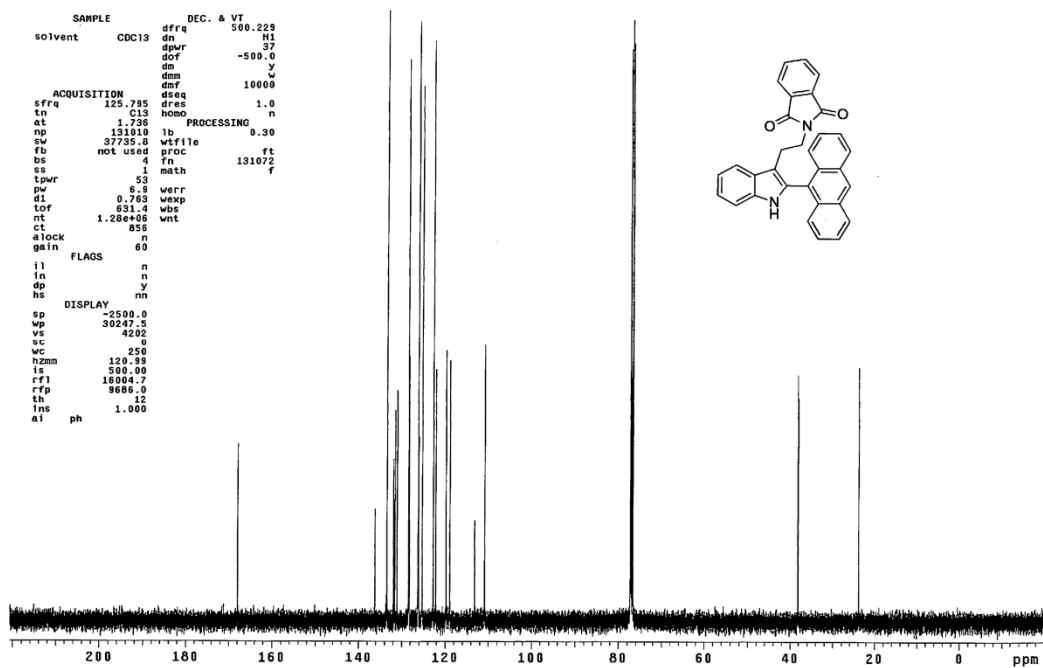
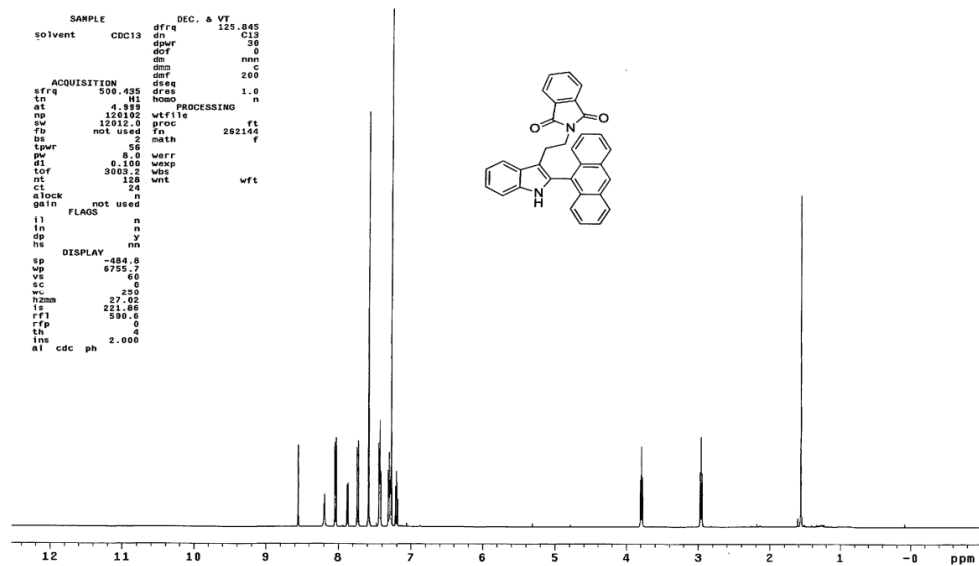
2-(2-(2-(4-(Trifluoromethyl)phenyl)-1H-indol-3-yl)ethyl)isoindoline-1,3-dione:
(Table 1, Entry 36f):



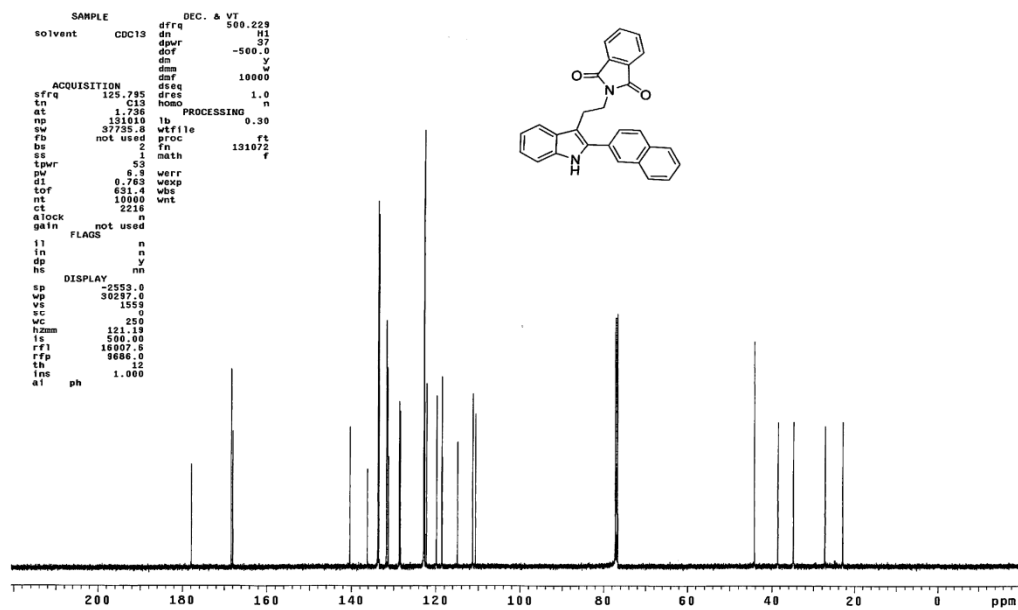
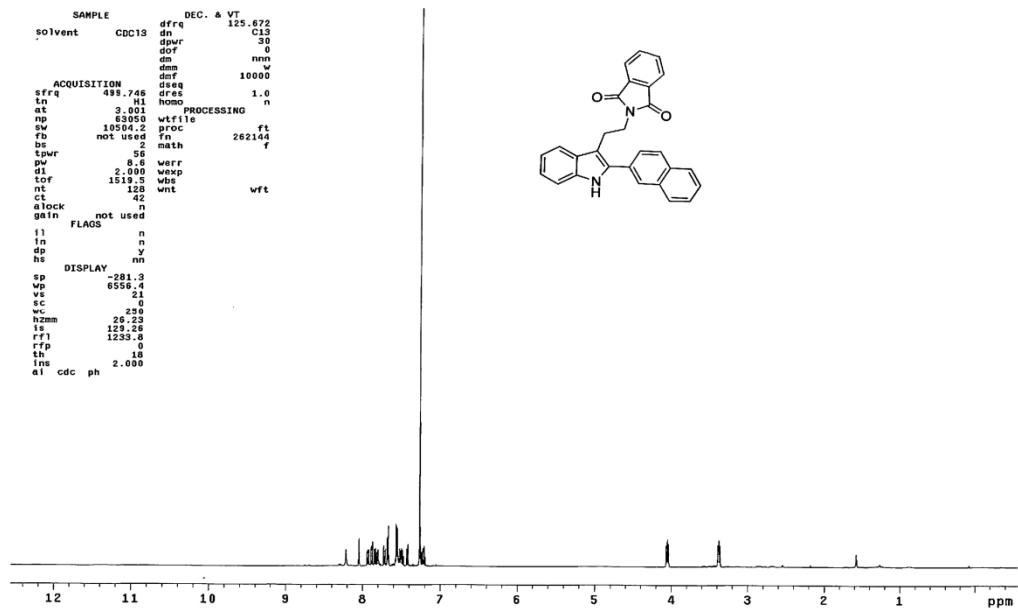
2-(2-(2-(Anthracen-1-yl)-1H-indol-3-yl)ethyl)isoindoline-1,3-dione: (Table 2, Entry 38a):



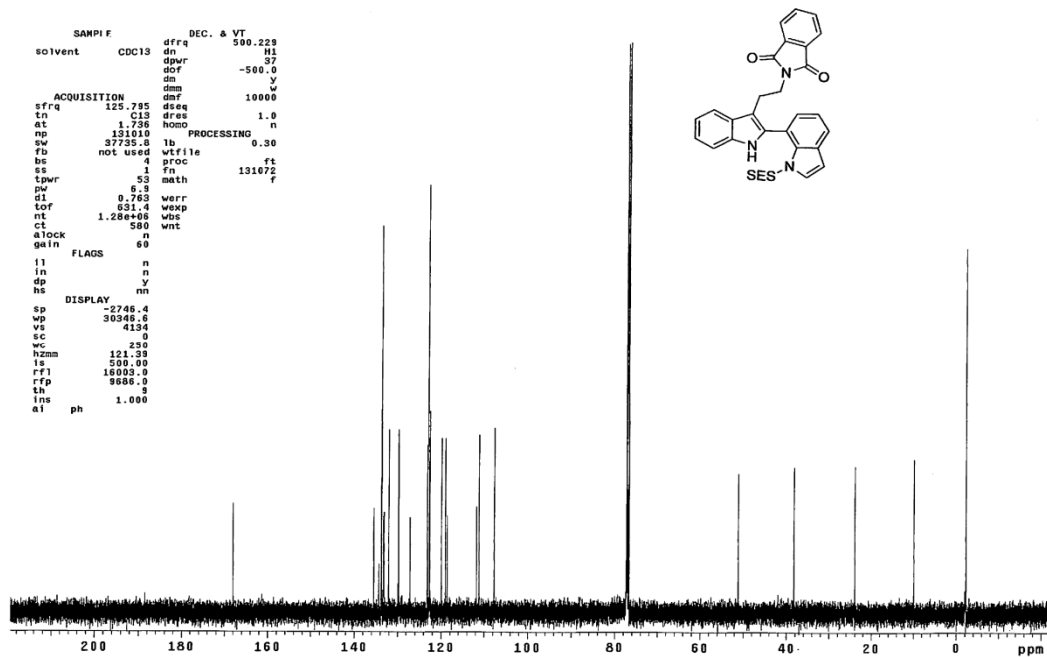
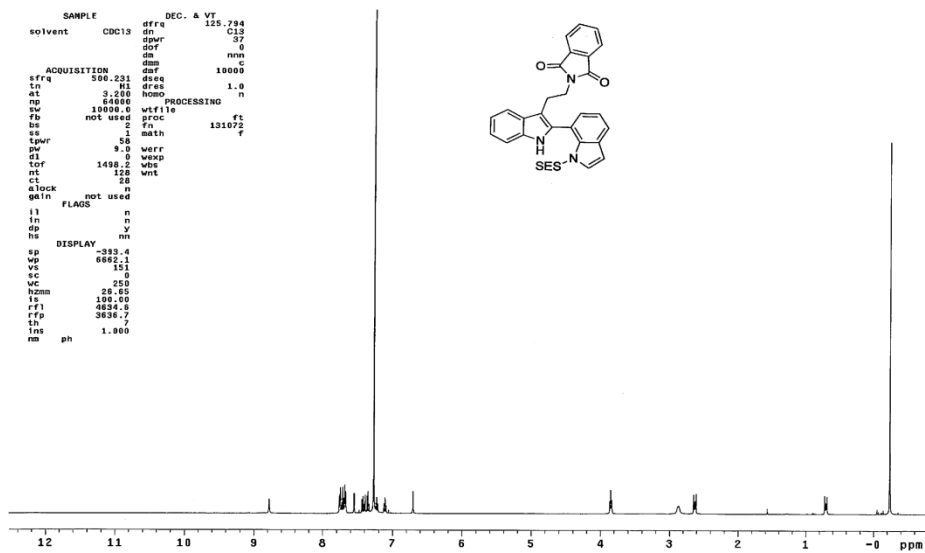
2-(2-(2-(Anthracen-9-yl)-1H-indol-3-yl)ethyl)isoindoline-1,3-dione: (Table 2, Entry 38b):

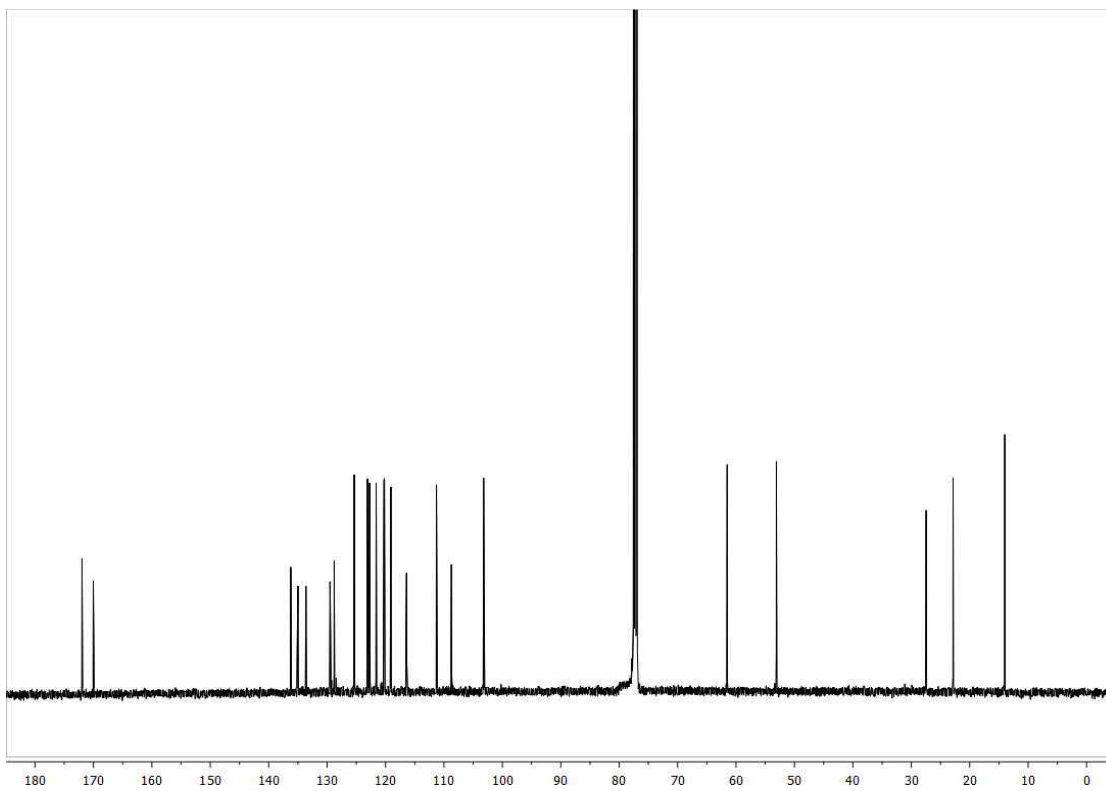
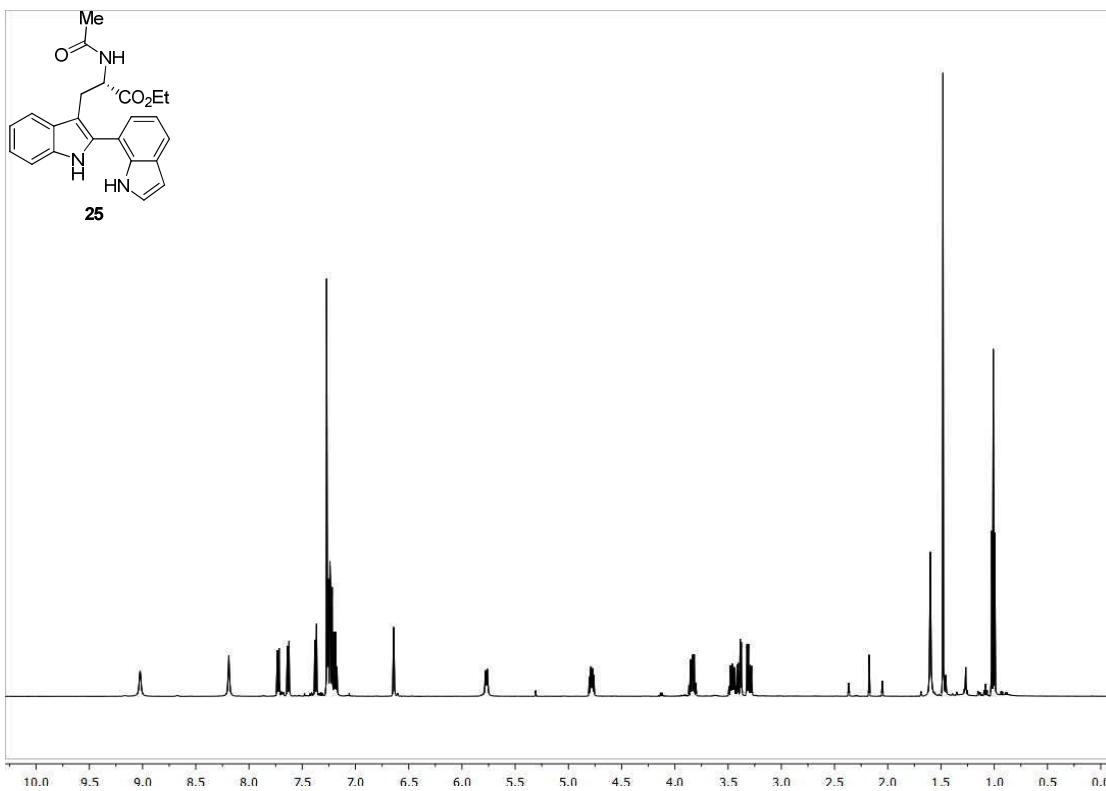


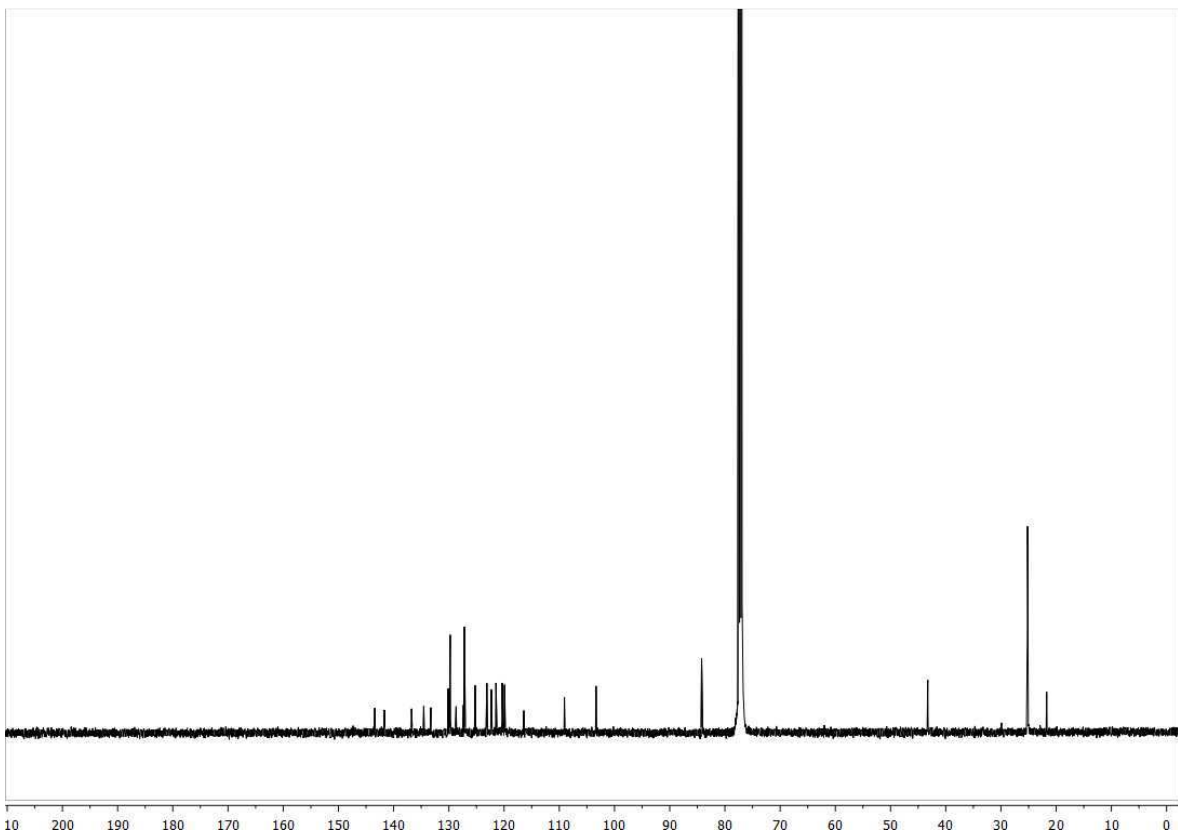
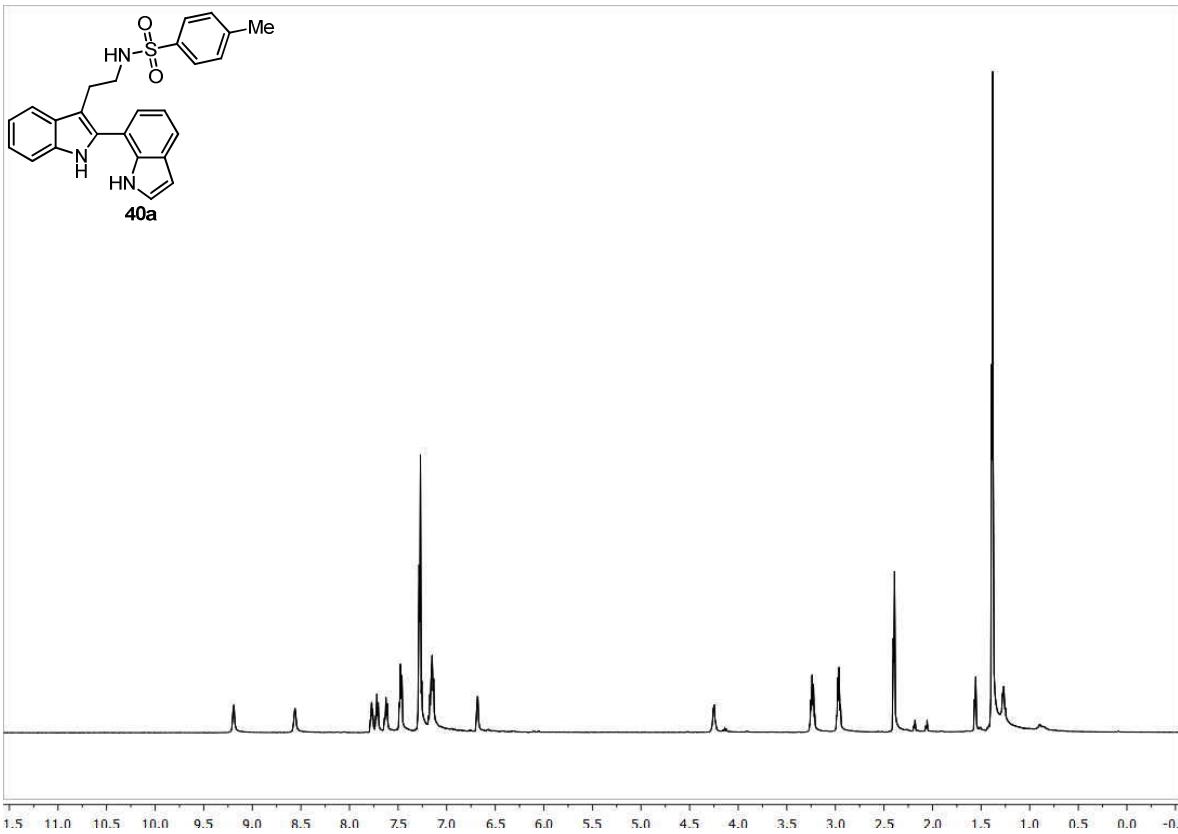
2-(2-(2-(Naphthalen-2-yl)-1H-indol-3-yl)ethyl)isoindoline-1,3-dione: (Table 2, Entry 38c):

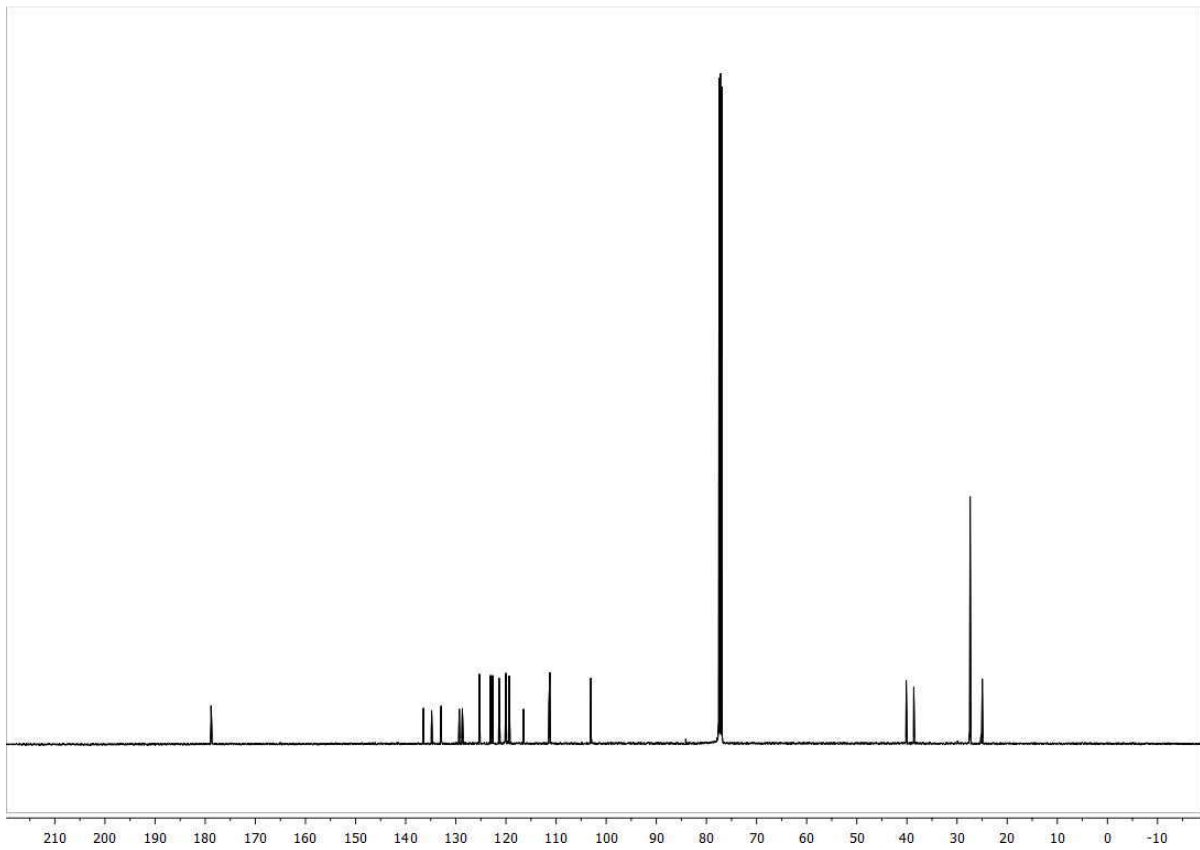
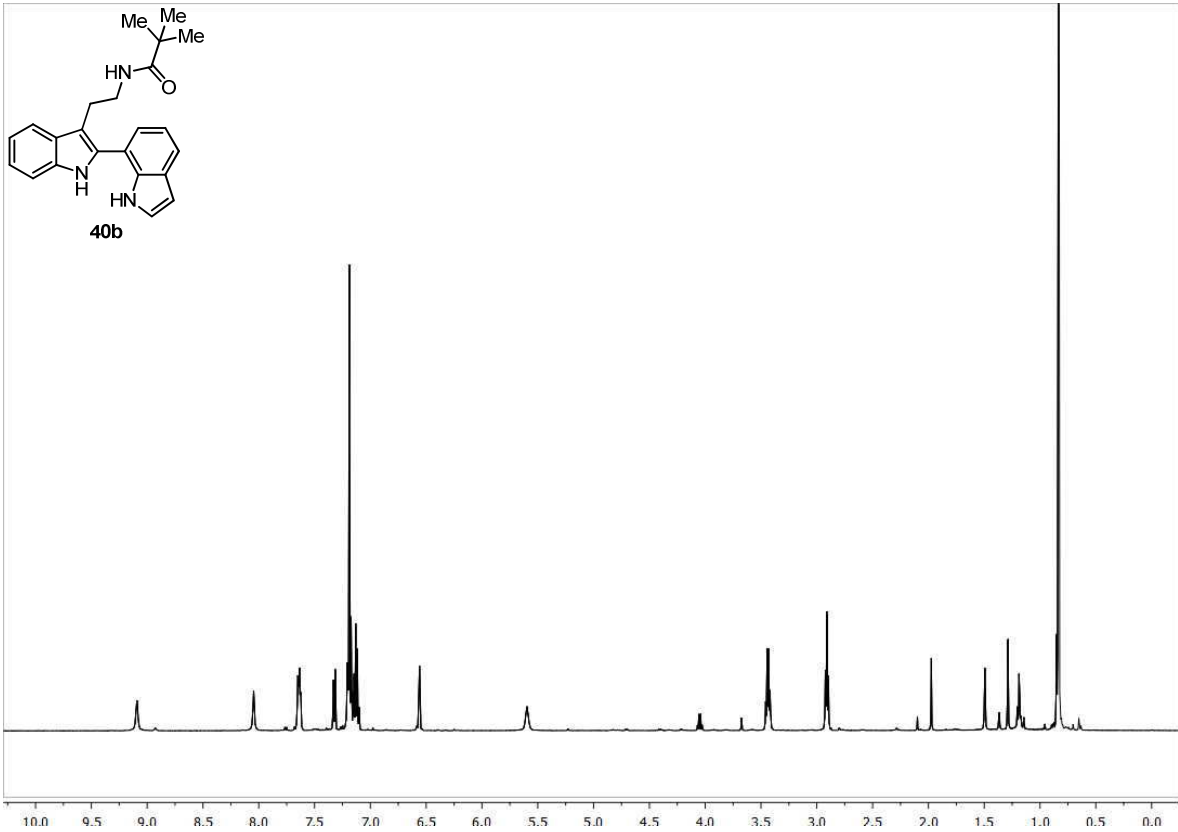


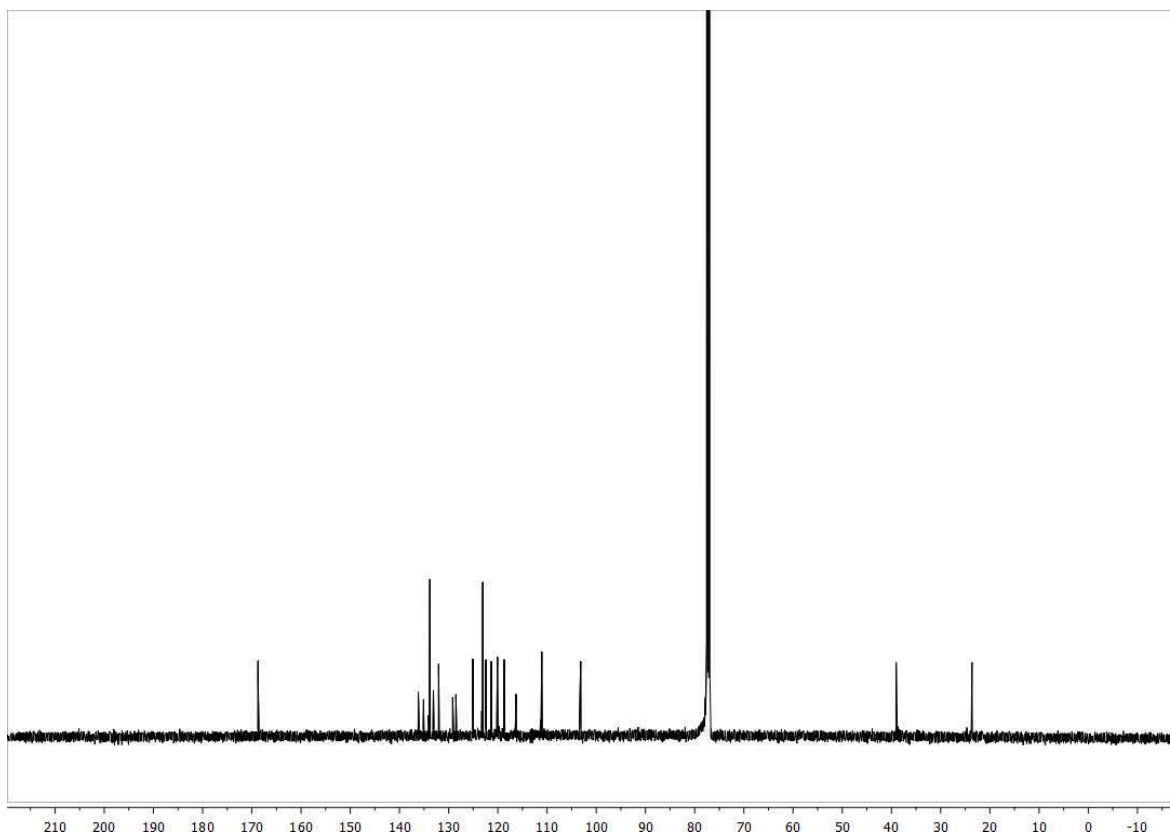
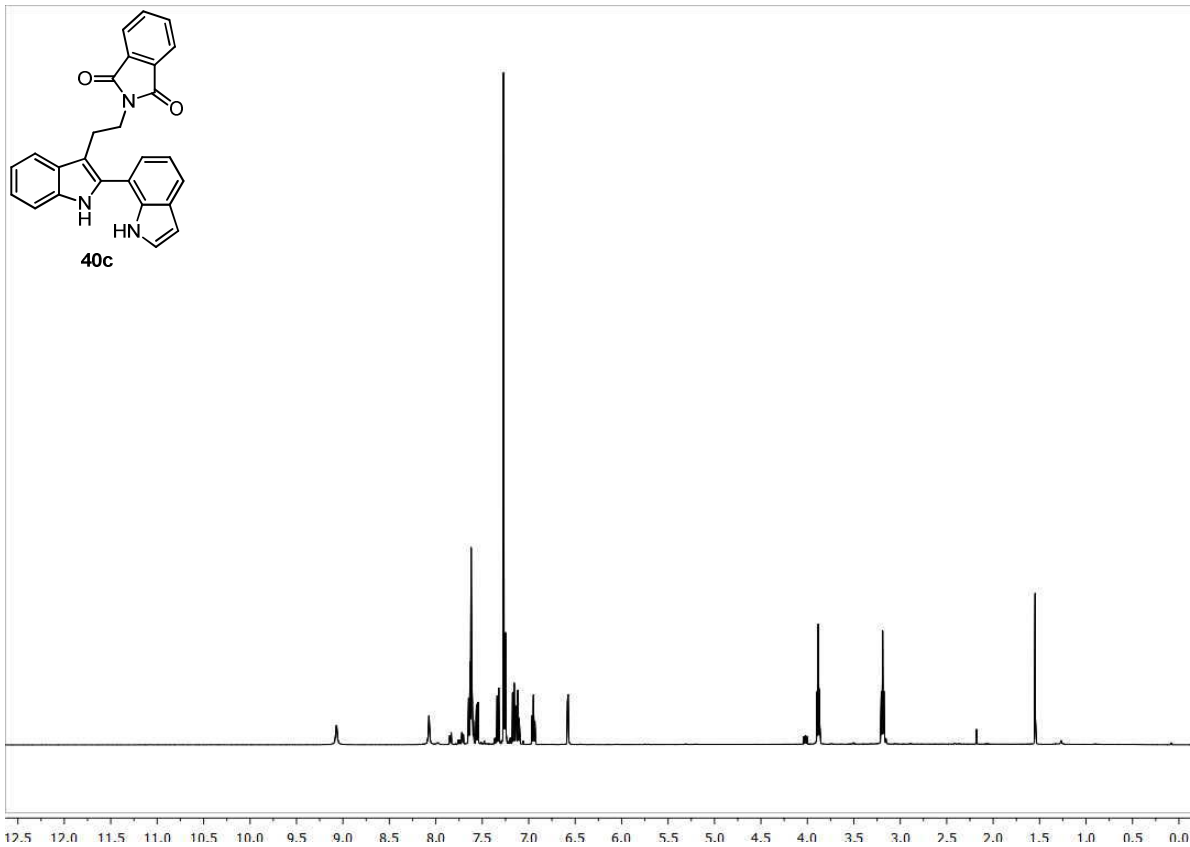
2-(2-(1'-((2-(Trimethylsilyl)ethyl)sulfonyl)-1*H*,1'*H*-[2,7'-bisindol]-3-yl)ethyl)isoindoline-1,3-dione: (Table 3, Entry 42):

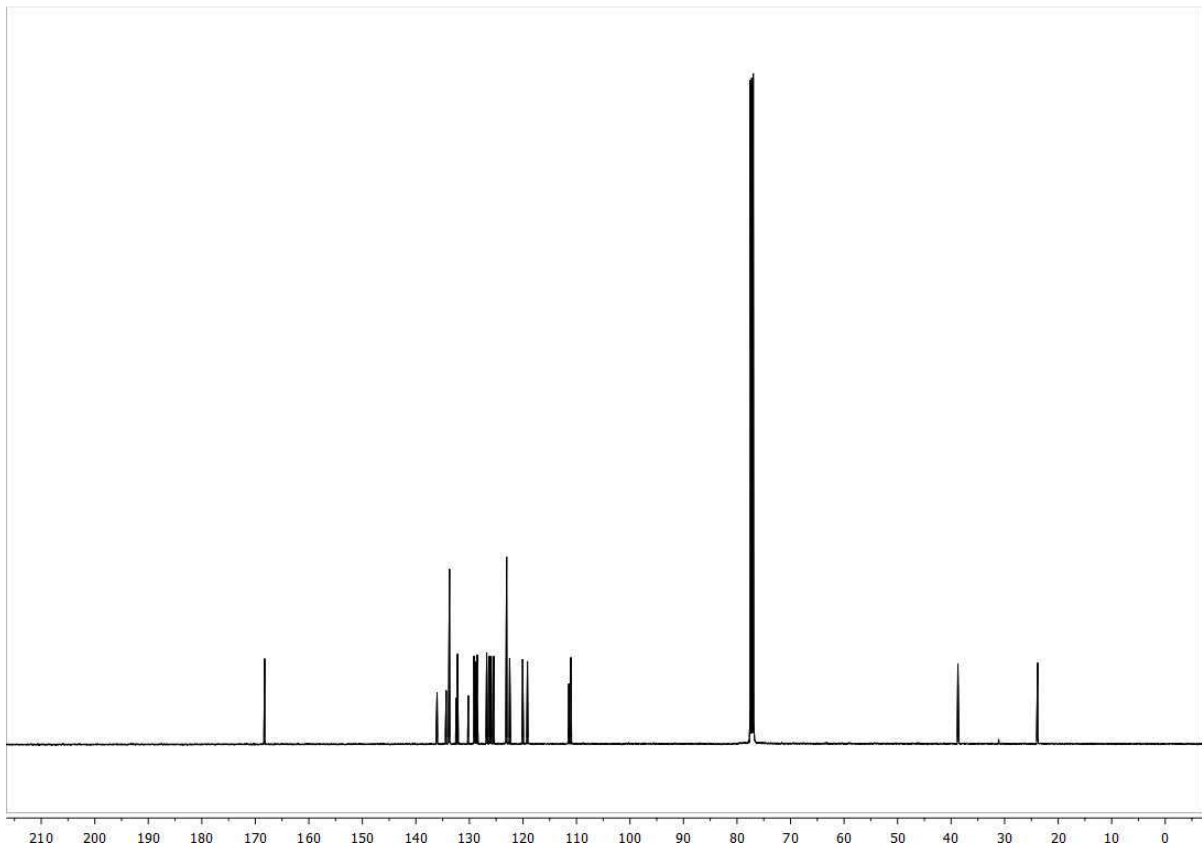
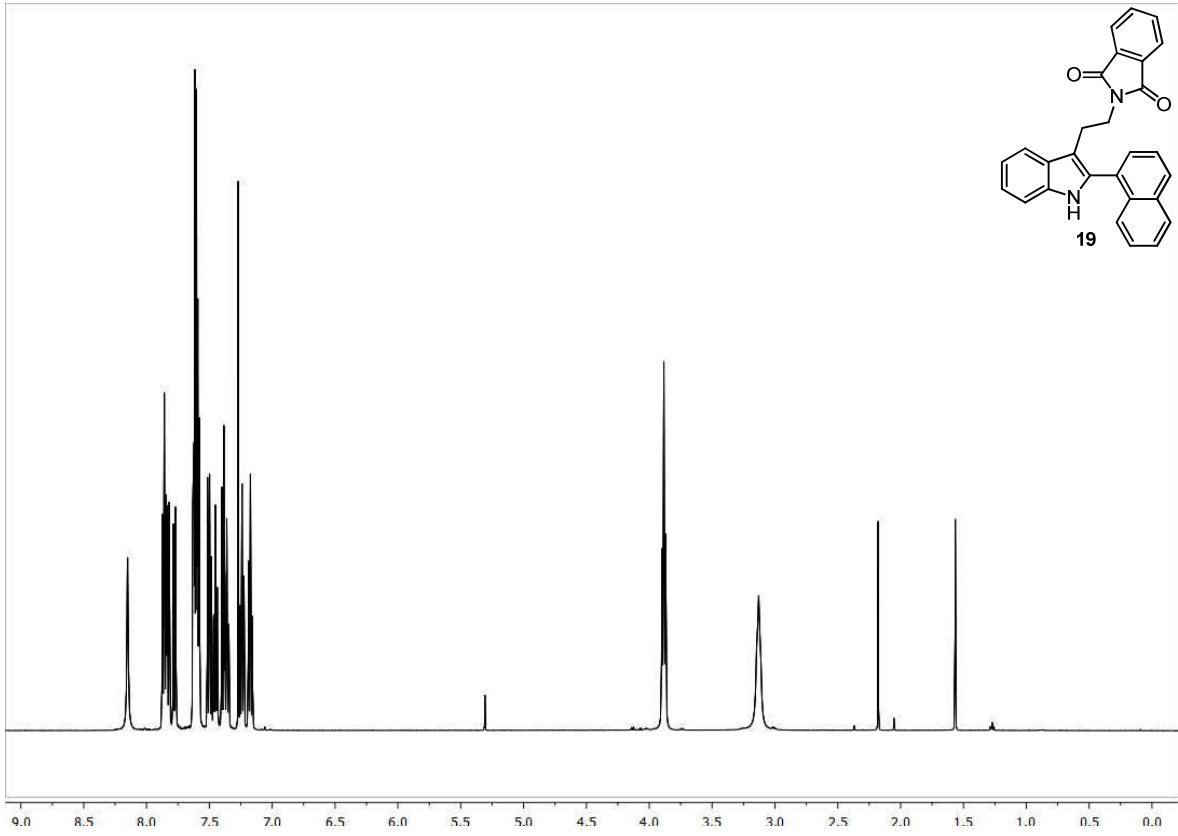


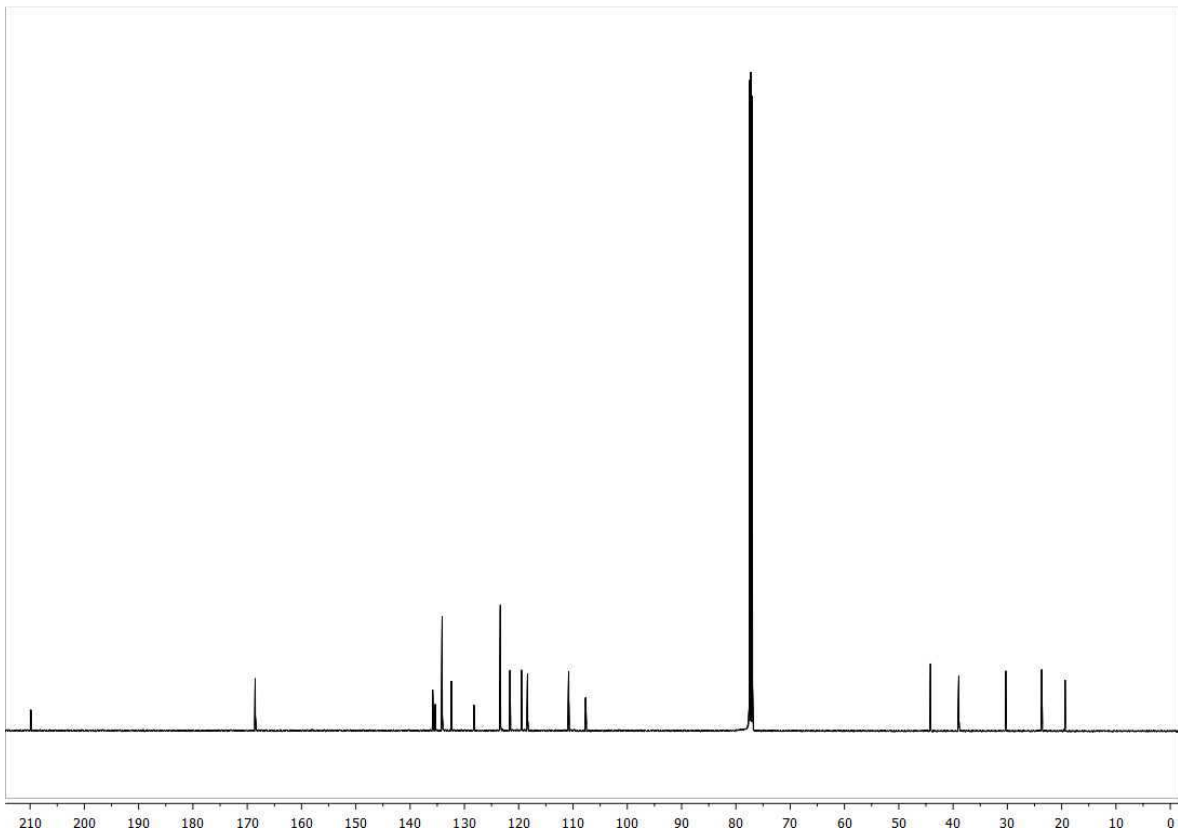
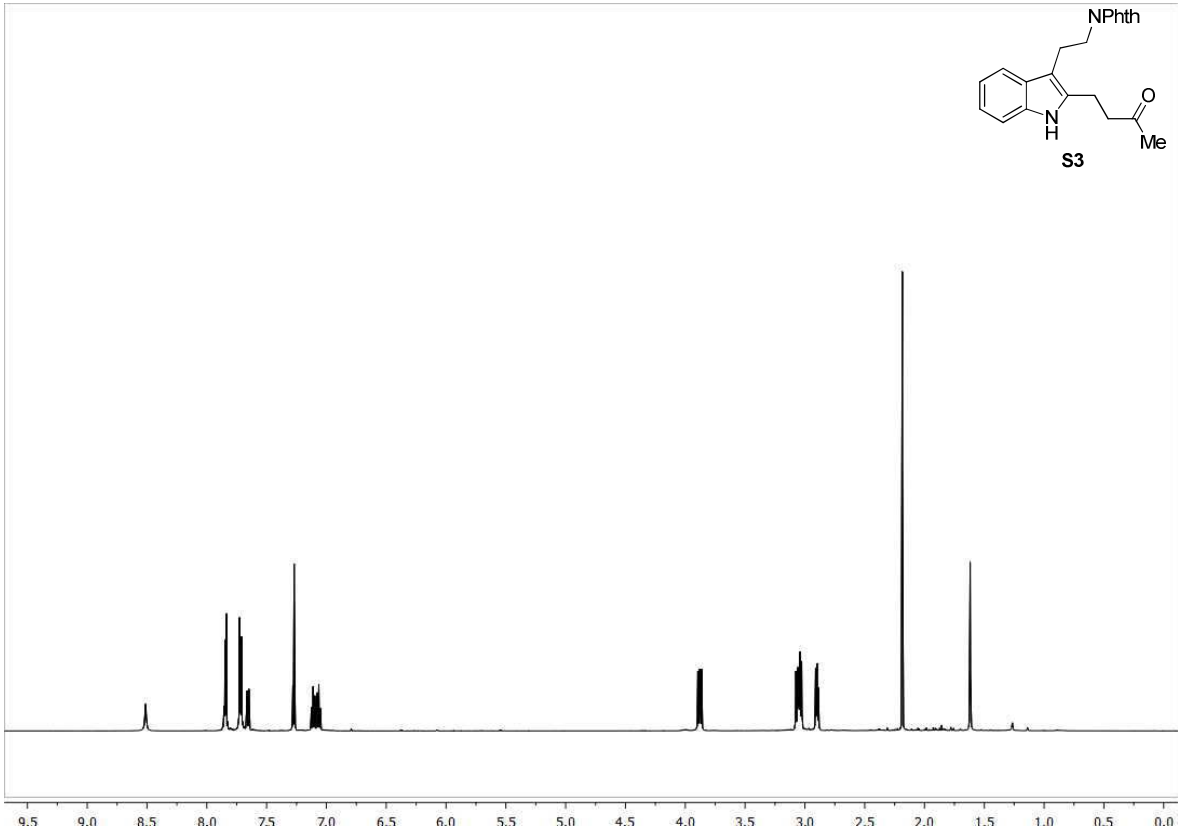


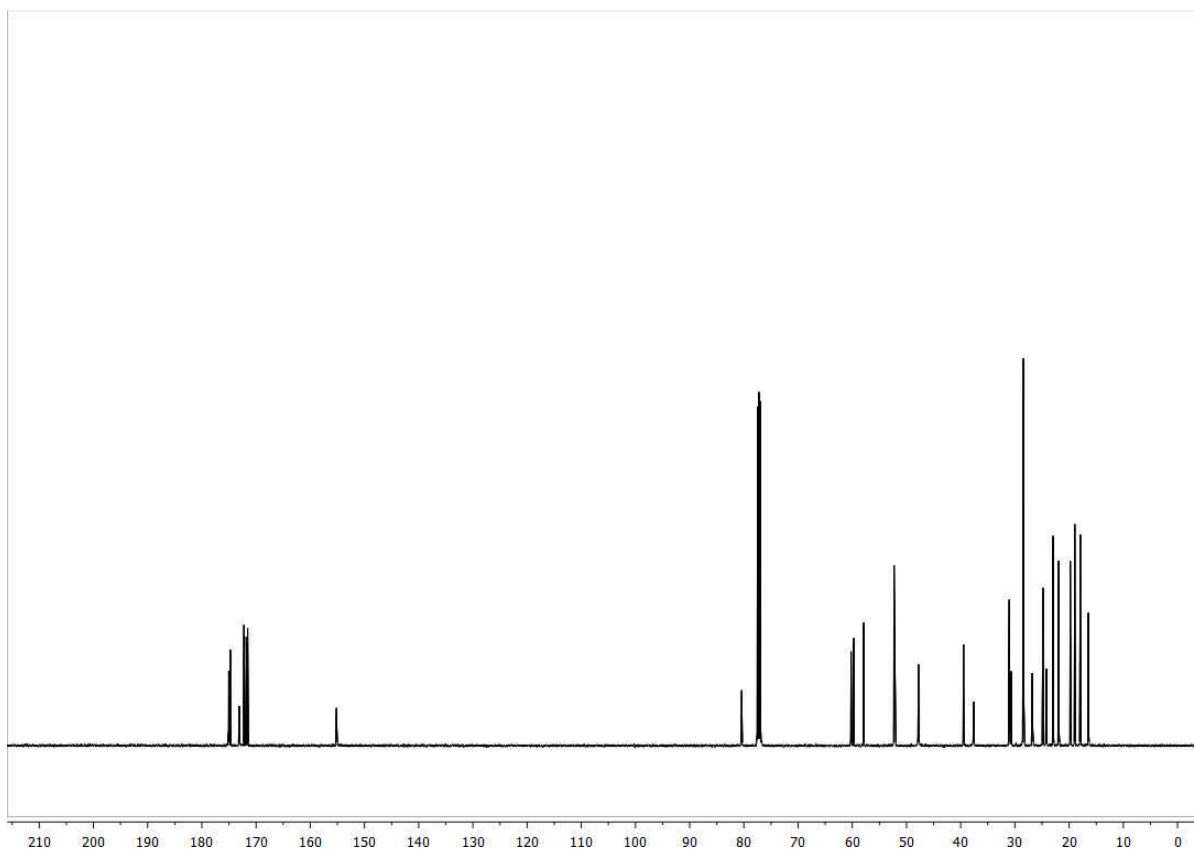
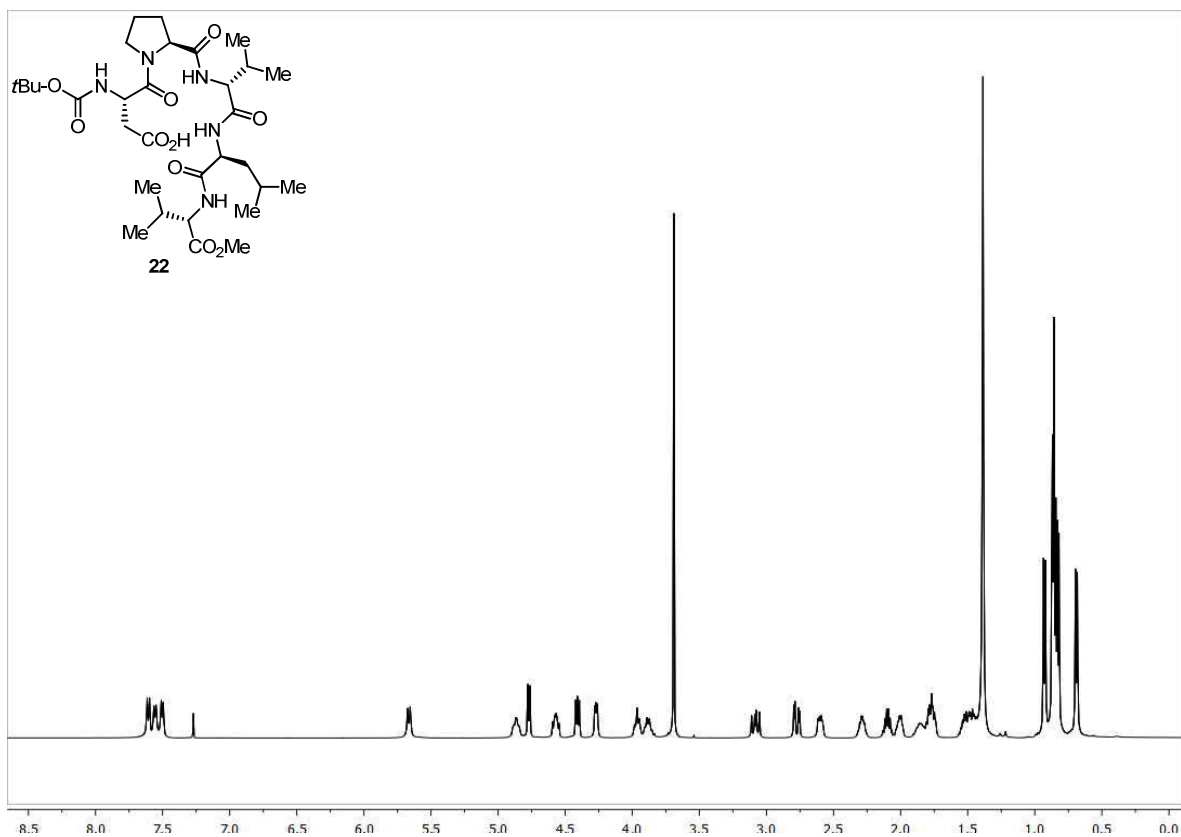


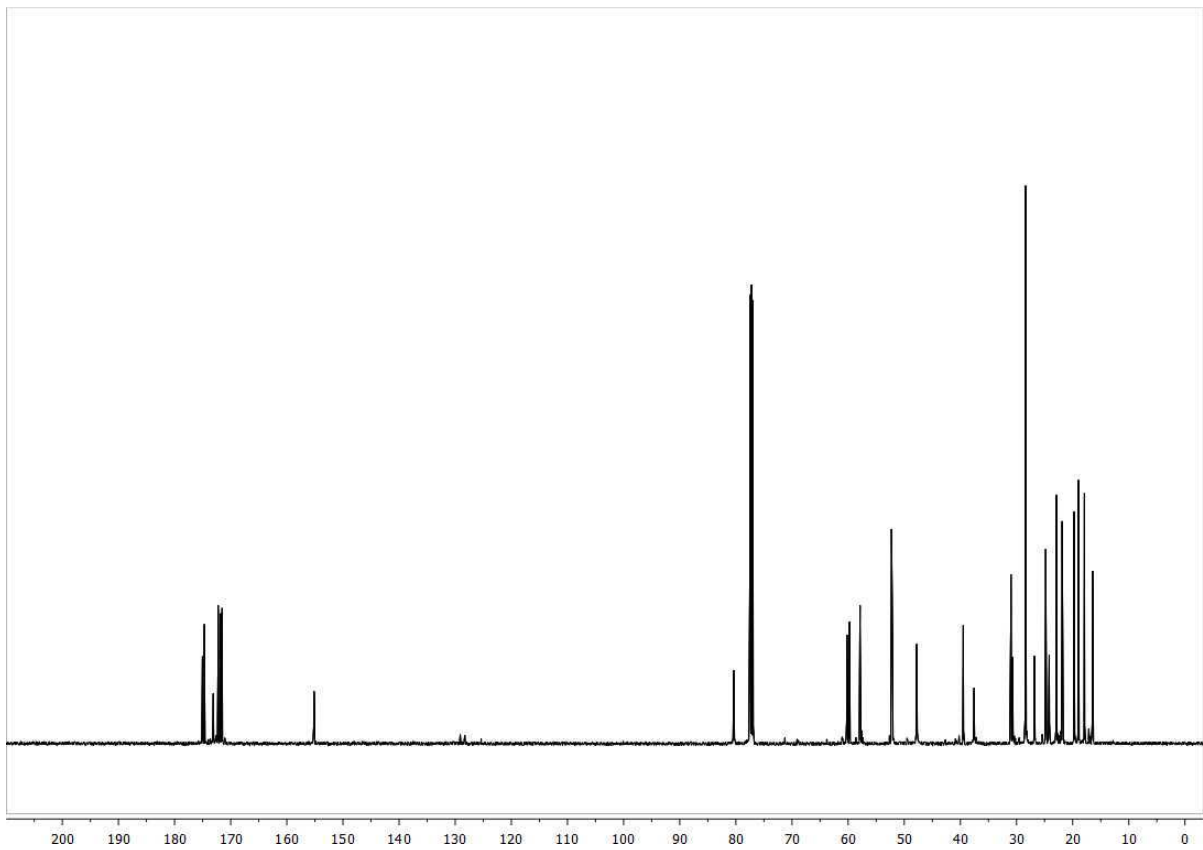
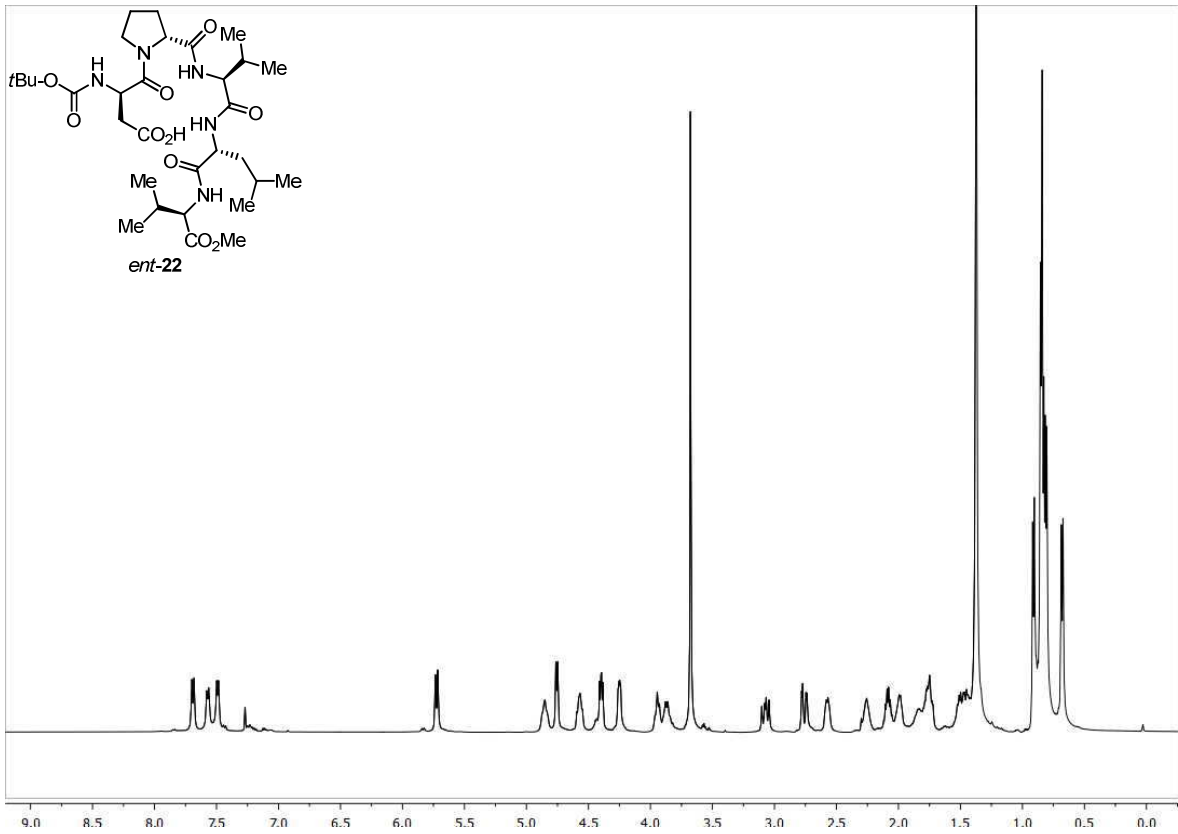


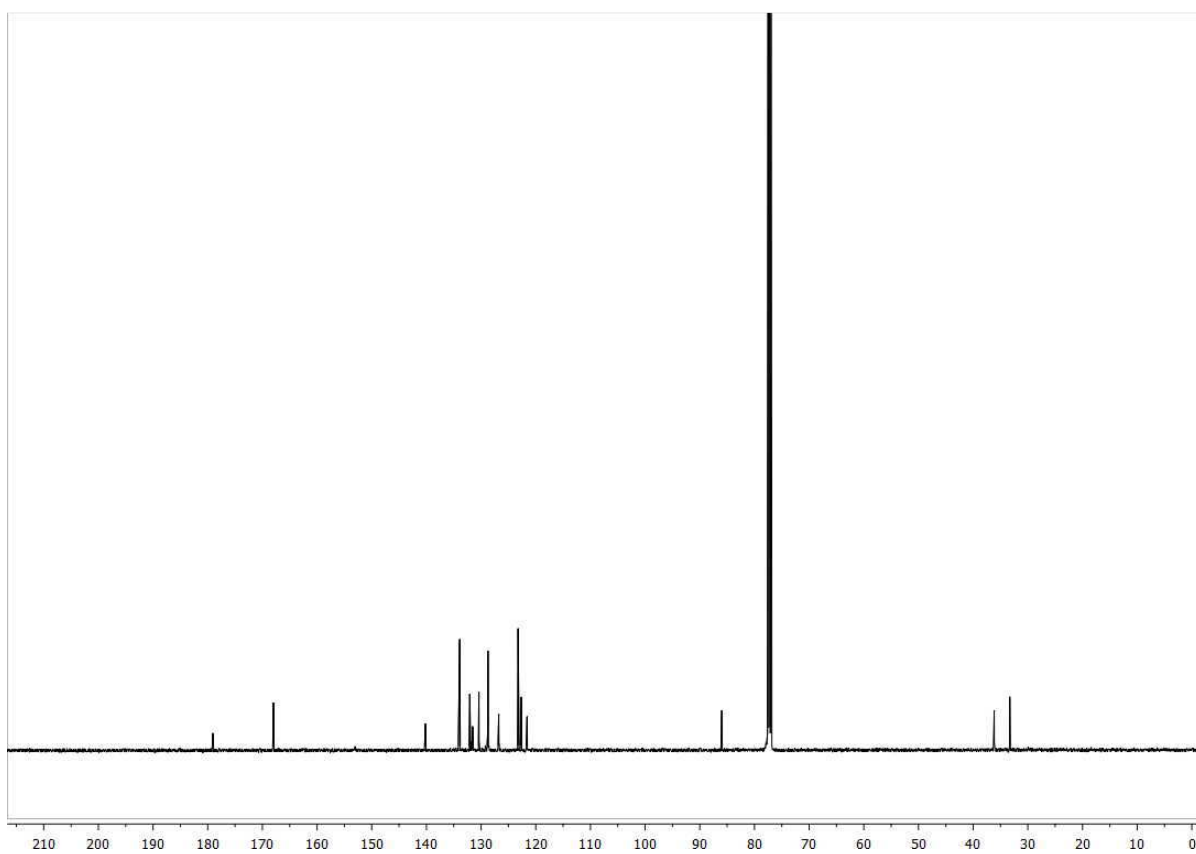
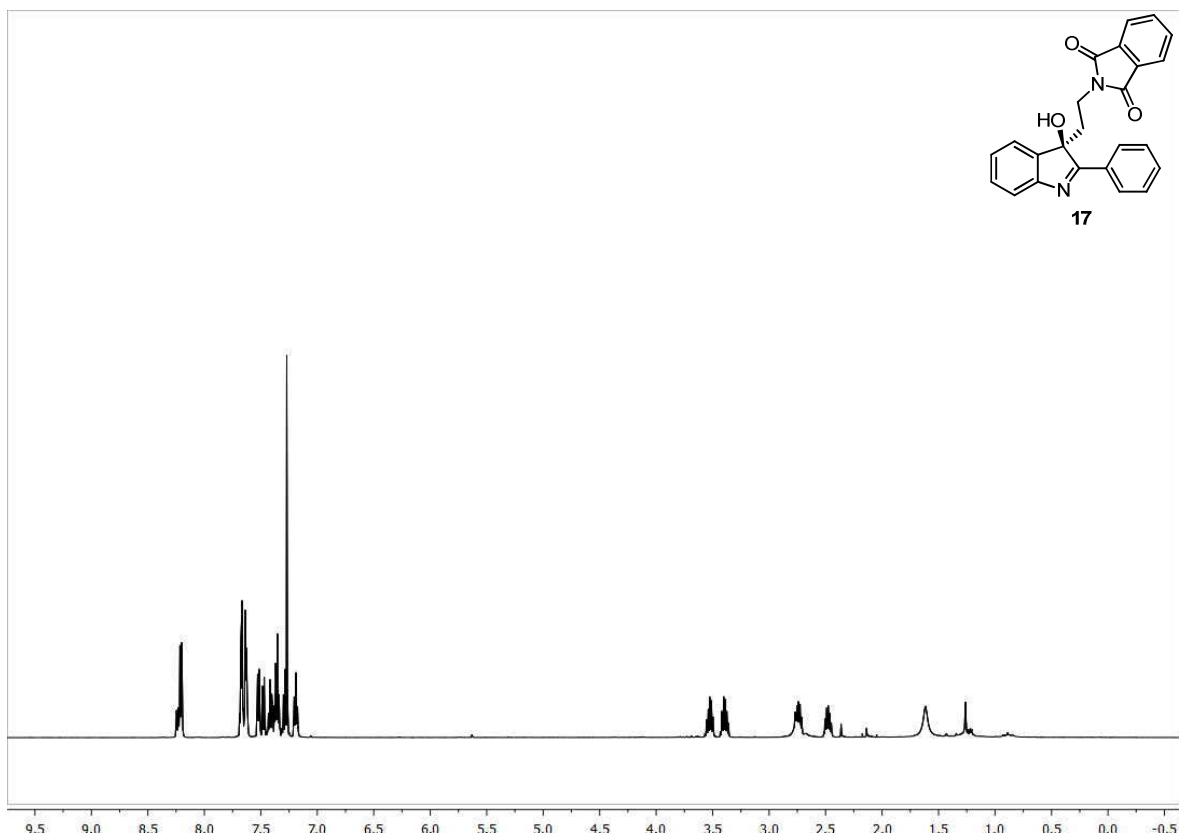
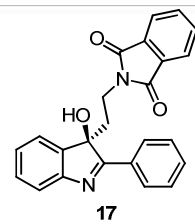


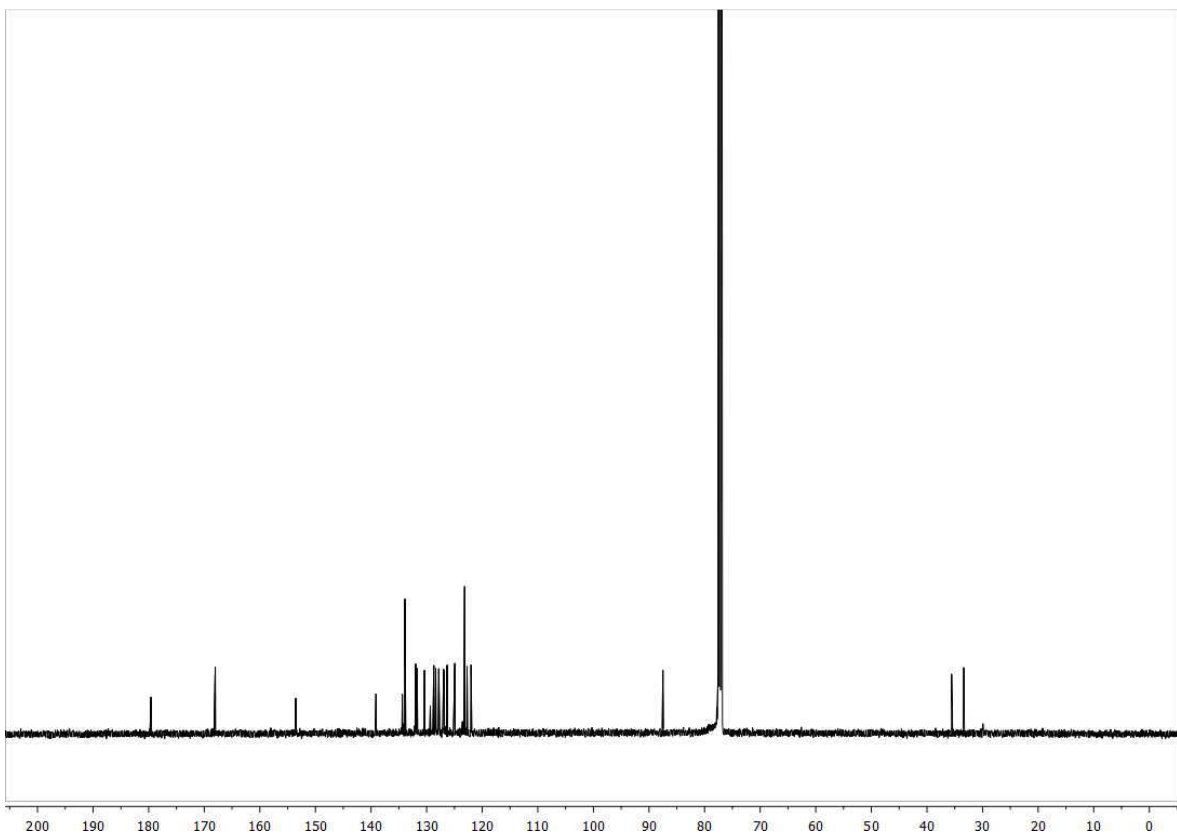
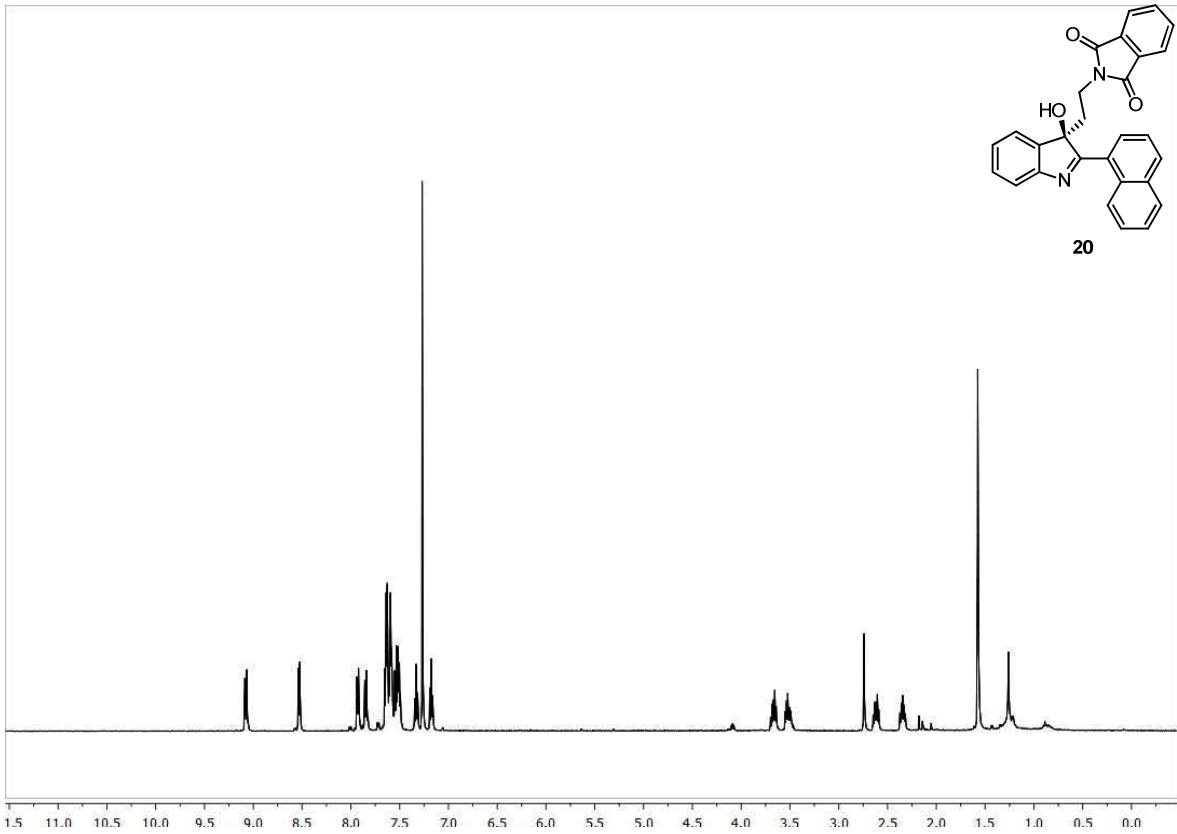


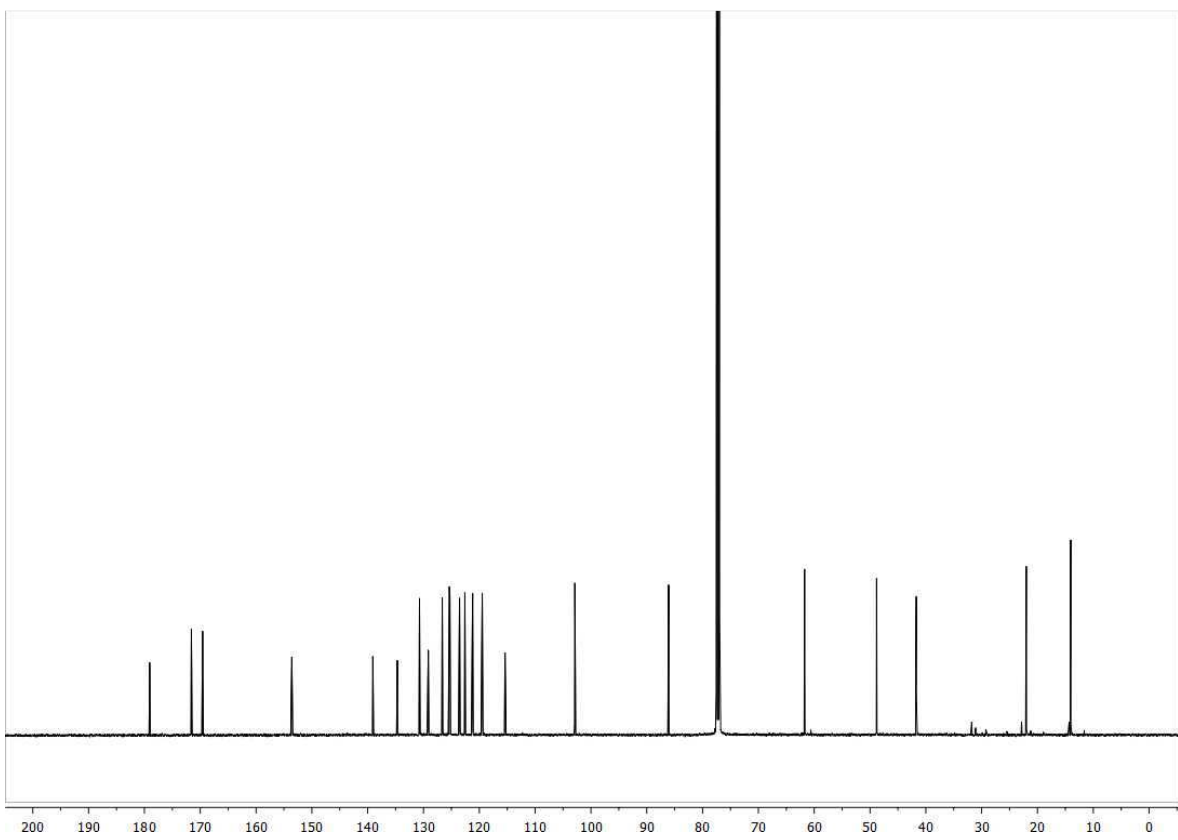
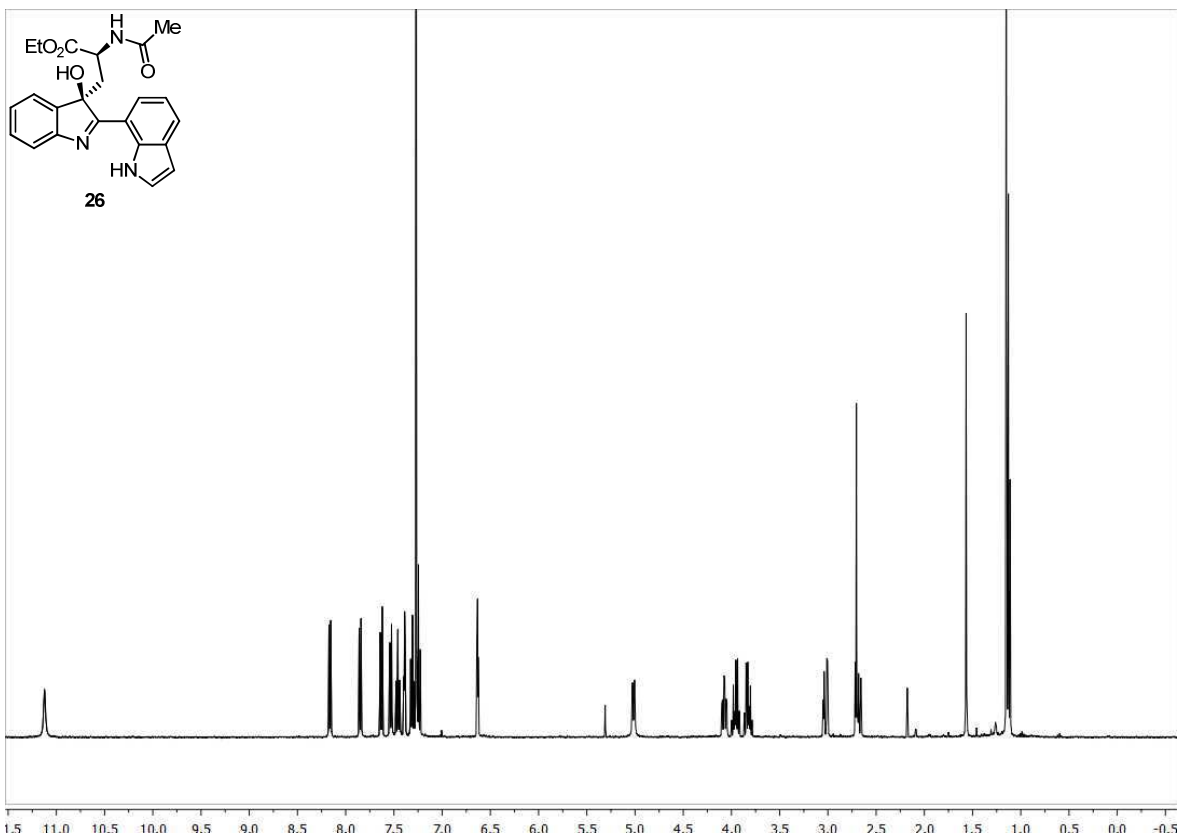


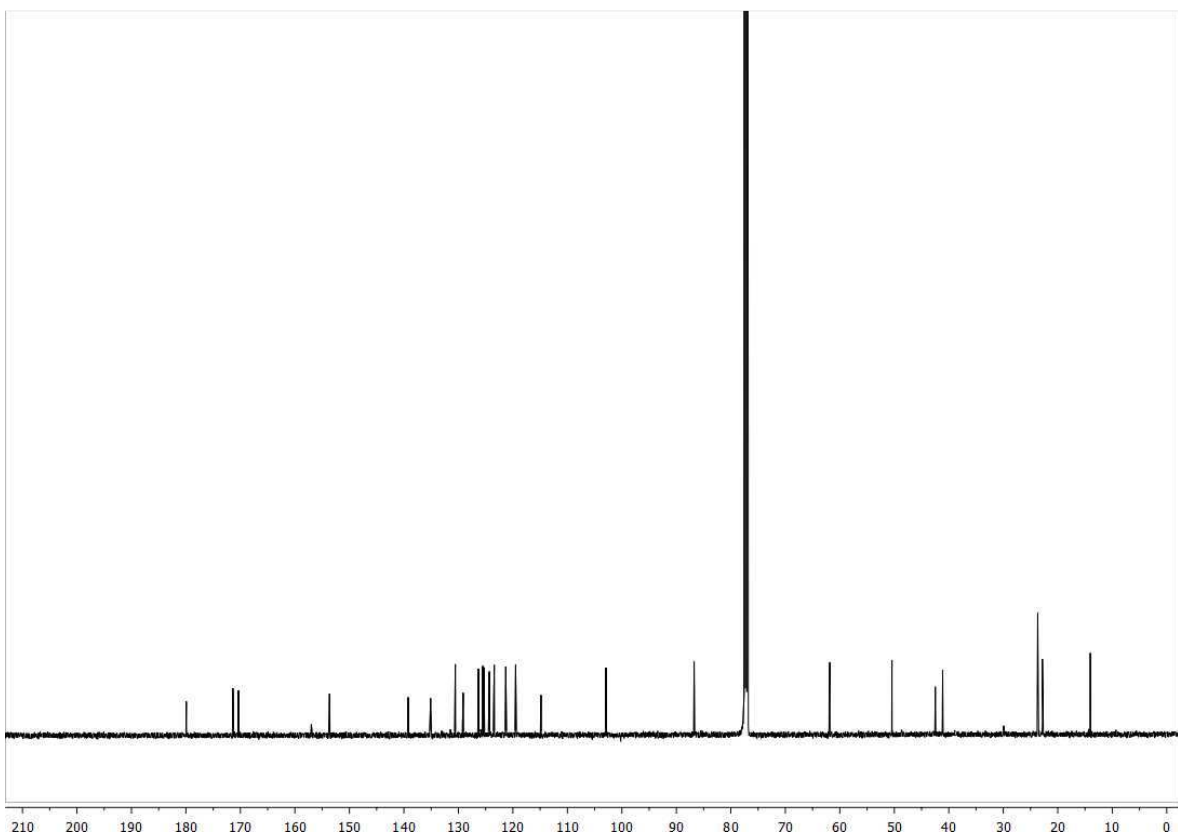
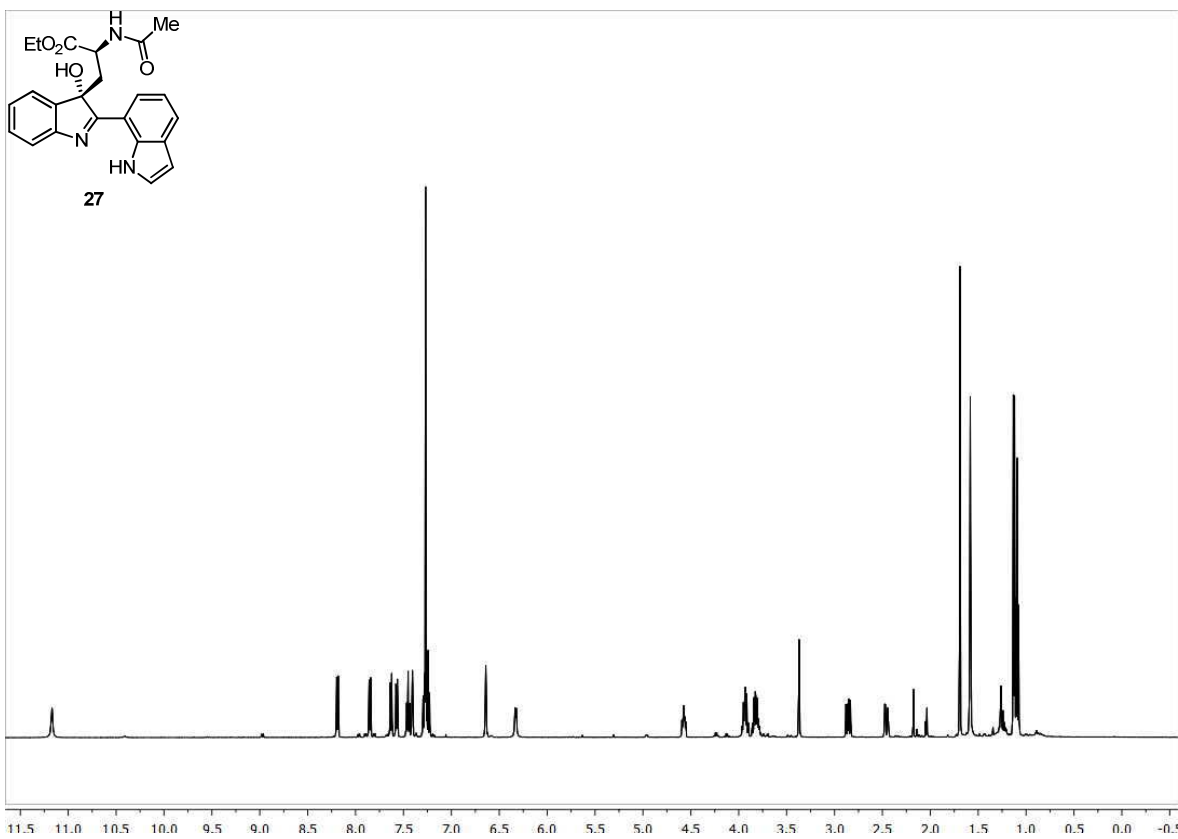


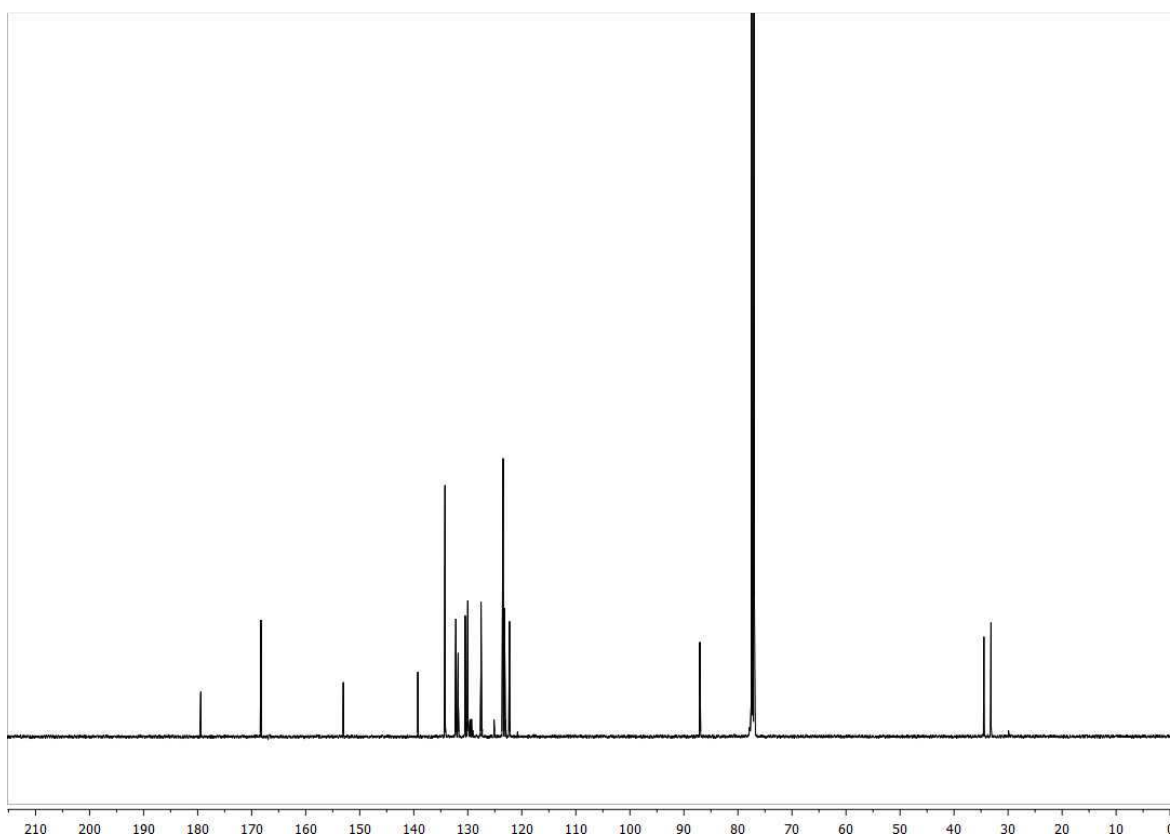
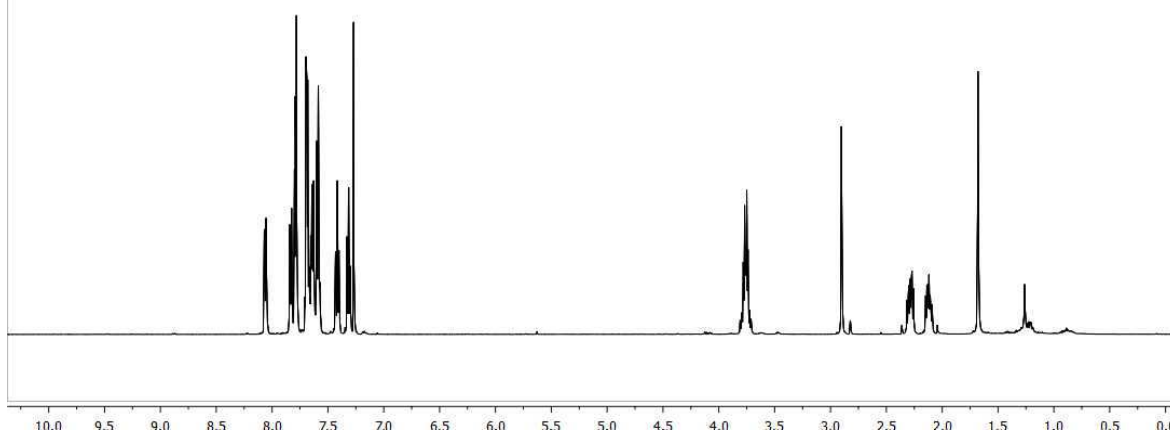
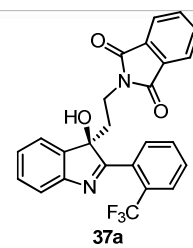


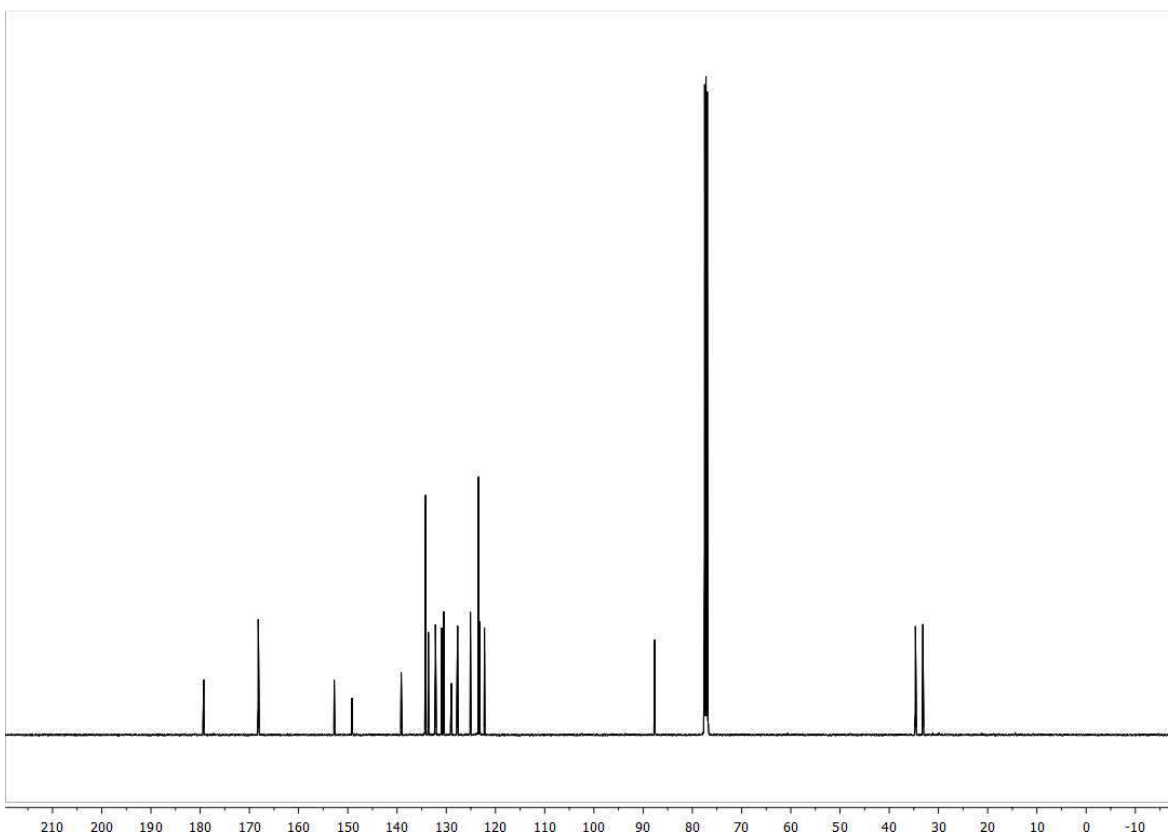
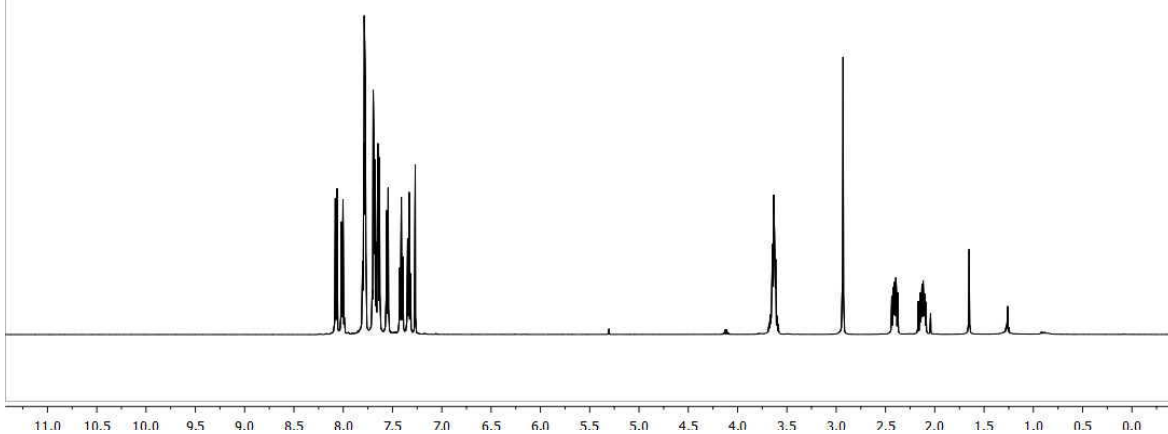
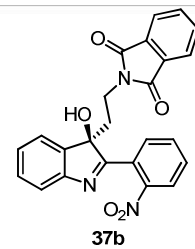


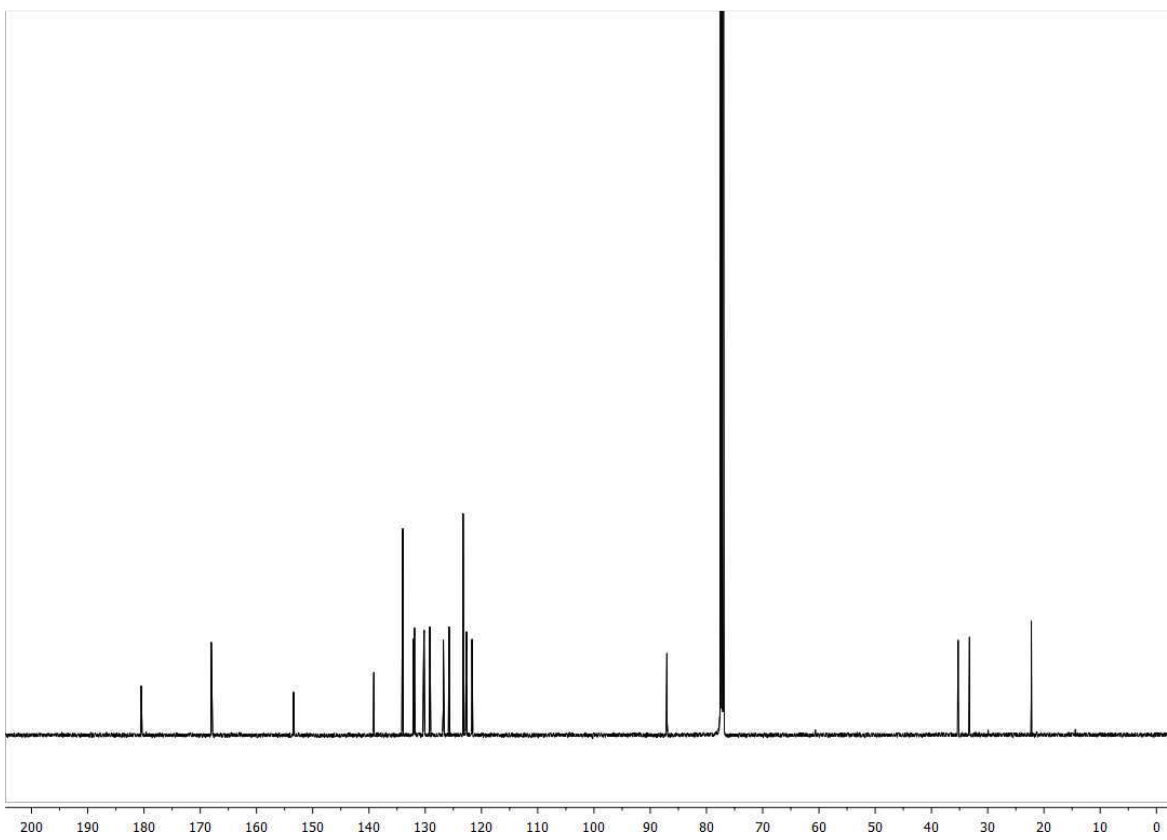
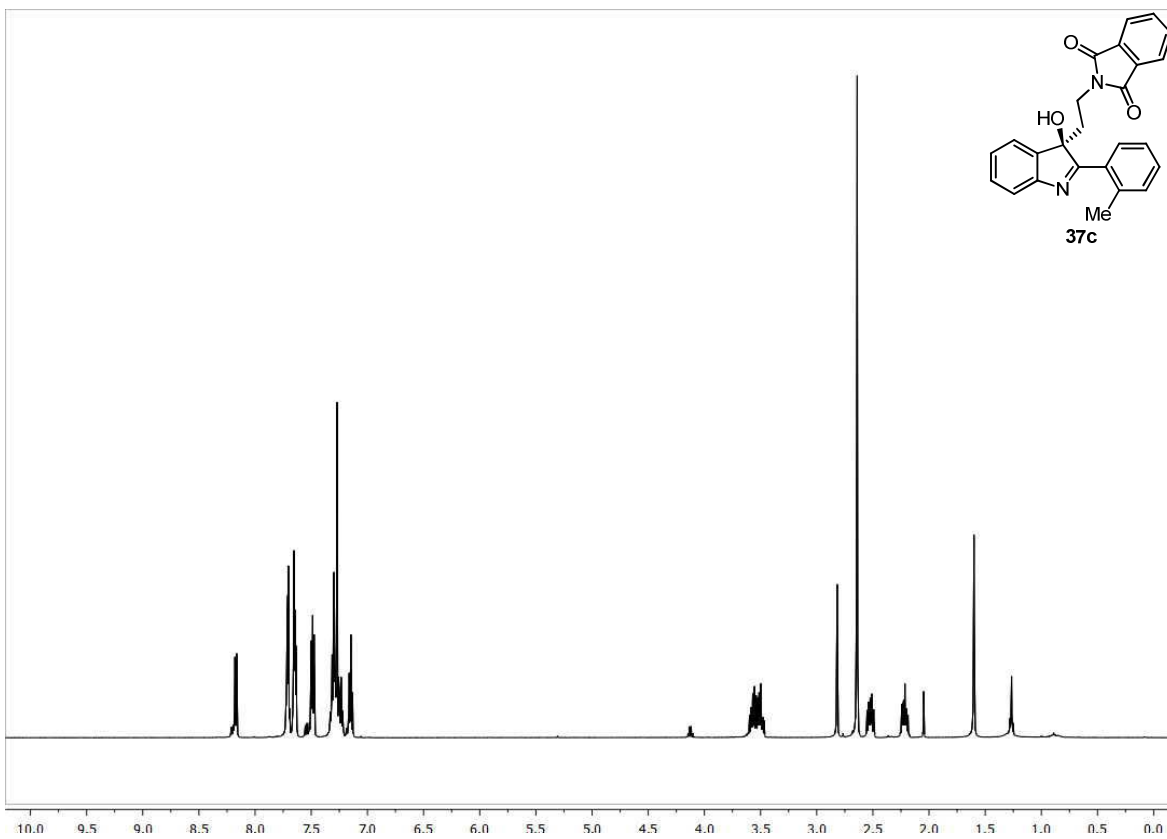


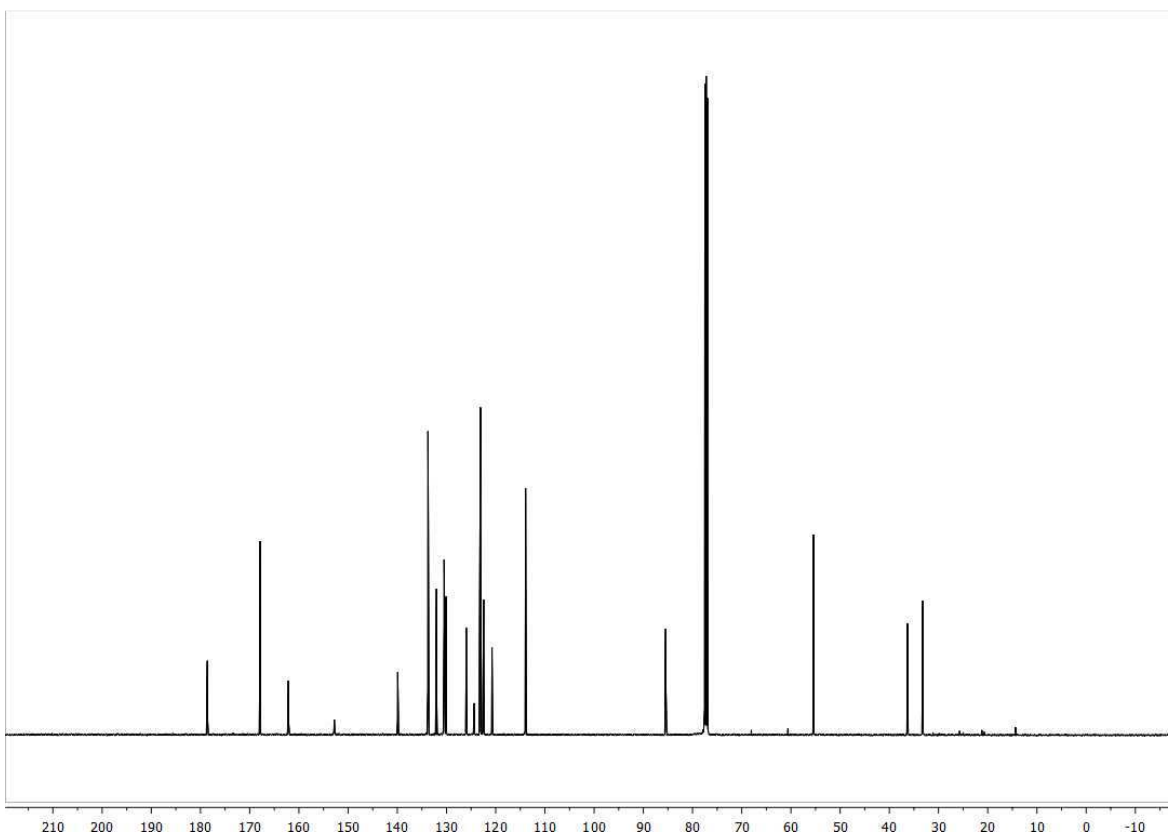
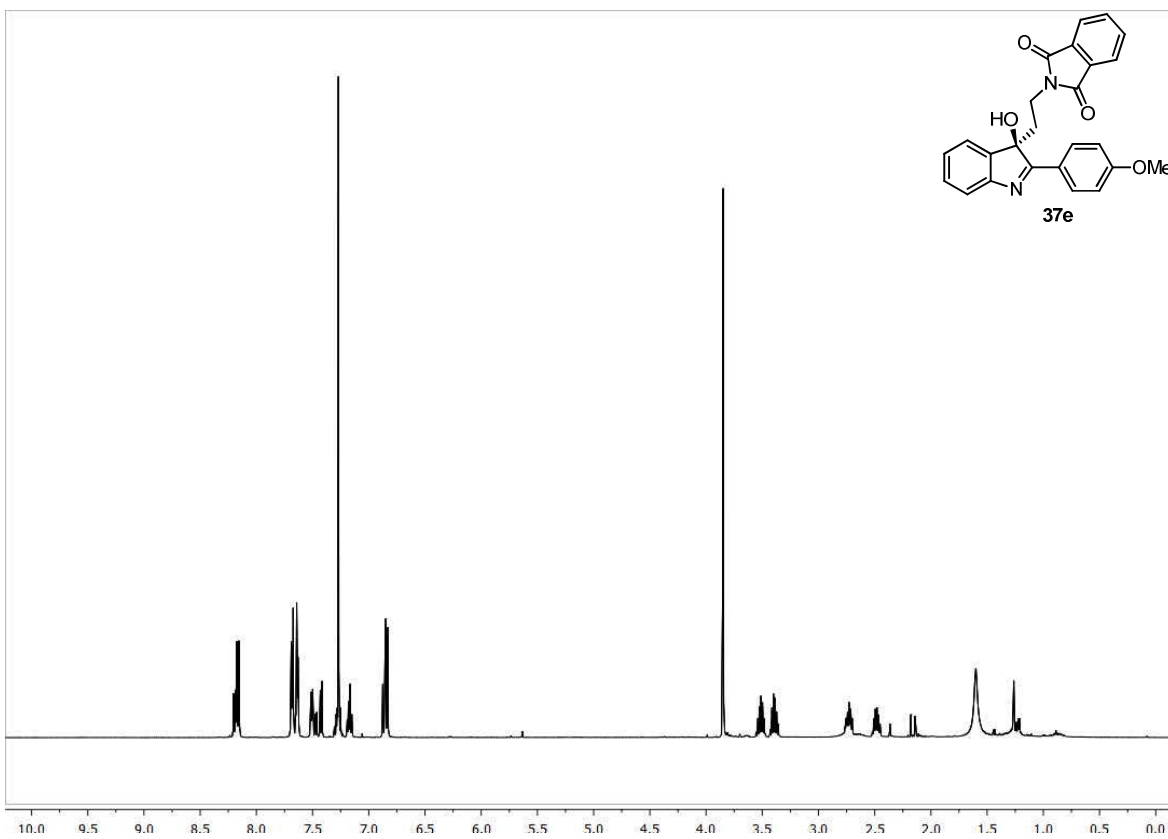


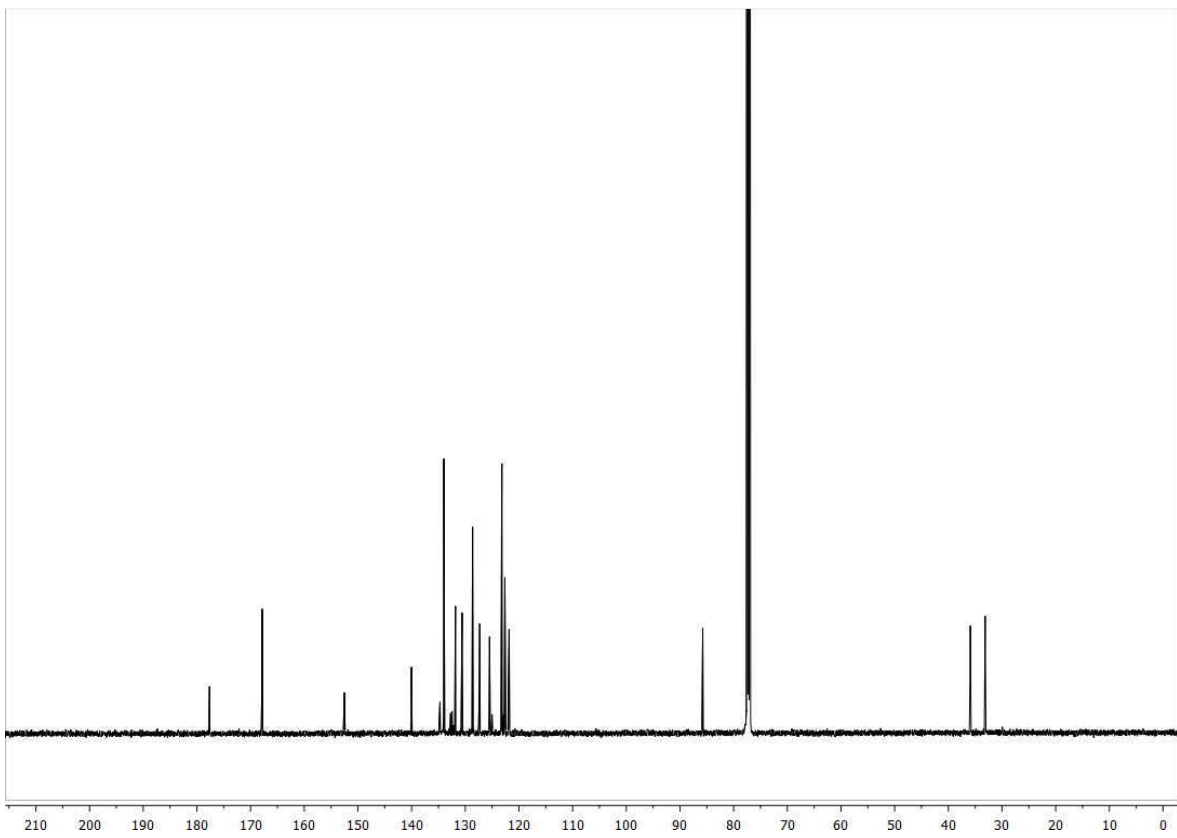
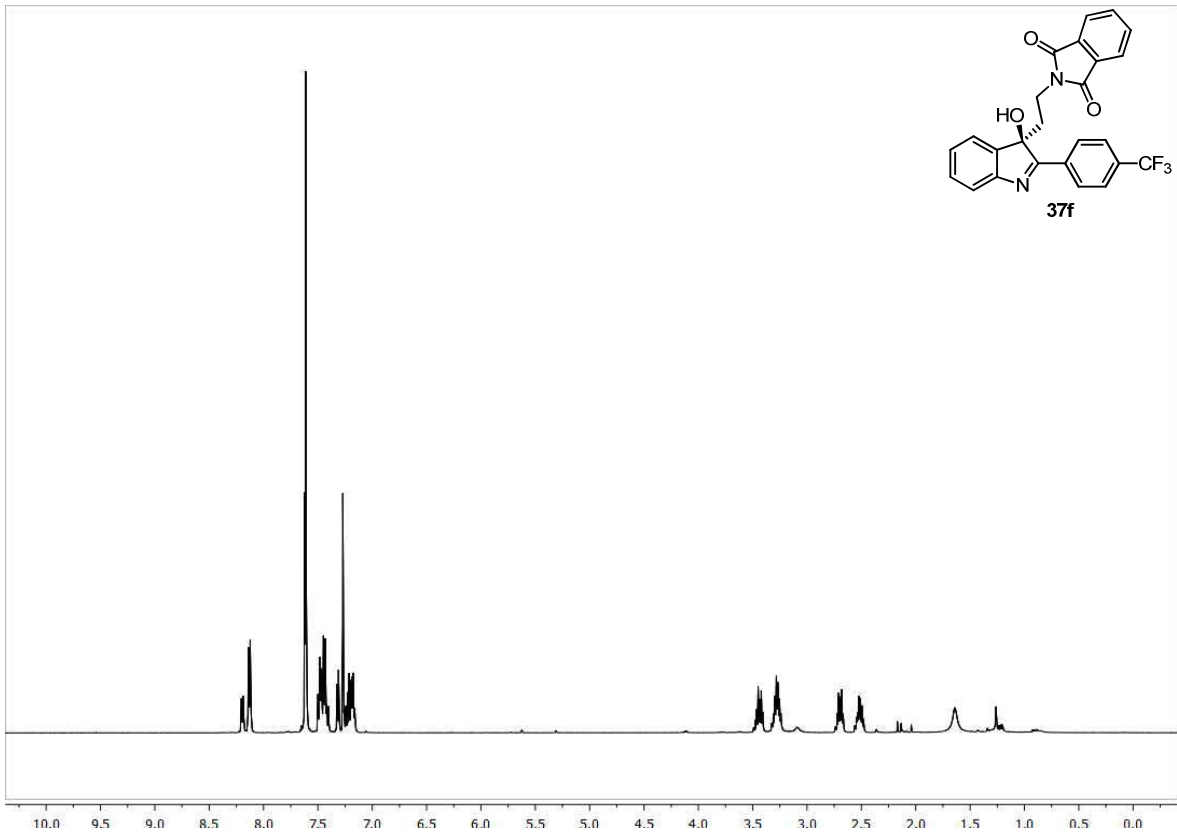


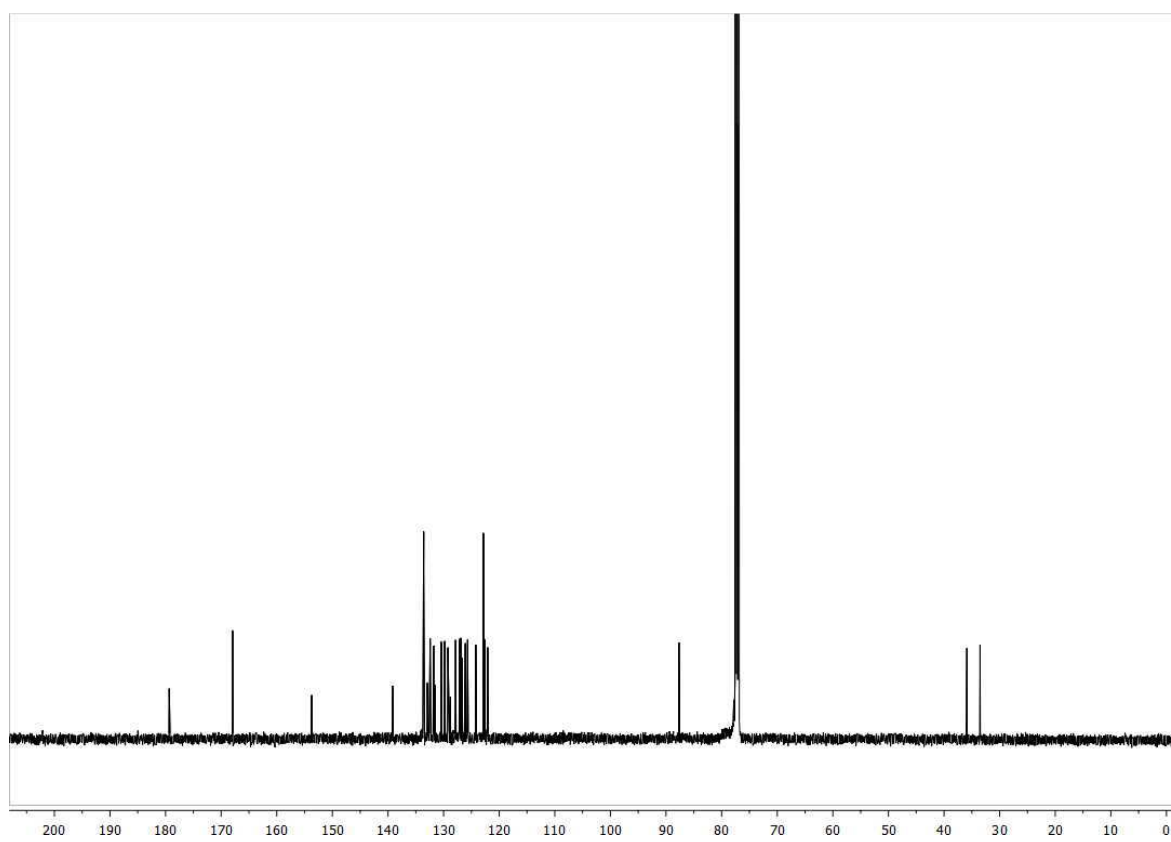
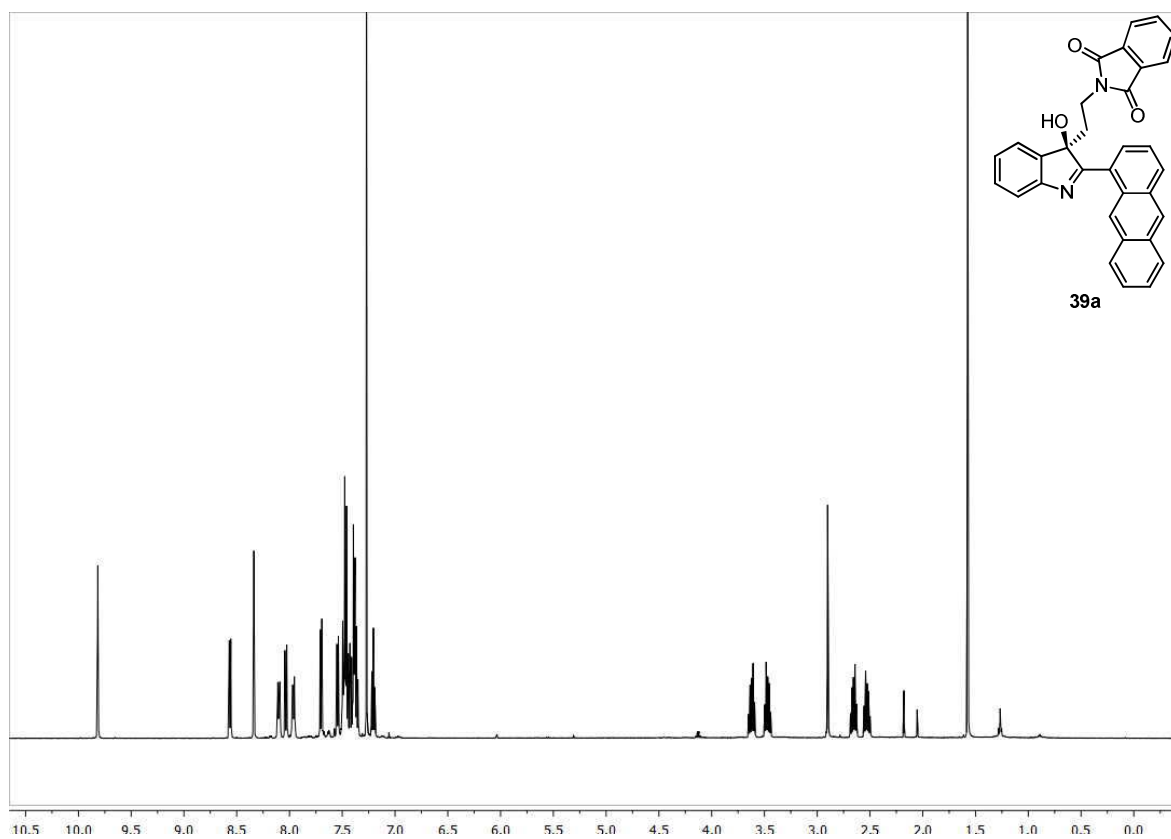


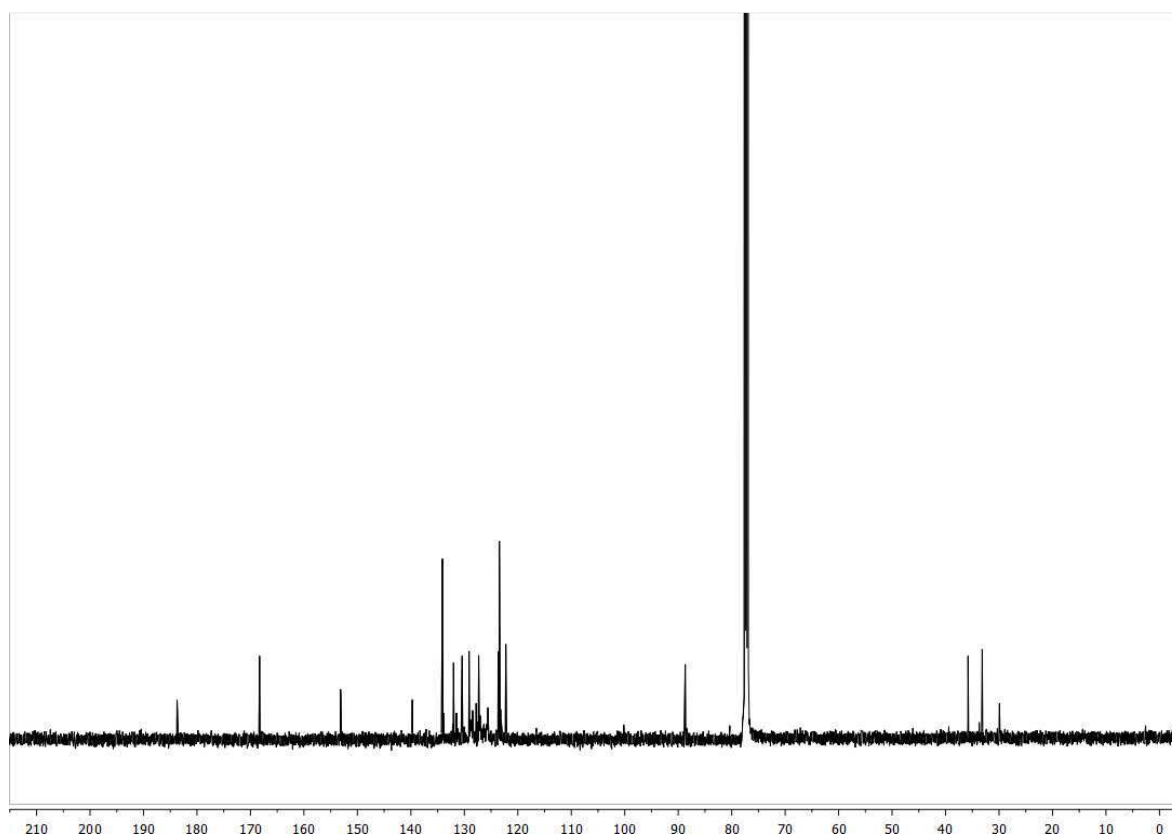
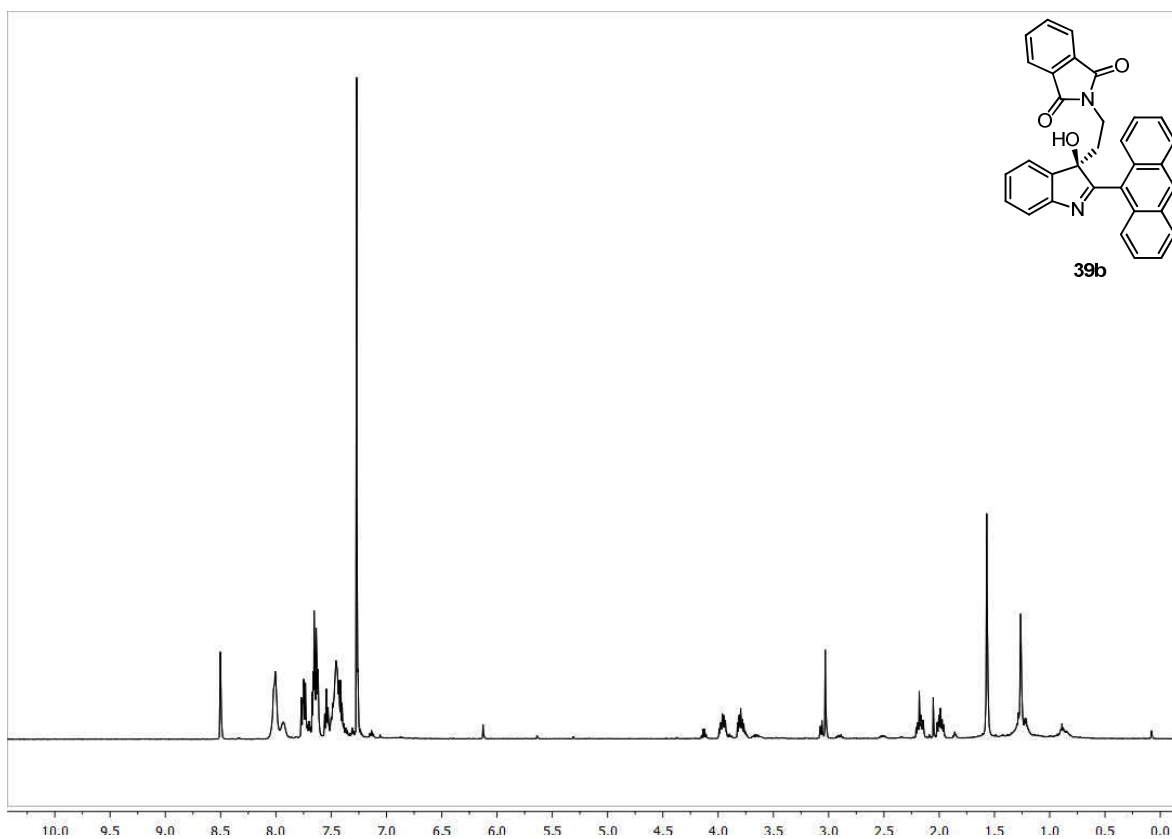


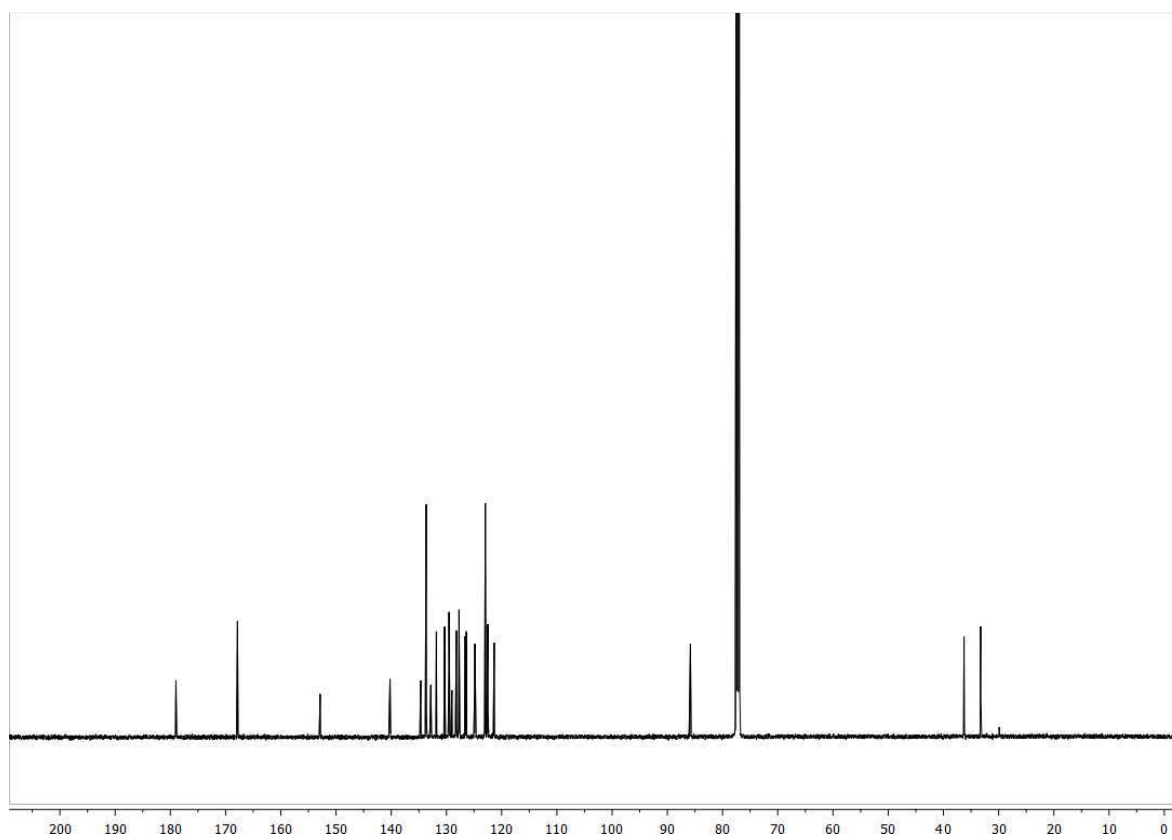
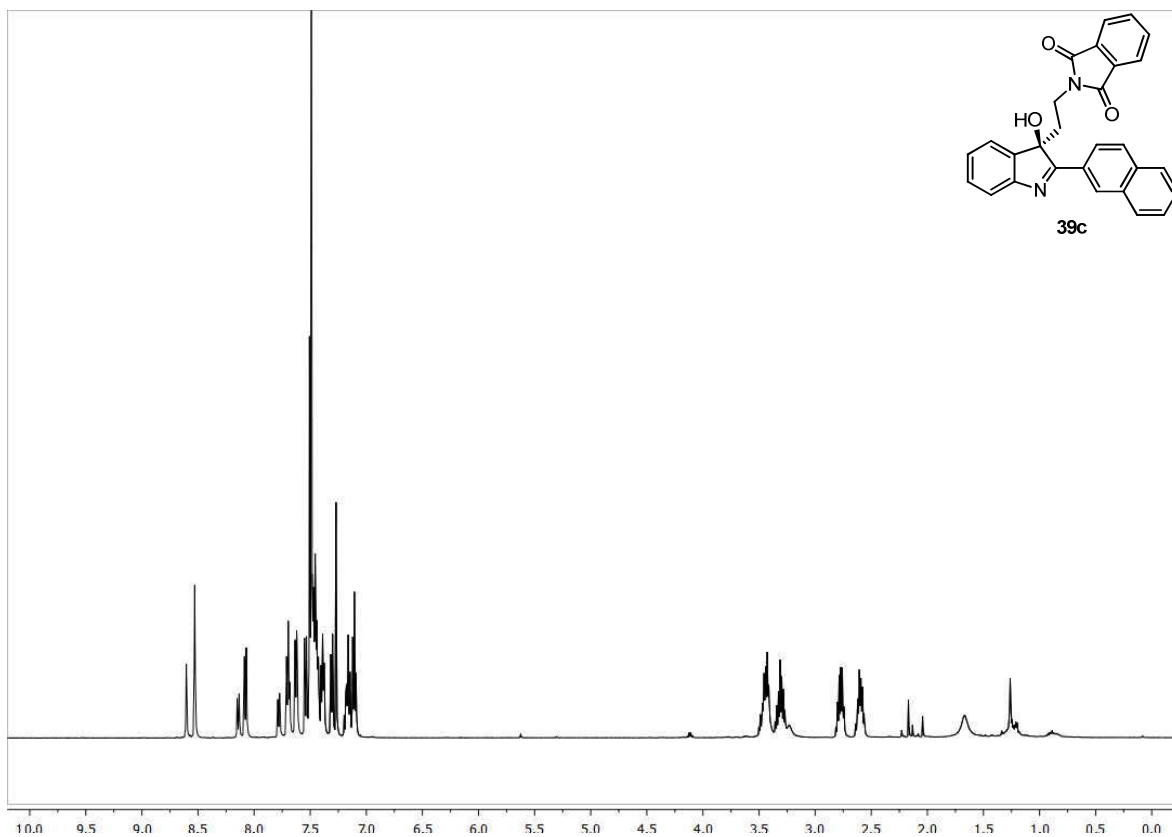


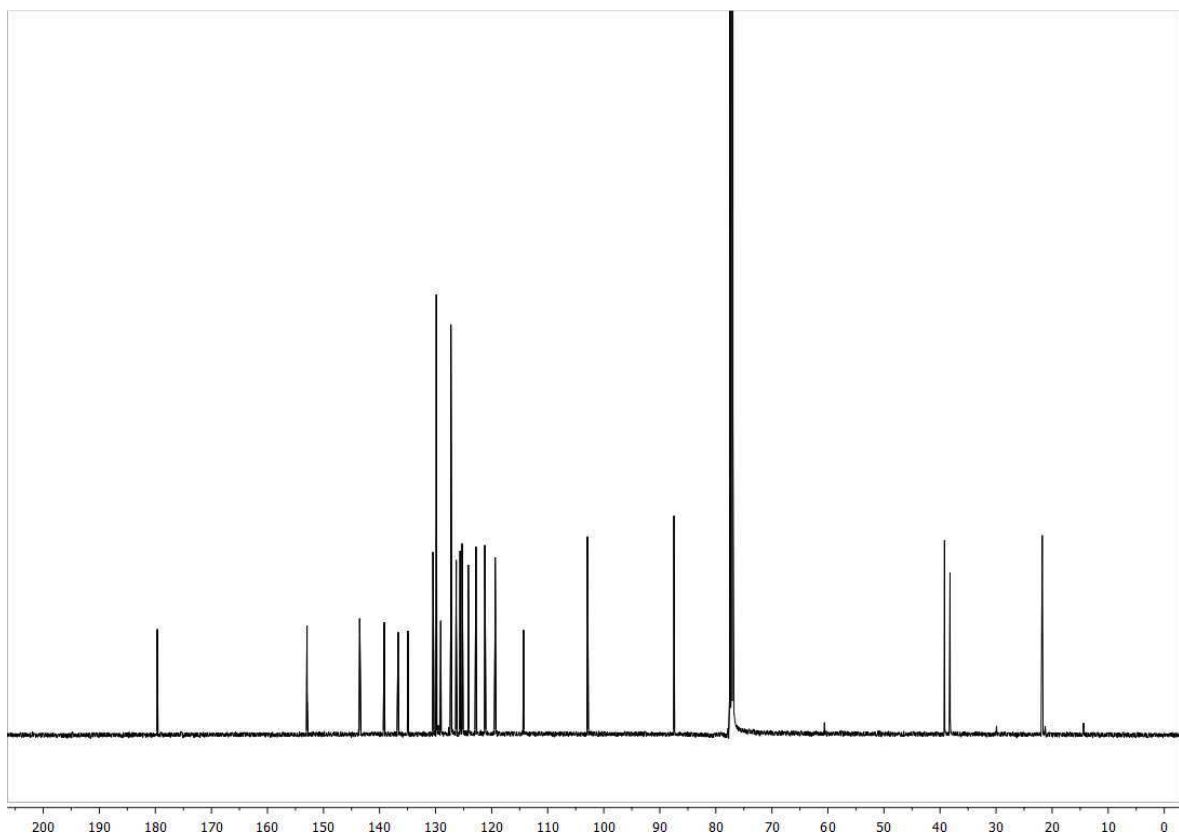
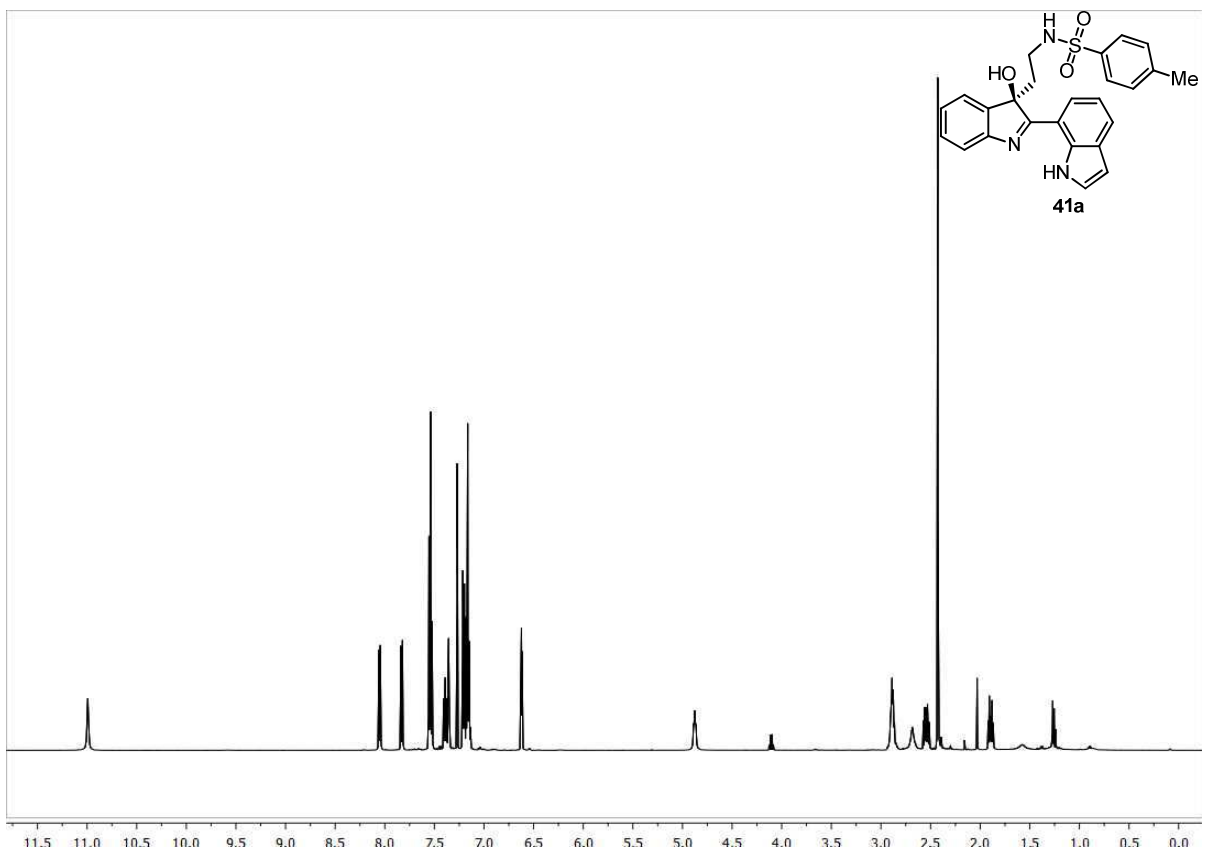


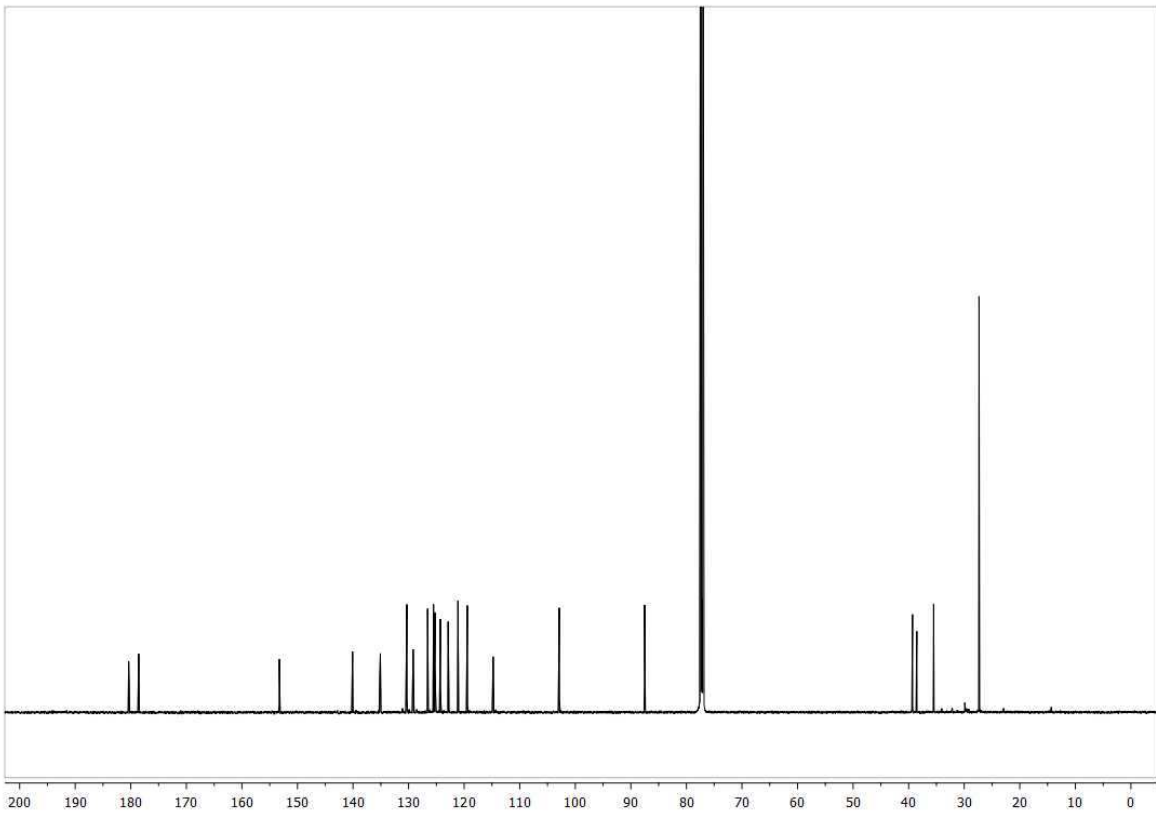
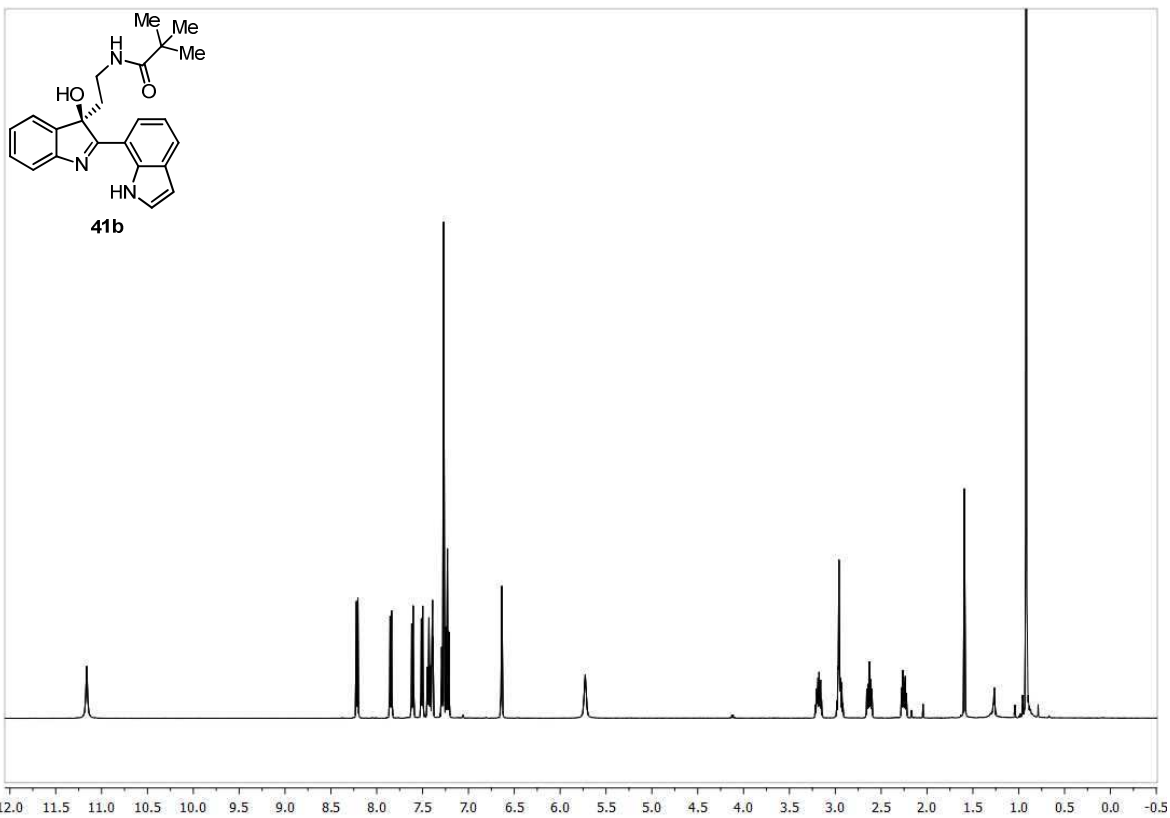


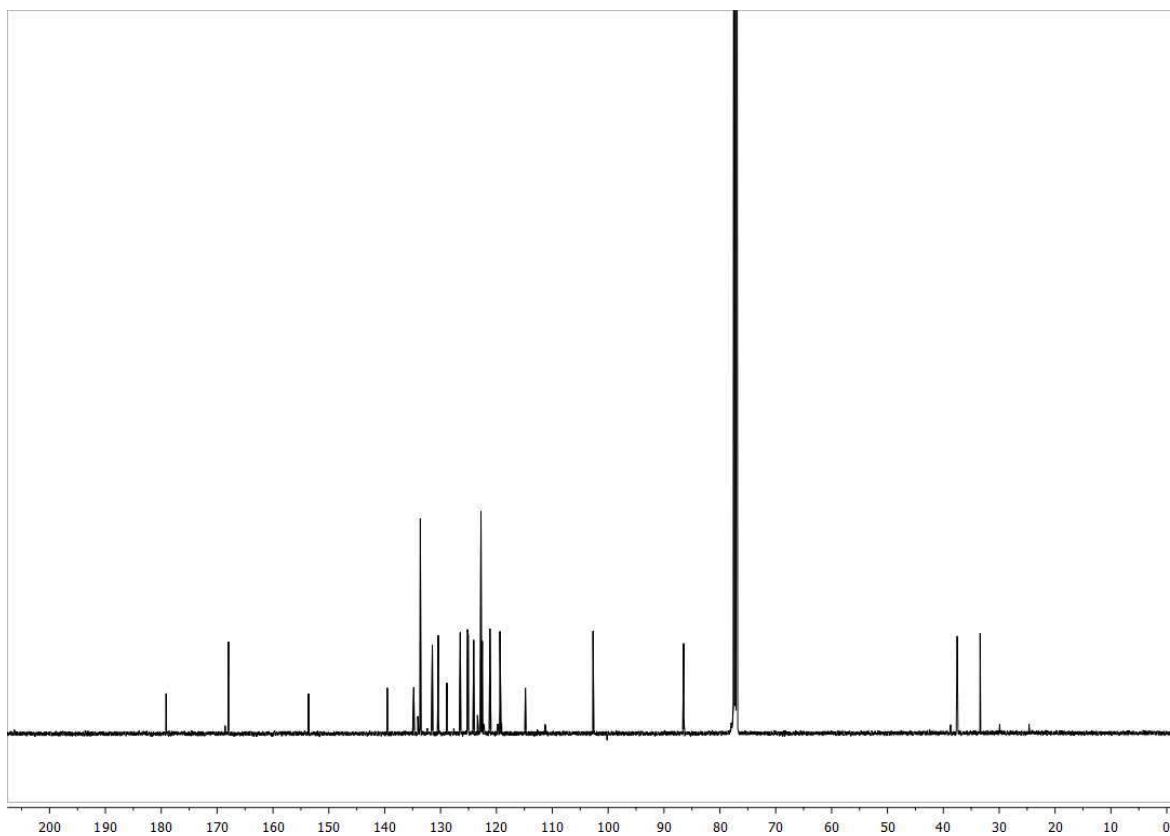
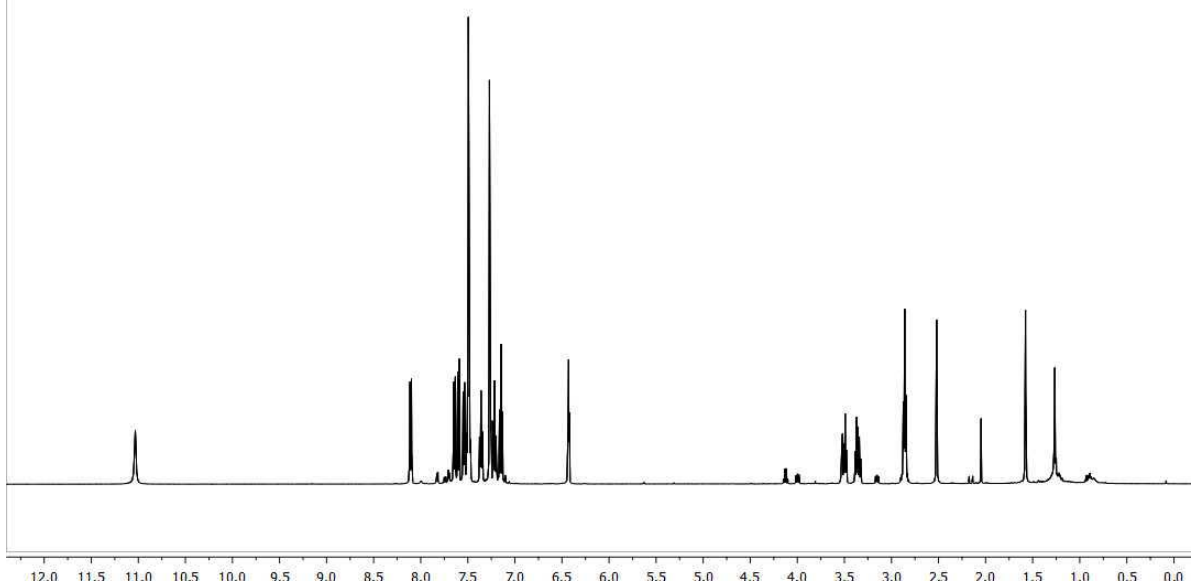
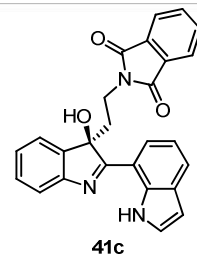


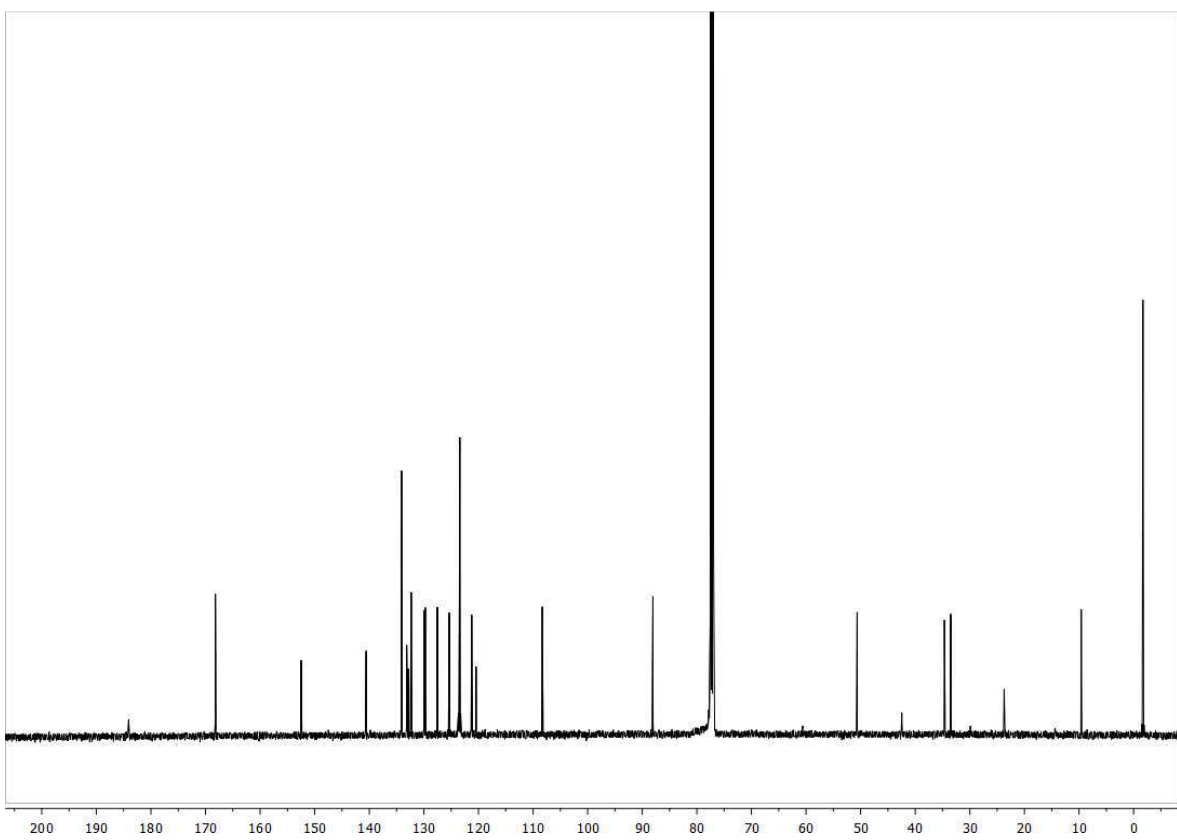
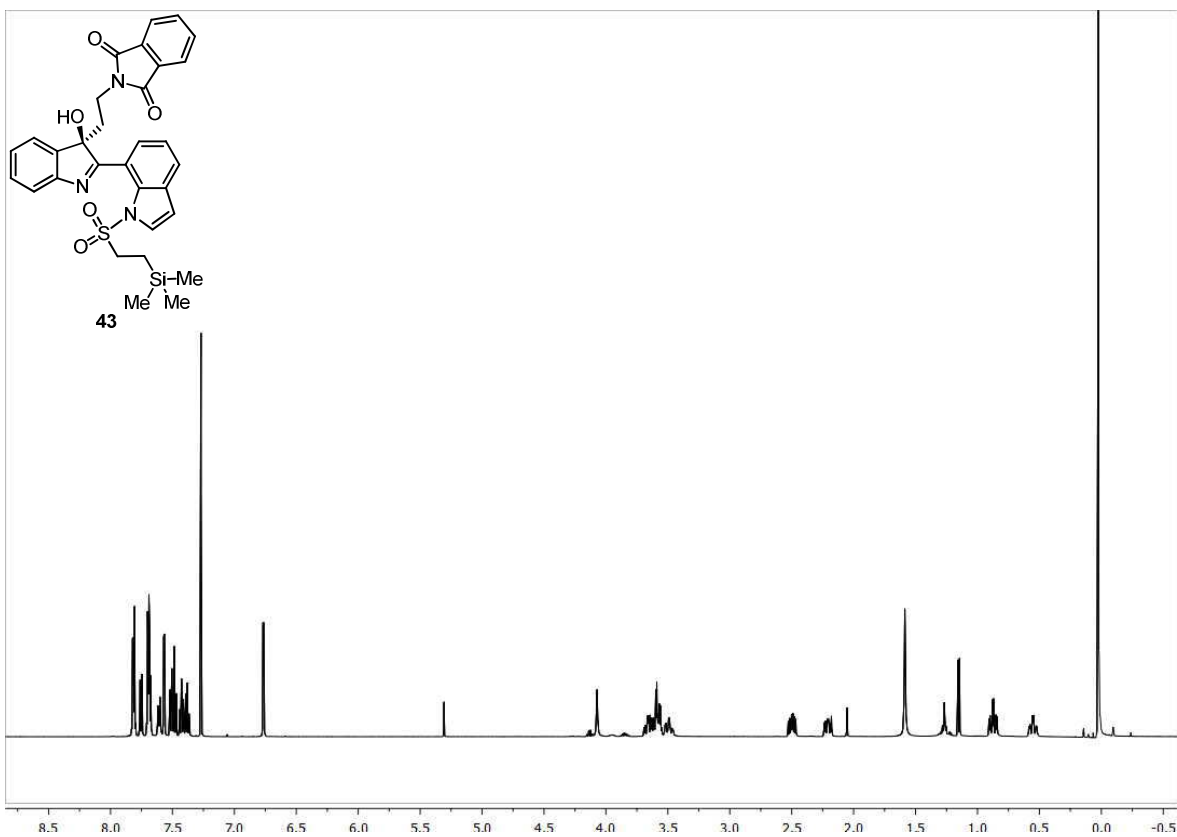


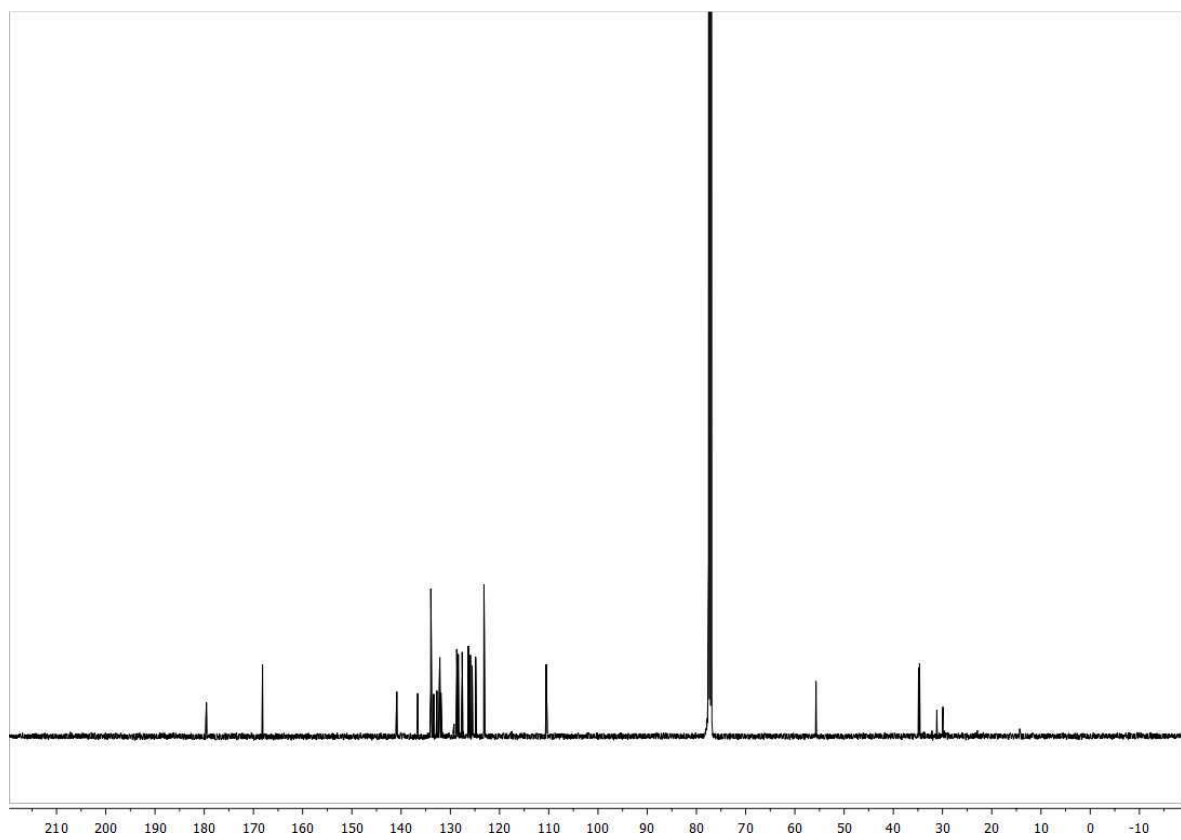
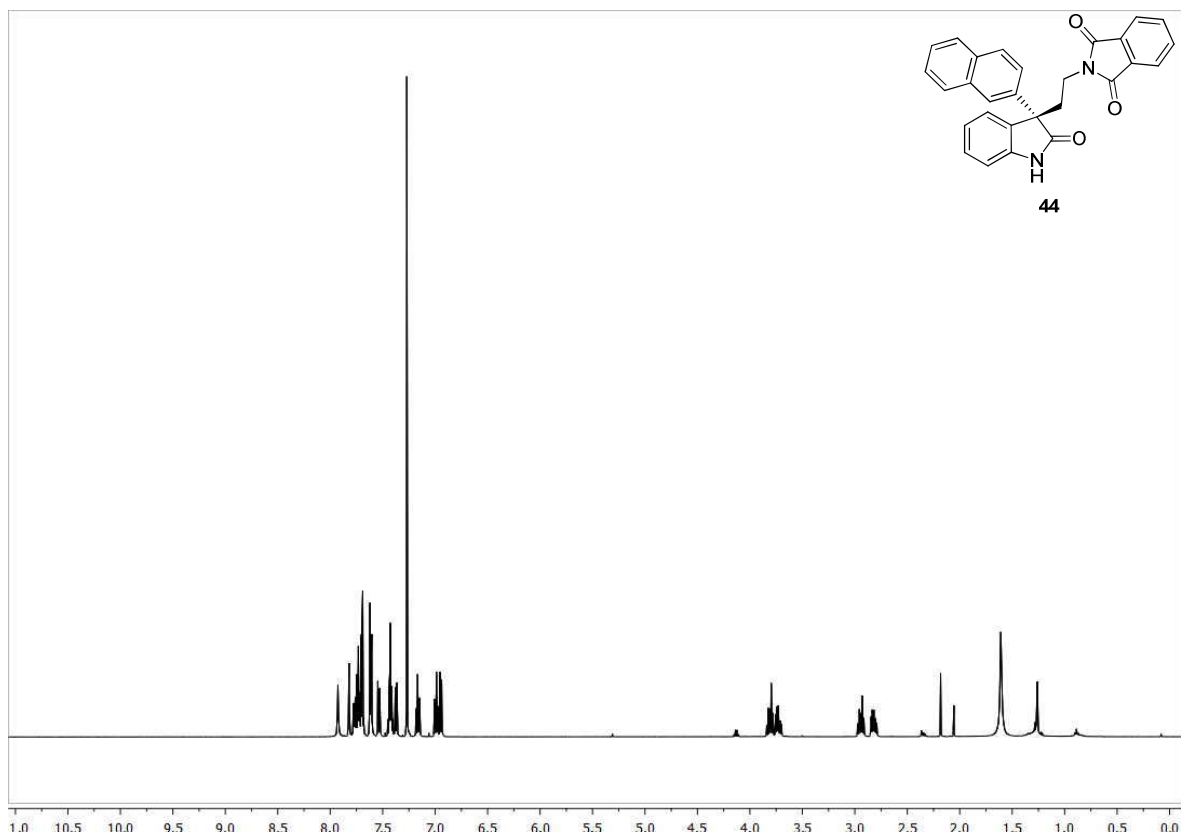


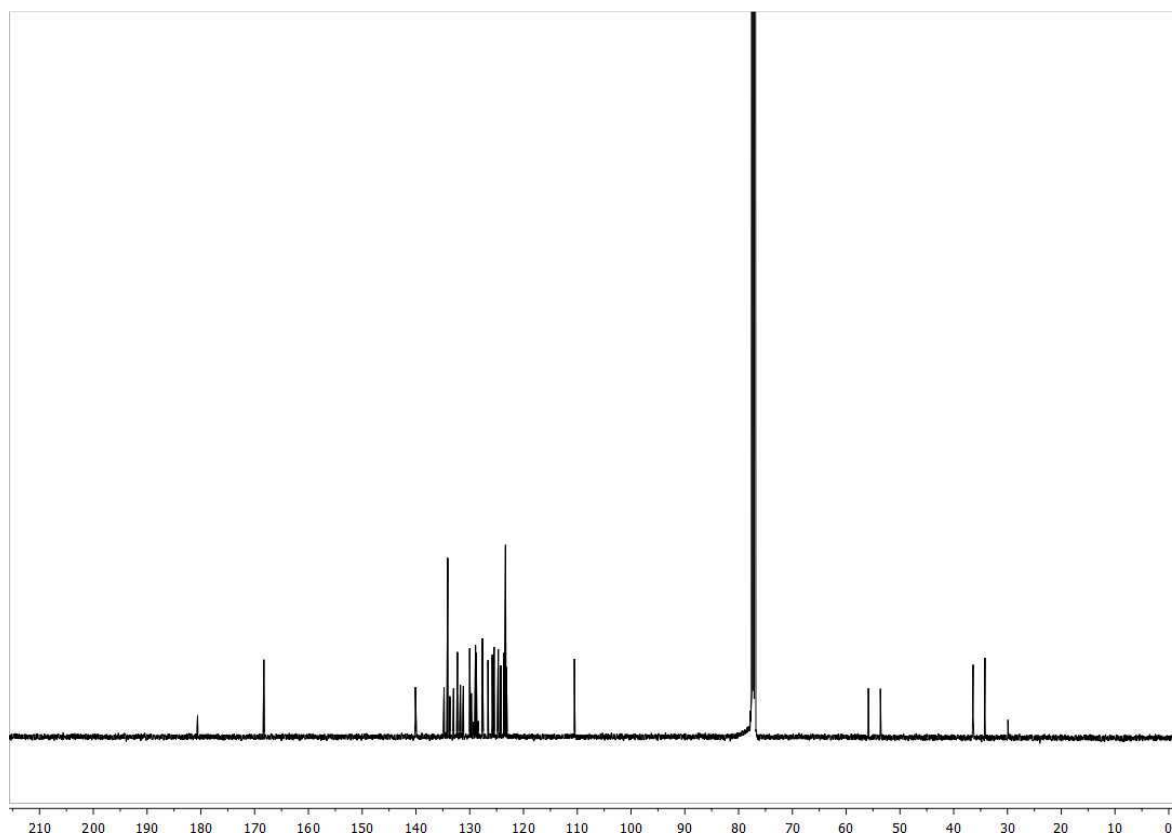
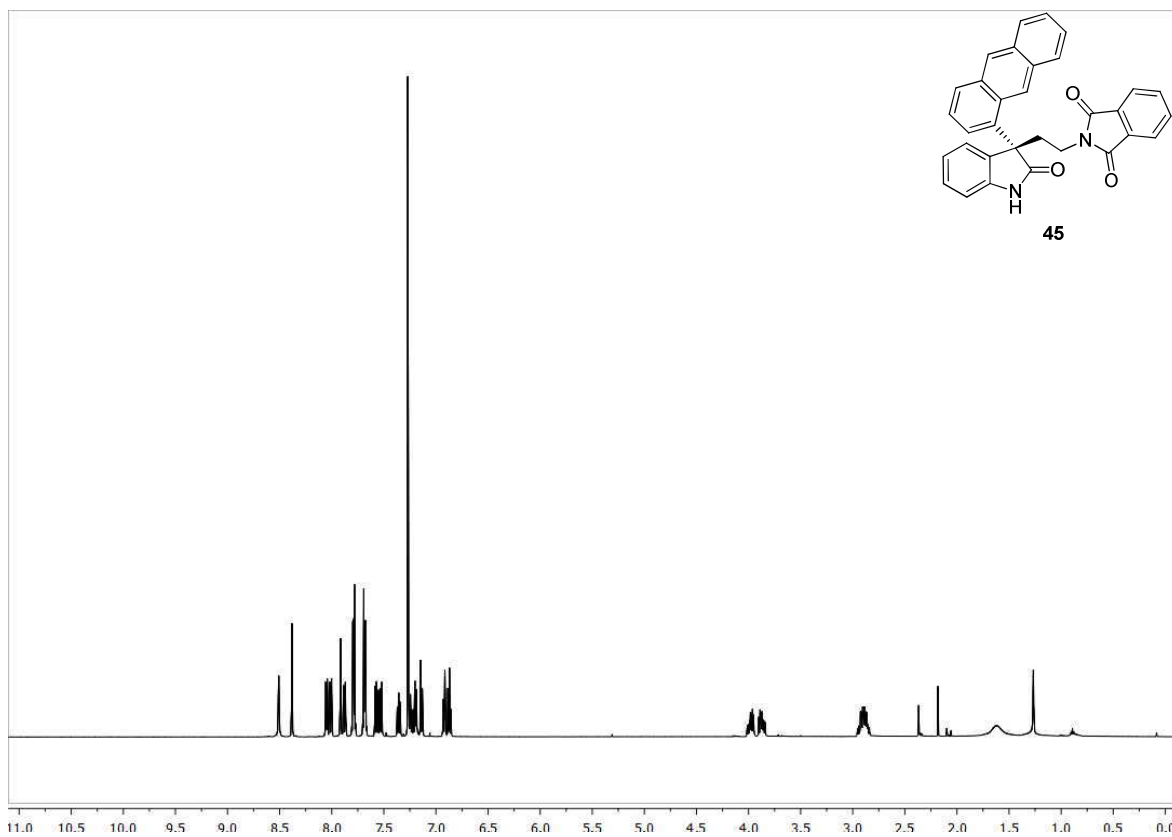


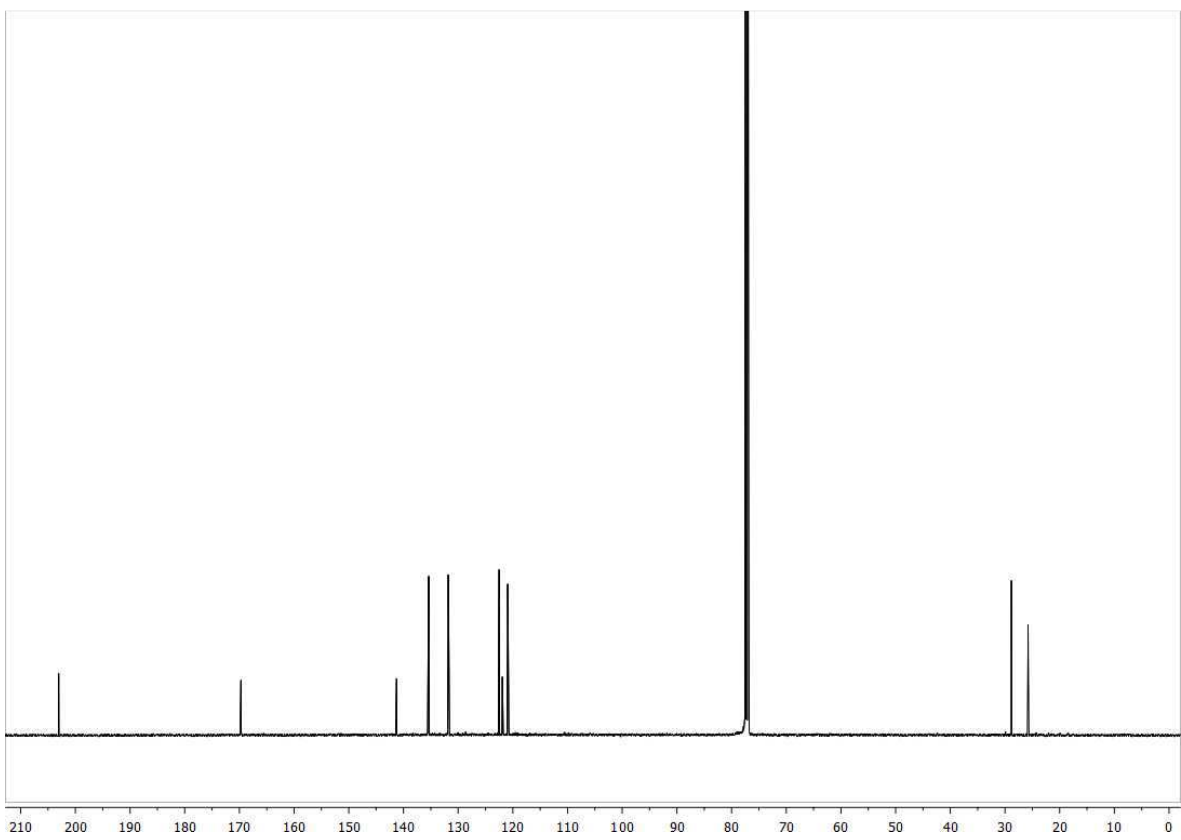
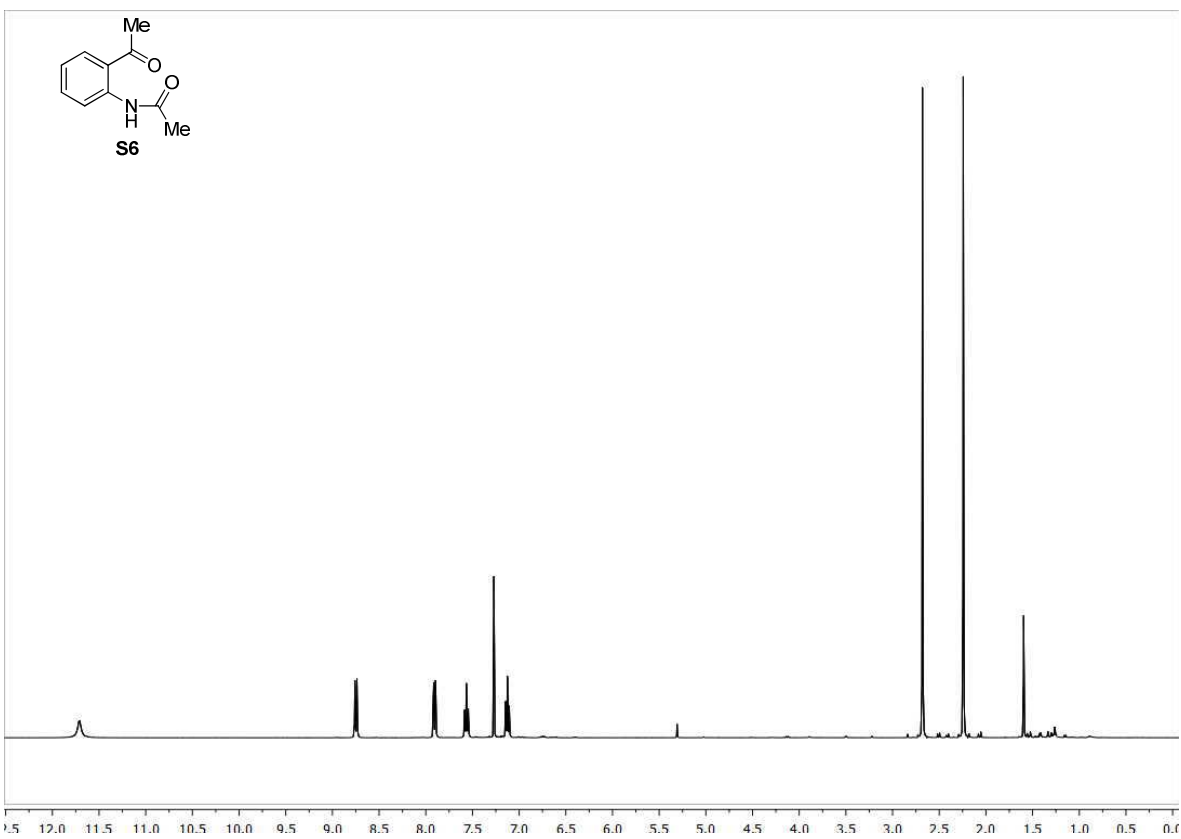


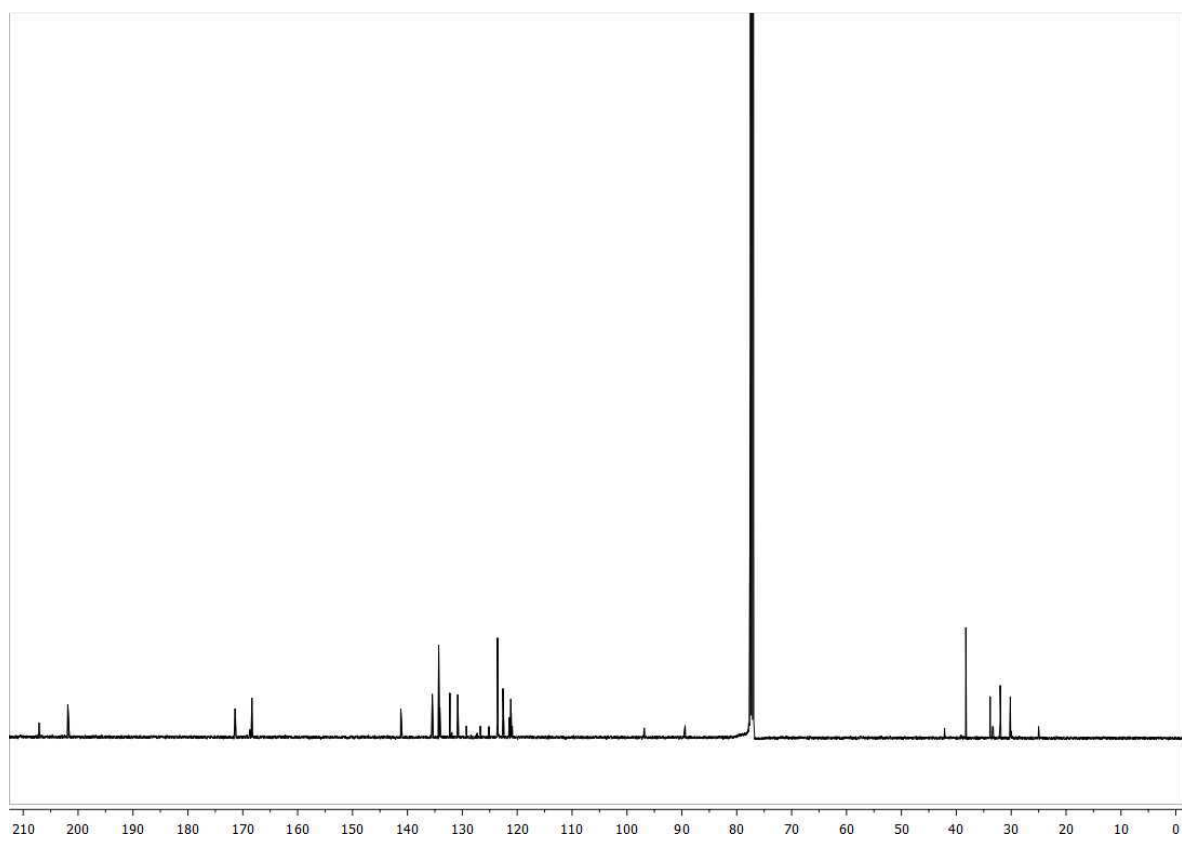
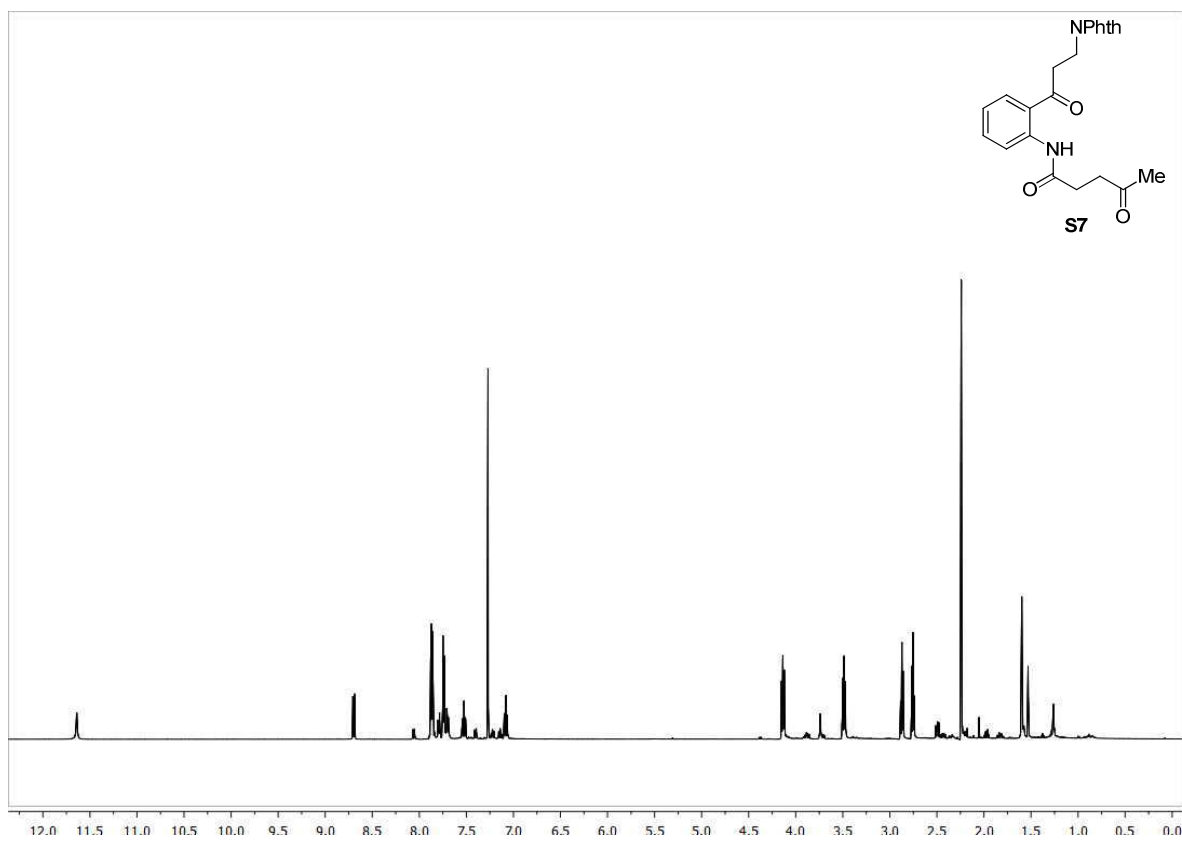












K. X-Ray Diffraction Report for Compound 41c.

Experimental

Data Collection

A colorless block crystal of $C_{26}H_{19}N_3O_3$ having approximate dimensions of $0.22 \times 0.22 \times 0.15$ mm was mounted in a loop. All measurements were made on a Rigaku RAXIS RAPID imaging plate area detector with graphite monochromated Cu-K α radiation.

Indexing was performed from 4 oscillations that were exposed for 180 seconds. The crystal-to-detector distance was 127.40 mm.

Cell constants and an orientation matrix for data collection corresponded to a primitive monoclinic cell with dimensions:

$$a = 7.62626(14) \text{ \AA}$$

$$b = 9.13104(17) \text{ \AA} \quad \beta = 92.6325(13)^\circ$$

$$c = 14.3228(3) \text{ \AA}$$

$$V = 996.33(3) \text{ \AA}^3$$

For $Z = 2$ and F.W. = 421.45, the calculated density is 1.405 g/cm^3 . Based on the systematic absences of:

$$0k0: k \pm 2n$$

packing considerations, a statistical analysis of intensity distribution, and the successful solution and refinement of the structure, the space group was determined to be: $P2_1$ (#4)

The data were collected at a temperature of $-180 \pm 1^\circ\text{C}$ to a maximum 2θ value of 136.5° . A total of 171 oscillation images were collected. A sweep of data was done using ω scans from 20.0 to 200.0° in 5.0° step, at $\chi=0.0^\circ$ and $\phi = 0.0^\circ$. The exposure rate was $36.0 \text{ [sec./}^\circ]$. A second sweep was performed using ω scans from 20.0 to 185.0° in 5.0° step, at $\chi=54.0^\circ$ and $\phi = 0.0^\circ$. The exposure rate was $36.0 \text{ [sec./}^\circ]$. Another sweep was performed using ω scans from 21.0 to 191.0° in 5.0° step, at $\chi=54.0^\circ$ and $\phi = 90.0^\circ$. The exposure rate was $36.0 \text{ [sec./}^\circ]$. Another sweep was performed using ω scans from 20.0 to 200.0° in 5.0° step, at $\chi=54.0^\circ$ and $\phi = 270.0^\circ$. The exposure rate was $36.0 \text{ [sec./}^\circ]$. Another sweep was performed using ω scans from 24.0 to 184.0° in 5.0° step, at $\chi=54.0^\circ$ and $\phi = 180.0^\circ$. The exposure rate was $36.0 \text{ [sec./}^\circ]$. The crystal-to-detector distance was 127.40 mm. Readout was performed in the 0.100 mm pixel mode.

Data Reduction

Of the 10803 reflections that were collected, 3418 were unique ($R_{\text{int}} = 0.030$).

The linear absorption coefficient, μ , for Cu-K α radiation is 7.601 cm^{-1} . An empirical absorption correction was applied which resulted in transmission factors ranging from 0.781 to 0.892. The data were corrected for Lorentz and polarization effects.

Structure Solution and Refinement

The structure was solved by direct methods⁷ and expanded using Fourier techniques⁸. The non-hydrogen atoms were refined anisotropically. Hydrogen atoms were refined using the riding model. The final cycle of full-matrix least-squares refinement⁹ on F^2 was based on 3413 observed reflections and 298 variable parameters and converged (largest parameter shift was 0.00 times its esd) with unweighted and weighted agreement factors of:

$$R1 = \sum ||F_o| - |F_c|| / \sum |F_o| = 0.0305$$

$$wR2 = [\sum (w (F_o^2 - F_c^2)^2) / \sum w(F_o^2)^2]^{1/2} = 0.0770$$

The standard deviation of an observation of unit weight¹⁰ was 1.08. Unit weights were used. The maximum and minimum peaks on the final difference Fourier map corresponded to 0.18 and -0.16 e⁻/Å³, respectively. The absolute structure was deduced based on Flack parameter, 0.24(17), using 1489 Friedel pairs.¹¹

Neutral atom scattering factors were taken from Cromer and Waber¹². Anomalous dispersion effects were included in F_{calc} ¹³; the values for $\Delta f'$ and $\Delta f''$ were those of Creagh and McAuley¹⁴. The values for the mass attenuation coefficients are those of Creagh and Hubbell¹⁵. All calculations were performed using the CrystalStructure¹⁶ crystallographic software package except for refinement, which was performed using SHELXL-97¹⁷.

Experimental Details

a. Crystal Data

Empirical Formula	C ₂₆ H ₁₉ N ₃ O ₃
Formula Weight	421.45
Crystal Color, Habit	colorless, block
Crystal Dimensions	0.22 × 0.22 × 0.15 mm
Crystal System	monoclinic
Lattice Type	Primitive
Indexing Images	4 oscillations @ 180.0 seconds
Detector Position	127.40 mm
Pixel Size	0.100 mm
Lattice Parameters	a = 7.62626(14) Å b = 9.13104(17) Å c = 14.3228(3) Å

$$\beta = 92.6325(13)^\circ$$

$$V = 996.33(3) \text{ \AA}^3$$

Space Group	P2 ₁ (#4)
Z value	2
D _{calc}	1.405 g/cm ³
F ₀₀₀	440.00
$\mu(\text{CuK}\alpha)$	7.601 cm ⁻¹

b. Intensity Measurements

Diffractionmeter Rigaku RAXIS-RAPID

Radiation CuK α ($\lambda = 1.54187 \text{ \AA}$)
graphite monochromated

Detector Aperture 280 mm \times 256 mm

Data Images 171 exposures

ω oscillation Range ($\chi=0.0, \varphi=0.0$) 20.0 - 200.0°

Exposure Rate 36.0 sec./°

ω oscillation Range ($\chi=54.0, \varphi=0.0$) 20.0 - 185.0°

Exposure Rate 36.0 sec./°

ω oscillation Range ($\chi=54.0, \varphi=90.0$) 21.0 - 191.0°

Exposure Rate 36.0 sec./°

ω oscillation Range ($\chi=54.0, \varphi=270.0$) 20.0 - 200.0°

Exposure Rate 36.0 sec./°

ω oscillation Range ($\chi=54.0, \varphi=180.0$) 24.0 - 184.0°

Exposure Rate 36.0 sec./°

Detector Position 127.40 mm

Pixel Size 0.100 mm

$2\theta_{\text{max}}$ 136.5°

No. of Reflections Measured Total: 10803

Unique: 3413 ($R_{\text{int}} = 0.030$)

Friedel pairs: 1489

Corrections Lorentz-polarization

Absorption

(trans. factors: 0.781 - 0.892)

c. Structure Solution and Refinement

Structure Solution Direct Methods (SHELX97)

Refinement Full-matrix least-squares on F^2

Function Minimized $\Sigma w (F_o^2 - F_c^2)^2$

Least Squares Weights $w = 1 / [\sigma^2(F_o^2) + (0.0399 \cdot P)^2 + 0.0984 \cdot P]$

where $P = (\text{Max}(F_o^2, 0) + 2F_c^2) / 3$

$2\theta_{\text{max}}$ cutoff 136.5°

Anomalous Dispersion All non-hydrogen atoms

No. Observations (All reflections) 3413

No. Variables 298

Reflection/Parameter Ratio 11.45

Residuals: R1 ($I > 2.00\sigma(I)$) 0.0305

Residuals: R (All reflections) 0.0323

Residuals: wR2 (All reflections) 0.0770

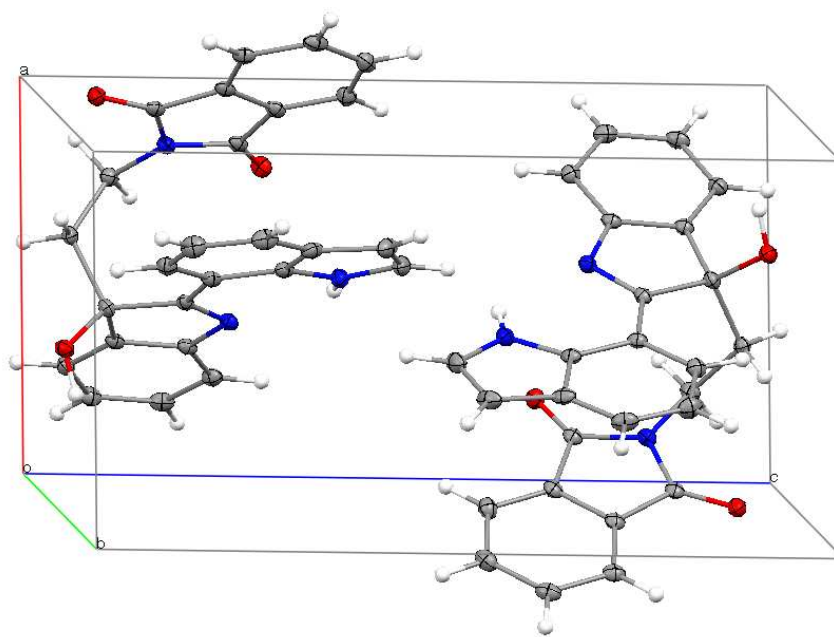
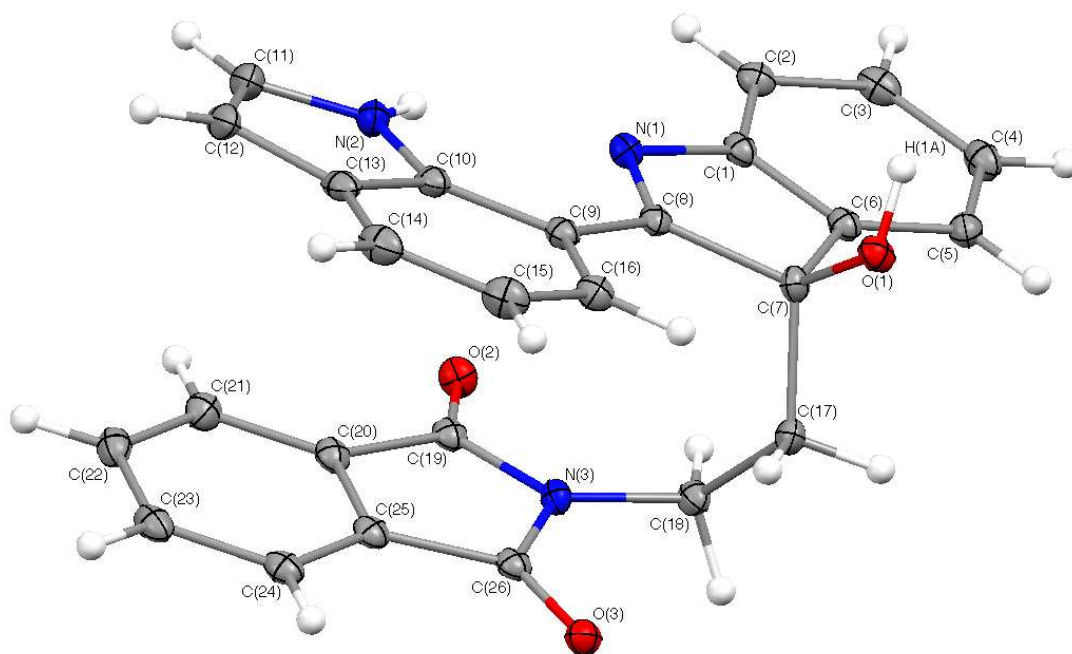
Goodness of Fit Indicator 1.080

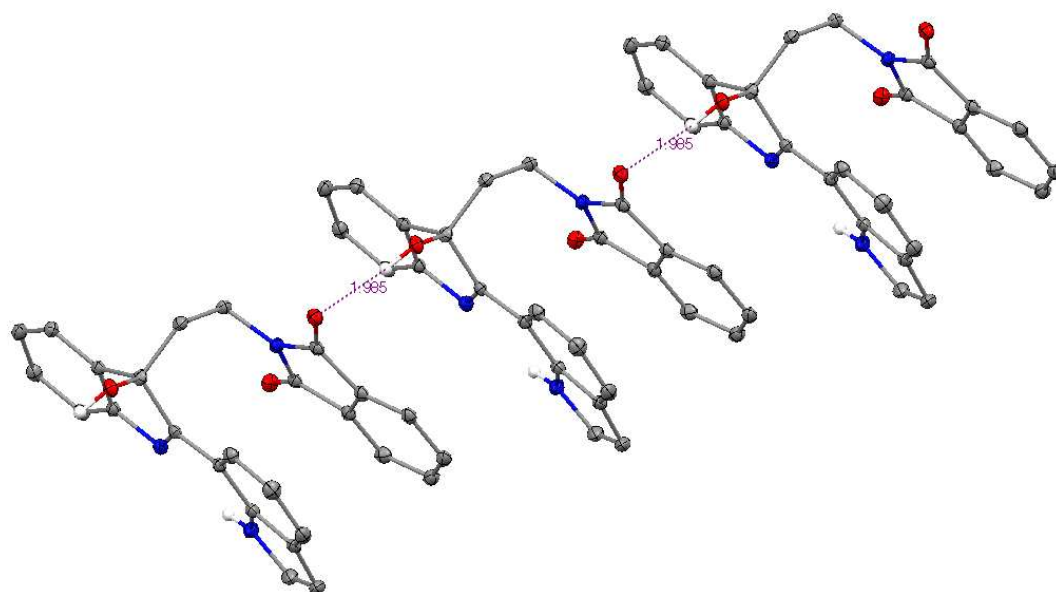
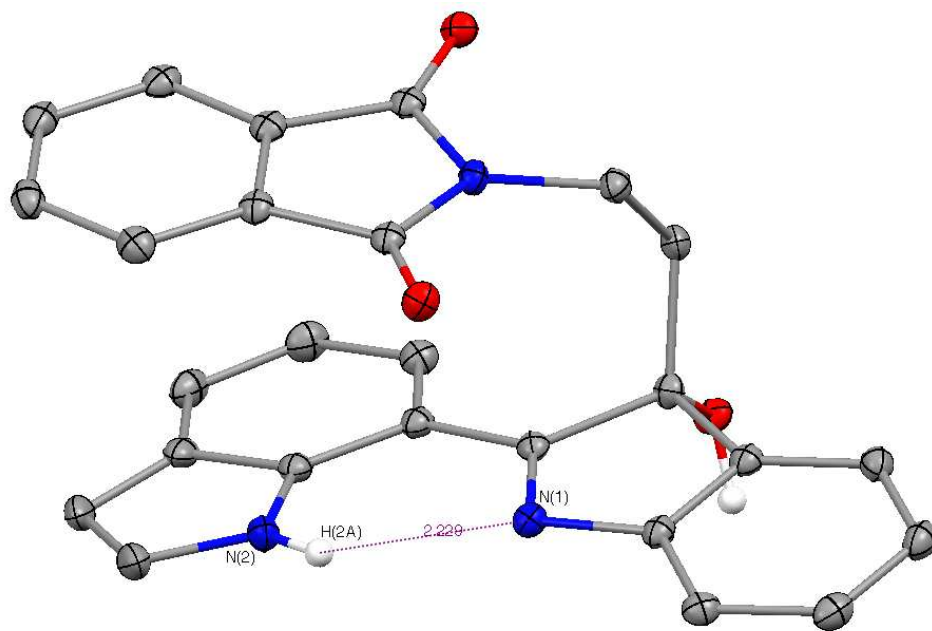
Flack Parameter 0.24(17)

Max Shift/Error in Final Cycle 0.001

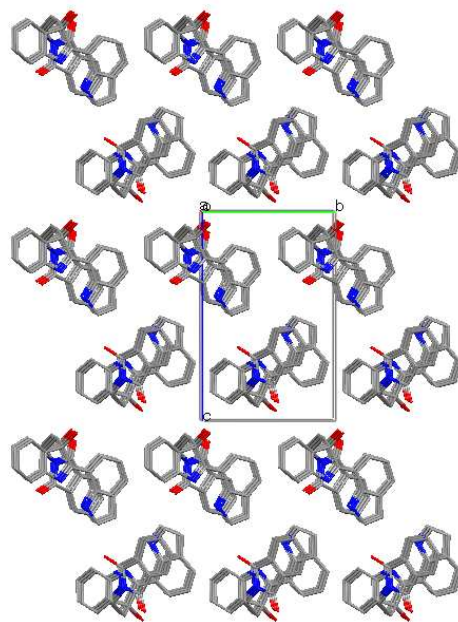
Maximum peak in Final Diff. Map $0.18 \text{ e}^-/\text{\AA}^3$

Minimum peak in Final Diff. Map $-0.16 \text{ e}^-/\text{\AA}^3$

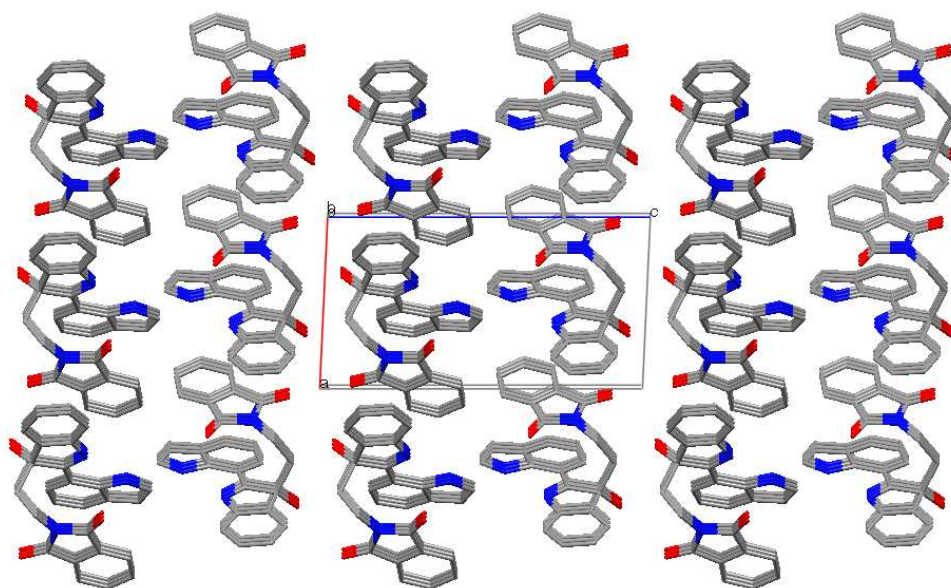




Packing diagram along the crystallographic a-axis



Packing diagram along the crystallographic b-axis



Packing diagram along the crystallographic c-axis

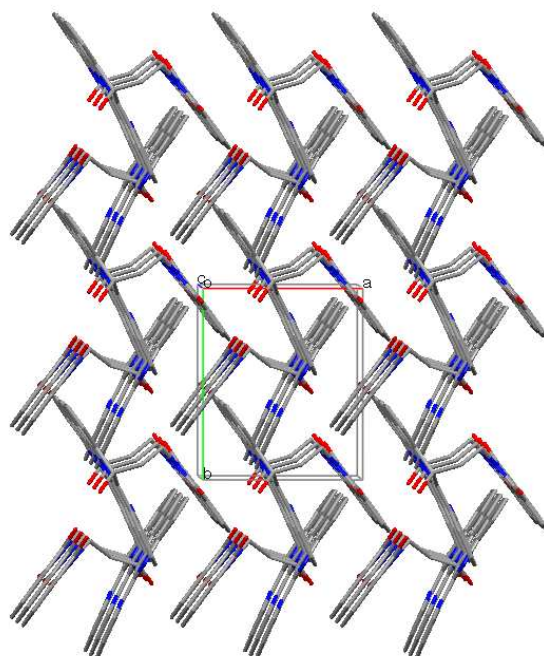


Table S3. Atomic coordinates and $B_{\text{iso}}/B_{\text{eq}}$

atom	x	y	z	B_{eq}
O(1)	0.67725(16)	0.54037(14)	0.94714(8)	1.86(2)
O(2)	0.25987(16)	0.29457(13)	0.65759(8)	1.94(2)
O(3)	0.04021(15)	0.54743(14)	0.90183(8)	1.90(2)
N(1)	0.62529(17)	0.43015(15)	0.71619(9)	1.55(2)
N(2)	0.4759(2)	0.62345(18)	0.58607(10)	1.88(2)
N(3)	0.18701(17)	0.40790(16)	0.79575(10)	1.53(2)
C(1)	0.7121(2)	0.30881(19)	0.76023(12)	1.58(2)
C(2)	0.8079(2)	0.20117(19)	0.71699(13)	1.79(3)
C(3)	0.8908(2)	0.0963(2)	0.77401(13)	2.03(3)
C(4)	0.8788(2)	0.0985(2)	0.87045(13)	2.14(3)

Table S3. Atomic coordinates and $B_{\text{iso}}/B_{\text{eq}}$ (continued)

C(5)	0.7831(2)	0.2073(2)	0.91289(12)	1.83(3)
C(6)	0.6999(2)	0.31174(19)	0.85742(12)	1.56(2)
C(7)	0.5913(2)	0.44398(19)	0.88092(11)	1.56(3)

C(8)	0.5630(2)	0.51060(18)	0.78191(11)	1.45(2)
C(9)	0.4767(2)	0.64975(19)	0.76040(12)	1.60(3)
C(10)	0.4430(2)	0.6947(2)	0.66736(12)	1.66(3)
C(11)	0.4259(2)	0.7110(2)	0.51166(13)	2.44(3)
C(12)	0.3579(2)	0.8380(2)	0.54324(14)	2.44(3)
C(13)	0.3679(2)	0.8319(2)	0.64334(13)	2.01(3)
C(14)	0.3238(2)	0.9278(2)	0.71549(14)	2.32(3)
C(15)	0.3525(2)	0.8845(2)	0.80680(14)	2.33(3)
C(16)	0.4273(2)	0.7480(2)	0.82939(13)	1.85(3)
C(17)	0.4182(2)	0.4009(2)	0.92477(11)	1.65(2)
C(18)	0.2862(2)	0.3148(2)	0.86318(12)	1.74(3)
C(19)	0.1846(2)	0.39121(19)	0.69811(12)	1.59(2)
C(20)	0.0738(2)	0.51268(19)	0.65939(12)	1.68(3)
C(21)	0.0312(2)	0.5514(2)	0.56793(12)	1.99(3)
C(22)	-0.0771(2)	0.6728(2)	0.55342(14)	2.12(3)
C(23)	-0.1394(2)	0.7516(2)	0.62804(13)	2.13(3)
C(24)	-0.0972(2)	0.71174(19)	0.72008(13)	1.85(3)
C(25)	0.0099(2)	0.59064(19)	0.73377(12)	1.66(3)
C(26)	0.0760(2)	0.51973(19)	0.82132(12)	1.61(3)

$$B_{\text{eq}} = 8/3 \pi^2 (U_{11}(\text{aa}^*)^2 + U_{22}(\text{bb}^*)^2 + U_{33}(\text{cc}^*)^2 + 2U_{12}(\text{aa}^*\text{bb}^*)\cos \gamma + 2U_{13}(\text{aa}^*\text{cc}^*)\cos \beta + 2U_{23}(\text{bb}^*\text{cc}^*)\cos \alpha)$$

Table S4. Atomic coordinates and B_{iso} involving hydrogens/ B_{eq}

atom	x	y	z	B_{eq}
H(1)	0.8166	0.1992	0.6511	2.15
H(2)	0.9573	0.0212	0.7463	2.43
H(3)	0.9364	0.0252	0.9076	2.57
H(4)	0.7752	0.2097	0.9789	2.20
H(5)	0.4372	0.6865	0.4478	2.93
H(6)	0.3121	0.9165	0.5060	2.93
H(7)	0.2748	1.0212	0.7013	2.79
H(8)	0.3211	0.9484	0.8556	2.80
H(9)	0.4448	0.7215	0.8933	2.21
H(10)	0.4471	0.3420	0.9814	1.98

H(11)	0.3602	0.4916	0.9453	1.98
H(12)	0.3494	0.2387	0.8287	2.09
H(13)	0.2029	0.2646	0.9034	2.09
H(14)	0.0741	0.4974	0.5170	2.39
H(15)	-0.1091	0.7023	0.4913	2.55
H(16)	-0.2124	0.8345	0.6159	2.55
H(17)	-0.1400	0.7652	0.7712	2.22
H(1A)	0.790(2)	0.552(2)	0.9342(13)	2.8(4)
H(2A)	0.522(2)	0.539(2)	0.5857(13)	2.6(4)

$$B_{\text{eq}} = 8/3 \pi^2 (U_{11}(\text{aa}^*)^2 + U_{22}(\text{bb}^*)^2 + U_{33}(\text{cc}^*)^2 + 2U_{12}(\text{aa}^*\text{bb}^*)\cos \gamma + 2U_{13}(\text{aa}^*\text{cc}^*)\cos \beta + 2U_{23}(\text{bb}^*\text{cc}^*)\cos \alpha)$$

Table S5. Anisotropic displacement parameters

atom	U ₁₁	U ₂₂	U ₃₃	U ₁₂	U ₁₃	U ₂₃		
O(1)	0.0206(6)		0.0266(6)		0.0232(6)	-0.0019(5)	-0.0013(5)	-0.0055(5)
O(2)	0.0265(6)		0.0209(6)		0.0260(6)	0.0019(5)	-0.0004(5)	-0.0042(5)
O(3)	0.0222(6)		0.0283(6)		0.0218(6)	-0.0020(5)	0.0008(5)	-0.0030(5)
N(1)	0.0193(7)		0.0180(7)		0.0215(7)	0.0001(6)	-0.0008(5)	-0.0021(6)
N(2)	0.0239(8)		0.0271(9)		0.0203(7)	0.0034(6)	0.0001(6)	0.0029(6)
N(3)	0.0197(7)		0.0168(7)		0.0214(7)	-0.0005(6)	-0.0026(5)	0.0007(6)
C(1)	0.0168(8)		0.0173(8)		0.0257(9)	-0.0016(6)	-0.0019(7)	0.0009(7)
C(2)	0.0217(8)		0.0202(9)		0.0261(9)	-0.0029(7)	0.0009(7)	-0.0033(7)
C(3)	0.0210(8)		0.0177(9)		0.0383(10)	-0.0002(7)	0.0008(7)	-0.0037(8)
C(4)	0.0229(9)		0.0213(9)		0.0369(10)	0.0015(7)	-0.0031(7)	0.0052(8)
C(5)	0.0216(8)		0.0245(9)		0.0233(9)	-0.0018(7)	-0.0015(7)	0.0034(8)
C(6)	0.0172(8)		0.0179(8)		0.0240(8)	-0.0024(6)	-0.0011(6)	0.0001(7)
C(7)	0.0208(8)		0.0209(9)		0.0175(8)	-0.0004(7)	-0.0028(6)	-0.0016(7)
C(8)	0.0159(8)		0.0185(8)		0.0205(8)	-0.0039(6)	-0.0016(6)	0.0006(7)
C(9)	0.0163(8)		0.0189(8)		0.0255(9)	-0.0017(6)	0.0006(7)	0.0011(7)
C(10)	0.0161(8)		0.0214(9)		0.0257(9)	-0.0022(7)	0.0020(6)	0.0027(8)
C(11)	0.0245(9)		0.0434(12)		0.0249(9)	0.0010(8)	0.0007(7)	0.0114(9)
C(12)	0.0229(9)		0.0345(11)		0.0355(10)	0.0026(8)	0.0021(8)	0.0187(9)
C(13)	0.0174(8)		0.0240(10)		0.0353(10)	-0.0010(7)	0.0054(7)	0.0090(8)
C(14)	0.0245(9)		0.0176(9)		0.0463(11)	0.0028(7)	0.0032(8)	0.0048(8)

C(15)	0.0295(10)	0.0200(9)	0.0393(11)	0.0021(7)	0.0046(8)	-0.0049(8)
C(16)	0.0230(8)	0.0210(9)	0.0260(9)	-0.0024(7)	0.0000(7)	-0.0029(8)
C(17)	0.0222(8)	0.0221(8)	0.0181(8)	0.0013(7)	-0.0005(6)	0.0037(7)
C(18)	0.0213(8)	0.0199(9)	0.0246(8)	-0.0008(7)	-0.0010(7)	0.0041(7)
C(19)	0.0178(8)	0.0180(9)	0.0242(8)	-0.0046(7)	-0.0016(6)	-0.0001(7)
C(20)	0.0193(8)	0.0192(9)	0.0250(9)	-0.0049(6)	-0.0034(7)	-0.0004(7)
C(21)	0.0270(9)	0.0247(9)	0.0238(9)	-0.0054(8)	-0.0015(7)	-0.0018(7)
C(22)	0.0271(9)	0.0240(10)	0.0288(9)	-0.0052(7)	-0.0058(7)	0.0065(8)
C(23)	0.0201(8)	0.0202(9)	0.0401(11)	-0.0027(7)	-0.0034(8)	0.0069(8)
C(24)	0.0191(8)	0.0184(9)	0.0328(10)	-0.0037(7)	0.0004(7)	-0.0013(8)
C(25)	0.0176(8)	0.0196(9)	0.0258(8)	-0.0055(6)	-0.0014(7)	-0.0018(7)
C(26)	0.0169(8)	0.0196(9)	0.0246(9)	-0.0053(6)	-0.0004(7)	-0.0007(7)

The general temperature factor expression: $\exp(-2\pi^2(a^{*2}U_{11}h^2 + b^{*2}U_{22}k^2 + c^{*2}U_{33}l^2 + 2a^*b^*U_{12}hk + 2a^*c^*U_{13}hl + 2b^*c^*U_{23}kl))$

Table S6. Bond lengths (Å)

atom	atom	distance	atom	atom	distance
O(1)	C(7)	1.431(2)	O(2)	C(19)	1.215(2)
O(3)	C(26)	1.224(2)	N(1)	C(1)	1.423(2)
N(1)	C(8)	1.301(2)	N(2)	C(10)	1.367(2)
N(2)	C(11)	1.372(2)	N(3)	C(18)	1.470(2)
N(3)	C(19)	1.406(2)	N(3)	C(26)	1.386(2)
C(1)	C(2)	1.387(2)	C(1)	C(6)	1.400(2)
C(2)	C(3)	1.392(2)	C(3)	C(4)	1.389(2)
C(4)	C(5)	1.389(2)	C(5)	C(6)	1.377(2)
C(6)	C(7)	1.511(2)	C(7)	C(8)	1.549(2)
C(7)	C(17)	1.539(2)	C(8)	C(9)	1.457(2)
C(9)	C(10)	1.407(2)	C(9)	C(16)	1.399(2)
C(10)	C(13)	1.414(2)	C(11)	C(12)	1.356(2)
C(12)	C(13)	1.433(2)	C(13)	C(14)	1.407(2)
C(14)	C(15)	1.374(2)	C(15)	C(16)	1.402(2)
C(17)	C(18)	1.526(2)	C(19)	C(20)	1.486(2)
C(20)	C(21)	1.381(2)	C(20)	C(25)	1.388(2)
C(21)	C(22)	1.393(2)	C(22)	C(23)	1.390(2)
C(23)	C(24)	1.391(2)	C(24)	C(25)	1.384(2)

C(25) C(26) 1.479(2)

Table S7. Bond lengths involving hydrogens (Å)

atom	atom	distance	atom	atom	distance
O(1)	H(1A)	0.89(2)	N(2)	H(2A)	0.85(2)
C(2)	H(1)	0.950	C(3)	H(2)	0.950
C(4)	H(3)	0.950	C(5)	H(4)	0.950
C(11)	H(5)	0.950	C(12)	H(6)	0.950
C(14)	H(7)	0.950	C(15)	H(8)	0.950
C(16)	H(9)	0.950	C(17)	H(10)	0.990
C(17)	H(11)	0.990	C(18)	H(12)	0.990
C(18)	H(13)	0.990	C(21)	H(14)	0.950
C(22)	H(15)	0.950	C(23)	H(16)	0.950
C(24)	H(17)	0.950			

Table S8. Bond angles (°)

atom	atom	atom	angle	atom	atom	atom	angle
C(1)	N(1)	C(8)	107.27(13)	C(10)	N(2)	C(11)	109.20(15)
C(18)	N(3)	C(19)	125.01(14)	C(18)	N(3)	C(26)	123.67(14)
C(19)	N(3)	C(26)	111.24(13)	N(1)	C(1)	C(2)	126.69(15)
N(1)	C(1)	C(6)	112.03(14)	C(2)	C(1)	C(6)	121.19(15)
C(1)	C(2)	C(3)	117.39(16)	C(2)	C(3)	C(4)	121.64(16)
C(3)	C(4)	C(5)	120.38(16)	C(4)	C(5)	C(6)	118.69(16)
C(1)	C(6)	C(5)	120.71(15)	C(1)	C(6)	C(7)	107.40(14)
C(5)	C(6)	C(7)	131.87(15)	O(1)	C(7)	C(6)	113.60(13)
O(1)	C(7)	C(8)	113.94(13)	O(1)	C(7)	C(17)	105.19(12)
C(6)	C(7)	C(8)	99.48(12)	C(6)	C(7)	C(17)	112.08(14)
C(8)	C(7)	C(17)	112.81(13)	N(1)	C(8)	C(7)	113.60(13)
N(1)	C(8)	C(9)	121.04(14)	C(7)	C(8)	C(9)	125.35(14)
C(8)	C(9)	C(10)	121.04(15)	C(8)	C(9)	C(16)	122.92(15)
C(10)	C(9)	C(16)	116.01(15)	N(2)	C(10)	C(9)	129.45(16)
N(2)	C(10)	C(13)	107.61(15)	C(9)	C(10)	C(13)	122.92(16)
N(2)	C(11)	C(12)	109.64(16)	C(11)	C(12)	C(13)	107.24(17)

C(10)	C(13)	C(12)	106.30(16)	C(10)	C(13)	C(14)	118.73(16)
C(12)	C(13)	C(14)	134.97(17)	C(13)	C(14)	C(15)	119.15(17)
C(14)	C(15)	C(16)	121.36(18)	C(9)	C(16)	C(15)	121.79(17)
C(7)	C(17)	C(18)	116.84(13)	N(3)	C(18)	C(17)	112.82(14)
O(2)	C(19)	N(3)	124.67(15)	O(2)	C(19)	C(20)	129.57(15)
N(3)	C(19)	C(20)	105.76(14)	C(19)	C(20)	C(21)	130.52(16)
C(19)	C(20)	C(25)	108.05(14)	C(21)	C(20)	C(25)	121.43(16)
C(20)	C(21)	C(22)	117.20(16)	C(21)	C(22)	C(23)	121.26(17)
C(22)	C(23)	C(24)	121.36(17)	C(23)	C(24)	C(25)	116.94(16)
C(20)	C(25)	C(24)	121.79(16)	C(20)	C(25)	C(26)	107.96(14)
C(24)	C(25)	C(26)	130.24(16)	O(3)	C(26)	N(3)	124.51(15)
O(3)	C(26)	C(25)	128.79(15)	N(3)	C(26)	C(25)	106.67(14)

Table S9. Bond angles involving hydrogens (°)

atom	atom	atom	angle	atom	atom	atom	angle
C(7)	O(1)	H(1A)	110.4(13)	C(10)	N(2)	H(2A)	122.0(13)
C(11)	N(2)	H(2A)	128.8(13)	C(1)	C(2)	H(1)	121.3
C(3)	C(2)	H(1)	121.3	C(2)	C(3)	H(2)	119.2
C(4)	C(3)	H(2)	119.2	C(3)	C(4)	H(3)	119.8
C(5)	C(4)	H(3)	119.8	C(4)	C(5)	H(4)	120.7
C(6)	C(5)	H(4)	120.7	N(2)	C(11)	H(5)	125.2
C(12)	C(11)	H(5)	125.2	C(11)	C(12)	H(6)	126.4
C(13)	C(12)	H(6)	126.4	C(13)	C(14)	H(7)	120.4
C(15)	C(14)	H(7)	120.4	C(14)	C(15)	H(8)	119.3
C(16)	C(15)	H(8)	119.3	C(9)	C(16)	H(9)	119.1
C(15)	C(16)	H(9)	119.1	C(7)	C(17)	H(10)	108.1
C(7)	C(17)	H(11)	108.1	C(18)	C(17)	H(10)	108.1
C(18)	C(17)	H(11)	108.1	H(10)	C(17)	H(11)	107.3
N(3)	C(18)	H(12)	109.0	N(3)	C(18)	H(13)	109.0
C(17)	C(18)	H(12)	109.0	C(17)	C(18)	H(13)	109.0
H(12)	C(18)	H(13)	107.8	C(20)	C(21)	H(14)	121.4
C(22)	C(21)	H(14)	121.4	C(21)	C(22)	H(15)	119.4
C(23)	C(22)	H(15)	119.4	C(22)	C(23)	H(16)	119.3

C(24)	C(23)	H(16)	119.3	C(23)	C(24)	H(17)	121.5
C(25)	C(24)	H(17)	121.5				

Table S10. Torsion Angles(°)

atom1	atom2	atom3	atom4	angle	atom1	atom2	atom3	atom4	angle
C(1)	N(1)	C(8)	C(7)	-4.10(18)	C(1)	N(1)	C(8)	C(9)	175.61(14)
C(8)	N(1)	C(1)	C(2)	-175.08(16)	C(8)	N(1)	C(1)	C(6)	1.49(18)
C(10)	N(2)	C(11)	C(12)	-1.0(2)	C(11)	N(2)	C(10)	C(9)	-178.15(17)
C(11)	N(2)	C(10)	C(13)	0.58(19)	C(18)	N(3)	C(19)	O(2)	2.5(2)
C(18)	N(3)	C(19)	C(20)	-177.70(14)	C(19)	N(3)	C(18)	C(17)	120.80(16)
C(18)	N(3)	C(26)	O(3)	-4.6(2)	C(18)	N(3)	C(26)	C(25)	177.11(14)
C(26)	N(3)	C(18)	C(17)	-62.6(2)	C(19)	N(3)	C(26)	O(3)	172.35(15)
C(19)	N(3)	C(26)	C(25)	-5.91(18)	C(26)	N(3)	C(19)	O(2)	-174.42(16)
C(26)	N(3)	C(19)	C(20)	5.37(18)	N(1)	C(1)	C(2)	C(3)	176.47(15)
N(1)	C(1)	C(6)	C(5)	-176.80(14)	N(1)	C(1)	C(6)	C(7)	1.69(18)
C(2)	C(1)	C(6)	C(5)	-0.0(2)	C(2)	C(1)	C(6)	C(7)	178.48(15)
C(6)	C(1)	C(2)	C(3)	0.2(2)	C(1)	C(2)	C(3)	C(4)	-0.04(19)
C(2)	C(3)	C(4)	C(5)	-0.3(2)	C(3)	C(4)	C(5)	C(6)	0.4(2)
C(4)	C(5)	C(6)	C(1)	-0.3(2)	C(4)	C(5)	C(6)	C(7)	-178.36(17)
C(1)	C(6)	C(7)	O(1)	-125.05(14)	C(1)	C(6)	C(7)	C(8)	-3.59(16)
C(1)	C(6)	C(7)	C(17)	115.88(14)	C(5)	C(6)	C(7)	O(1)	53.2(2)
C(5)	C(6)	C(7)	C(8)	174.67(17)	C(5)	C(6)	C(7)	C(17)	-65.9(2)
O(1)	C(7)	C(8)	N(1)	126.09(15)	O(1)	C(7)	C(8)	C(9)	-53.6(2)
O(1)	C(7)	C(17)	C(18)	171.62(13)	C(6)	C(7)	C(8)	N(1)	4.87(17)
C(6)	C(7)	C(8)	C(9)	-174.81(15)	C(6)	C(7)	C(17)	C(18)	-64.47(18)
C(8)	C(7)	C(17)	C(18)	46.8(2)	C(17)	C(7)	C(8)	N(1)	-114.05(16)
C(17)	C(7)	C(8)	C(9)	66.3(2)	N(1)	C(8)	C(9)	C(10)	6.2(2)
N(1)	C(8)	C(9)	C(16)	-171.88(15)	C(7)	C(8)	C(9)	C(10)	-174.14(15)
C(7)	C(8)	C(9)	C(16)	7.8(2)	C(8)	C(9)	C(10)	N(2)	1.9(2)
C(8)	C(9)	C(10)	C(13)	-176.68(15)	C(8)	C(9)	C(16)	C(15)	176.69(16)
C(10)	C(9)	C(16)	C(15)	-1.5(2)	C(16)	C(9)	C(10)	N(2)	-179.92(16)
C(16)	C(9)	C(10)	C(13)	1.5(2)	N(2)	C(10)	C(13)	C(12)	0.03(14)
N(2)	C(10)	C(13)	C(14)	-179.16(15)	C(9)	C(10)	C(13)	C(12)	178.86(15)

C(9)	C(10)	C(13)	C(14)	-0.3(2)	N(2)	C(11)	C(12)	C(13)	1.0(2)
C(11)	C(12)	C(13)	C(10)	-0.63(19)	C(11)	C(12)	C(13)	C(14)	178.36(19)
C(10)	C(13)	C(14)	C(15)	-1.0(2)	C(12)	C(13)	C(14)	C(15)	-179.87(19)
C(13)	C(14)	C(15)	C(16)	1.0(2)	C(14)	C(15)	C(16)	C(9)	0.2(2)
C(7)	C(17)	C(18)	N(3)	-78.36(18)	O(2)	C(19)	C(20)	C(21)	-2.4(3)
O(2)	C(19)	C(20)	C(25)	177.15(17)	N(3)	C(19)	C(20)	C(21)	177.87(17)
N(3)	C(19)	C(20)	C(25)	-2.63(18)	C(19)	C(20)	C(21)	C(22)	-179.95(13)
C(19)	C(20)	C(25)	C(24)	179.56(15)	C(19)	C(20)	C(25)	C(26)	-0.84(18)
C(21)	C(20)	C(25)	C(24)	-0.9(2)	C(21)	C(20)	C(25)	C(26)	178.72(15)

Table S11. Torsion angles (°) (continued)

atom1	atom2	atom3	atom4	angle	atom1	atom2	atom3	atom4	angle
C(25)	C(20)	C(21)	C(22)	0.6(2)	C(20)	C(21)	C(22)	C(23)	0.1(2)
C(21)	C(22)	C(23)	C(24)	-0.5(2)	C(22)	C(23)	C(24)	C(25)	0.2(2)
C(23)	C(24)	C(25)	C(20)	0.4(2)	C(23)	C(24)	C(25)	C(26)	-179.07(17)
C(20)	C(25)	C(26)	O(3)	-174.09(17)	C(20)	C(25)	C(26)	N(3)	4.07(18)
C(24)	C(25)	C(26)	O(3)	5.5(3)	C(24)	C(25)	C(26)	N(3)	-176.38(17)

The sign is positive if when looking from atom 2 to atom 3 a clock-wise motion of atom 1 would superimpose it on atom 4.

Table S12. Distances beyond the asymmetric unit out to 3.60 Å

atom	atom	distance	atom	atom	distance
O(1)	O(3) ¹⁾	2.8727(16)	O(1)	N(1)	3.4629(17)
O(1)	C(1)	3.431(2)	O(1)	C(5)	3.190(2)
O(1)	C(6)	2.462(2)	O(1)	C(8)	2.4988(19)
O(1)	C(9)	3.180(2)	O(1)	C(16)	3.127(2)
O(1)	C(17)	2.360(2)	O(2)	N(1)	3.1286(17)
O(2)	N(2)	3.597(2)	O(2)	N(3)	2.3230(18)
O(2)	C(8)	3.4681(19)	O(2)	C(11) ²⁾	3.569(2)
O(2)	C(14) ³⁾	3.479(2)	O(2)	C(18)	2.948(2)
O(2)	C(20)	2.446(2)	O(2)	C(21)	3.159(2)
O(2)	C(22) ⁴⁾	3.453(2)	O(2)	C(25)	3.510(2)
O(2)	C(26)	3.463(2)	O(3)	O(1) ⁵⁾	2.8727(16)

O(3)	N(3)	2.3111(18)	O(3)	C(4) ⁶⁾	3.324(2)
O(3)	C(5) ⁶⁾	3.265(2)	O(3)	C(6) ⁵⁾	3.409(2)
O(3)	C(7) ⁵⁾	3.551(2)	O(3)	C(17)	3.181(2)
O(3)	C(18)	2.904(2)	O(3)	C(19)	3.472(2)
O(3)	C(20)	3.508(2)	O(3)	C(24)	3.142(2)
O(3)	C(25)	2.440(2)	N(1)	O(1)	3.4629(17)
N(1)	O(2)	3.1286(17)	N(1)	N(2)	2.773(2)
N(1)	N(3)	3.5863(18)	N(1)	C(2)	2.512(2)
N(1)	C(6)	2.341(2)	N(1)	C(7)	2.389(2)
N(1)	C(9)	2.402(2)	N(1)	C(10)	2.858(2)
N(1)	C(17)	3.453(2)	N(1)	C(18)	3.568(2)
N(1)	C(19)	3.378(2)	N(1)	C(24) ¹⁾	3.329(2)
N(1)	C(25) ¹⁾	3.278(2)	N(2)	O(2)	3.597(2)
N(2)	N(1)	2.773(2)	N(2)	C(8)	3.033(2)
N(2)	C(9)	2.508(2)	N(2)	C(12)	2.231(2)
N(2)	C(12) ²⁾	3.471(2)	N(2)	C(13)	2.244(2)
N(2)	C(14)	3.564(2)	N(2)	C(19)	3.512(2)
N(2)	C(20)	3.439(2)	N(2)	C(21)	3.452(2)
N(2)	C(22) ¹⁾	3.491(2)	N(2)	C(23) ¹⁾	3.190(2)
N(3)	O(2)	2.3230(18)	N(3)	O(3)	2.3111(18)
N(3)	N(1)	3.5863(18)	N(3)	C(2) ⁵⁾	3.591(2)
N(3)	C(7)	3.280(2)	N(3)	C(8)	3.032(2)
N(3)	C(9)	3.181(2)	N(3)	C(17)	2.496(2)
N(3)	C(20)	2.307(2)	N(3)	C(25)	2.299(2)
C(1)	O(1)	3.431(2)	C(1)	C(3)	2.374(2)
C(1)	C(4)	2.759(2)	C(1)	C(5)	2.413(2)

Table S12. Distances beyond the asymmetric unit out to 3.60 Å (continued)

atom	atom	distance	atom	atom	distance
C(1)	C(7)	2.347(2)	C(1)	C(8)	2.195(2)
C(1)	C(9)	3.594(2)	C(1)	C(17)	3.431(2)
C(1)	C(25) ¹⁾	3.465(2)	C(1)	C(26) ¹⁾	3.457(2)
C(2)	N(1)	2.512(2)	C(2)	N(3) ¹⁾	3.591(2)

C(2)	C(4)	2.428(2)	C(2)	C(5)	2.821(2)
C(2)	C(6)	2.428(2)	C(2)	C(8)	3.535(2)
C(2)	C(19) ¹⁾	3.377(2)	C(3)	C(1)	2.374(2)
C(3)	C(5)	2.410(2)	C(3)	C(6)	2.753(2)
C(3)	C(24) ⁷⁾	3.597(2)	C(4)	O(3) ⁸⁾	3.324(2)
C(4)	C(1)	2.759(2)	C(4)	C(2)	2.428(2)
C(4)	C(6)	2.380(2)	C(5)	O(1)	3.190(2)
C(5)	O(3) ⁸⁾	3.265(2)	C(5)	C(1)	2.413(2)
C(5)	C(2)	2.821(2)	C(5)	C(3)	2.410(2)
C(5)	C(7)	2.637(2)	C(5)	C(17)	3.308(2)
C(6)	O(1)	2.462(2)	C(6)	O(3) ¹⁾	3.409(2)
C(6)	N(1)	2.341(2)	C(6)	C(2)	2.428(2)
C(6)	C(3)	2.753(2)	C(6)	C(4)	2.380(2)
C(6)	C(8)	2.335(2)	C(6)	C(17)	2.530(2)
C(6)	C(18)	3.160(2)	C(6)	C(26) ¹⁾	3.498(2)
C(7)	O(3) ¹⁾	3.551(2)	C(7)	N(1)	2.389(2)
C(7)	N(3)	3.280(2)	C(7)	C(1)	2.347(2)
C(7)	C(5)	2.637(2)	C(7)	C(9)	2.671(2)
C(7)	C(16)	3.120(2)	C(7)	C(18)	2.611(2)
C(8)	O(1)	2.4988(19)	C(8)	O(2)	3.4681(19)
C(8)	N(2)	3.033(2)	C(8)	N(3)	3.032(2)
C(8)	C(1)	2.195(2)	C(8)	C(2)	3.535(2)
C(8)	C(6)	2.335(2)	C(8)	C(10)	2.493(2)
C(8)	C(16)	2.509(2)	C(8)	C(17)	2.572(2)
C(8)	C(18)	3.039(2)	C(8)	C(19)	3.262(2)
C(8)	C(24) ¹⁾	3.329(2)	C(8)	C(25) ¹⁾	3.584(2)
C(9)	O(1)	3.180(2)	C(9)	N(1)	2.402(2)
C(9)	N(2)	2.508(2)	C(9)	N(3)	3.181(2)
C(9)	C(1)	3.594(2)	C(9)	C(7)	2.671(2)
C(9)	C(13)	2.478(2)	C(9)	C(14)	2.855(2)
C(9)	C(15)	2.448(2)	C(9)	C(17)	3.317(2)
C(9)	C(19)	3.339(2)	C(9)	C(20)	3.563(2)
C(9)	C(24) ¹⁾	3.374(2)	C(9)	C(26)	3.428(2)

Table S12. Distances beyond the asymmetric unit out to 3.60 Å (continued)

atom	atom	distance	atom	atom	distance
C(10)	N(1)	2.858(2)	C(10)	C(8)	2.493(2)
C(10)	C(11)	2.233(2)	C(10)	C(12)	2.278(2)
C(10)	C(14)	2.427(2)	C(10)	C(15)	2.756(2)
C(10)	C(16)	2.380(2)	C(10)	C(19)	3.441(2)
C(10)	C(20)	3.267(2)	C(10)	C(23) ¹⁾	3.300(2)
C(10)	C(24) ¹⁾	3.556(2)	C(11)	O(2) ⁹⁾	3.569(2)
C(11)	C(10)	2.233(2)	C(11)	C(13)	2.247(2)
C(11)	C(21)	3.472(2)	C(12)	N(2)	2.231(2)
C(12)	N(2) ⁹⁾	3.471(2)	C(12)	C(10)	2.278(2)
C(12)	C(14)	2.624(2)	C(13)	N(2)	2.244(2)
C(13)	C(9)	2.478(2)	C(13)	C(11)	2.247(2)
C(13)	C(15)	2.398(2)	C(13)	C(16)	2.790(2)
C(14)	O(2) ¹⁰⁾	3.479(2)	C(14)	N(2)	3.564(2)
C(14)	C(9)	2.855(2)	C(14)	C(10)	2.427(2)
C(14)	C(12)	2.624(2)	C(14)	C(16)	2.421(2)
C(15)	C(9)	2.448(2)	C(15)	C(10)	2.756(2)
C(15)	C(13)	2.398(2)	C(16)	O(1)	3.127(2)
C(16)	C(7)	3.120(2)	C(16)	C(8)	2.509(2)
C(16)	C(10)	2.380(2)	C(16)	C(13)	2.790(2)
C(16)	C(14)	2.421(2)	C(16)	C(17)	3.453(2)
C(16)	C(26)	3.392(2)	C(17)	O(1)	2.360(2)
C(17)	O(3)	3.181(2)	C(17)	N(1)	3.453(2)
C(17)	N(3)	2.496(2)	C(17)	C(1)	3.431(2)
C(17)	C(5)	3.308(2)	C(17)	C(6)	2.530(2)
C(17)	C(8)	2.572(2)	C(17)	C(9)	3.317(2)
C(17)	C(16)	3.453(2)	C(17)	C(26)	3.135(2)
C(18)	O(2)	2.948(2)	C(18)	O(3)	2.904(2)
C(18)	N(1)	3.568(2)	C(18)	C(6)	3.160(2)
C(18)	C(7)	2.611(2)	C(18)	C(8)	3.039(2)
C(18)	C(19)	2.551(2)	C(18)	C(26)	2.518(2)
C(19)	O(3)	3.472(2)	C(19)	N(1)	3.378(2)

C(19)	N(2)	3.512(2)	C(19)	C(2) ⁵⁾	3.377(2)
C(19)	C(8)	3.262(2)	C(19)	C(9)	3.339(2)
C(19)	C(10)	3.441(2)	C(19)	C(18)	2.551(2)
C(19)	C(21)	2.604(2)	C(19)	C(25)	2.327(2)
C(19)	C(26)	2.305(2)	C(20)	O(2)	2.446(2)
C(20)	O(3)	3.508(2)	C(20)	N(2)	3.439(2)

Table S12. Distances beyond the asymmetric unit out to 3.60 Å (continued)

atom	atom	distance	atom	atom	distance
C(20)	N(3)	2.307(2)	C(20)	C(9)	3.563(2)
C(20)	C(10)	3.267(2)	C(20)	C(22)	2.367(2)
C(20)	C(23)	2.746(2)	C(20)	C(24)	2.422(2)
C(20)	C(26)	2.319(2)	C(21)	O(2)	3.159(2)
C(21)	N(2)	3.452(2)	C(21)	C(11)	3.472(2)
C(21)	C(19)	2.604(2)	C(21)	C(23)	2.425(2)
C(21)	C(24)	2.837(2)	C(21)	C(25)	2.415(2)
C(22)	O(2) ¹¹⁾	3.453(2)	C(22)	N(2) ⁵⁾	3.491(2)
C(22)	C(20)	2.367(2)	C(22)	C(24)	2.425(2)
C(22)	C(25)	2.742(2)	C(23)	N(2) ⁵⁾	3.190(2)
C(23)	C(10) ⁵⁾	3.300(2)	C(23)	C(20)	2.746(2)
C(23)	C(21)	2.425(2)	C(23)	C(25)	2.365(2)
C(24)	O(3)	3.142(2)	C(24)	N(1) ⁵⁾	3.329(2)
C(24)	C(3) ¹²⁾	3.597(2)	C(24)	C(8) ⁵⁾	3.329(2)
C(24)	C(9) ⁵⁾	3.374(2)	C(24)	C(10) ⁵⁾	3.556(2)
C(24)	C(20)	2.422(2)	C(24)	C(21)	2.837(2)
C(24)	C(22)	2.425(2)	C(24)	C(26)	2.597(2)
C(25)	O(2)	3.510(2)	C(25)	O(3)	2.440(2)
C(25)	N(1) ⁵⁾	3.278(2)	C(25)	N(3)	2.299(2)
C(25)	C(1) ⁵⁾	3.465(2)	C(25)	C(8) ⁵⁾	3.584(2)
C(25)	C(19)	2.327(2)	C(25)	C(21)	2.415(2)
C(25)	C(22)	2.742(2)	C(25)	C(23)	2.365(2)
C(26)	O(2)	3.463(2)	C(26)	C(1) ⁵⁾	3.457(2)
C(26)	C(6) ⁵⁾	3.498(2)	C(26)	C(9)	3.428(2)
C(26)	C(16)	3.392(2)	C(26)	C(17)	3.135(2)

C(26) C(18) 2.518(2) C(26) C(19) 2.305(2)

C(26) C(20) 2.319(2) C(26) C(24) 2.597(2)

Symmetry Operators:

(1) X+1,Y,Z (2) -X+1,Y+1/2-1,-Z+1

(3) X,Y-1,Z (4) -X,Y+1/2-1,-Z+1

(5) X-1,Y,Z (6) -X+1,Y+1/2,-Z+2

(7) X+1,Y-1,Z (8) -X+1,Y+1/2-1,-Z+2

(9) -X+1,Y+1/2,-Z+1 (10) X,Y+1,Z

(11) -X,Y+1/2,-Z+1 (12) X-1,Y+1,Z

Table S13. Distances beyond the asymmetric unit out to 3.60 Å involving hydrogens

atom	atom	distance	atom	atom	distance
O(1)	H(3) ¹⁾	3.531	O(1)	H(4)	3.139
O(1)	H(8) ²⁾	2.947	O(1)	H(9)	2.519
O(1)	H(10)	2.585	O(1)	H(10) ³⁾	3.102
O(1)	H(11)	2.457	O(1)	H(13) ³⁾	3.070
O(1)	H(17) ⁴⁾	3.582	O(2)	H(1) ⁵⁾	3.488
O(2)	H(5) ⁶⁾	2.985	O(2)	H(7) ⁷⁾	2.574
O(2)	H(12)	2.565	O(2)	H(13)	3.578
O(2)	H(14)	3.038	O(2)	H(15) ⁸⁾	2.521
O(2)	H(2A)	3.20(2)	O(3)	H(3) ³⁾	2.734
O(3)	H(4) ³⁾	2.622	O(3)	H(9)	3.478
O(3)	H(11)	2.542	O(3)	H(13)	2.865
O(3)	H(17)	3.018	O(3)	H(1A) ⁵⁾	1.99(2)
N(1)	H(1)	2.752	N(1)	H(5) ⁶⁾	3.254
N(1)	H(6) ⁶⁾	3.241	N(1)	H(12)	3.224
N(1)	H(1A)	3.492(19)	N(1)	H(2A)	2.23(2)
N(2)	H(5)	2.071	N(2)	H(6)	3.147
N(2)	H(6) ⁶⁾	2.849	N(2)	H(14)	3.378
N(2)	H(15) ⁴⁾	3.574	N(2)	H(16) ⁴⁾	3.075
N(3)	H(10)	3.299	N(3)	H(11)	2.580
N(3)	H(12)	2.022	N(3)	H(13)	2.022
C(1)	H(1)	2.049	C(1)	H(2)	3.235

C(1)	H(4)	3.274	C(1)	H(5) ⁶⁾	3.333
C(1)	H(12)	3.045	C(1)	H(1A)	3.37(2)
C(1)	H(2A)	3.52(2)	C(2)	H(2)	2.032
C(2)	H(3)	3.279	C(2)	H(5) ⁶⁾	2.946
C(3)	H(1)	2.053	C(3)	H(3)	2.035
C(3)	H(4)	3.271	C(3)	H(7) ⁹⁾	3.226
C(3)	H(13) ⁴⁾	3.325	C(3)	H(16) ⁹⁾	3.361
C(3)	H(17) ⁹⁾	3.032	C(4)	H(1)	3.287
C(4)	H(2)	2.029	C(4)	H(4)	2.044
C(4)	H(11) ²⁾	3.418	C(4)	H(13) ⁴⁾	2.919
C(4)	H(17) ⁹⁾	3.360	C(5)	H(2)	3.262
C(5)	H(3)	2.036	C(5)	H(9) ²⁾	3.345
C(5)	H(10)	3.045	C(5)	H(11) ²⁾	3.067
C(5)	H(12)	3.480	C(5)	H(13) ⁴⁾	3.252
C(5)	H(1A)	3.16(2)	C(6)	H(1)	3.290
C(6)	H(3)	3.239	C(6)	H(4)	2.033

Table S13. Distances beyond the asymmetric unit out to 3.60 Å involving hydrogens (continued)

atom	atom	distance	atom	atom	distance
C(6)	H(10)	2.695	C(6)	H(11)	3.361
C(6)	H(12)	2.767	C(6)	H(1A)	2.53(2)
C(7)	H(4)	2.886	C(7)	H(9)	2.779
C(7)	H(10)	2.072	C(7)	H(11)	2.072
C(7)	H(12)	2.711	C(7)	H(13)	3.414
C(7)	H(1A)	1.93(2)	C(8)	H(9)	2.683
C(8)	H(10)	3.398	C(8)	H(11)	2.868
C(8)	H(12)	3.060	C(8)	H(17) ⁴⁾	3.254
C(8)	H(1A)	2.75(2)	C(8)	H(2A)	2.824(19)
C(9)	H(8)	3.295	C(9)	H(9)	2.038
C(9)	H(11)	3.178	C(9)	H(17) ⁴⁾	3.105
C(9)	H(1A)	3.49(2)	C(9)	H(2A)	2.74(2)
C(10)	H(5)	3.144	C(10)	H(6)	3.198
C(10)	H(7)	3.291	C(10)	H(9)	3.244

C(10)	H(16) ⁴⁾	3.042	C(10)	H(17) ⁴⁾	3.508
C(10)	H(2A)	1.96(2)	C(11)	H(1) ¹⁰⁾	2.910
C(11)	H(6)	2.067	C(11)	H(6) ⁶⁾	3.367
C(11)	H(14)	3.321	C(11)	H(15) ⁴⁾	3.573
C(11)	H(16) ⁴⁾	3.276	C(11)	H(2A)	2.02(2)
C(11)	H(2A) ¹⁰⁾	3.33(2)	C(12)	H(1) ¹⁰⁾	3.284
C(12)	H(5)	2.056	C(12)	H(5) ¹⁰⁾	3.544
C(12)	H(7)	2.907	C(12)	H(16) ⁴⁾	3.393
C(12)	H(2A)	3.06(2)	C(12)	H(2A) ¹⁰⁾	2.79(2)
C(13)	H(5)	3.166	C(13)	H(6)	2.138
C(13)	H(7)	2.058	C(13)	H(8)	3.256
C(13)	H(16) ⁴⁾	3.243	C(13)	H(2A)	3.05(2)
C(14)	H(2) ¹¹⁾	2.975	C(14)	H(6)	3.000
C(14)	H(8)	2.017	C(14)	H(9)	3.266
C(14)	H(12) ¹²⁾	3.271	C(15)	H(2) ¹¹⁾	3.340
C(15)	H(7)	2.028	C(15)	H(9)	2.041
C(15)	H(10) ³⁾	3.355	C(15)	H(12) ¹²⁾	3.250
C(16)	H(4) ³⁾	3.231	C(16)	H(7)	3.279
C(16)	H(8)	2.043	C(16)	H(10) ³⁾	2.959
C(16)	H(11)	2.928	C(16)	H(17) ⁴⁾	3.444
C(16)	H(1A)	3.57(2)	C(17)	H(4)	3.297
C(17)	H(4) ³⁾	3.495	C(17)	H(9)	2.971
C(17)	H(9) ²⁾	3.212	C(17)	H(12)	2.072

Table S13. Distances beyond the asymmetric unit out to 3.60 Å involving hydrogens (continued)

atom	atom	distance	atom	atom	distance
C(17)	H(13)	2.072	C(17)	H(1A)	3.15(2)
C(18)	H(7) ⁷⁾	3.543	C(18)	H(8) ⁷⁾	3.358
C(18)	H(10)	2.061	C(18)	H(11)	2.061
C(19)	H(1) ⁵⁾	3.351	C(19)	H(7) ⁷⁾	3.447
C(19)	H(12)	2.608	C(19)	H(13)	3.157
C(19)	H(14)	2.859	C(19)	H(15) ⁸⁾	3.244
C(19)	H(2A)	3.38(2)	C(20)	H(1) ⁵⁾	3.469

C(20)	H(14)	2.044	C(20)	H(15)	3.229
C(20)	H(15) ⁸⁾	3.580	C(20)	H(17)	3.283
C(21)	H(6) ⁸⁾	3.040	C(21)	H(15)	2.035
C(21)	H(15) ⁸⁾	3.358	C(21)	H(16)	3.275
C(22)	H(6) ⁸⁾	3.045	C(22)	H(14)	2.054
C(22)	H(14) ¹³⁾	3.132	C(22)	H(16)	2.032
C(22)	H(17)	3.288	C(22)	H(2A) ⁵⁾	3.34(2)
C(23)	H(2) ¹¹⁾	3.059	C(23)	H(14)	3.287
C(23)	H(14) ¹³⁾	3.114	C(23)	H(15)	2.033
C(23)	H(17)	2.054	C(23)	H(2A) ⁵⁾	3.26(2)
C(24)	H(2) ¹¹⁾	2.878	C(24)	H(15)	3.274
C(24)	H(16)	2.033	C(24)	H(1A) ⁵⁾	3.54(2)
C(25)	H(14)	3.277	C(25)	H(16)	3.228
C(25)	H(17)	2.047	C(25)	H(1A) ⁵⁾	3.41(2)
C(26)	H(4) ³⁾	3.491	C(26)	H(9)	3.479
C(26)	H(11)	2.750	C(26)	H(12)	3.305
C(26)	H(13)	2.765	C(26)	H(17)	2.854
C(26)	H(1A) ⁵⁾	2.79(2)	H(1)	O(2) ⁴⁾	3.488
H(1)	N(1)	2.752	H(1)	C(1)	2.049
H(1)	C(3)	2.053	H(1)	C(4)	3.287
H(1)	C(6)	3.290	H(1)	C(11) ⁶⁾	2.910
H(1)	C(12) ⁶⁾	3.284	H(1)	C(19) ⁴⁾	3.351
H(1)	C(20) ⁴⁾	3.469	H(1)	H(2)	2.348
H(1)	H(5) ⁶⁾	2.348	H(1)	H(6) ⁶⁾	3.123
H(1)	H(14) ⁶⁾	3.173	H(1)	H(15) ⁶⁾	3.090
H(1)	H(16) ⁹⁾	3.374	H(2)	C(1)	3.235
H(2)	C(2)	2.032	H(2)	C(4)	2.029
H(2)	C(5)	3.262	H(2)	C(14) ⁹⁾	2.975
H(2)	C(15) ⁹⁾	3.340	H(2)	C(23) ⁹⁾	3.059
H(2)	C(24) ⁹⁾	2.878	H(2)	H(1)	2.348

Table S13. Distances beyond the asymmetric unit out to 3.60 Å involving hydrogens (continued)

atom	atom	distance	atom	atom	distance
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H(2)	H(3)	2.324	H(2)	H(7) ⁹⁾	2.534
H(2)	H(8) ⁹⁾	3.193	H(2)	H(16) ⁹⁾	2.801
H(2)	H(17) ⁹⁾	2.483	H(3)	O(1) ¹⁴⁾	3.531
H(3)	O(3) ²⁾	2.734	H(3)	C(2)	3.279
H(3)	C(3)	2.035	H(3)	C(5)	2.036
H(3)	C(6)	3.239	H(3)	H(2)	2.324
H(3)	H(4)	2.346	H(3)	H(8) ⁹⁾	3.139
H(3)	H(11) ²⁾	3.176	H(3)	H(13) ⁴⁾	2.987
H(3)	H(17) ⁹⁾	3.113	H(3)	H(1A) ¹⁴⁾	3.020
H(4)	O(1)	3.139	H(4)	O(3) ²⁾	2.622
H(4)	C(1)	3.274	H(4)	C(3)	3.271
H(4)	C(4)	2.044	H(4)	C(6)	2.033
H(4)	C(7)	2.886	H(4)	C(16) ²⁾	3.231
H(4)	C(17)	3.297	H(4)	C(17) ²⁾	3.495
H(4)	C(26) ²⁾	3.491	H(4)	H(3)	2.346
H(4)	H(8) ²⁾	3.327	H(4)	H(9) ²⁾	2.542
H(4)	H(10)	2.780	H(4)	H(11) ²⁾	2.513
H(4)	H(13) ⁴⁾	3.518	H(4)	H(1A)	3.190
H(5)	O(2) ¹⁰⁾	2.985	H(5)	N(1) ¹⁰⁾	3.254
H(5)	N(2)	2.071	H(5)	C(1) ¹⁰⁾	3.333
H(5)	C(2) ¹⁰⁾	2.946	H(5)	C(10)	3.144
H(5)	C(12)	2.056	H(5)	C(12) ⁶⁾	3.544
H(5)	C(13)	3.166	H(5)	H(1) ¹⁰⁾	2.348
H(5)	H(6)	2.467	H(5)	H(6) ⁶⁾	3.171
H(5)	H(7) ⁶⁾	3.477	H(5)	H(14)	3.448
H(5)	H(15) ⁴⁾	3.491	H(5)	H(2A)	2.455
H(5)	H(2A) ¹⁰⁾	3.270	H(6)	N(1) ¹⁰⁾	3.241
H(6)	N(2)	3.147	H(6)	N(2) ¹⁰⁾	2.849
H(6)	C(10)	3.198	H(6)	C(11)	2.067
H(6)	C(11) ¹⁰⁾	3.367	H(6)	C(13)	2.138
H(6)	C(14)	3.000	H(6)	C(21) ¹³⁾	3.040
H(6)	C(22) ¹³⁾	3.045	H(6)	H(1) ¹⁰⁾	3.123
H(6)	H(5)	2.467	H(6)	H(5) ¹⁰⁾	3.171
H(6)	H(7)	2.982	H(6)	H(14) ¹³⁾	3.040

H(6)	H(15) ¹³⁾	3.036	H(6)	H(2A) ¹⁰⁾	2.173
H(7)	O(2) ¹²⁾	2.574	H(7)	C(3) ¹¹⁾	3.226
H(7)	C(10)	3.291	H(7)	C(12)	2.907

Table S13. Distances beyond the asymmetric unit out to 3.60 Å involving hydrogens (continued)

atom	atom	distance	atom	atom	distance
H(7)	C(13)	2.058	H(7)	C(15)	2.028
H(7)	C(16)	3.279	H(7)	C(18) ¹²⁾	3.543
H(7)	C(19) ¹²⁾	3.447	H(7)	H(2) ¹¹⁾	2.534
H(7)	H(5) ¹⁰⁾	3.477	H(7)	H(6)	2.982
H(7)	H(8)	2.320	H(7)	H(12) ¹²⁾	2.740
H(7)	H(15) ¹³⁾	3.409	H(8)	O(1) ³⁾	2.947
H(8)	C(9)	3.295	H(8)	C(13)	3.256
H(8)	C(14)	2.017	H(8)	C(16)	2.043
H(8)	C(18) ¹²⁾	3.358	H(8)	H(2) ¹¹⁾	3.193
H(8)	H(3) ¹¹⁾	3.139	H(8)	H(4) ³⁾	3.327
H(8)	H(7)	2.320	H(8)	H(9)	2.330
H(8)	H(10) ³⁾	3.022	H(8)	H(12) ¹²⁾	2.689
H(8)	H(13) ¹²⁾	3.110	H(8)	H(1A) ³⁾	3.302
H(9)	O(1)	2.519	H(9)	O(3)	3.478
H(9)	C(5) ³⁾	3.345	H(9)	C(7)	2.779
H(9)	C(8)	2.683	H(9)	C(9)	2.038
H(9)	C(10)	3.244	H(9)	C(14)	3.266
H(9)	C(15)	2.041	H(9)	C(17)	2.971
H(9)	C(17) ³⁾	3.212	H(9)	C(26)	3.479
H(9)	H(4) ³⁾	2.542	H(9)	H(8)	2.330
H(9)	H(10) ³⁾	2.230	H(9)	H(11)	2.328
H(9)	H(1A)	3.087	H(10)	O(1)	2.585
H(10)	O(1) ²⁾	3.102	H(10)	N(3)	3.299
H(10)	C(5)	3.045	H(10)	C(6)	2.695
H(10)	C(7)	2.072	H(10)	C(8)	3.398
H(10)	C(15) ²⁾	3.355	H(10)	C(16) ²⁾	2.959
H(10)	C(18)	2.061	H(10)	H(4)	2.780

H(10)	H(8) ²⁾	3.022	H(10)	H(9) ²⁾	2.230
H(10)	H(11)	1.595	H(10)	H(12)	2.466
H(10)	H(13)	2.242	H(10)	H(1A)	3.334
H(10)	H(1A) ²⁾	3.458	H(11)	O(1)	2.457
H(11)	O(3)	2.542	H(11)	N(3)	2.580
H(11)	C(4) ³⁾	3.418	H(11)	C(5) ³⁾	3.067
H(11)	C(6)	3.361	H(11)	C(7)	2.072
H(11)	C(8)	2.868	H(11)	C(9)	3.178
H(11)	C(16)	2.928	H(11)	C(18)	2.061
H(11)	C(26)	2.750	H(11)	H(3) ³⁾	3.176

Table S13. Distances beyond the asymmetric unit out to 3.60 Å involving hydrogens (continued)

atom	atom	distance	atom	atom	distance
H(11)	H(4) ³⁾	2.513	H(11)	H(9)	2.328
H(11)	H(10)	1.595	H(11)	H(12)	2.848
H(11)	H(13)	2.456	H(11)	H(1A)	3.333
H(12)	O(2)	2.565	H(12)	N(1)	3.224
H(12)	N(3)	2.022	H(12)	C(1)	3.045
H(12)	C(5)	3.480	H(12)	C(6)	2.767
H(12)	C(7)	2.711	H(12)	C(8)	3.060
H(12)	C(14) ⁷⁾	3.271	H(12)	C(15) ⁷⁾	3.250
H(12)	C(17)	2.072	H(12)	C(19)	2.608
H(12)	C(26)	3.305	H(12)	H(7) ⁷⁾	2.740
H(12)	H(8) ⁷⁾	2.689	H(12)	H(10)	2.466
H(12)	H(11)	2.848	H(12)	H(13)	1.600
H(13)	O(1) ²⁾	3.070	H(13)	O(2)	3.578
H(13)	O(3)	2.865	H(13)	N(3)	2.022
H(13)	C(3) ⁵⁾	3.325	H(13)	C(4) ⁵⁾	2.919
H(13)	C(5) ⁵⁾	3.252	H(13)	C(7)	3.414
H(13)	C(17)	2.072	H(13)	C(19)	3.157
H(13)	C(26)	2.765	H(13)	H(3) ⁵⁾	2.987
H(13)	H(4) ⁵⁾	3.518	H(13)	H(8) ⁷⁾	3.110
H(13)	H(10)	2.242	H(13)	H(11)	2.456
H(13)	H(12)	1.600	H(13)	H(1A) ²⁾	3.030

H(14)	O(2)	3.038	H(14)	N(2)	3.378
H(14)	C(11)	3.321	H(14)	C(19)	2.859
H(14)	C(20)	2.044	H(14)	C(22)	2.054
H(14)	C(22) ⁸⁾	3.132	H(14)	C(23)	3.287
H(14)	C(23) ⁸⁾	3.114	H(14)	C(25)	3.277
H(14)	H(1) ¹⁰⁾	3.173	H(14)	H(5)	3.448
H(14)	H(6) ⁸⁾	3.040	H(14)	H(15)	2.354
H(14)	H(15) ⁸⁾	2.711	H(14)	H(16) ⁸⁾	2.671
H(14)	H(2A)	3.534	H(15)	O(2) ¹³⁾	2.521
H(15)	N(2) ⁵⁾	3.574	H(15)	C(11) ⁵⁾	3.573
H(15)	C(19) ¹³⁾	3.244	H(15)	C(20)	3.229
H(15)	C(20) ¹³⁾	3.580	H(15)	C(21)	2.035
H(15)	C(21) ¹³⁾	3.358	H(15)	C(23)	2.033
H(15)	C(24)	3.274	H(15)	H(1) ¹⁰⁾	3.090
H(15)	H(5) ⁵⁾	3.491	H(15)	H(6) ⁸⁾	3.036
H(15)	H(7) ⁸⁾	3.409	H(15)	H(14)	2.354

Table S13. Distances beyond the asymmetric unit out to 3.60 Å involving hydrogens (continued)

atom	atom	distance	atom	atom	distance
H(15)	H(14) ¹³⁾	2.711	H(15)	H(16)	2.322
H(15)	H(2A) ⁵⁾	3.508	H(16)	N(2) ⁵⁾	3.075
H(16)	C(3) ¹¹⁾	3.361	H(16)	C(10) ⁵⁾	3.042
H(16)	C(11) ⁵⁾	3.276	H(16)	C(12) ⁵⁾	3.393
H(16)	C(13) ⁵⁾	3.243	H(16)	C(21)	3.275
H(16)	C(22)	2.032	H(16)	C(24)	2.033
H(16)	C(25)	3.228	H(16)	H(1) ¹¹⁾	3.374
H(16)	H(2) ¹¹⁾	2.801	H(16)	H(14) ¹³⁾	2.671
H(16)	H(15)	2.322	H(16)	H(17)	2.353
H(16)	H(2A) ⁵⁾	3.389	H(17)	O(1) ⁵⁾	3.582
H(17)	O(3)	3.018	H(17)	C(3) ¹¹⁾	3.032
H(17)	C(4) ¹¹⁾	3.360	H(17)	C(8) ⁵⁾	3.254
H(17)	C(9) ⁵⁾	3.105	H(17)	C(10) ⁵⁾	3.508
H(17)	C(16) ⁵⁾	3.444	H(17)	C(20)	3.283
H(17)	C(22)	3.288	H(17)	C(23)	2.054

H(17)	C(25)	2.047	H(17)	C(26)	2.854
H(17)	H(2) ¹¹⁾	2.483	H(17)	H(3) ¹¹⁾	3.113
H(17)	H(16)	2.353	H(17)	H(1A) ⁵⁾	3.107
H(1A)	O(3) ⁴⁾	1.99(2)	H(1A)	N(1)	3.492(19)
H(1A)	C(1)	3.37(2)	H(1A)	C(5)	3.16(2)
H(1A)	C(6)	2.53(2)	H(1A)	C(7)	1.93(2)
H(1A)	C(8)	2.75(2)	H(1A)	C(9)	3.49(2)
H(1A)	C(16)	3.57(2)	H(1A)	C(17)	3.15(2)
H(1A)	C(24) ⁴⁾	3.54(2)	H(1A)	C(25) ⁴⁾	3.41(2)
H(1A)	C(26) ⁴⁾	2.79(2)	H(1A)	H(3) ¹⁾	3.020
H(1A)	H(4)	3.190	H(1A)	H(8) ²⁾	3.302
H(1A)	H(9)	3.087	H(1A)	H(10)	3.334
H(1A)	H(10) ³⁾	3.458	H(1A)	H(11)	3.333
H(1A)	H(13) ³⁾	3.030	H(1A)	H(17) ⁴⁾	3.107
H(2A)	O(2)	3.20(2)	H(2A)	N(1)	2.23(2)
H(2A)	C(1)	3.52(2)	H(2A)	C(8)	2.824(19)
H(2A)	C(9)	2.74(2)	H(2A)	C(10)	1.96(2)
H(2A)	C(11)	2.02(2)	H(2A)	C(11) ⁶⁾	3.33(2)
H(2A)	C(12)	3.06(2)	H(2A)	C(12) ⁶⁾	2.79(2)
H(2A)	C(13)	3.05(2)	H(2A)	C(19)	3.38(2)
H(2A)	C(22) ⁴⁾	3.34(2)	H(2A)	C(23) ⁴⁾	3.26(2)
H(2A)	H(5)	2.455	H(2A)	H(5) ⁶⁾	3.270

Table S13. Distances beyond the asymmetric unit out to 3.60 Å involving hydrogens (continued)

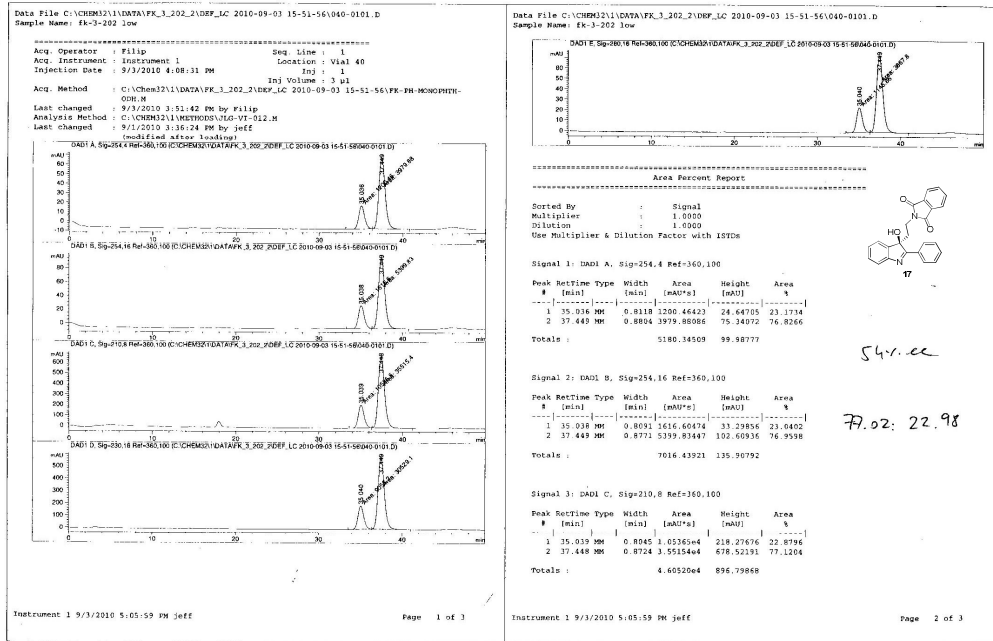
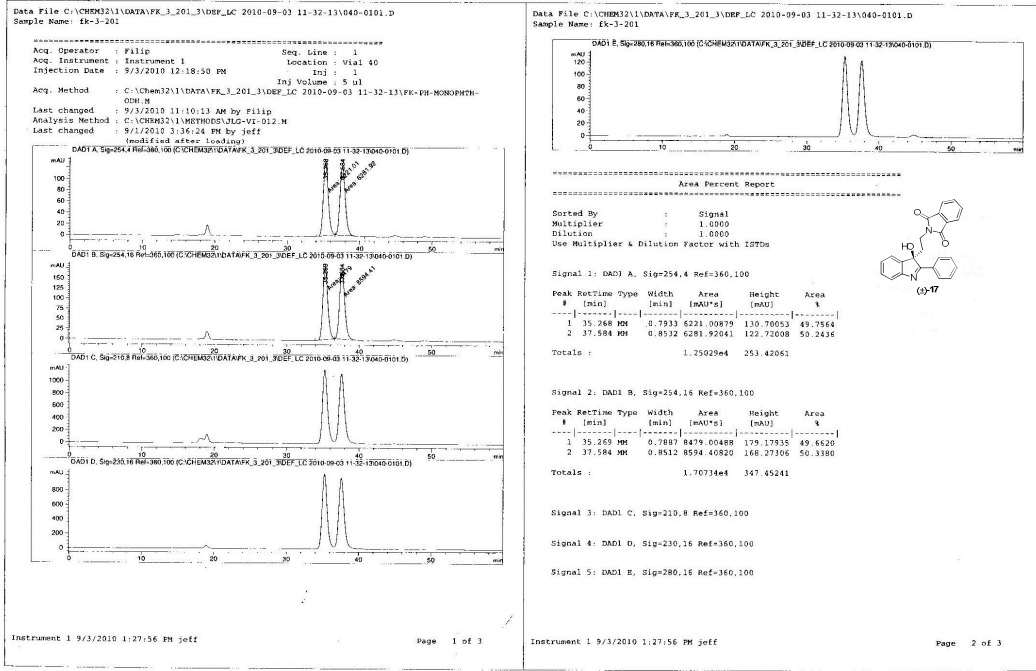
atom	atom	distance	atom	atom	distance
H(2A)	H(6) ⁶⁾	2.173	H(2A)	H(14)	3.534
H(2A)	H(15) ⁴⁾	3.508	H(2A)	H(16) ⁴⁾	3.389

Symmetry Operators:

(1)	-X+2,Y+1/2,-Z+2	(2)	-X+1,Y+1/2-1,-Z+2
(3)	-X+1,Y+1/2,-Z+2	(4)	X+1,Y,Z
(5)	X-1,Y,Z	(6)	-X+1,Y+1/2-1,-Z+1
(7)	X,Y-1,Z	(8)	-X,Y+1/2-1,-Z+1
(9)	X+1,Y-1,Z	(10)	-X+1,Y+1/2,-Z+1
(11)	X-1,Y+1,Z	(12)	X,Y+1,Z
(13)	-X,Y+1/2,-Z+1	(14)	-X+2,Y+1/2-1,-Z+2

L. Chromatograms.

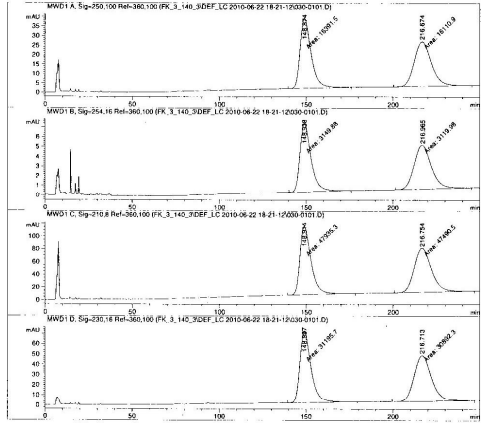
0.0-4, 0.2 ml/min, 80:20 Hex: EtOH



Data File C:\CHEM2\1\DATA\FK_3_140_3\DEF_LC 2010-06-22 18-21-12\030-0101.D
 Sample Name: FK-3-140

Acq. Operator : Filip
 Acq. Instrument : Instrument 2
 Injection Date : 6/22/2010 5:28:49 PM
 Seq. Line : 1
 Location : Vial 30
 Inj Volume : 15 µl

Acq. Method : C:\Chem2\1\DATA\FK_3_140_3\DEF_LC 2010-06-22 18-21-12\FK-NPHTH-PHTH.M
 Last changed : 6/22/2010 5:19:47 PM by Filip
 Analysis Method : C:\CHEM2\1\DATA\FK_3_140_3\DEF_LC 2010-06-22 18-21-12\030-0101.D\DA.M (FK-NPHTH-PHTH.M)
 Last changed : 6/22/2010 5:19:47 PM by Filip
 Method Info : chiralcol.D



Data File C:\CHEM2\1\DATA\FK_3_140_3\DEF_LC 2010-06-22 18-21-12\030-0101.D
 Sample Name: FK-3-140

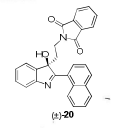
MWD1 E. Sig=250.16 Ref=360.100 (FK_3_140_3\DEF_LC 2010-06-22 18-21-12\030-0101.D)

Area Percent Report

Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDs

Signal 1: MWD1 A. Sig=250.100 Ref=360.100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	148.974	PK	7.4108	1.63515e4	36.86395	50.4337
2	216.874	PK	11.3478	1.61209e4	21.60274	49.5683
Totals :				3.25024e4	60.52669	



Signal 2: MWD1 B. Sig=254.16 Ref=360.100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	148.938	PK	7.4293	3349.87964	7.07585	50.2384
2	216.965	PK	11.4595	3319.87974	4.53770	49.7616
Totals :				6269.85937	11.61355	

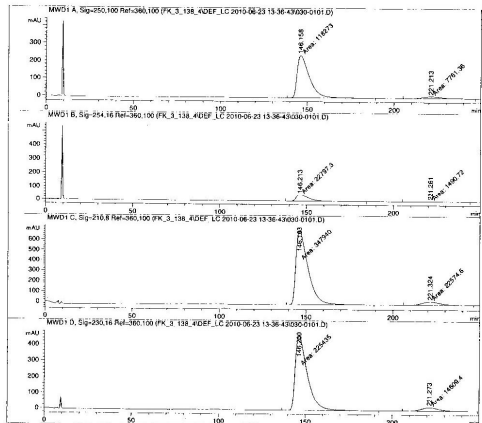
Signal 3: MWD1 C. Sig=210.8 Ref=360.100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	148.934	PK	7.4173	4.79353e4	107.71022	50.2330
2	216.754	PK	11.4414	4.74905e4	69.17962	49.7670
Totals :				9.54258e4	176.88984	

Data File C:\CHEM2\1\DATA\FK_3_138_4\DEF_LC 2010-06-23 13-36-43\030-0101.D
 Sample Name: FK-3-138

Acq. Operator : Filip
 Acq. Instrument : Instrument 2
 Injection Date : 6/23/2010 1:43:56 PM
 Seq. Line : 1
 Location : Vial 30
 Inj Volume : 15 µl

Acq. Method : C:\Chem2\1\DATA\FK_3_138_4\DEF_LC 2010-06-23 13-36-43\FK-NPHTH-PHTH.M
 Last changed : 6/22/2010 5:19:47 PM by Filip
 Analysis Method : C:\CHEM2\1\DATA\FK_3_138_4\DEF_LC 2010-06-23 13-36-43\030-0101.D\DA.M (FK-NPHTH-PHTH.M)
 Last changed : 6/22/2010 5:19:47 PM by Filip
 Method Info : chiralcol.D



Data File C:\CHEM2\1\DATA\FK_3_138_4\DEF_LC 2010-06-23 13-36-43\030-0101.D
 Sample Name: FK-3-138

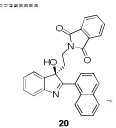
MWD1 E. Sig=250.16 Ref=360.100 (FK_3_138_4\DEF_LC 2010-06-23 13-36-43\030-0101.D)

Area Percent Report

Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDs

Signal 1: MWD1 A. Sig=250.100 Ref=360.100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	146.158	PK	8.1471	1.18973e5	241.96290	93.8418
2	221.213	PK	11.6939	7761.36377	11.00487	6.1582
Totals :				1.26036e5	253.01476	



Signal 2: MWD1 B. Sig=254.16 Ref=360.100

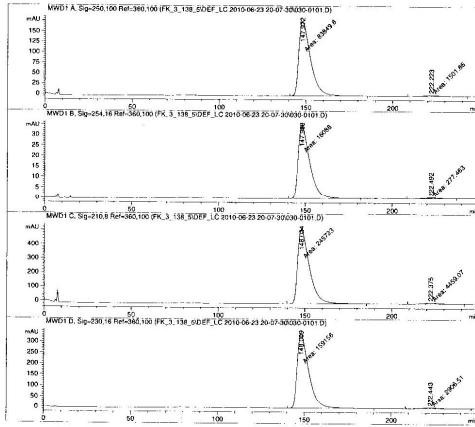
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	146.213	PK	8.0653	2.27973e4	47.10973	93.8623
2	221.261	PK	11.6748	1490.72253	2.12813	6.1377
Totals :				2.42880e4	49.23785	

Signal 3: MWD1 C. Sig=210.8 Ref=360.100

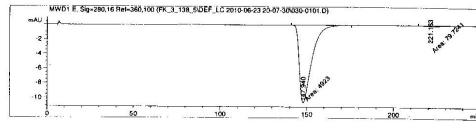
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	146.163	PK	8.1080	3.47940e5	715.14539	93.8072
2	221.354	PK	11.6944	2.25746e4	32.17287	6.0928
Totals :				3.70515e5	747.33826	

Data File C:\CHEM32\DATA\FK_3_138_5\DEF_LC 2010-06-23 20-07-30\030-0101.D
 Sample Name: FK-3-138 recrystallized

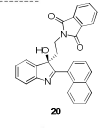
=====
 Acq. Operator : Filip Seq. Line : 1
 Acq. Instrument : Instrument 2 Location : Vial 10
 Injection Date : 6/23/2010 8:14:32 PM Inj : 1
 Inj Volume : 15 µl
 Acq. Method : C:\Chem32\DATA\FK_3_138_5\DEF_LC 2010-06-23 20-07-30\FK-NPHTH-PHTH.M
 Last changed : 6/22/2010 5:19:47 PM by Filip
 Analysis Method : C:\CHEM32\DATA\FK_3_138_5\DEF_LC 2010-06-23 20-07-30\030-0101.D\DA.M (FK-NPHTH-PHTH.M)
 Last changed : 6/22/2010 5:19:47 PM by Filip
 Method Info : chemical CD



Data File C:\CHEM32\DATA\FK_3_138_5\DEF_LC 2010-06-23 20-07-30\030-0101.D
 Sample Name: FK-3-138 recrystallized



=====
 Area Percent Report
 =====
 Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with 15TD6



Signal 1: MMD1 A, Sig=250.100 Ref=360.100

Peak #	RetTime (min)	Type	Width (min)	Area (mAU*s)	Height (mAU)	Area %
1	147.972	PK	7.7946	8.38498e4	179.28931	98.2404
2	222.223	PK	10.8480	3501.85876	2.30743	1.7596
Totals :				8.53516e4	181.59673	

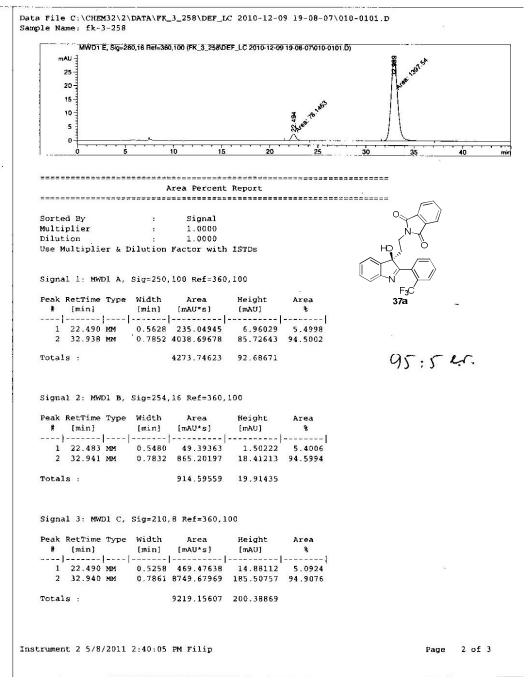
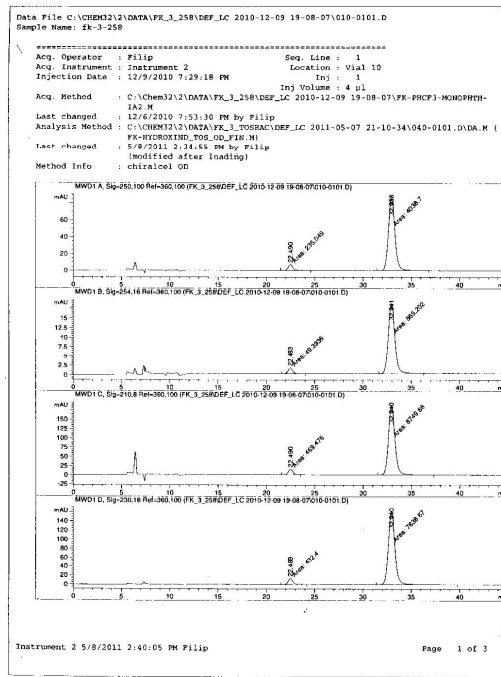
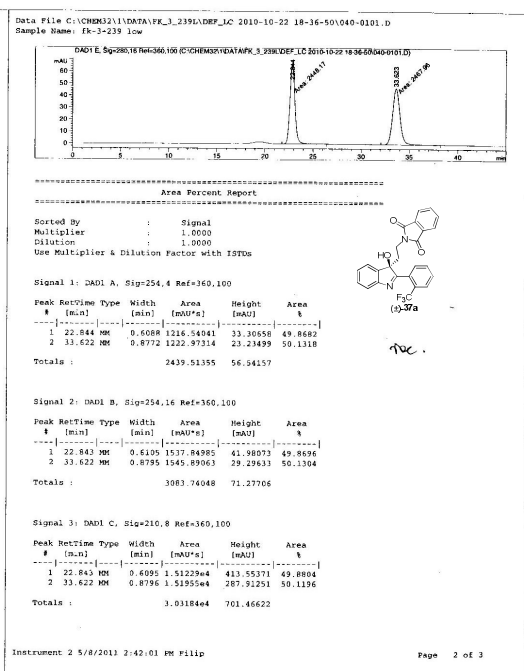
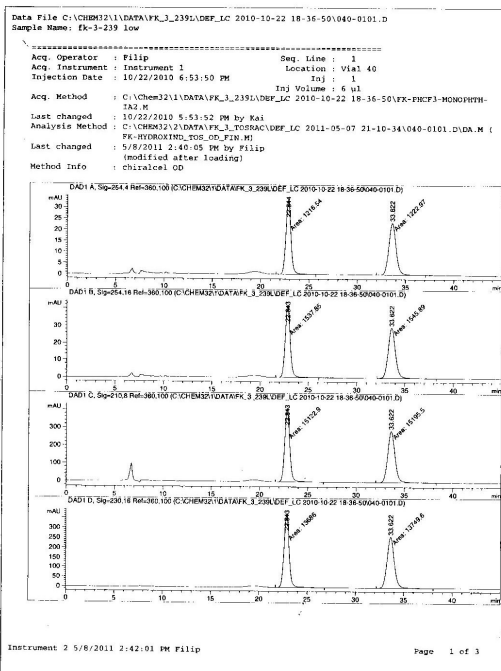
Signal 2: MMD1 B, Sig=254.16 Ref=360.100

Peak #	RetTime (min)	Type	Width (min)	Area (mAU*s)	Height (mAU)	Area %
1	147.988	PK	7.7823	1.60890e4	34.54107	98.3946
2	222.492	PK	10.6765	277.46274	4.33120e-1	1.6954
Totals :				1.63654e4	34.97619	

Signal 3: MMD1 C, Sig=210.8 Ref=360.100

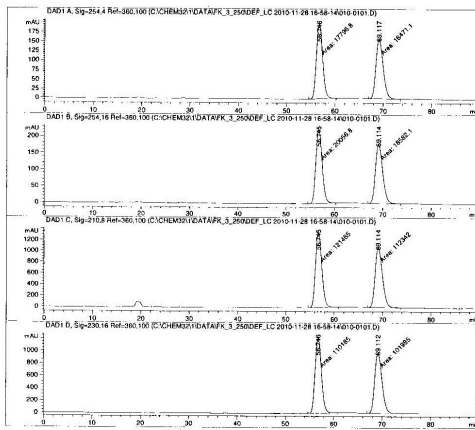
Peak #	RetTime (min)	Type	Width (min)	Area (mAU*s)	Height (mAU)	Area %
1	148.034	PK	7.7705	2.45723e5	527.04474	98.2173
2	222.375	PK	11.1815	4459.07080	6.64647	1.7823
Totals :				2.50282e5	533.69121	

98.5 : 1.5



Data File C:\CHEM321\DATA\FK_3_250\DEF_LC 2010-11-28 16:58-14\010-0101.D
 Sample Name: FK-3-250

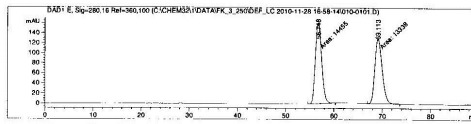
=====
 Acq. Operator : Filip Seq. Line : 1
 Acq. Instrument : Instrument 1 Location : Vial 10
 Injection Date : 11/28/2010 5:15:20 PM Inj : 1
 Inj Volume : 7 µl
 Acq. Method : C:\Chem321\DATA\FK_3_250\DEF_LC 2010-11-28 16:58-14\FK-PH-MONOPHTH-CON-H
 Last changed : 11/28/2010 3:06:27 PM by Filip
 Analysis Method : C:\CHEM321\METHODS\FK-MONOPHTH-CP1_FCN.M
 Last changed : 11/24/2010 4:34:13 PM by Filip
 Method Info : chr1xal02-H



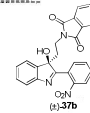
Instrument 1 11/29/2010 11:53:00 AM Filip

Page 1 of 3

Data File C:\CHEM321\DATA\FK_3_250\DEF_LC 2010-11-28 16:58-14\010-0101.D
 Sample Name: FK-3-250



=====
 Area Percent Report
 =====
 Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDs



Signal 1: DAD1 A, Sig=254.4 Ref=360.100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	56.745	HM	1.5021	1.77958e4	197.47076	51.9343
2	69.117	HM	1.7377	1.64711e4	157.98227	48.0657
Totals : 3.42680e4 355.45303						

Signal 2: DAD1 B, Sig=254.16 Ref=360.100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	56.745	HM	1.5032	2.00560e4	222.38306	51.9083
2	69.114	HM	1.7398	1.81821e4	178.00981	48.0917
Totals : 3.86390e4 400.39287						

Signal 3: DAD1 C, Sig=210.8 Ref=360.100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	56.745	HM	1.5100	1.21465e5	1340.66772	51.9508
2	69.114	HM	1.7408	1.12342e5	1075.58105	48.0492
Totals : 2.33807e5 2416.24878						

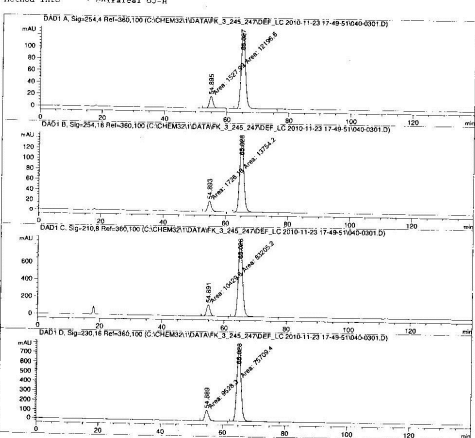
Instrument 1 11/29/2010 11:53:00 AM Filip

Page 2 of 3

OD-H, 0.2 µl/min, 80 lux: 20 EtOH

Data File C:\CHEM321\DATA\FK_3_245_247\DEF_LC 2010-11-23 17:49-51\040-0301.D
 Sample Name: FK-3-247 low

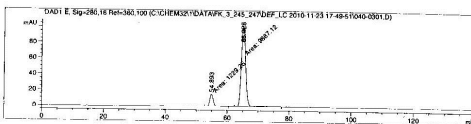
=====
 Acq. Operator : Filip Seq. Line : 3
 Acq. Instrument : Instrument 1 Location : Vial 40
 Injection Date : 11/23/2010 10:35:13 PM Inj : 1
 Inj Volume : 4 µl
 Acq. Method : C:\Chem321\DATA\FK_3_245_247\DEF_LC 2010-11-23 17:49-51\FK-PH-MONOPHTH-CON-H
 Last changed : 11/23/2010 5:28:14 PM by Filip
 Analysis Method : C:\CHEM321\METHODS\FK-MONOPHTH-CP1_FCN.M
 Last changed : 11/22/2010 11:19:35 AM by Filip
 Method Info : chr1xal02-H



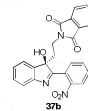
Instrument 1 11/24/2010 11:32:40 AM Filip

Page 1 of 3

Data File C:\CHEM321\DATA\FK_3_245_247\DEF_LC 2010-11-23 17:49-51\040-0301.D
 Sample Name: FK-3-247 low



=====
 Area Percent Report
 =====
 Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDs



Signal 1: DAD1 A, Sig=254.4 Ref=360.100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	54.893	HM	1.3062	1527.98547	19.49644	11.1332
2	65.027	HM	1.6030	1.21860e4	126.80947	88.8668
Totals : 1.37246e4 146.30591						

Signal 2: DAD1 B, Sig=254.16 Ref=360.100

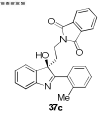
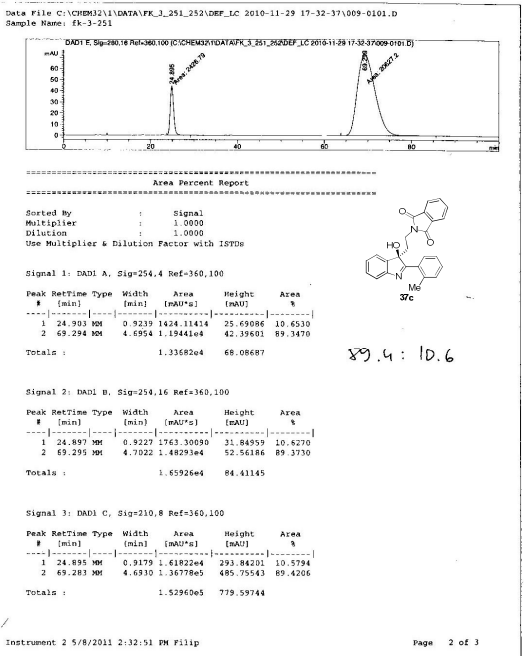
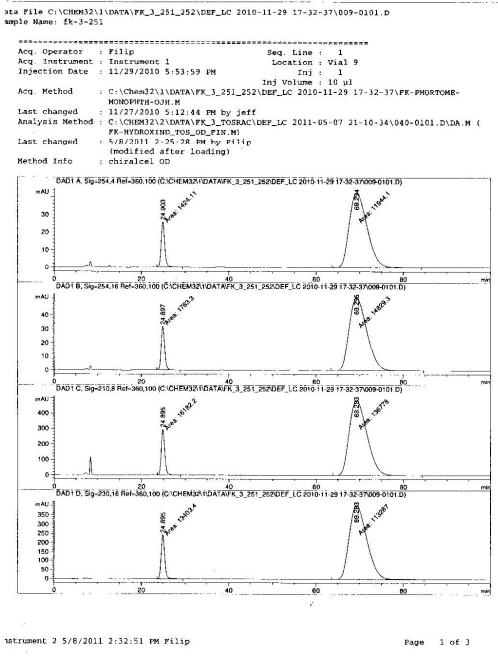
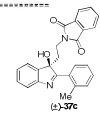
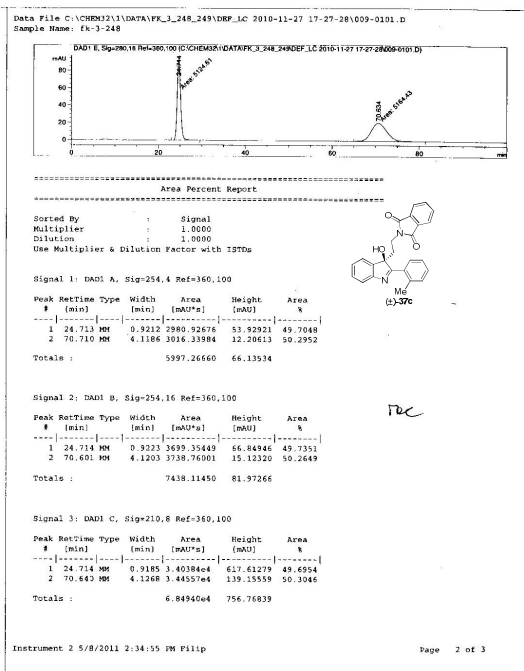
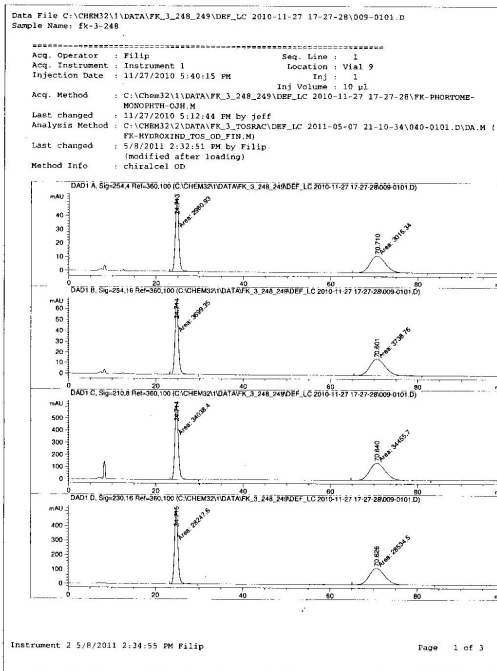
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	54.893	HM	1.3052	1726.18286	21.97551	11.1508
2	65.028	HM	1.6048	1.37542e4	142.84019	88.8492
Totals : 1.54803e4 164.81571						

Signal 3: DAD1 C, Sig=210.8 Ref=360.100

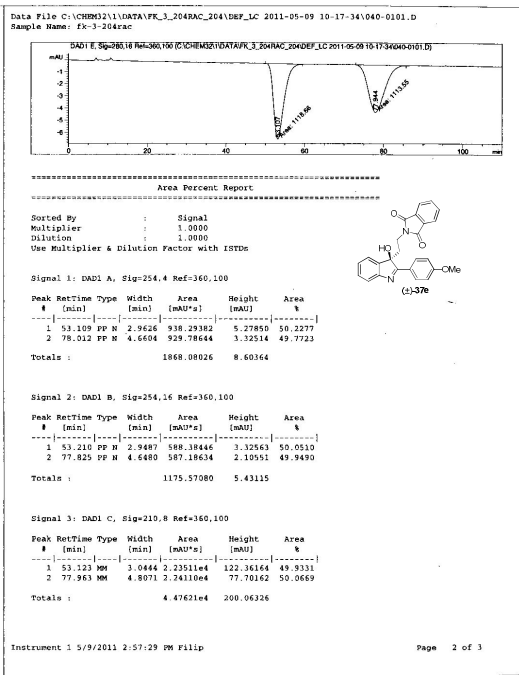
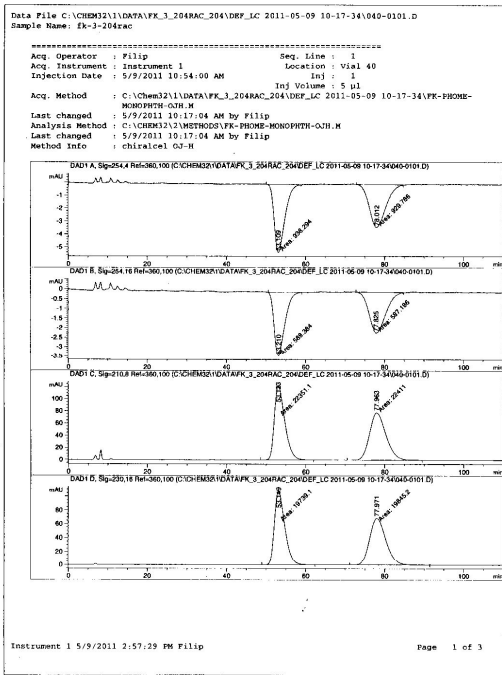
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	54.893	HM	1.3102	1.04298e4	132.67923	11.1398
2	65.028	HM	1.6046	8.32352e4	864.25287	88.8612
Totals : 9.36350e4 996.93210						

Instrument 1 11/24/2010 11:32:40 AM Filip

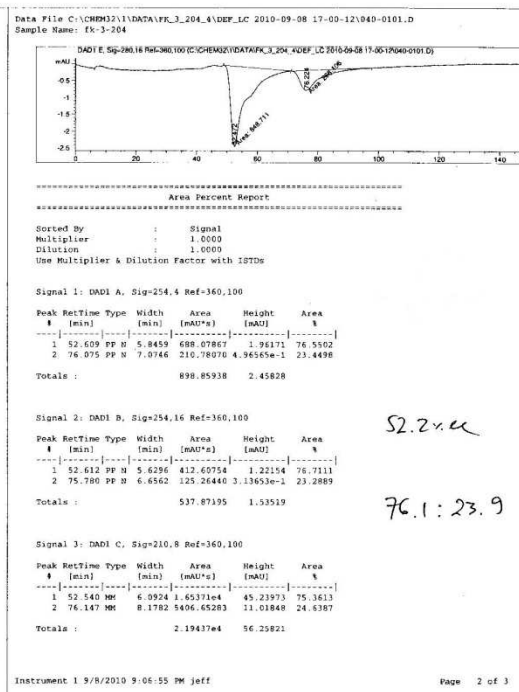
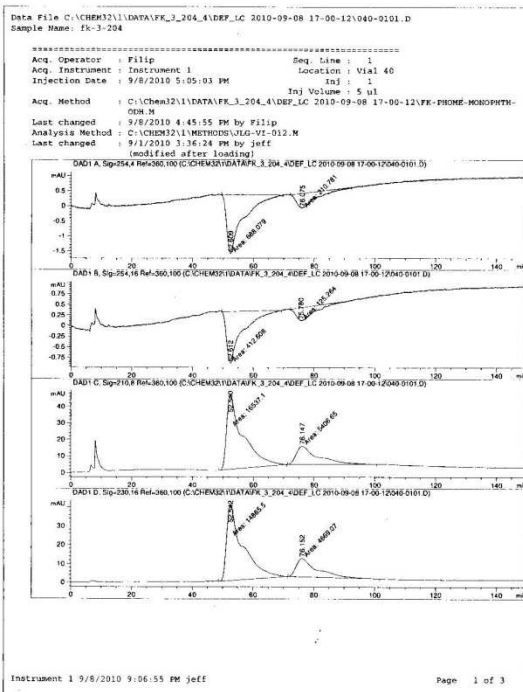
Page 2 of 3



89.9 : 10.6



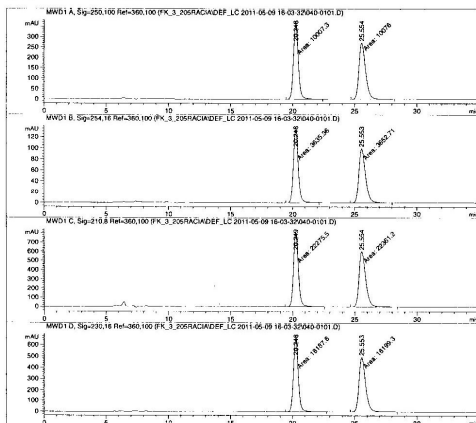
0.5-4, 0.5 mL/min, 75-25 Hex: EtOH



Data File C:\CHEM32\DATA\FK_3_205RACIA\DEF_LC 2011-05-09 16-03-32\040-0101.D
 Sample Name: FK-3-205rac

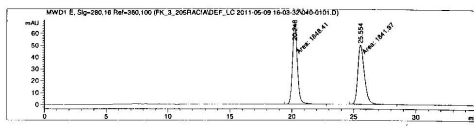
Acq. Operator : Filip Filip Seq. Line : 1
 Acq. Instrument : INSTRUMENT 2 Location : Vial 40
 Injection Date : 5/9/2011 4:14:49 PM Inj : 1
 Inj Volume : 5 µl

Acq. Method : C:\Chem32\DATA\FK_3_205RACIA\DEF_LC 2011-05-09 16-03-32\FK-PRCP3-MONOHPFH-COLSCR.M
 Last changed : 5/9/2011 3:23:45 PM by Filip
 Analysis Method : C:\CHEM32\DATA\FK_3_205RACIA\DEF_LC 2011-05-09 16-03-32\040-0101.D\DA.M
 (FK-PRCP3-MONOHPFH-COLSCR.M)
 Last changed : 5/9/2011 3:23:45 PM by Filip
 Method Info : chiralcol 1A



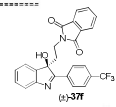
Instrument 2 5/10/2011 9:40:39 AM Filip Page 1 of 3

Data File C:\CHEM32\DATA\FK_3_205RACIA\DEF_LC 2011-05-09 16-03-32\040-0101.D
 Sample Name: FK-3-205rac



Area Percent Report

Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDs



Signal 1: MWDI A, Sig=250.100 Ref=360.100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.248	MM	0.4607	1.00073e4	362.00296	49.8291
2	25.354	MM	0.6202	1.0076e4	270.29772	50.1709
Totals : 2.00833e4 632.76068						

Signal 2: MWDI B, Sig=234.16 Ref=360.100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.248	MM	0.4557	3635.36108	132.95705	49.8810
2	25.553	MM	0.6158	3652.70874	98.86414	50.1190
Totals : 7288.06982 231.82119						

Signal 3: MWDI C, Sig=210.8 Ref=360.100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.248	MM	0.4607	2.22755e4	805.84265	49.9041
2	25.354	MM	0.6381	2.2182e4	601.94212	50.0959
Totals : 4.4637e4 1407.78497						

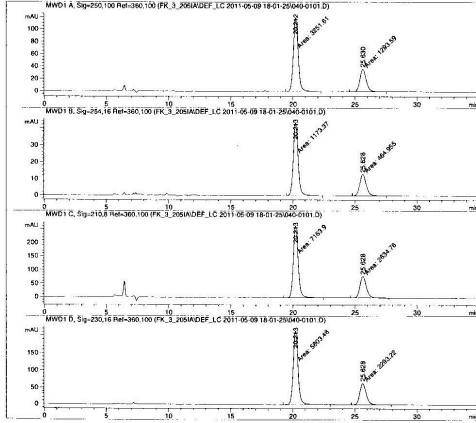
Instrument 2 5/10/2011 9:40:39 AM Filip Page 2 of 3

1A, 0.5 ml/min, 80:20 Hex:IPA

Data File C:\CHEM32\DATA\FK_3_205IA\DEF_LC 2011-05-09 18-01-25\040-0101.D
 Sample Name: FK-3-205

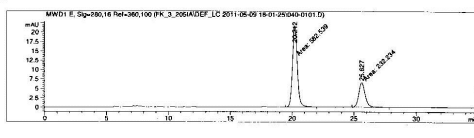
Acq. Operator : Filip Filip Seq. Line : 1
 Acq. Instrument : INSTRUMENT 2 Location : Vial 40
 Injection Date : 5/9/2011 7:12:38 PM Inj : 1
 Inj Volume : 3 µl

Acq. Method : C:\Chem32\DATA\FK_3_205IA\DEF_LC 2011-05-09 18-01-25\FK-PRCP3-MONOHPFH-COLSCR.M
 Last changed : 5/9/2011 6:00:16 PM by Filip
 Analysis Method : C:\CHEM32\DATA\FK_3_205IA\DEF_LC 2011-05-09 18-01-25\040-0101.D\DA.M (FK-PRCP3-MONOHPFH-COLSCR.M)
 Last changed : 5/9/2011 6:00:16 PM by Filip
 Method Info : chiralcol 1A



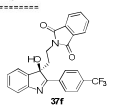
Instrument 2 5/10/2011 9:38:52 AM Filip Page 1 of 3

Data File C:\CHEM32\DATA\FK_3_205IA\DEF_LC 2011-05-09 18-01-25\040-0101.D
 Sample Name: FK-3-205



Area Percent Report

Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDs



Signal 1: MWDI A, Sig=250.100 Ref=360.100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.212	MM	0.4532	1251.61450	119.57166	71.5395
2	25.630	MM	0.5957	1293.58972	36.19318	28.4605
Totals : 4545.20422 155.76284						

Signal 2: MWDI B, Sig=254.16 Ref=360.100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.212	MM	0.4526	1173.36516	43.20894	71.6200
2	25.628	MM	0.5931	464.99477	23.05622	28.3800
Totals : 1638.32013 56.27516						

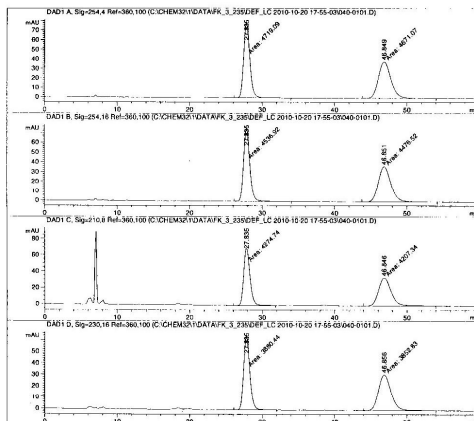
Signal 3: MWDI C, Sig=210.8 Ref=360.100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.212	MM	0.4527	7163.90234	263.74695	71.6486
2	25.628	MM	0.5936	2834.75854	79.59386	28.3514
Totals : 9998.66089 343.34081						

Instrument 2 5/10/2011 9:38:52 AM Filip Page 2 of 3

Data File C:\CHEM32\1\DATA\FK_3_235\DEF_IC 2010-10-20 17:55-03\040-0101.D
 Sample Name: FK-3-235 low

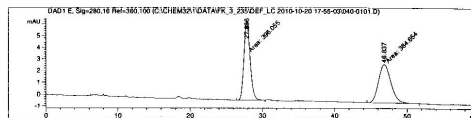
Acq. Operator : Filip Seq. Line : 1
 Acq. Instrument : Instrument 1 Location : Vial 40
 Injection Date : 10/20/2010 5:58:34 PM Inj. Inj : 1
 Inj. Volume : 5 µl
 Acq. Method : C:\Chem32\1\DATA\FK_3_235\DEF_IC 2010-10-20 17:55-03\FK-ANTRAC-PHTR.M
 Last changed : 10/20/2010 4:48:10 PM by Filip
 Analysis Method : C:\CHEM32\1\DATA\FK_3_235\DEF_IC 2011-05-07 21-10-34\040-0101.D\DA.M (FK-HYDROXIND_TOS_OD_FIN.M)
 Last changed : 5/8/2011 2:23:37 PM by Filip (modified after loading)
 Method Info : chiralcol OD



Instrument 2 5/8/2011 2:25:28 PM Filip

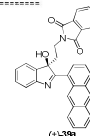
Page 1 of 3

Data File C:\CHEM32\1\DATA\FK_3_235\DEF_IC 2010-10-20 17:55-03\040-0101.D
 Sample Name: FK-3-235 low



Area Percent Report

Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDs



Signal 1: DAD1 A, Sig=254.4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	27.835	PK	1.0056	4719.09473	78.21732	50.2557
2	46.849	PK	2.0232	4671.06787	38.49893	49.7483
Totals :				9390.16260	116.71626	

Signal 2: DAD1 B, Sig=254.16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	27.835	PK	1.0084	4536.92334	74.98431	50.3351
2	46.851	PK	2.0226	4476.51855	36.88670	49.6649
Totals :				9013.44189	111.87102	

Signal 3: DAD1 C, Sig=210.8 Ref=360,100

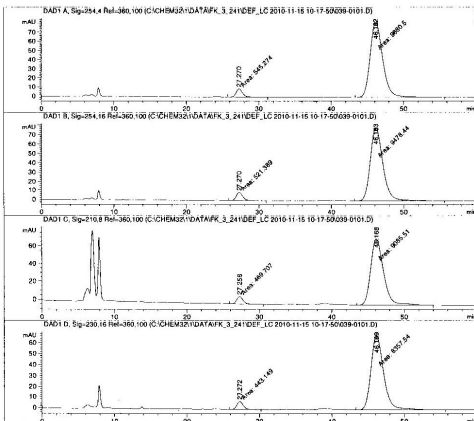
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	27.835	PK	1.0118	4274.73975	70.27375	50.3973
2	46.846	PK	2.0458	4207.34033	34.27620	49.6027
Totals :				8482.08008	104.54955	

Instrument 2 5/8/2011 2:25:28 PM Filip

Page 2 of 3

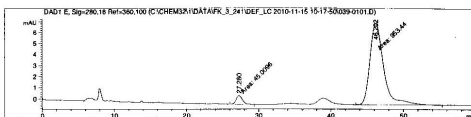
Data File C:\CHEM32\1\DATA\FK_3_241\DEF_IC 2010-11-15 10:17:50\039-0101.D
 Sample Name: FK-3-241 low all

Acq. Operator : Filip Seq. Line : 1
 Acq. Instrument : Instrument 1 Location : Vial 39
 Injection Date : 11/15/2010 10:29:57 AM Inj. Inj : 1
 Inj. Volume : 10 µl
 Acq. Method : C:\Chem32\1\DATA\FK_3_241\DEF_IC 2010-11-15 10-17-50\FK-ANTRAC-PHTR.M
 Last changed : 11/15/2010 10:06:09 AM by Filip
 Analysis Method : C:\CHEM32\1\DATA\FK_3_241\DEF_IC 2011-05-07 21-10-34\040-0101.D\DA.M (FK-HYDROXIND_TOS_OD_FIN.M)
 Last changed : 5/8/2011 2:20:45 PM by Filip (modified after loading)
 Method Info : chiralcol OD



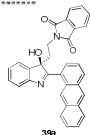
Instrument 2 5/8/2011 2:23:36 PM Filip

Data File C:\CHEM32\1\DATA\FK_3_241\DEF_IC 2010-11-15 10:17:50\039-0101.D
 Sample Name: FK-3-241 low all



Area Percent Report

Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDs



Signal 1: DAD1 A, Sig=254.4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	27.270	PK	1.0170	545.27380	8.93626	5.2301
2	46.162	PK	2.0001	9880.49805	82.33318	94.7699
Totals :				1.0425864	91.26945	

Signal 2: DAD1 B, Sig=254.16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	27.270	PK	1.0155	521.98979	8.35710	5.2140
2	46.163	PK	2.0020	9478.44434	78.50980	94.7860
Totals :				999.83313	87.46690	

Signal 3: DAD1 C, Sig=210.8 Ref=360,100

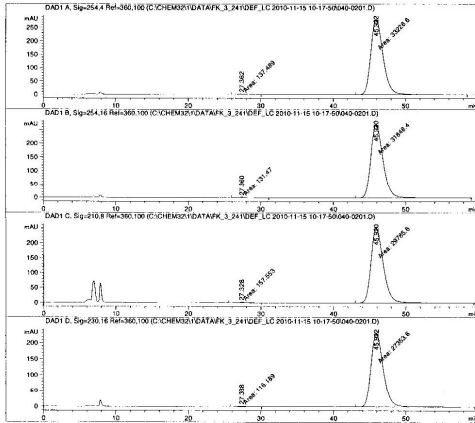
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	27.258	PK	0.9941	469.70676	7.87520	4.9157
2	46.168	PK	2.0382	9085.51455	74.29472	95.0843
Totals :				955.22141	82.16991	

Instrument 2 5/8/2011 2:23:36 PM Filip

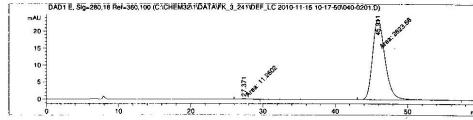
Page 2 of 3

Data File C:\CHEM32\1\DATA\FK_3_241\DEF_LC 2010-11-15 10-17-50\040-0201.D
 Sample Name: FK-3-241 low cryst

Acq. Operator : Filip Filip Seq. Line : 2
 Acq. Instrument : Instrument 1 Location : Vial 40
 Injection Date : 11/25/2010 11:51:24 AM Inj 1
 Inj Volume : 10 µl
 Acq. Method : C:\Chem32\1\DATA\FK_3_241\DEF_LC 2010-11-15 10-17-50\FK-ANTRAC-PRTH.M
 Last changed : 11/25/2010 10:05:09 AM by Filip
 Analysis Method : C:\CHEM32\1\DATA\FK_3_241\DEF_LC 2011-05-07 21-10-34\040-0101.D\DA.M (FK-HYDROXIND_T05_OD_FIN.M)
 Last changed : 5/8/2011 2:11:45 PM by Filip (modified after loading)
 Method Info : chiralcel OD



Data File C:\CHEM32\1\DATA\FK_3_241\DEF_LC 2010-11-15 10-17-50\040-0201.D
 Sample Name: FK-3-241 low cryst

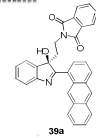


Area Percent Report

Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=254.4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	27.362	NM	1.0812	137.48908	2.11930	0.4121
2	45.922	NM	1.9638	3.3236e4	282.04535	99.5879
Totals:				3.33661e4	284.16465	



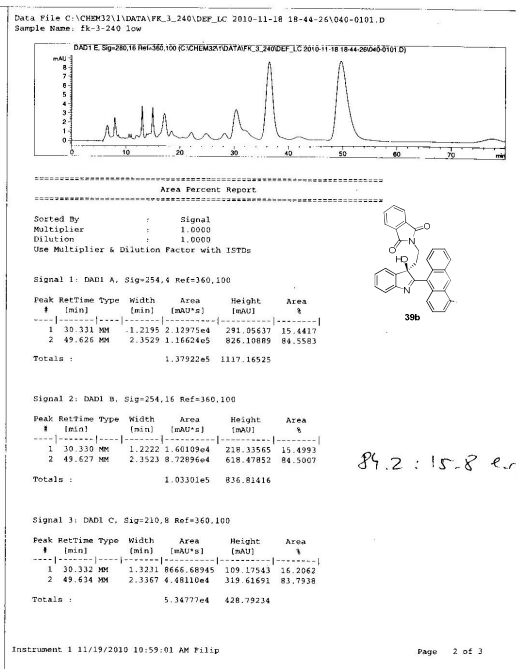
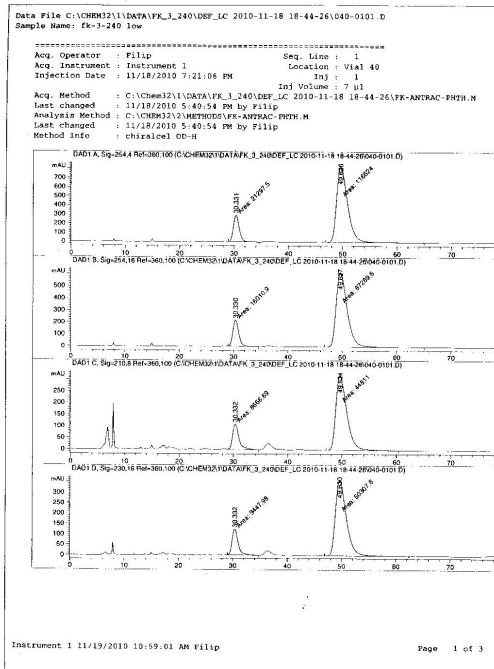
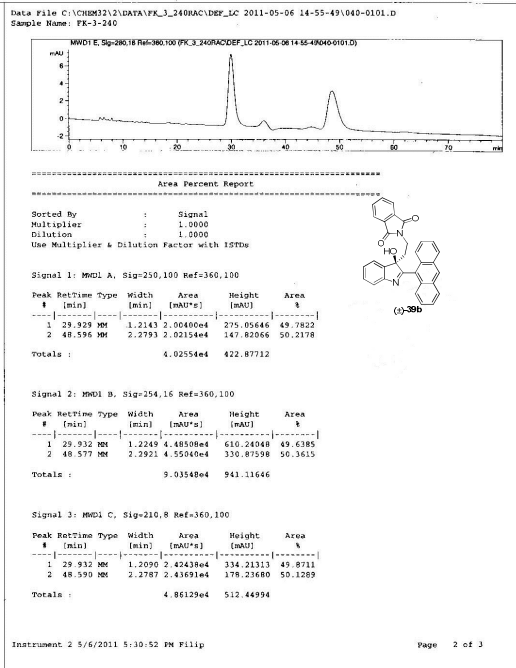
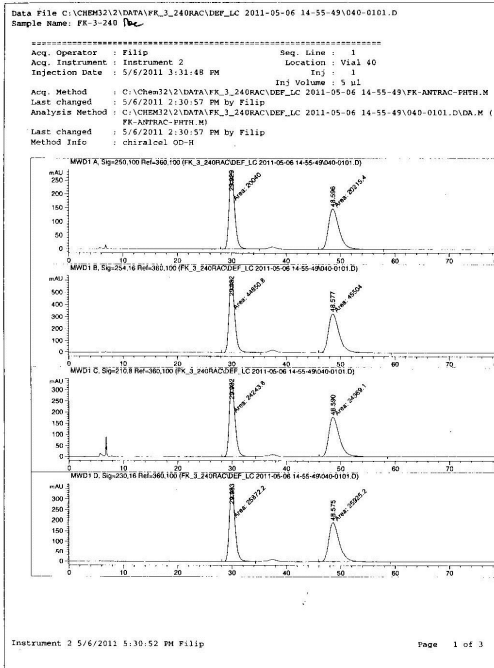
Signal 2: DAD1 B, Sig=254.16 Ref=360,100

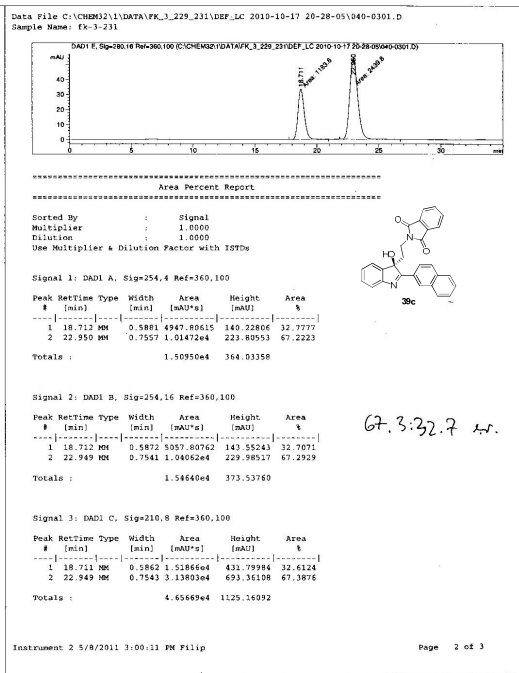
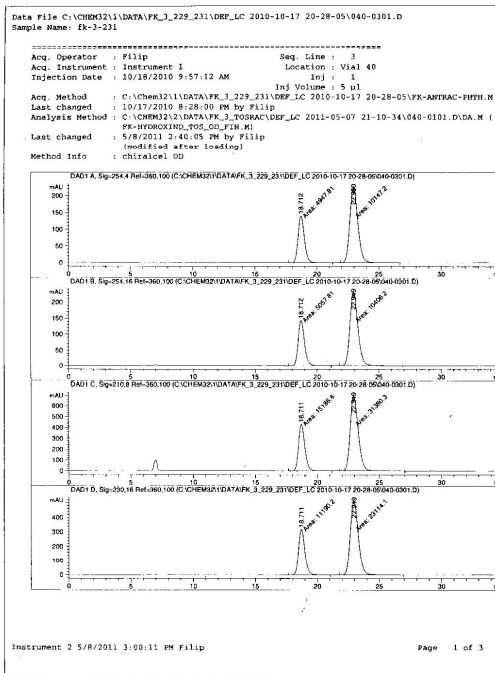
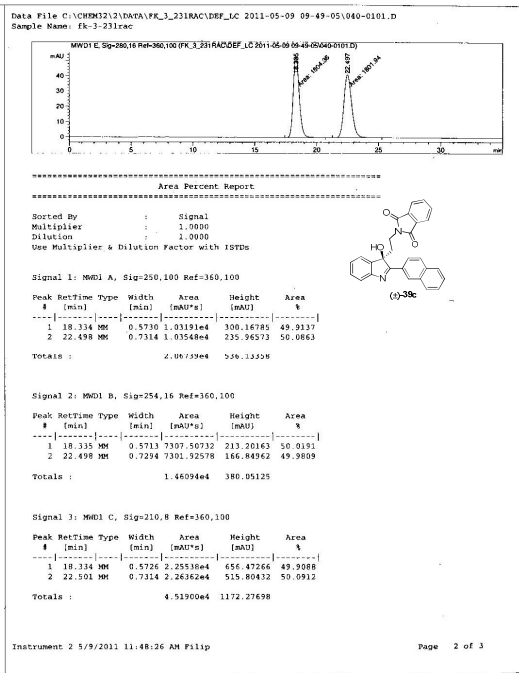
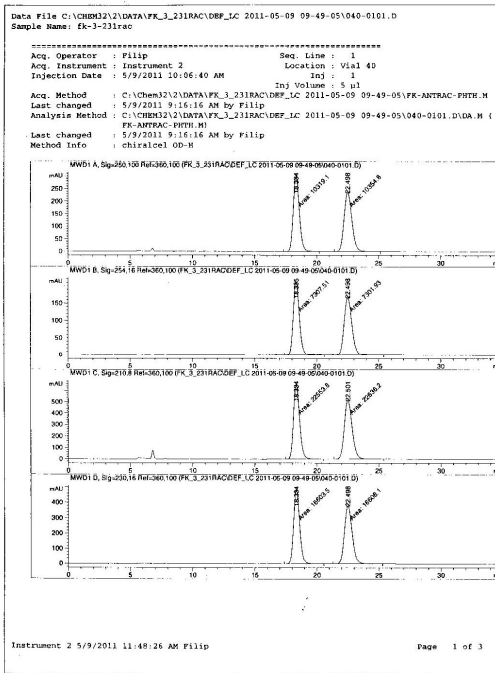
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	27.360	NM	1.0809	131.47032	2.02712	0.4111
2	45.920	NM	1.9651	3.18484e4	270.18004	99.5889
Totals:				3.19799e4	272.14516	

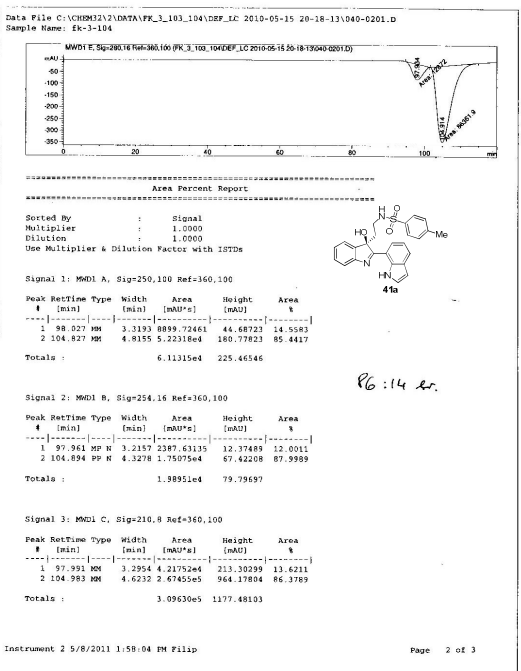
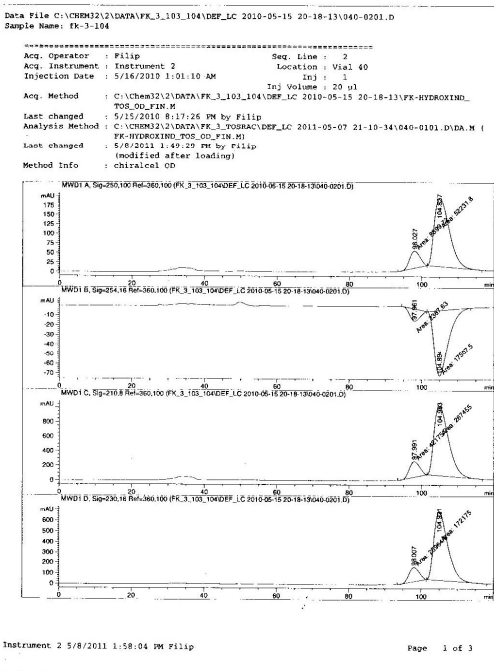
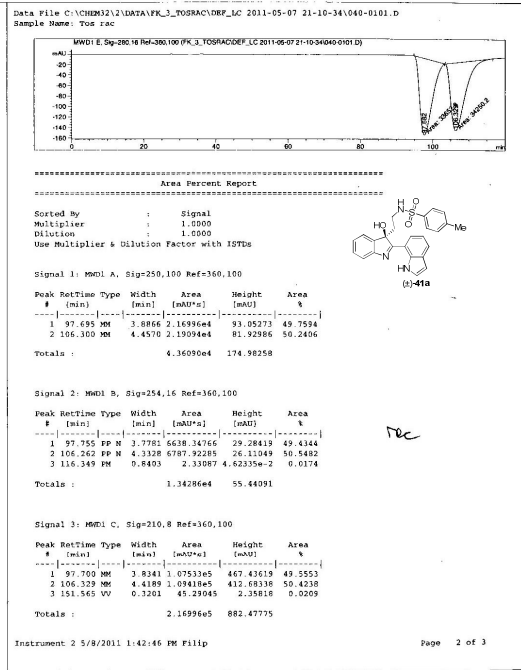
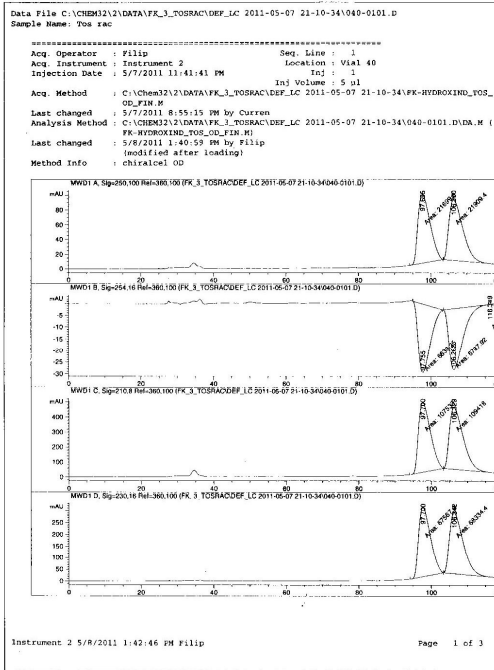
99.6:0.4

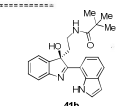
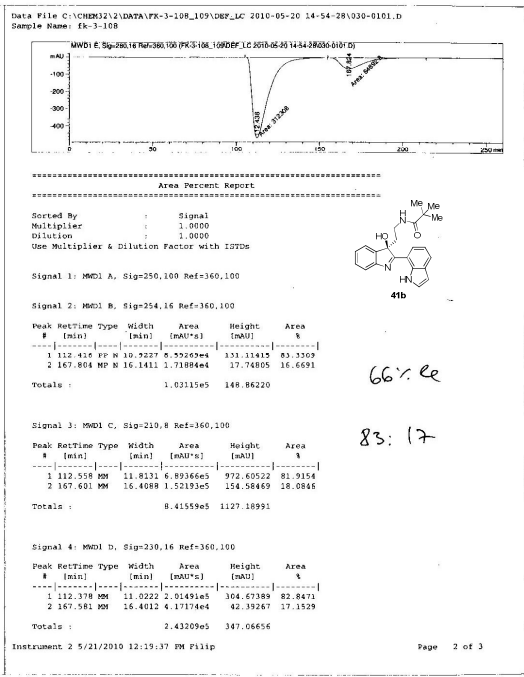
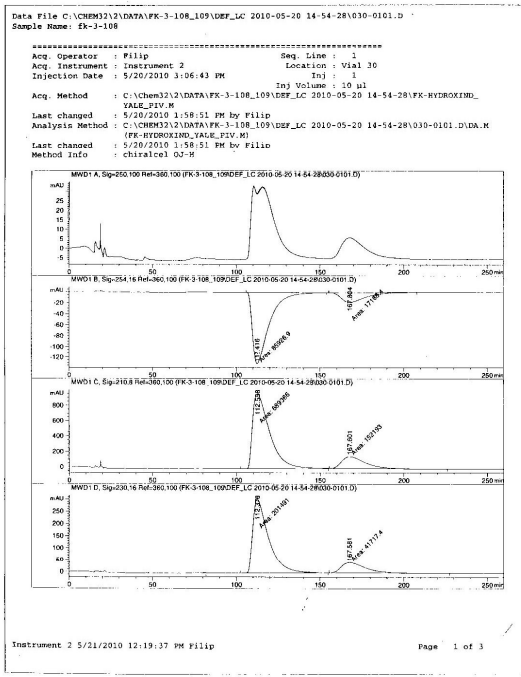
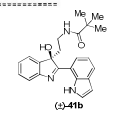
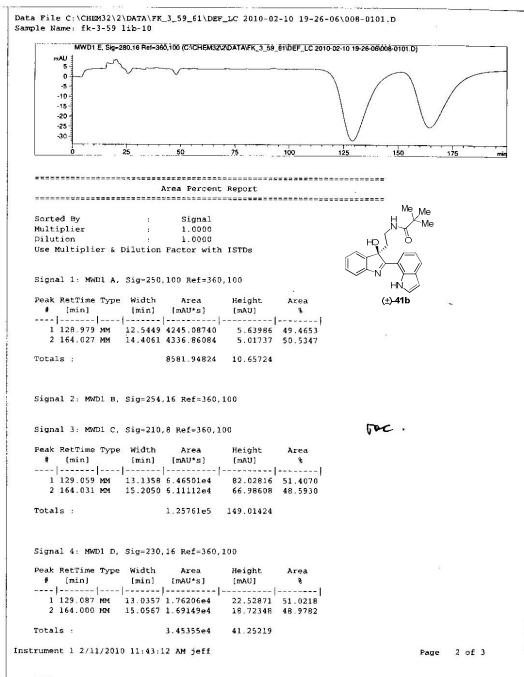
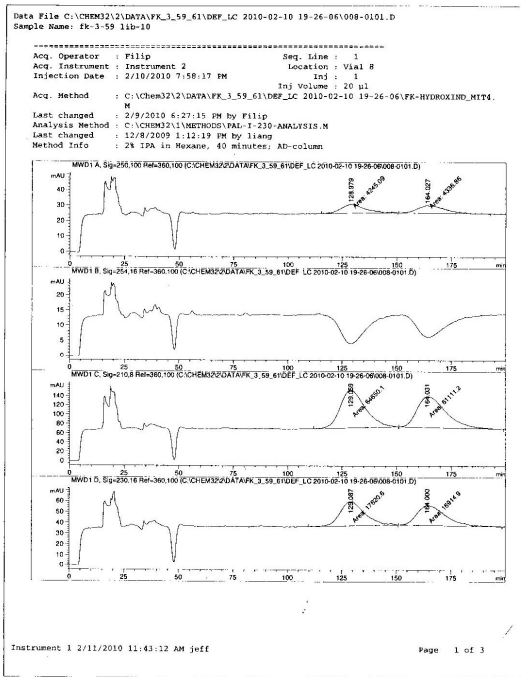
Signal 3: DAD1 C, Sig=210.8 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	27.328	NM	1.2821	157.55267	2.04815	0.5262
2	45.920	NM	1.9688	2.9785e4	252.15018	99.4738
Totals:				2.99431e4	254.19632	





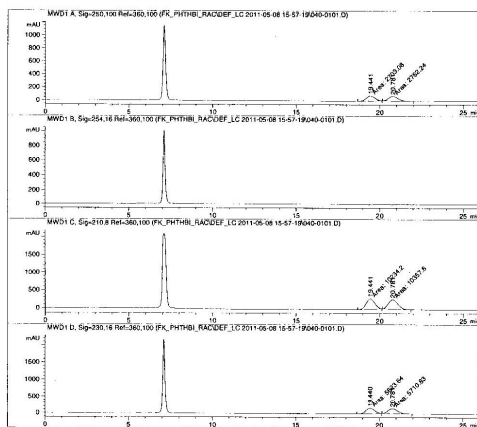




66% ee
83:17

Data File C:\CHEM32\2\DATA\FK_PHTHBI_RAC\DEF_IC 2011-05-08 15:57-19\040-0101.D
 Sample Name: Phtb rac

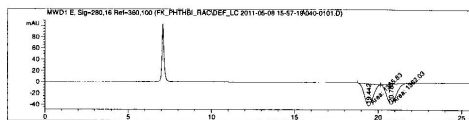
Acq. Operator : Filip Sep. Line : 1
 Acq. Instrument : Instrument 2 Location : Vial 40
 Injection Date : 5/8/2011 4:01:39 PM Inj : 1
 Inj Volume : 10 µl
 Acq. Method : C:\Chem32\2\DATA\FK_PHTHBI_RAC\DEF_IC 2011-05-08 15:57-19\FK-MONO-PHTHBI-ODH.M
 Last changed : 5/8/2011 3:10:27 PM by Filip
 Analysis Method : C:\CHEM32\2\DATA\FK_PHTHBI_RAC\DEF_IC 2011-05-08 15:57-19\040-0101.D\DA.M
 (FK-MONO-PHTHBI-ODH.M)
 Last changed : 5/8/2011 3:10:27 PM by Filip
 Method Info : chiralcel OD-H



Instrument 2 5/8/2011 4:41:41 PM Filip

Page 1 of 3

Data File C:\CHEM32\2\DATA\FK_PHTHBI_RAC\DEF_IC 2011-05-08 15:57-19\040-0101.D
 Sample Name: Phtb rac



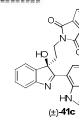
Area Percent Report

Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDs

Signal 1: MWD1 A, Sig=250.100 Ref=360.100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	19.441	MM	0.5919	2703.07861	76.10938	49.4587
2	20.781	MM	0.6558	2762.24292	70.19650	50.5413

Totals : 5465.32153 146.30585



Signal 2: MWD1 B, Sig=254.16 Ref=360.100

Signal 3: MWD1 C, Sig=210.8 Ref=360.100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	19.441	MM	0.5894	1.02342e4	289.41443	49.7002
2	20.781	MM	0.6518	1.03776e4	264.85392	50.2998

Totals : 2.05918e4 554.26925

Signal 4: MWD1 D, Sig=230.16 Ref=360.100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	19.440	MM	0.5880	5623.64111	159.39807	49.6154
2	20.781	MM	0.6514	5710.82813	146.11992	50.3846

Totals : 1.13345e4 305.51799

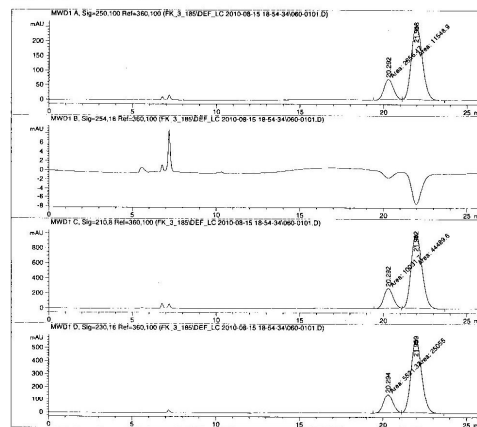
Instrument 2 5/8/2011 4:41:41 PM Filip

rac

4c

Data File C:\CHEM32\2\DATA\FK_3_185\DEF_IC 2010-08-15 18:54-34\060-0101.D
 Sample Name: FK-3-185

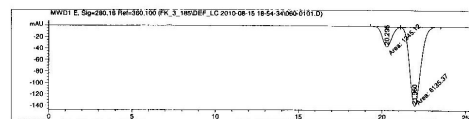
Acq. Operator : Filip Sep. Line : 1
 Acq. Instrument : Instrument 2 Location : Vial 60
 Injection Date : 8/15/2010 7:25:38 PM Inj : 1
 Inj Volume : 3 µl
 Acq. Method : C:\CHEM32\2\DATA\FK_3_185\DEF_IC 2010-08-15 18:54-34\FK-MONO-PHTHBI-ODH.M
 Last changed : 8/15/2010 6:53:38 PM by Filip
 Analysis Method : C:\CHEM32\2\DATA\FK_3_185\DEF_IC 2010-08-15 18:54-34\040-0101.D\DA.M (FK-HYDROXIND_TOS_ODH.M)
 Last changed : 5/8/2011 1:58:14 PM by Filip
 (modified after loading)
 Method Info : chiralcel OD



Instrument 2 5/8/2011 2:11:45 PM Filip

Page 1 of 3

Data File C:\CHEM32\2\DATA\FK_3_185\DEF_IC 2010-08-15 18:54-34\060-0101.D
 Sample Name: FK-3-185



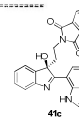
Area Percent Report

Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDs

Signal 1: MWD1 A, Sig=250.100 Ref=360.100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.292	MM	0.6420	2656.40729	68.94424	18.7005
2	21.958	MM	0.7622	1.15489e4	252.54510	81.2995

Totals : 1.42053e4 321.50935



Signal 2: MWD1 B, Sig=254.16 Ref=360.100

Signal 3: MWD1 C, Sig=210.8 Ref=360.100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.292	MM	0.6398	1.00317e4	261.54788	18.3986
2	21.962	MM	0.7580	4.44896e4	978.21533	81.6014

Totals : 5.45213e4 1239.76321

Signal 4: MWD1 D, Sig=230.16 Ref=360.100

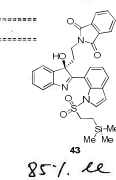
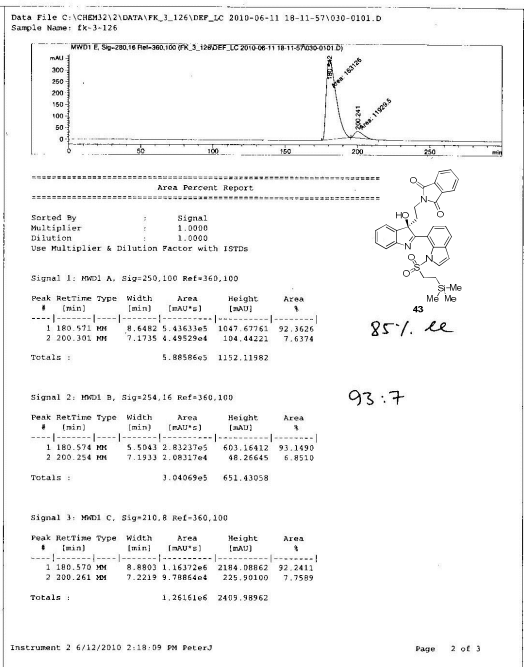
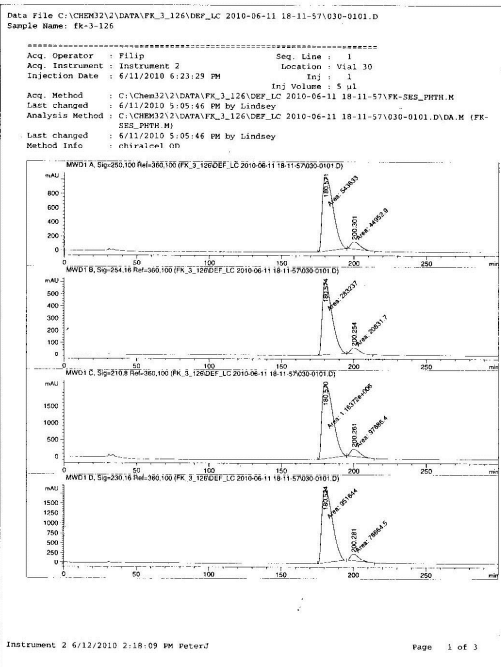
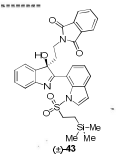
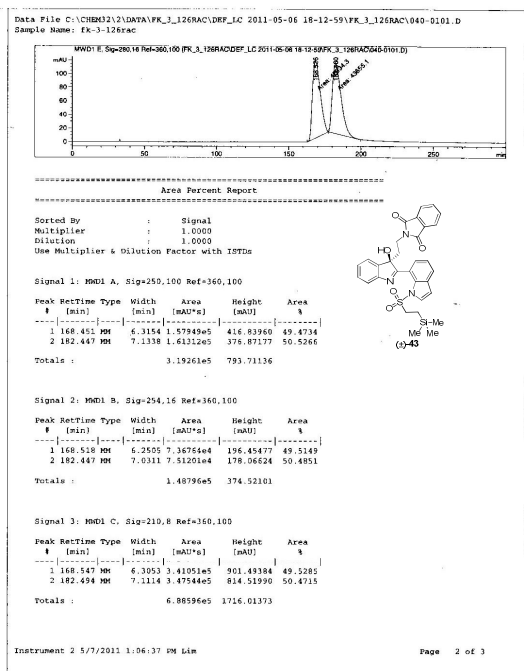
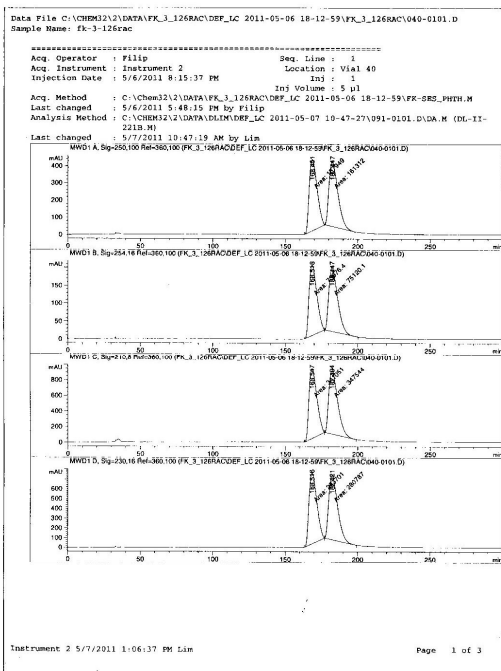
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.294	MM	0.6388	5211.32520	144.05894	18.0574
2	21.959	MM	0.7404	2.50550e4	563.99811	81.9425

Totals : 3.05763e4 708.05705

Instrument 2 5/8/2011 2:11:45 PM Filip

82.2 : 17.8

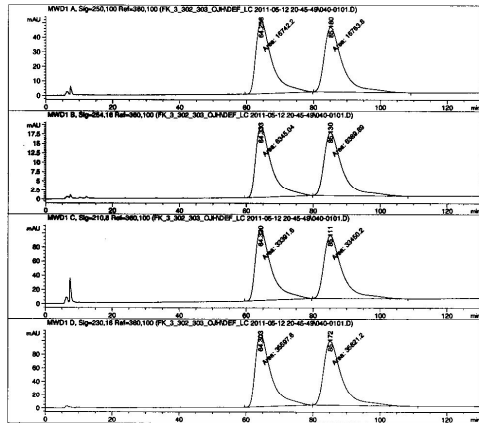
Page 2 of 3



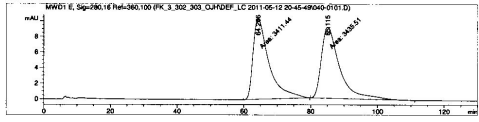
93:7

Data File C:\CHEM32\1\DATA\FK_3_302_303_OJH\DEF_LC 2011-05-12 20:45:49\040-0101.D
 Sample Name: FK-3-302

 Acq. Operator : Filip Seq. Line : 1
 Acq. Instrument : Instrument 2 Location : Vial 40
 Injection Date : 5/12/2011 9:57:24 PM Inj : 1
 Inj Volume : 6 ul
 Acq. Method : C:\Chem32\1\DATA\FK_3_302_303_OJH\DEF_LC 2011-05-12 20:45:49\FK-
 NAFFTYLEBARANO.M
 Last changed : 5/12/2011 8:00:23 PM by Filip
 Analysis Method : C:\CHEM32\1\DATA\FK_3_302_303_OJH\DEF_LC 2011-05-12 20:45:49\040-0101.D\
 DA.M (FK-NAFFTYLEBARANO.M)
 Last changed : 5/12/2011 8:00:23 PM by Filip
 Method Info : chiralcol OJ-H

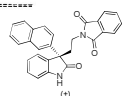


Data File C:\CHEM32\1\DATA\FK_3_302_303_OJH\DEF_LC 2011-05-12 20:45:49\040-0101.D
 Sample Name: FK-3-302



Area Percent Report

Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with %TMS



Signal 1: MWD1 A, Sig=250.100 Ref=360.100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	64.258	NM	5.5448	1.67422e4	50.32381	49.9678
2	85.180	NM	6.2804	2.67638e4	44.48713	50.0322
Totals:				3.35060e4		94.81094

Signal 2: MWD1 B, Sig=254.16 Ref=360.100

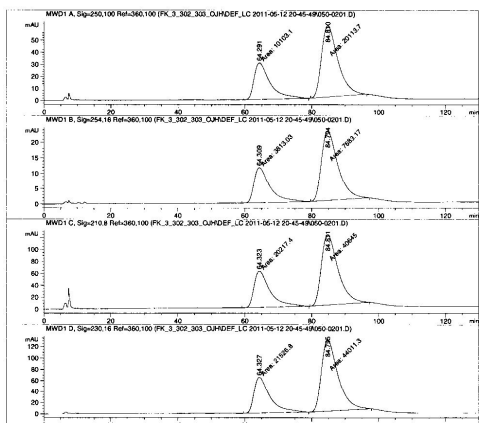
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	64.333	NM	5.5521	6345.03613	19.04701	49.8239
2	85.130	NM	6.3108	6389.88574	16.87547	50.1761
Totals:				1.27349e4		35.9248

Signal 3: MWD1 C, Sig=210.8 Ref=360.100

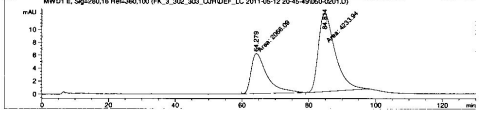
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	64.320	NM	5.5345	3.33916e4	100.53446	49.9562
2	85.111	NM	6.2597	3.3452e4	89.66238	50.0438
Totals:				6.68418e4		100.61584

Data File C:\CHEM32\1\DATA\FK_3_302_303_OJH\DEF_LC 2011-05-12 20:45:49\050-0201.D
 Sample Name: FK-3-303

 Acq. Operator : Filip Seq. Line : 2
 Acq. Instrument : Instrument 2 Location : Vial 50
 Injection Date : 5/13/2011 2:57:57 AM Inj : 1
 Inj Volume : 6 ul
 Acq. Method : C:\Chem32\1\DATA\FK_3_302_303_OJH\DEF_LC 2011-05-12 20:45:49\FK-
 NAFFTYLEBARANO.M
 Last changed : 5/12/2011 8:00:23 PM by Filip
 Analysis Method : C:\CHEM32\1\DATA\FK_3_302_303_OJH\DEF_LC 2011-05-12 20:45:49\050-0201.D\
 DA.M (FK-NAFFTYLEBARANO.M)
 Last changed : 5/13/2011 8:03:20 AM by Filip
 (modified after loading)
 Method Info : chiralcol OJ-H

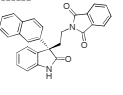


Data File C:\CHEM32\1\DATA\FK_3_302_303_OJH\DEF_LC 2011-05-12 20:45:49\050-0201.D
 Sample Name: FK-3-303



Area Percent Report

Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with %TMS



Signal 1: MWD1 A, Sig=250.100 Ref=360.100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	64.293	NM	5.5758	1.01032e4	30.99999	33.4353
2	84.810	NM	5.6723	2.01137e4	59.09898	66.5647
Totals:				3.02168e4		89.2907

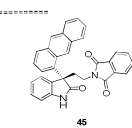
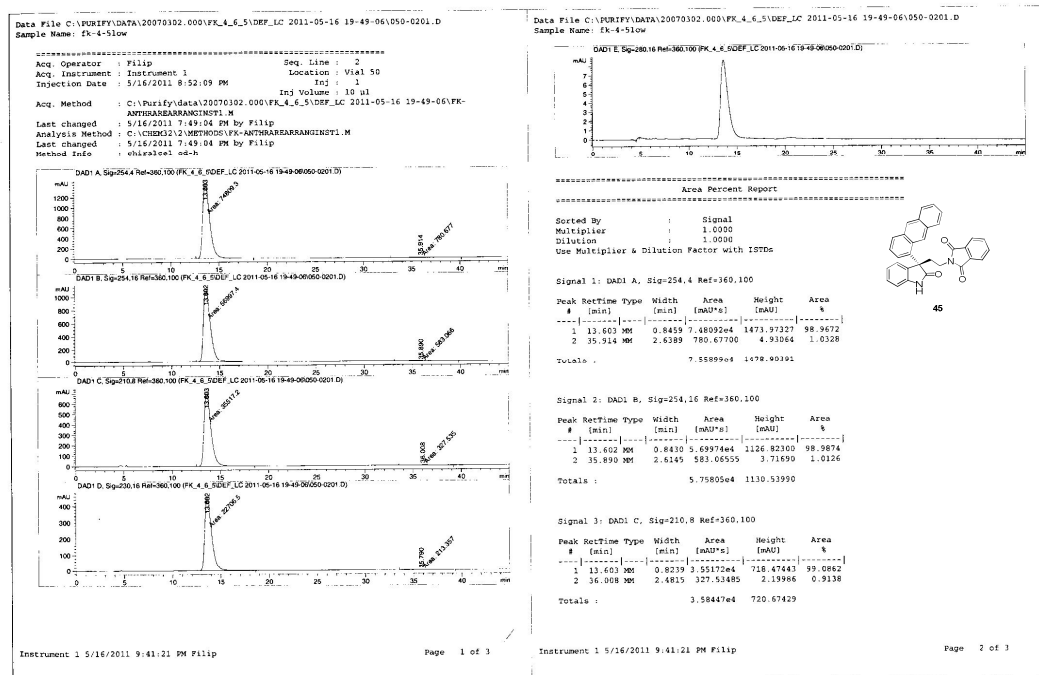
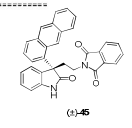
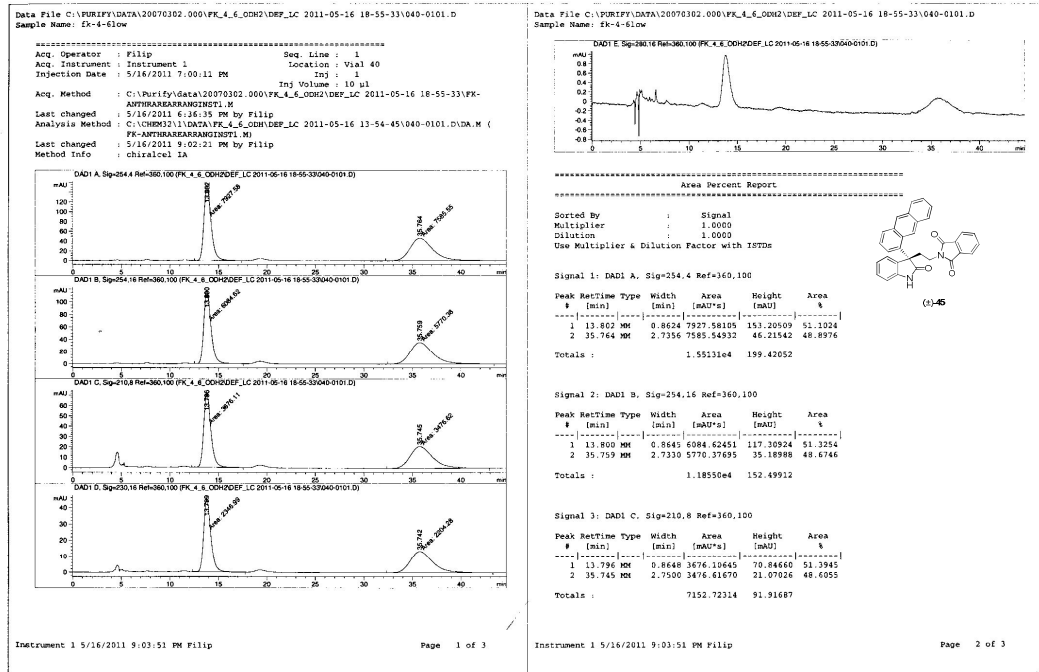
Signal 2: MWD1 B, Sig=254.16 Ref=360.100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	64.309	NM	5.5597	3813.02808	11.43055	33.1677
2	84.794	NM	5.7139	7683.16748	22.41063	66.8323
Totals:				1.14962e4		33.84138

Signal 3: MWD1 C, Sig=210.8 Ref=360.100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	64.322	NM	5.5708	2.02174e4	60.48874	33.2282
2	84.811	NM	5.7259	4.06450e4	118.20669	66.7818
Totals:				6.08625e4		178.79244

0.7 ml/min, 55:45 Hex:EtOH



M. References.

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9. Least Squares function minimized: (SHELXL97)

$$\Sigma w(F_o^2 - F_c^2)^2 \quad \text{where } w = \text{Least Squares weights.}$$

10. Standard deviation of an observation of unit weight:

$$[\Sigma w(F_o^2 - F_c^2)^2 / (N_o - N_v)]^{1/2}$$

where N_o = number of observations

N_v = number of variables

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