Supplementary Material

Structural Basis for the Function of Tim50 in the Mitochondrial Presequence Translocase Qian, X. *et al.*

Generation and synthesis of Tim50 mutant forms

Site-directed mutants were created by using the QuickChange Site-Directed Mutagenesis Kit (Stratagene, Santa Clara, CA). Tim 50_{1-361} mutants were amplified and fused to a SP6 promotor by polymerase chain reactions, transcribed with the mMESSAGE mMACHINE Kit (Ambion, Austin, TX) and subsequently translated by using rabbit reticulocyte lysate (Promega, Madison, WI) in the presence of [³⁵S]methionine.³¹

Import of proteins into mitochondria

Mitochondria were isolated from a yeast strain expressing Tim23-protein A^{15} and the corresponding wild-type strain grown in YPG (1% yeast extract, 2% bacto peptone, 3% glycerol) medium at 24°C. Mitochondria were preincubated for 2 min in import buffer (10 mM MOPS/KOH, pH 7.2, 3% [w/v] bovine serum albumin, 250 mM sucrose, 80 mM KCl, 5 mM MgCl₂, 2 mM KH₂PO₄, 5 mM methionine) at 25°C. Import reactions were started by the addition of radiolabeled precursor proteins and stopped by adding AVO-mix (8 µM antimycin A, 1 µM valinomycin, 20 µM oligomycin) on ice to dissipate the membrane potential.³¹ Protease treatment was performed by adding 50 µg/ml proteinase K and stopped after 15 min on ice by the addition of 2 mM PMSF. Mitochondria were reisolated by centrifugation, washed with SEM buffer (10 mM MOPS/KOH, pH 7.2, 250 mM sucrose, 1 mM EDTA) and analyzed by SDS-PAGE and digital autoradiography with a Storm 820 image analyzer (GE Healthcare).

Affinity chromatography

For protein complex isolation, mitochondria were solubilized in solubilization buffer (20 mM Tris/HCl, pH 7.4, 50 mM NaCl, 0.1 mM EDTA, 10% glycerol) containing 1% digitonin.

Protease inhibitors (2 mM PMSF, 2 mM Pefablock SC [Roche] and 4 µg/ml leupeptine) were added and the samples were incubated for 30 min at 4°C. 30 µl 50% IgG sepharose slurry per column was washed two times with 0.5 ml acetate buffer (0.5 M HAc/NH₄Ac, pH 3.5), two times with 2-fold solubilization buffer without digitonin and two times with solubilization buffer (see above). Sepharose beads were incubated with solubilized mitochondria and shaken for 90 min at 4°C. Columns were washed ten times with washing buffer (20 mM Tris/HCl, pH 7.4, 60 mM NaCl, 0.5 mM EDTA, 10% glycerol, 0.3% digitonin and 2 mM PMSF). Bound proteins were eluted with 50 µl SDS loading buffer without β -mercaptoethanol, incubated for 15 min at 37°C and subjected to SDS-PAGE. Gels were analyzed by immunoblotting or by digital autoradiography.

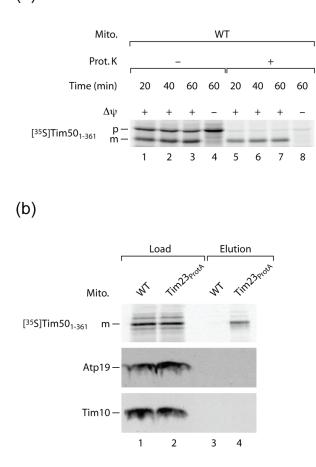
Supplementary Reference

 Stojanovski, D., Pfanner, N. & Wiedemann, N. (2007). Import of proteins into mitochondria. *Methods Cell Biol.* 80, 783-806.

Supplementary Figures

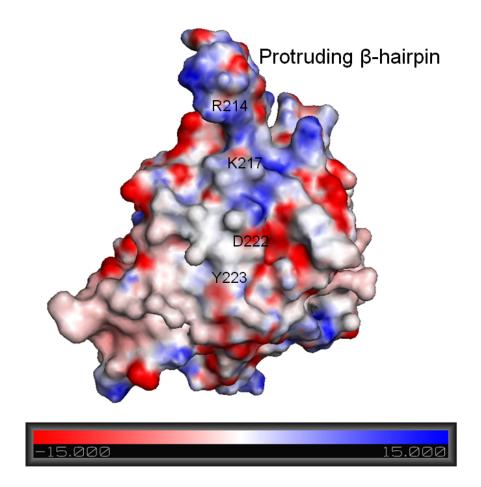
Supplementary Fig. S1. Sequence alignment of the Tim50 family members. The program ClustalW was utilized to align the Tim50 sequences from ten species. The conserved regions are highlighted in blue color. The fragment used for crystallization of Tim50_{IMS} is marked by a black bar. The α -helices A1 to A5 and the β -strands B1 to B9 are labeled by red bars and green arrows, respectively. The protruding β -hairpin is formed by B2, B3 and the connecting loop.

S.cerevisiae C.elegans H.sapiens D.melanogaster N.crassa D.rerio M.musculus B.taurus P.pastoris D.hansenii	1 -MLSILRNSVRLNSRALRVVP20 1 -MSLSKLTQTFSRHQA16 1 -MSLSKLTQTFSPRVENPFIKSLTP 1 -MSLSKLTGDVAVGNTISVRIQPPNGSGMTFSPKVLHFEKNKKFQWKGKLLFEGLFDGEHVFELIDNGDGTLFKQSE109 1 -MLRN - TRLLTRT112	
S.cerevisiae C.elegans H.sapiens D.melanogaster N.crassa D.rerio M.musculus B.taurus P.pastoris D.hansenii	21 SAANTLTSVQASRRLLTSY SSFLQK ETKDDKPKSILTDDMLFKAGVDVDEKGQGKNEETSGE82 17 KTFIRLYSSDFKSLLGPPAVANPYADNGRTRFAPIVPINHGNVFASIKLPINETQEAIAFKSEVEEAPKVEKLEVESPKIEAEKVLSSPPPAPAPTSSAIDELNSLKDSLE127 6 AVFSRLRSG LRLGSRGLCTR LATPPRRAPDQAAEIGSRGSTKAQ 84 EPAESEKAEQKQQQQQQQ FTLLHHQQYLRLFTCTALPAAAPALFSILHTARGYSSTTKQEAGATGPNDAPEVAPNAPLLAKLFP113 64 EPAESEKAEQKQQQQQQQ FTPAESEPEPEIDLSKLPDLRGGIPTTLEYEMAQKEAGKKPVAGEAETQAEGAE136 6 VYPMCVRASR GLRLRQGARGLCTR LAPPPRTPEQVTEIANRGGSKAQ49 6 ALFSRLRSG LRVGARGLCTR LAPPPRTPEQVTEIANRGGSKAQ49 6 AVFLRLRSG LRVGARGLCTR LAPPPRAPDQAAEIGSRGSTKAQ49 10 TFNGIFVSGRAT LAPPPRAPDQAAEIGSRAGTKAQ49 LAVGARGLCTR 13 SSSLRFAATKNT RLPVQH KFYSKK TDKKAEEPQSILTDDLLAKAGFEDPNEPKEKSEQQESE74	
S.cerevisiae C.elegans H.sapiens D.melanogaster N.crassa D.rerio M.musculus B.taurus P.pastoris D.hansenii	83 GGEDKNEPSSKSE KTRRKRQ TSTDIKREKYANWFYIFSLSALTGTAIYMARDWEPQESEELKK DIDHGYTLSLMYKRFKARFNSMFTYFQEP14 128 KLESAASKSSSSGGSSDN SDPGNAEEIEARRKRMERNTRIGAYVLFGGSIIGFISFCFYYGRAQRDEFGNVISD EFSG.SFLAPFYRIANSFKLWRDYV P229 50 - GPQQPQSEGP SYAKKVALVLAGLGAGGTVS VYIFGNNPVDENGAKIPD EFDNDPILVQQLRTYKYFKDYRQMIIEP129 114 QTSPEVDSNAEQE RKKREEEEEKNERAWKRMKLGFA FIGGSAVAAGFWAVYEFGKPEVDPNGQPIED EFTHKPLVQQLRTYKYFKDYRQMIIEP129 137 GPEAATSGSGGG RKKQDPDSYSTEKRRQKMANWAFIAAGLALVGGTIYLGREWDEEELEKNHD IFNGWGLGUWKRAKARAMTGTVSYY QRMIQEP210 137 GPEAATSGSGGG RKKQDEQDSAYVSSTEKRRQKMANWAFIAAGIALVGGTIYLGREWDEEELEKNHD IFNGWGLGUWKRAKARAMTGTVSYY QRMIQEP220 59 ERLQQQQKSGEQPPPEGEDSGHKQDEQGEDKKQKENTAYAKKMVLRLAGIMGLGGTVG IYIFGNNVDENGTKIPD IFDNDVDVQQRRTFKYFKDYRQMIIEP129 59 GRLQQQRSSEGP SYAKKVALWLAGLLGAGGTVS IYIFGNNVDENGTKIPD IFDDNDVVQQRRTFKYFKDYRQMIIEP164 50 TGGPQQQRSSEGP SYAKKVALWLAGLLGAGGTVS IYIFGNNVDENGTKIPD IFDNDVVQQRRTFKYFKDYRQMIIEP164 50 TGGPQQQRSSEGP SYAKKVALWLAGLLGAGGTVS IYIFGNNAVDENGTKIPD IFDSDPILVQQLRTYKYFKDYRQMIIEP164 50 TGGPQQQRSSEGP SYAKKVALWLAGLLGAGGTVS IYIFGNNAVDENGTKIPD IFDNDVVIQLRTYKYFKDYRQMIIEP164 50 TGGPQQQRSSEGP SYAKVALWLAGLLGAGGTVS IYIFGNNAVDENGAKIPD IFDNDVVIQLRTYKYFKDYR	
S.cerevisiae C.elegans H.sapiens D.melanogaster N.crassa D.rerio M.musculus B.taurus P.pastoris D.hansenii	175 P F P D LL P P P P P Y Q R P - L T L V I T L E D F , V H S EWS QKH GWR TAK R P G A D Y F L GYL - SQY Y E I V L E'S NYMMYS D K I A E KL D P I HA F VS Y N L F K E H C V Y K D G V H I KD L S K L N 233 230 A R E Q LL P D P L P A Y L Q P K Y T I V I E L K N I L V H P EWT Y K T GYR F L K R P A L D Y F L D V I GYP N F E V V I Y S S E SMMT A A P V V D S F D P K Q - R I M Y K L F R D C T K YMN G H H V KD L S K L N 339 130 A R E Q LL P D P L Q E P Y Q Q P Y T L V L E L T G V L H P EWS LAT GWR F K K R P G I E T L F Q L - A P L Y E I V I F T S E T GMT A F P L I D S V D P H G - F I S Y R L F R D A T R YMD G H H V KD I S C L N 238 211 S R A K LL P D P L V Q P Y T L V L E M KD V L V H P D WT Y Q T GWR F K K R P G V D H F L A C - A KD F E I V V F T A E Q GMT V F P I LD A L D P H G - Y I M YR L V R D A T H F V D G H H V KH D H N 19 231 A F E K LL P D P L N P Y V Q P R Y T L V L E M M V N S E W T R D H GWR F K K R P G V D H F L A C - A KD F E I V V F T A E Q GMT V F P I LD A L D P H G - Y I M YR L V R D A T H F V D G H H V KH D H N 19 233 A F E K LL P D P L N P Y V Q P R Y T L V L E M M D W N T G W R F K K R P G V D F H L Y L S Q Y E I V L F T S V F A A A E P I V P K M D Y R - F I M YR L Y R D A T R YM E G H H V KD V S C L N 238 165 T S P K LL P D P L R P Y Y Q P P Y T L V L E L T G V L H P E W S L A T GWR F K K R P G I E T L F Q L - A P L Y E I V I F T S E T GMT A F P L I D S V D P H G - F I S YR L F R D A T R YM E G H H V KD I S C L N 238 130 T S P C L L P D P L R P Y Y Q P P Y T L V L E L T G V L H P EWS L A T GWR F K K R P G I E T L F Q L - A P L Y E I V I F T S E T GMT A F P L I D S V D P H G - F I S YR L F R D A T R YM E G H H V KD I S C	
S.cerevisiae C.elegans H.sapiens D.melanogaster N.crassa D.rerio M.musculus B.taurus P.pastoris D.hansenii	284 RD L SK VI I I D T D P N S Y KL OP E H A I P ME P WN GE - AD D KL V RL I P EL EY L A T O O T KD V R P I L N S FED K KNL A E E FD H R V K KL K D K F Y GD H K	
S.cerevisiae C.elegans H.sapiens D.melanogaster N.crassa D.rerio M.musculus B.taurus P.pastoris D.hansenii	375 NWAMTALGLG - NSLGGSTKFPLDLIHEEGQKNYLMFMKMIEEEKEKIRIQQEQMGGQT - FTLKDYVEGNLPSPEEQMKIQLEKQKEVDALFEEEKKKKIAESK 476 434 S - MLKRYSGRLFGSRRHVNA - 353 333 NKQNLFLGSLTSRLWPRSKQP - 353 412 SKTKPMVKQWSRNILGR - 428 450 HPMAMEGEEDPSEAFAKGKMIQDIARERGMRNYLAMEEEIKKNGEMWLKMEQEAQEKAQKEMMKNMQSSVFGWFGGAPSGEQQSG - ESEKKA	



Supplementary Fig. S2. Import of truncated Tim50 into isolated yeast mitochondria and interaction with Tim23. (a) Tim50₁₋₃₆₁ was synthesized in reticulocyte lysate and labeled with [35 S]methionine. Mitochondria were isolated from yeast wild-type (WT) cells. Tim50₁₋₃₆₁ was imported into the isolated mitochondria and processed to the mature form in a membrane potential ($\Delta\psi$)-dependent manner (lanes 1-3). Mature Tim50₁₋₃₆₁ was protected against externally added proteinase K (lanes 5-7). The mitochondria were separated by SDS-PAGE and radiolabeled Tim50₁₋₃₆₁ was detected by digital autoradiography. p, precursor; m, mature. (b) Tim50₁₋₃₆₁ was imported into control mitochondria (WT). After the import reaction, the mitochondria were lysed with digitonin and subjected to affinity purification with IgG-Sepharose. The eluate containing Tim23 and associated proteins is shown in lane 4. The samples were separated by SDS-PAGE and mitochondrial Tim23. Control proteins, Atp19 and Tim10, were detected by Western blotting. Load, 10%; elution, 100%.

(a)



Supplementary Fig. S3. Electrostatic surface potential drawing of Tim50_{IMS}. The Tim50 molecule is rotated along the vertical axis by ~90° from the orientation in Fig. 1a. The negatively charged region is shown in red and the positively charged region is shown in blue. The protruding β -hairpin is labeled. The residues R214 and K217 that are involved in binding Tim23 are labeled. Residues D222 andY223, which were replaced for mutagenesis studies, are also labeled.