

**Nickel–Catalyzed, Carbonyl–Ene–Type Reactions: Selective for Alpha Olefins and
More Efficient with Electron–Rich Aldehydes**

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Supporting Information

Experimental Procedures, Analytical and Spectroscopic Data for Compounds.

General Aspect.

Unless otherwise indicated, all reactions were performed under an oxygen–free atmosphere of nitrogen or argon with rigid exclusion of moisture from reagents and glassware. Bis(cyclooctadienyl)nickel(0) (Ni(cod)₂) and triphenylphosphine were purchased from Strem Chemicals, Inc., stored under nitrogen atmosphere and used without further purification. Ethyl diphenylphosphinite was purchased from Aldrich Chemical Co. and was used as received. All alkenes and aldehydes were used as received without further purification. Triethylsilyl trifluoromethanesulfonate (TESOTf), triethylamine and toluene were distilled over calcium hydride before use.

Analytical thin layer chromatography (TLC) was performed using EM Science silica gel 60 F254 plates. The developed chromatogram was analyzed by UV lamp (254 nm), ethanolic phosphomolybdic acid (PMA) or potassium permanganate (KMnO₄). Liquid chromatography was performed using a forced flow (flash chromatography) of the indicated solvent system on Silicycle Silica Gel (230–400 mesh). ¹H and ¹³C NMR spectra were recorded on Bruker 400 MHz or Varian 500 MHz spectrometers in CDCl₃. Chemical shifts in ¹H NMR spectra are reported in ppm on the δ scale from an internal

standard of residual chloroform (7.27 ppm). Data are reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, br = broad), coupling constant in hertz (Hz), and integration. Chemical shifts of ^{13}C NMR spectra are reported in ppm from the central peak of CDCl_3 (77.23 ppm) on the δ scale. Infrared (IR) spectra were recorded on a Perkin–Elmer 2000 FT–IR. High resolution mass spectra (HRMS) were obtained on a Bruker Daltonics APEXII 3 Tesla Fourier Transform Mass Spectrometer by Dr. Li Li of the Massachusetts Institute of Technology, Department of Chemistry Instrument Facility.

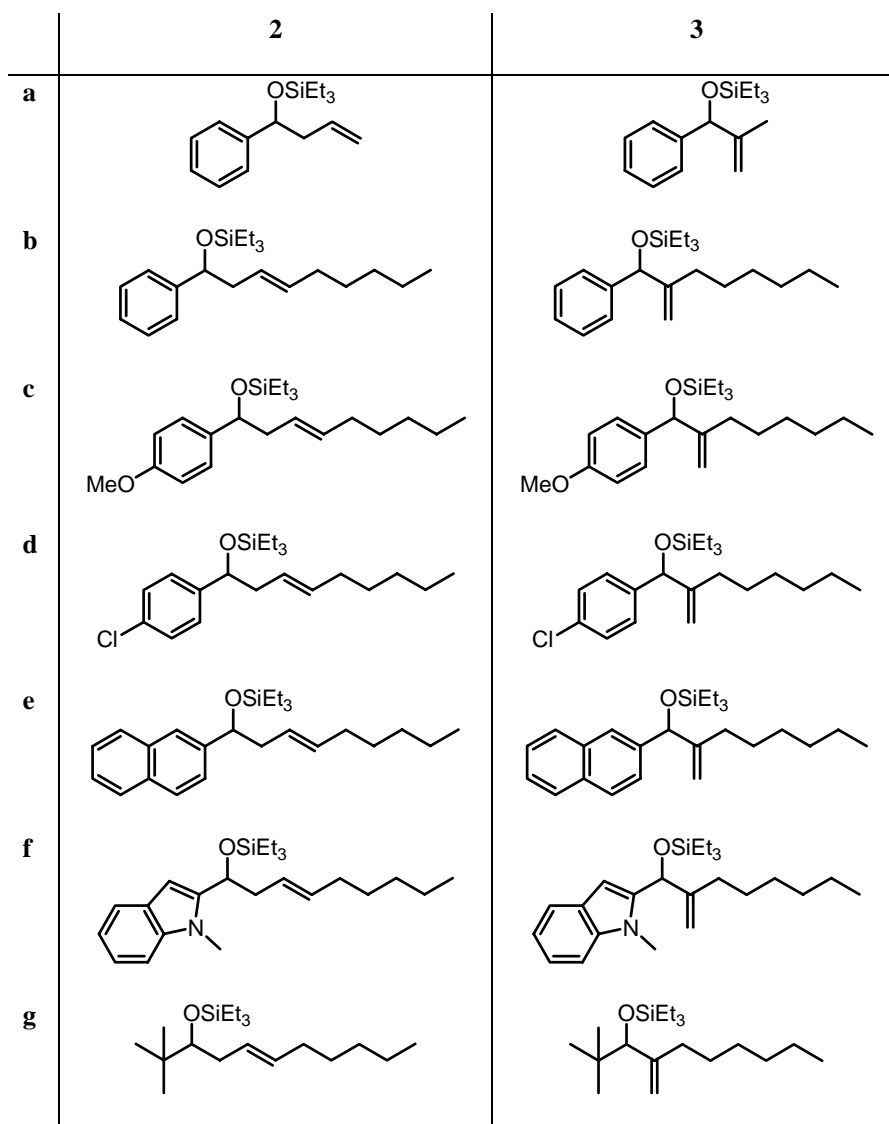
General Procedure for the Nickel–Catalyzed Coupling Reaction.

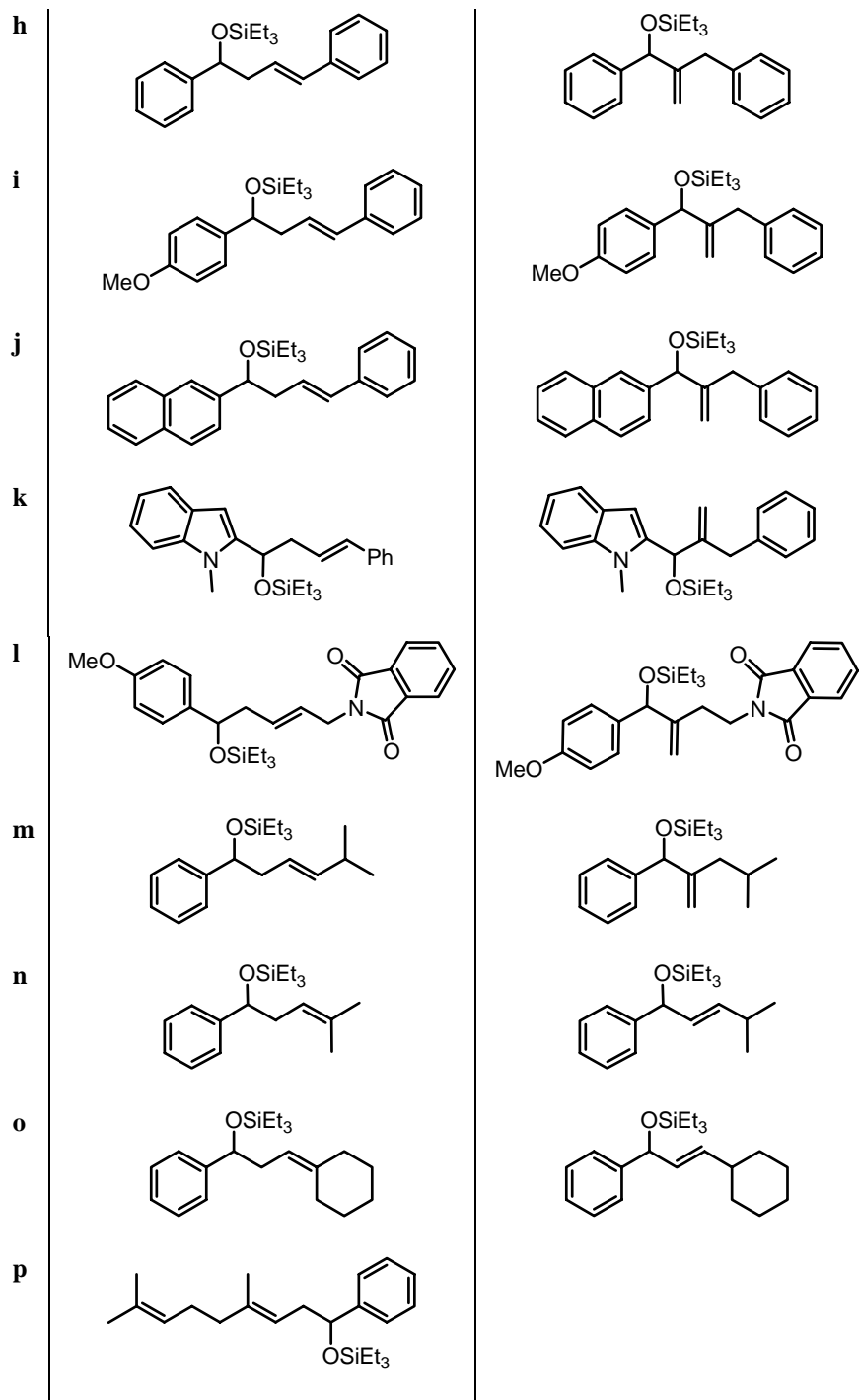
A 10 mL test tube and a stir bar were oven–dried and brought into a glove box. $\text{Ni}(\text{cod})_2$ (0.1 mmol, 20 mol%) and phosphine ligand (0.2 mmol, 40 mol%) were added to the tube. The tube was sealed with a septum, and was brought out of the glove box and connected to an argon line. The catalyst mixture was dissolved in degassed toluene (2.5 mL) under argon and stirred at room temperature. The alkene (1.5 mmol, 300 mol% to 0.5 mL), triethylamine (3 mmol, 600 mol%), aldehyde (0.5 mmol, 100 mol%) were added sequentially to the reaction mixture. Triethylsilyltriflate (0.875 mmol, 175 mol%) was added finally to the reaction mixture in dropwise fraction and the mixture was stirred at room temperature or at indicated temperature for 48 h. The mixture was diluted with ether and filtered through a plug of silica gel. Solvent was removed under reduced pressure. Purification via flash chromatography on silica gel (1% ethyl acetate in hexane, unless otherwise indicated) afforded the coupling product.

In some cases, the coupling product overlapped with the starting materials,

characterization data of the corresponding alcohol was reported instead. The corresponding alcohol product was obtained by treating the mixture with 1 M TBAF in THF at 0 °C and allowed to stir at room temperature for 2 hours, followed by flash column chromatography on silica (15% ethyl acetate in hexane, unless otherwise indicated). E:Z selectivity for the compound **2** was determined by comparing the TES group deprotected alcohol product with the literature.^{1,2}

Figure 1. Structures of the Coupling Products





Compound Characterization

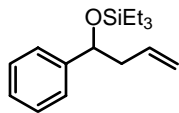


Table 1, entry 1: **2a**:

The standard procedure was followed, except that a balloon filled with propene (**1a**, 1 atm) was used in place of Ar. Yield: 65%.

^1H NMR (400 MHz, CDCl_3 , δ): 7.27–7.38 (m, 5H); 5.78–5.89 (m, 1H); 5.05–5.10 (m, 2H); 4.74 (dd, $J = 7.2, 5.5$ Hz, 1H); 2.42–2.59 (m, 2H); 0.94 (t, $J = 7.9$ Hz, 9H); 0.59 (dq, $J = 2.6, 7.9$ Hz, 6H).

^{13}C NMR (100 MHz, CDCl_3 , δ): 145.3, 135.4, 128.2, 127.2, 126.1, 117.0, 75.1, 45.6, 7.0, 5.0.

IR (NaCl, thin film): 3029, 2956, 2912, 2877, 1641, 1493, 1454, 1414, 1239, 1089, 1068, 1006, 914, 839, 744, 699.

HRMS–ESI (m/z): $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{16}\text{H}_{26}\text{OSiNa}$, 285.1651; found, 285.1633.

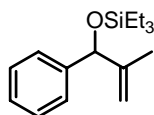


Table 1, entry 1: **3a**:

The standard procedure was followed, except that a balloon filled with propene (**1a**, 1 atm) was used in place of Ar. Yield: 8%.

^1H NMR (400 MHz, CDCl_3 , δ): 7.24–7.39 (m, 5H); 5.15 (m, 2H); 4.86 (s, 1H); 1.56 (s, 3H); 0.94 (t, $J = 7.8$ Hz, 9H); 0.61 (q, $J = 7.8$ Hz, 6H). ^{13}C NMR (100 MHz, CDCl_3 , δ): 148.1, 143.5, 128.1, 127.0, 126.3, 111.0, 78.4, 17.4, 7.0, 5.0.

IR (NaCl, thin film): 2955, 2913, 2877, 1451, 1237, 1091, 1066, 1005, 899, 853, 740, 698.

HRMS–ESI (m/z): [M+Na]⁺ calcd for C₁₆H₂₆OSiNa, 285.1645; found, 285.1651.

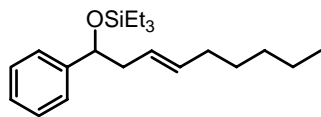


Table 1, entry 2 and 3: **2b'**

The standard procedure was followed (entry 2). Yield: 81%. Isolated as a 75:25 mixture of E and Z isomers, respectively.

The standard procedure was followed, except that the reaction time was 18 h (entry 3). Yield: 68%. Isolated as a 75:25 mixture of E and Z isomers, respectively.

¹H NMR (400 MHz, CDCl₃, δ): 7.20–7.40 (m, 5H); 5.30–5.50 (m, 2H); 4.63 (dd, *J* = 5.6, 7.2 Hz, 1H); 2.45 (quintet, *J* = 6.1 Hz, 1H); 2.35 (quintet, *J* = 5.9 Hz, 1H); 1.33 (m, 2H); 0.92 (t, *J* = 7.8 Hz, 12H); 0.56 (dq, *J* = 2.4, 7.6 Hz, 6H).

¹³C NMR (100 MHz, CDCl₃, δ): 145.6, 133.3, 128.1, 127.1, 126.6, 126.2, 75.6, 44.5, 32.8, 31.6, 29.3, 22.8, 14.2, 7.0, 5.1.

HRMS–ESI (m/z): [M+Na]⁺ calcd for C₂₁H₃₆OSi, 355.243; found, 355.244.

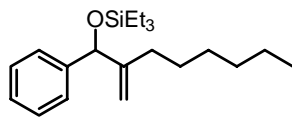


Table 1, entry 2 and 3: **3b'**

The standard procedure was followed (entry 2). Yield: 4%.

The standard procedure was followed, except that the reaction time was 18 h (entry 3). Yield: 4%.

¹H NMR (400 MHz, CDCl₃, δ): 7.36 (d, *J* = 7.0 Hz, 2H); 7.31 (t, *J* = 7.1 Hz, 2H); 7.24 (t, *J* = 7.2, 1H); 5.22 (brs, 1H); 5.15 (brs, 1H); 5.25 (d, *J* = 5.9 Hz, 1H); 4.87 (s, 1H); 1.96 (pentet, *J* = 7.8 Hz, 1H); 1.76 (pentet, *J* = 8.0 Hz, 1H); 1.15–1.40 (m, 8H); 0.93 (t, *J* = 8.0

Hz, 9H); 0.87 (t, $J = 6.8$ Hz, 3H); 0.60 (dq, $J = 1.6, 7.9$ Hz, 6H).

^{13}C NMR (100 MHz, CDCl_3 , δ): 152.3, 143.8, 128.1, 127.1, 126.6, 109.5, 78.3, 32.0, 30.8, 29.4, 28.0, 22.8, 14.3, 7.0, 5.1.

IR (NaCl, thin film): 2956, 2876, 1647, 1456, 1089, 1066, 742.

HRMS–ESI (m/z): $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{21}\text{H}_{36}\text{OSi}$, 355.243; found, 355.242.

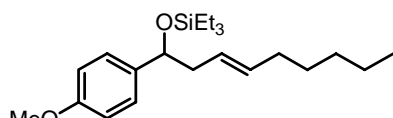


Table 1, entry 4: **2c**:

The standard procedure was followed, except that the reaction time was 18 h. Yield: 81%. Isolated as a 75:25 mixture of E and Z isomers, respectively.

^1H NMR (400 MHz, CDCl_3 , δ): 7.22 (d, $J = 8.6$ Hz, 2H); 6.84 (d, $J = 8.6$ Hz, 2H); 5.33–5.43 (m, 2H); 4.58 (dd, $J = 6.1$ Hz, 6.1 Hz, 1H); 3.81 (s, 3H); 2.27–2.42 (m, 2H); 1.93–1.98 (m, 2H); 1.22–1.60 (m, 6H); 0.95 (t, $J = 8.0$ Hz, 3H); 0.88 (t, $J = 7.8$ Hz, 9H); 0.53 (q, $J = 7.8$ Hz, 6H). ^{13}C NMR (100 MHz, CDCl_3 , δ): 158.7, 137.9, 133.2, 127.3, 126.7, 113.4, 75.2, 55.4, 44.5, 32.8, 31.6, 29.3, 22.8, 14.3, 7.0, 5.0.

IR (NaCl, thin film): 2955, 2876, 1613, 1512, 1459, 1302, 1247, 1172, 1078, 1005, 972, 830, 742.

HRMS–ESI (m/z): $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{22}\text{H}_{38}\text{O}_2\text{SiNa}$, 385.2539; found, 385.2537.

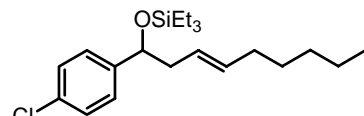


Table 1, entry 5: **2d**:

The standard procedure was followed, except that the reaction time was 18 h. Yield: 35%. Isolated as a 74:26 mixture of E and Z isomers, respectively.

^1H NMR (400 MHz, CDCl_3 , δ): 7.28–7.24 (m, 4H); 5.30–5.41 (m, 2H); 4.61 (dd, $J = 6.1$ Hz, 6.1 Hz, 1H); 2.26–2.40 (m, 2H); 1.89–1.97 (m, 2H); 1.21–1.59 (m, 6H); 0.94 (t, $J = 8.0$ Hz, 3H); 0.89 (t, $J = 7.8$ Hz, 9H); 0.54 (q, $J = 7.8$ Hz, 6H).

^{13}C NMR (100 MHz, CDCl_3 , δ): 144.1, 133.7, 132.5, 128.2, 127.5, 126.0, 74.8, 44.4, 32.8, 31.5, 29.2, 22.7, 14.3, 7.0, 4.9.

IR (NaCl, thin film): 2956, 2876, 1490, 1458, 1412, 1238, 1080, 1014, 971, 741.

HRMS–ESI (m/z): $[\text{M}+\text{Na}-\text{C}_6\text{H}_{15}\text{SiOCl}]^+$ calcd for $\text{C}_{15}\text{H}_{20}\text{Na}$, 223.1463; found, 223.1305.

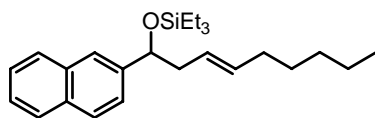


Table 1, entry 6: **2e**:

The standard procedure was followed, except that the reaction was heated at 35 °C. Yield: 84%. Isolated as a 70:30 mixture of E and Z isomers, respectively.

^1H NMR (400 MHz, CDCl_3 , δ): 7.83–7.92 (m, 3H); 7.80 (s, 1H); 7.48–7.59 (m, 3H); 5.43–5.53 (m, 2H); 4.89 (dd, $J = 6.9, 13.2$ Hz, 1H); 2.45–2.68 (m, 2H); 1.98–2.05 (m, 2H); 1.26–1.39 (m, 6H); 0.97 (t, $J = 8.0$ Hz, 9H); 0.94 (t, $J = 7.6$ Hz, 3H); 0.63 (q, $J = 4.1, 8.0$ Hz, 6H).

^{13}C NMR (100 MHz, CDCl_3 , δ): 143.1, 133.4, 133.0, 132.3, 128.1, 127.9, 126.5, 126.0, 125.6, 125.6, 124.7, 124.7, 75.7, 44.4, 32.8, 31.7, 29.3, 22.8, 14.3, 7.0, 5.1.

IR (NaCl, thin film): 2956, 2929, 2875, 1458, 1414, 1377, 1239, 1086, 1005, 972, 744.

HRMS–ESI (m/z): $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{25}\text{H}_{38}\text{OSiNa}$, 405.2590; found, 405.2584

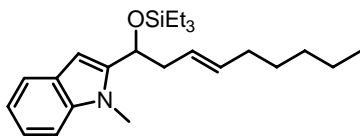


Table 1, entry 7: **2f**:

The standard procedure was followed, except that the product was purified with 10% ethyl acetate in hexane buffered with 1% triethylamine. Yield: 53%. Isolated as a 83:17 mixture of E and Z isomers, respectively.

^1H NMR (400 MHz, CDCl_3 , δ): 7.65 (d, $J = 7.8$ Hz, 1H); 7.37 (d, $J = 8.2$ Hz, 1H); 7.27 (t, $J = 7.1$ Hz, 1H); 7.17 (t, $J = 7.1$ Hz, 1H); 6.40 (s, 1H); 5.43–5.59 (m, 2H); 4.96 (dd, $J = 6.5, 7.4$ Hz, 1H); 3.92 (s, 3H); 2.56–2.71 (m, 2H); 2.01–2.07 (m, 2H); 1.29–1.42 (m, 6H); 0.97 (t, $J = 8.0$ Hz, 9H); 0.95 (t, $J = 4.0$ Hz, 3H); 0.63 (dq, $J = 1.1, 8.0$ Hz, 6H).

^{13}C NMR (100 MHz, CDCl_3 , δ): 142.3, 138.4, 133.7, 127.7, 126.2, 121.3, 120.7, 119.4, 109.1, 100.2, 70.6, 42.2, 32.8, 31.6, 31.0, 29.3, 22.8, 14.3, 7.0, 5.0.

IR (NaCl, thin film): 2954, 2927, 2874, 1466, 1339, 1236, 1072, 1010, 731.

HRMS–ESI (m/z): $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{24}\text{H}_{39}\text{ONSiNa}$, 408.2693; found, 408.2695.

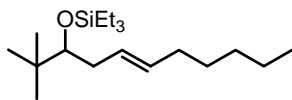


Table 1, entry 8: **2g**:

The standard procedure was followed, except that the reaction was heated at 35 °C. Yield: 61%. Isolated as a 78:22 mixture of E and Z isomers, respectively.

^1H NMR (400 MHz, CDCl_3 , δ): 5.37–5.53 (m, 2H); 3.37 (dd, $J = 3.8, 7.4$ Hz, 1H); 2.30–2.36 (m, 1H); 1.99–2.12 (m, 3H); 1.27–1.42 (m, 6H); 0.99 (t, $J = 8.0$ Hz, 9H); 0.92 (t, $J = 6.8$ Hz, 3H); 0.90 (s, 9H); 0.63 (q, $J = 8.0$ Hz, 6H).

^{13}C NMR (100 MHz, CDCl_3 , δ): 130.6, 128.5, 81.2, 36.2, 31.8, 31.4, 29.5, 27.6, 26.5, 22.8, 14.2, 7.3, 5.7.

IR (NaCl, thin film): 2956, 2876, 1466, 1238, 1096, 1009, 737.

HRMS–ESI (m/z): $[M+Na]^+$ calcd for $C_{19}H_{40}OSiNa$, 335.2746; found, 335.2741.

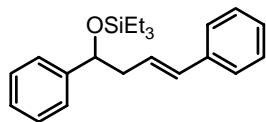


Table 1, entry 9: **2h**:

The standard procedure was followed. Yield: 79%. Isolated as a 95:5 mixture of E and Z isomers, respectively.

1H NMR (400 MHz, $CDCl_3$, δ): 7.30–7.50 (m, 10H); 6.51 (d, $J = 15.9$ Hz, 1H); 6.34 (dt, $J = 7.2, 15.9$ Hz 1H); 4.89 (dd, $J = 5.3, 7.2$ Hz, 1H); 2.64–2.81 (m, 2H); 1.03 (t, $J = 7.9$ Hz, 9H); 0.68 (dq, $J = 2.0, 7.9$ Hz, 6H).

^{13}C NMR (100 MHz, $CDCl_3$, δ): 145.5, 138.0, 132.4, 128.8, 128.4, 127.4, 127.4, 127.2, 126.3, 126.2, 75.5, 45.0, 7.1, 5.2.

IR (NaCl, thin film): 3062, 3028, 2955, 2911, 2876, 1600, 1494, 1453, 1414, 1239, 1088, 1070, 1006, 965, 830, 742, 700.

HRMS–ESI (m/z): $[M+Na]^+$ calcd for $C_{22}H_{30}OSiNa$, 361.1964; found, 361.1974.

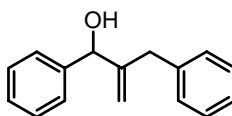


Table 1, entry 9: **3h** (TES group deprotected):

The standard procedure was followed, except that the coupling product was first purified with 1% ethyl acetate in hexane, followed by TES group deprotection of the coupling product **3h** with TBAF and finally purified with 10% ethyl acetate in hexane in order to separate it from the starting materials. Yield 7%.

1H NMR (400 MHz, $CDCl_3$, δ): 7.39 (m, 4H), 7.29–7.35 (m, 3H), 7.22–7.24 (m, 1H),

7.13–7.15 (m, 2H), 5.37 (s, 1H); 5.15 (s, 1H); 4.93 (s, 1H); 3.38 (d, $J = 15.5$ Hz, 1H); 3.13 (d, $J = 15.5$ Hz, 1H); 1.24 (brs, 1H).

^{13}C NMR (100 MHz, CDCl_3 , δ): 150.6, 142.0, 139.3, 129.4, 128.7, 128.5, 128.1, 127.0, 126.4, 112.4, 76.7, 39.2.

IR (NaCl, thin film): 3377, 3061, 3028, 2919, 1494, 1453, 1025, 909, 750, 699.

HRMS–ESI (m/z): $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{16}\text{H}_{16}\text{ONa}$, 247.1099; found, 247.1101.

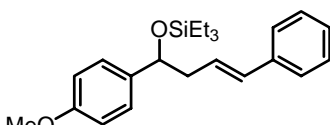


Table 1, entry 10 and 11: **2i**:

The standard procedure was followed (entry 10). Yield: 91%. Isolated as a 95:5 mixture of E and Z isomers, respectively.

The standard procedure was followed, except that the reaction was carried out in five fold larger scale at reaction was heated at 35 °C and using 9 mL toluene (entry 11). Yield: 90%. Isolated as a 95:5 mixture of E and Z isomers, respectively.

^1H NMR (400 MHz, CDCl_3 , δ): 7.29–7.49 (m, 7H); 7.00 (d, $J = 8.6$ Hz, 2H); 6.52 (d, $J = 15.9$ Hz, 1H); 6.35 (dt, $J = 7.2, 15.9$ Hz, 1H); 4.85 (dd, $J = 6.4, 6.4$ Hz, 1H); 3.89 (s, 3H); 2.63–2.81 (m, 2H); 1.04 (t, $J = 7.8$ Hz, 9H); 0.70 (q, $J = 7.8$ Hz, 6H).

^{13}C NMR (100 MHz, CDCl_3 , δ): 159.0, 138.1, 137.7, 132.3, 128.7, 127.5, 127.3, 127.2, 126.3, 113.7, 75.0, 55.4, 45.1, 7.1, 5.2.

IR (NaCl, thin film): 3027, 2954, 2910, 2875, 1612, 1511, 1414, 1302, 1248, 1171, 1081, 1005, 966, 836, 743, 693.

HRMS–ESI (m/z): $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{23}\text{H}_{32}\text{O}_2\text{SiNa}$, 391.2069; found, 391.2057.

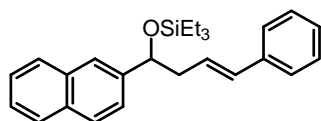


Table 1, entry 12: **2j**:

The standard procedure was followed, except that the reaction was heated at 35 °C. Yield: 84%. Isolated as a 95:5 mixture of E and Z isomers, respectively.

¹H NMR (400 MHz, CDCl₃, δ): 7.90–7.96 (m, 4H); 7.67 (d, *J* = 1.6 Hz, 1H); 7.60–7.65 (m, 2H); 7.30–7.59 (m, 5H); 6.54 (d, *J* = 15.9 Hz, 1H); 6.36 (dt, *J* = 7.2, 15.9 Hz, 1H); 5.05 (dd, *J* = 5.4, 7.2 Hz, 1H); 2.74–2.89 (m, 2H); 1.03 (t, *J* = 8.0 Hz, 9H); 0.70 (dq, *J* = 2.9, 8.0 Hz, 6H).

¹³C NMR (100 MHz, CDCl₃, δ): 142.9, 137.9, 133.4, 133.1, 132.4, 128.7, 128.1, 128.1, 127.9, 127.1, 126.2, 126.1, 125.7, 124.6, 75.5, 44.9, 7.0, 5.1.

IR (NaCl, thin film): 3026, 2954, 2910, 2875, 1507, 1496, 1457, 1239, 1123, 1083, 1005, 965, 819, 744.

HRMS–ESI (*m/z*): [M+Na]⁺ calcd for C₂₆H₃₂OSiNa, 411.2120; found, 411.2167.

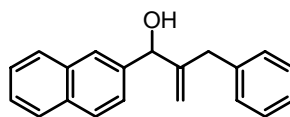


Table 1, entry 12: **3j** (TES group deprotected):

The standard procedure was followed, except that the coupling product was first purified with 1% ethyl acetate in hexane, followed by TES group deprotection of the coupling product **3j** with TBAF and finally purified with 10% ethyl acetate in hexane in order to separate it from the starting materials. Yield 4%.

¹H NMR (400 MHz, CDCl₃, δ): 7.86–7.88 (m, 4H); 7.48–7.55 (m, 3H); 7.20–7.36 (m, 3H); 7.13–7.16 (m, 2H); 5.43 (s, 1H); 5.32 (s, 1H); 4.97 (s, 1H); 3.41 (d, *J* = 15.6 Hz, 1H), 3.16 (d, *J* = 15.6 Hz, 1H), 2.02 (brs, 1H).

^{13}C NMR (100 MHz, CDCl_3 , δ): 150.5, 149.2, 139.4, 139.3, 133.4, 133.3, 129.4, 128.6, 128.2, 127.9, 126.4, 126.4, 126.2, 126.0, 124.9, 112.8, 77.4, 39.2.

IR (NaCl, thin film): 3365, 3058, 2923, 1495, 1453, 1031, 908, 819, 745, 700.

HRMS–ESI (m/z): $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{20}\text{H}_{18}\text{ONa}$, 297.1255; found, 297.1260.

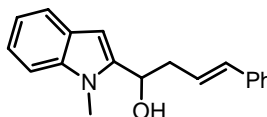


Table 1, entry 13: **2k** (TES group deprotected):

The standard procedure was followed, except that the coupling product was first purified with 10% ethyl acetate in hexane buffered with 1% triethylamine, followed by TES group deprotection of the coupling product **2k** with TBAF and finally purified with 25% ethyl acetate in hexane buffered with 1% triethylamine in order to separate it from the starting materials. Yield: 54%. Isolated as a 95:5 mixture of E and Z isomers, respectively.

^1H NMR (400 MHz, CDCl_3 , δ): 7.63 (d, $J = 7.8$ Hz, 1H); 7.20–7.41 (m, 8H); 7.14 (t, $J = 7.8$ Hz, 1H); 6.62 (d, $J = 15.8$ Hz, 1H); 6.55 (s, 1H); 6.34 (dt, $J = 7.3, 15.8$ Hz, 1H); 5.01 (m, 1H); 3.86 (s, 3H); 2.93–2.99 (m, 2H); 1.93 (brs, 1H).

^{13}C NMR (100 MHz, CDCl_3 , δ): 149.2, 141.3, 138.1, 137.2, 133.8, 128.7, 127.6, 126.4, 125.7, 122.1, 121.0, 119.8, 109.3, 99.4, 66.9, 40.2, 30.4.

IR (NaCl, thin film): 3640, 3026, 2953, 2910, 2875, 1467, 1339, 1237, 1073, 1006, 966, 744.

HRMS–ESI (m/z): $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{19}\text{H}_{19}\text{ONNa}$, 300.1364; found, 300.1365.

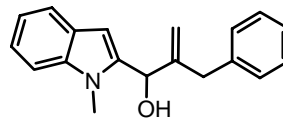


Table 1, entry 13: **3k** (TES group deprotected):

The standard procedure was followed, except that the coupling product was first purified with 10% ethyl acetate in hexane buffered with 1% triethylamine, followed by TES group deprotection of the coupling product **3k** with TBAF and finally purified with 25% ethyl acetate in hexane buffered with 1% triethylamine in order to separate it from the starting materials. Yield 3%.

^1H NMR (400 MHz, CDCl_3 , δ): 7.63 (d, 2H); 7.12–7.38 (m, 10H); 6.49 (s, 1H); 5.38 (s, 1H); 5.31 (s, 1H); 5.14 (s, 1H); 3.70 (s, 3H); 3.54 (d, $J = 15.3$ Hz, 1H); 3.33 (d, $J = 15.3$ Hz, 1H); 1.98 (d, $J = 5.1$ Hz, 1H).

^{13}C NMR (100 MHz, CDCl_3 , δ): 148.7, 139.6, 139.1, 138.4, 129.3, 128.6, 127.3, 126.6, 122.0, 121.0, 119.7, 113.2, 109.3, 101.5, 69.6, 40.2, 30.3.

IR (NaCl, thin film): 3349, 3059, 3027, 2923, 1649, 1601, 1494, 1468, 1453, 1318, 1234, 1030, 968, 907, 751, 737, 700.

HRMS–ESI (m/z): $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{19}\text{H}_{19}\text{NONa}$, 300.1364; found, 300.1369.

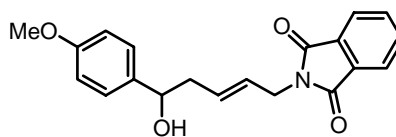


Table 1, entry 14: **2l** (TES group deprotected):

The standard procedure was followed, except that the reaction was carried out at 35 °C and the coupling product was first purified with 40% hexane in CH_2Cl_2 , followed by TES group deprotection of the coupling product **2l** with TBAF and finally purified with 15% hexane in CH_2Cl_2 . Yield: 72%. Isolated as a 83:17 mixture of E and Z isomers, respectively.

^1H NMR (400 MHz, CDCl_3 , δ): 7.86 (dd, $J = 3.1, 5.4$ Hz, 2H); 7.73 (dd, $J = 3.1, 5.4$ Hz, 2H); 7.25 (d, $J = 8.7$ Hz, 2H); 6.84 (d, $J = 8.7$ Hz, 2H); 5.73 (dt, $J = 6.0, 15.4$ Hz, 1H); 5.62 (dt, $J = 6.0, 15.4$ Hz, 1H); 4.68 (dd, $J = 6.4, 6.4$ Hz, 1H); 4.25–4.33 (m, 2H); 3.78 (s, 3H); 2.46 (m, 2H), 2.09 (brs, 1H).

^{13}C NMR (100 MHz, CDCl_3 , δ): 168.2, 159.1, 136.1, 134.1, 132.3, 130.8, 127.2, 127.1, 123.5, 113.9, 73.1, 55.4, 42.3, 39.7.

IR (NaCl, thin film): 3466, 2929, 1770, 1711, 1611, 1512, 1395, 1249, 1174, 1034, 833, 720.

HRMS–ESI (m/z): $[\text{M} - \text{OH}]^+$ calcd for $\text{C}_{20}\text{H}_{18}\text{O}_3\text{N}$, 320.1287; found, 320.1297.

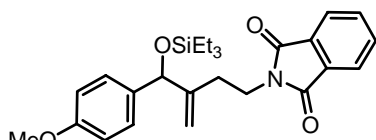


Table 1, entry 14: **3l**:

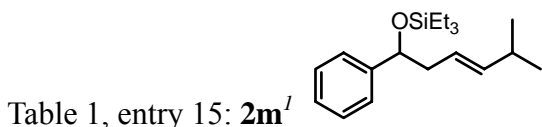
The standard procedure was followed, except that the reaction was carried out at 35 °C and the coupling product was purified with 40% hexane in CH_2Cl_2 . Yield 4%.

^1H NMR (400 MHz, CDCl_3 , δ): 7.81 (dd, 2H); 7.70 (dd, 2H); 7.26 (d, $J = 8.7$ Hz, 2H); 6.79 (d, $J = 8.7$ Hz, 2H); 5.27 (s, 1H); 5.15 (s, 1H); 4.99 (s, 1H); 3.66–3.86 (m, 2H); 3.78 (s, 3H); 2.33–2.40 (m, 1H); 2.16–2.23 (m, 1H); 0.90 (t, $J = 7.9$ Hz, 9H); 0.57 (q, $J = 7.9$ Hz, 6H).

^{13}C NMR (100 MHz, CDCl_3 , δ): 168.4, 158.8, 148.6, 135.2, 134.0, 132.3, 127.7, 123.3, 113.5, 111.8, 77.6, 55.3, 37.2, 29.8, 7.0, 5.0.

IR (NaCl, thin film): 2954, 2876, 1773, 1715, 1511, 1467, 1431, 1395, 1354, 1247, 1078, 952, 719.

HRMS–ESI (m/z): $[\text{M} + \text{Na}]^+$ calcd for $\text{C}_{26}\text{H}_{33}\text{O}_4\text{SiNa}$, 474.2066; found, 474.2071.

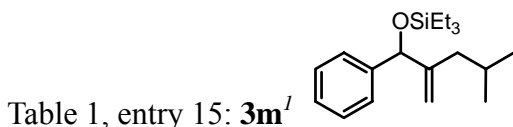


The standard procedure was followed. Yield: 78%. Isolated as a 81:29 mixture of E and Z isomers, respectively.

¹H NMR (500 MHz, CDCl₃, δ): 7.30 (m, 5H); 5.40 (m, 2H); 4.63 (dd, *J* = 5.3, 7.3 Hz, 1H); 2.41 (quintet, *J* = 5.3 Hz, 1H); 2.30 (quintet, *J* = 5.5 Hz, 1H); 2.24 (septet, *J* = 6.7 Hz, 1H); 2.00 (m, 2H); 0.95 (dd, *J* = 6.7, 7.6 Hz, 6H); 0.89 (t, *J* = 7.9 Hz, 9H); 0.62 (q, *J* = 7.9 Hz, 6H).

¹³C NMR (125 MHz, CDCl₃, δ): 145.6, 140.2, 128.1, 127.0, 126.1, 123.7, 75.7, 44.5, 31.3, 22.6, 7.01, 5.0.

HRMS–ESI (*m/z*): [M+Na]⁺ calcd for C₁₉H₃₂OSi, 327.212; found, 327.212.



The standard procedure was followed. Yield: 5%.

¹H NMR (400 MHz, CDCl₃, δ): 7.36 (d, *J* = 7.8 Hz, 2H); 7.32 (t, *J* = 7.1 Hz, 2H); 7.25 (t, *J* = 7.1, 1H); 5.30 (brs, 1H); 5.12 (brs, 1H); 4.87 (brs, 1H); 1.65–1.85 (m, 3H); 0.93 (t, *J* = 8.0 Hz, 9H); 0.84 (d, *J* = 6.4 Hz, 3H); 0.82 (d, *J* = 6.2 Hz, 3H); 0.60 (dq, *J* = 1.3, 8.3 Hz, 6H).

¹³C NMR (100 MHz, CDCl₃, δ): 150.5, 143.7, 128.1, 127.1, 126.7, 110.7, 77.9, 41.1, 26.3, 23.0, 22.6, 7.0, 5.0.

IR (NaCl, thin film): 2955, 2877, 1646, 1454, 1088, 1067, 743.

HRMS–ESI (*m/z*): [M+Na]⁺ calcd for C₁₉H₃₂OSi, 327.211; found, 327.212.

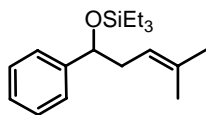


Table 1, entry 16: **2n**:

The standard procedure was followed. Yield: 82%.

^1H NMR (400 MHz, CDCl_3 , δ): 7.27–7.43 (m, 5H); 5.19–5.24 (m, 1H); 4.68 (dd, $J = 5.8$, 7.2 Hz, 1H); 2.36–2.54 (m, 2H); 1.74 (d, $J = 0.8$ Hz, 3H); 1.58 (s, 3H); 0.95 (t, $J = 7.8$ Hz, 9H); 0.60 (dq, $J = 3.4$, 7.8 Hz, 6H).

^{13}C NMR (100 MHz, CDCl_3 , δ): 145.8, 133.6, 128.1, 127.0, 126.1, 121.0, 75.4, 40.0, 26.0, 18.0, 7.0, 5.0.

IR (NaCl, thin film): 3028, 2956, 2877, 2912, 1454, 1414, 1377, 1239, 1089, 1069, 1005, 941, 744, 699.

HRMS–ESI (m/z): $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{18}\text{H}_{30}\text{OSiNa}$, 313.1964; found, 313.1966.

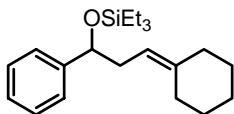


Table 1, entry 17: **2o**:

The standard procedure was followed. Isolated as a mixture of **2o** and **3o**.

2o: Yield 74%. ^1H NMR (400 MHz, CDCl_3 , δ): 7.24–7.42 (m, 5H); 5.14 (t, $J = 7.4$ Hz, 1H); 4.66 (t, $J = 6.4$ Hz, 1H); 2.37–2.52 (m, 2H); 2.00–2.11 (m, 3H); 1.50–1.78 (m, 3H); 1.03–1.48 (m, 4H); 0.94 (t, $J = 7.9$ Hz, 9H); 0.59 (dq, $J = 2.8$, 7.9 Hz, 6H).

^{13}C NMR (100 MHz, CDCl_3 , δ): 145.7, 141.6, 128.0, 127.0, 126.2, 117.5, 75.6, 39.0, 37.5, 29.0, 28.7, 27.8, 27.1, 7.0, 5.0.

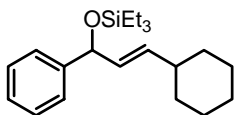


Table 1, entry 17: **3o**:

The standard procedure was followed. Isolated as a mixture of **2o** and **3o**.

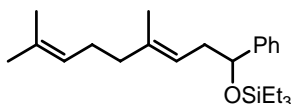
3o: Yield 25%.

^1H NMR (400 MHz, CDCl_3 , δ): 7.24–7.42 (m, 5H); 5.69 (dd, $J = 6.5, 15.4$ Hz, 1H); 5.56 (dd, $J = 7.0, 15.4$ Hz, 1H); 5.18 (d, $J = 7.0$ Hz, 1H); 2.00–2.11 (m, 3H); 1.63–1.78 (m, 1H); 1.50–1.78 (m, 3H); 1.03–1.48 (m, 4H); 1.00 (t, $J = 8.0$ Hz, 9H); 0.67 (dq, $J = 2.3, 8.0$ Hz, 6H).

^{13}C NMR (100 MHz, CDCl_3 , δ): 144.8, 136.9, 131.2, 128.2, 126.9, 126.1, 75.9, 40.4, 33.0, 32.9, 26.4, 26.2, 7.1, 5.2.

IR (NaCl, thin film): 2954, 2928, 2876, 2853, 1449, 1414, 1238, 1086, 1067, 1007, 969, 829, 744, 699.

HRMS–ESI (m/z): $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{21}\text{H}_{34}\text{OSiNa}$, 353.2277; found, 353.2267.



Scheme 1: **2p**:

The standard procedure was followed. Yield: 75%. Isolated as a 71:29 mixture of E and Z isomers, respectively.

^1H NMR (400 MHz, CDCl_3 , δ): 7.25–7.36 (m, 5H); 5.18–5.20 (m, 1H); 5.12–5.12 (m, 1H); 4.64–4.68 (m, 1H); 2.37–2.48 (m, 2H); 2.01–2.10 (m, 4H); 1.73 (s, 4H); 1.64 (s, 3H); 1.55 (s, 2H); 0.92 (t, $J = 6.9$ Hz, 9H); 0.57 (dq, $J = 3.0, 6.9$ Hz, 6H).

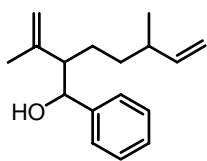
^{13}C NMR (100 MHz, CDCl_3 , δ): 145.8, 145.8, 137.2, 137.1, 131.7, 131.5, 128.1, 128.0, 127.0, 127.0, 126.1, 126.1, 124.6, 124.5, 121.8, 120.7, 75.5, 75.3, 40.0, 39.8, 39.7, 32.3, 26.8, 26.7, 25.9, 25.9, 23.6, 17.8, 17.8, 16.3, 7.0, 7.0, 5.0, 5.0.

IR (NaCl, thin film): 2955, 2876, 1454, 1376, 1239, 1088, 1068, 1006, 829, 743, 700.

HRMS–ESI (m/z): [M+Na]⁺ calcd for C₂₃H₂₈OSiNa, 381.2590; found, 381.2583.

Procedure for the preparation of **4**³

β–Citronellene (0.5 mmol) in anhydrous CH₂Cl₂ (5 mL) was added Me₂AlCl (1.0 M in hexane, 1.1 mL) at 0 °C and stirred at room temperature for 24 hrs. The reaction was quenched by dilution with ether followed by slow addition of water until gas evolution ceased. The organic layer was separated, and the aqueous layer was extracted with ether twice. The combined organic layers were washed with brine, dried and evaporated in vacuo. Purification via flash chromatography on silica gel afforded the coupling product.



Scheme 1: **4**:

Yield: 40%. Isolated as a mixture of diastereomers.

¹H NMR (400 MHz, CDCl₃, δ): 7.28–7.39 (m, 5H); 5.45–5.61 (m, 1H); 5.09 (s, 1H); 5.00 (s, 1H); 4.79–4.94 (m, 2H); 4.38 (dd, *J* = 0.7, 8.5 Hz, 1H); 2.19–2.35 (m, 1H); 1.89–2.05 (m, 1H); 1.73, 1.75 (two s, 3H); 1.67 (brs, 1H); 0.91–1.27 (m, 4H); 0.87, 0.84 (two d, *J* = 6.8 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃, δ): 145.1, 144.3, 142.9, 128.5, 127.9, 127.4, 127.3, 126.0, 116.5, 116.4, 113.1, 112.4, 75.5, 75.4, 56.5, 56.4, 37.8, 37.5, 34.2, 34.2, 31.2, 26.3, 21.0, 19.5, 18.3, 18.1.

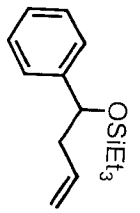
References:

1. Ng, S. S.; Jamison, T. F. *J. Am. Chem. Soc.* **2005**, *127*, 14194–14195.
2. E:Z selectivity for the compound **2** was determined by comparing ¹H NMR of the TES group deprotected alcohol product with the literature, (a) Padwa, A.; Rodriguez, A.; Tohidi, M.; Fukunaga, T. *J. Am. Chem. Soc.* **1983**, *105*, 933–943. (b) Craig, D.; Smith, A. M. *Tetrahedron Lett.* **1990**, *31*, 2631–2632. (c) Yanagisawa, A.; Habaue, S.; Yasue, K.; Yamamoto, H. *J. Am. Chem. Soc.* **1994**, *116*, 6130–6141. (d) Okuma, K.; Tanaka, Y.-i.; Hirabayashi, S. i.; Shioji, K.; Matsuyama, H. *Heterocycles* **1997**, *45*, 1385–1390. (e) Sumida, S. i.; Ohga, M.; Mitani, J.; Nokami, J. *J. Am. Chem. Soc.* **2000**, *122*, 1310–1313. (f) Loh, T. P.; Hu, Q. Y.; Chok, Y. K.; Tan, K. T. *Tetrahedron Lett.* **2001**, *42*, 9277–9280.
3. (a) Snider, B. B.; Rodini, D. J.; Kirk, T. C.; Cordova, R. *J. Am. Chem. Soc.* **1982**, *104*, 555–563. (b) Snider, B. B.; Phillips, G. B. *J. Org. Chem.* **1983**, *48*, 464–469.

NY-8-89 H pdt

propene, H pdt

2a



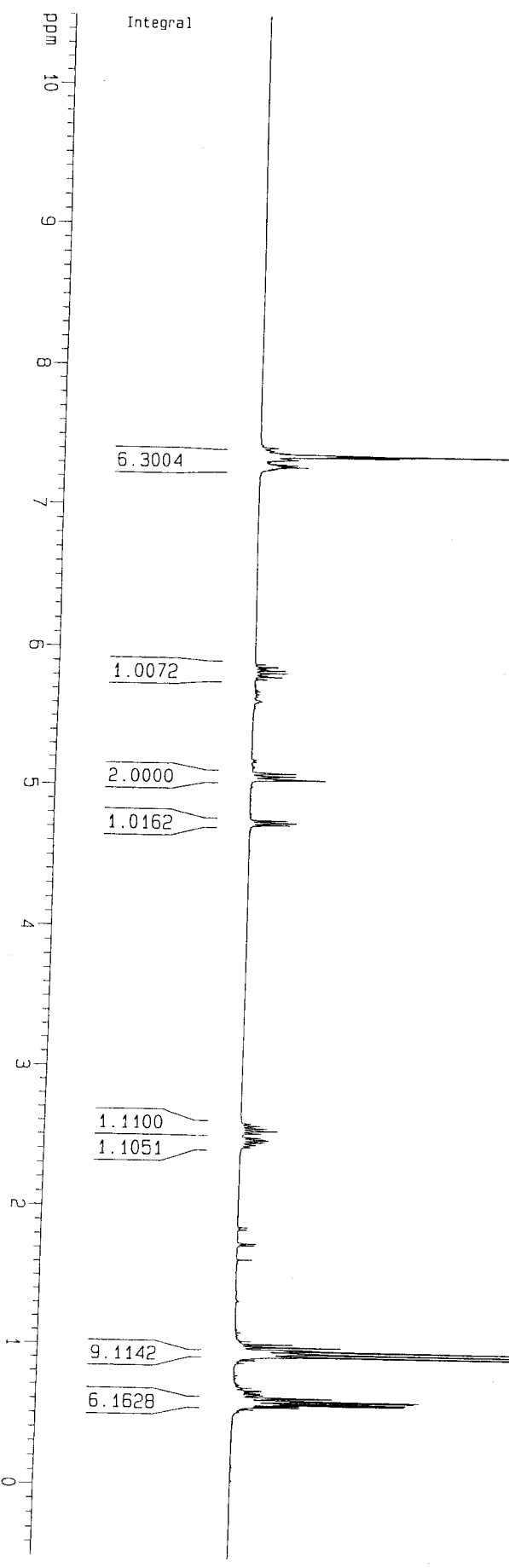
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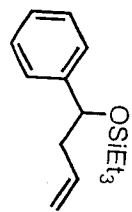
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propene, H-pdt

2a



ppm

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- 128.182
- 127.199
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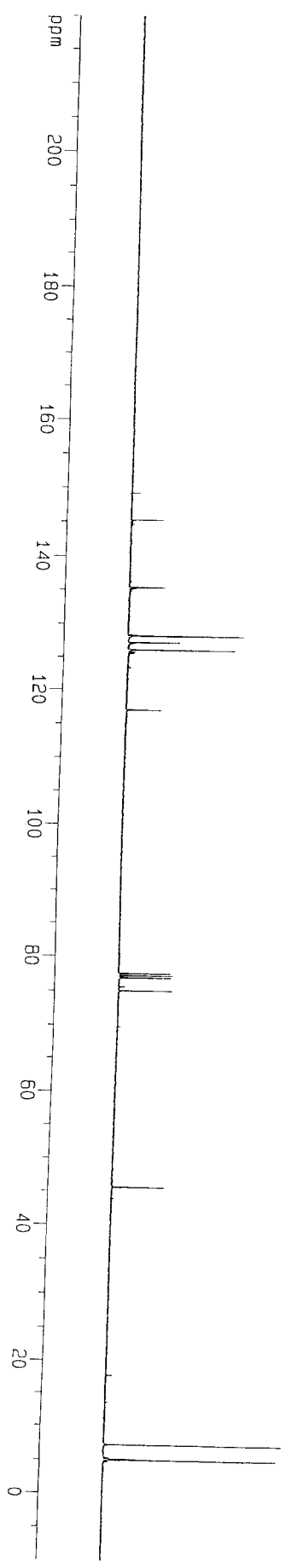
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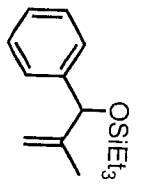
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HCY - 8-89 Addt.

propene, A-pdt



3a

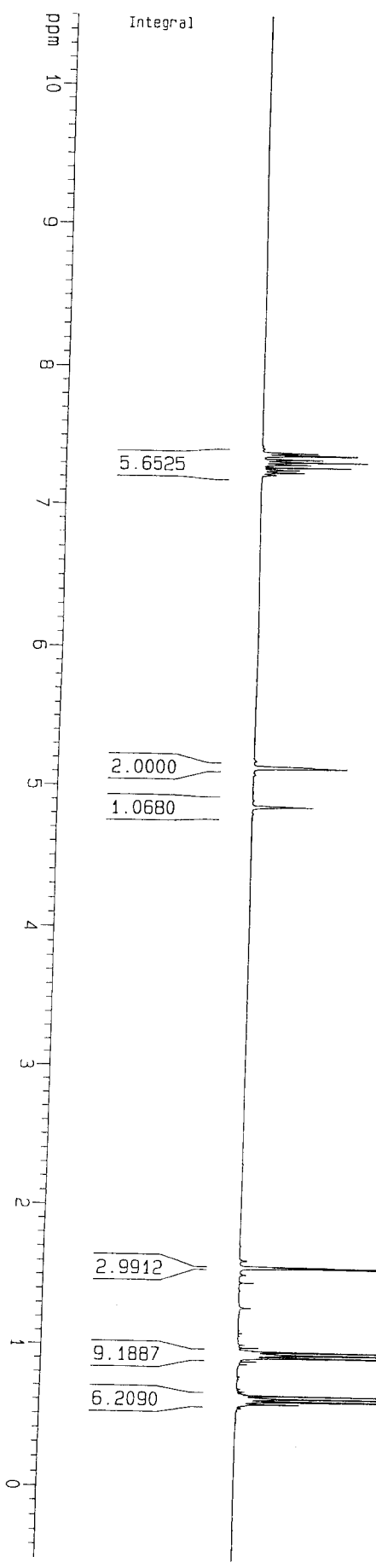
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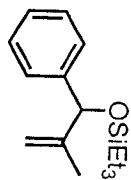
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Propene, Apdt

3a



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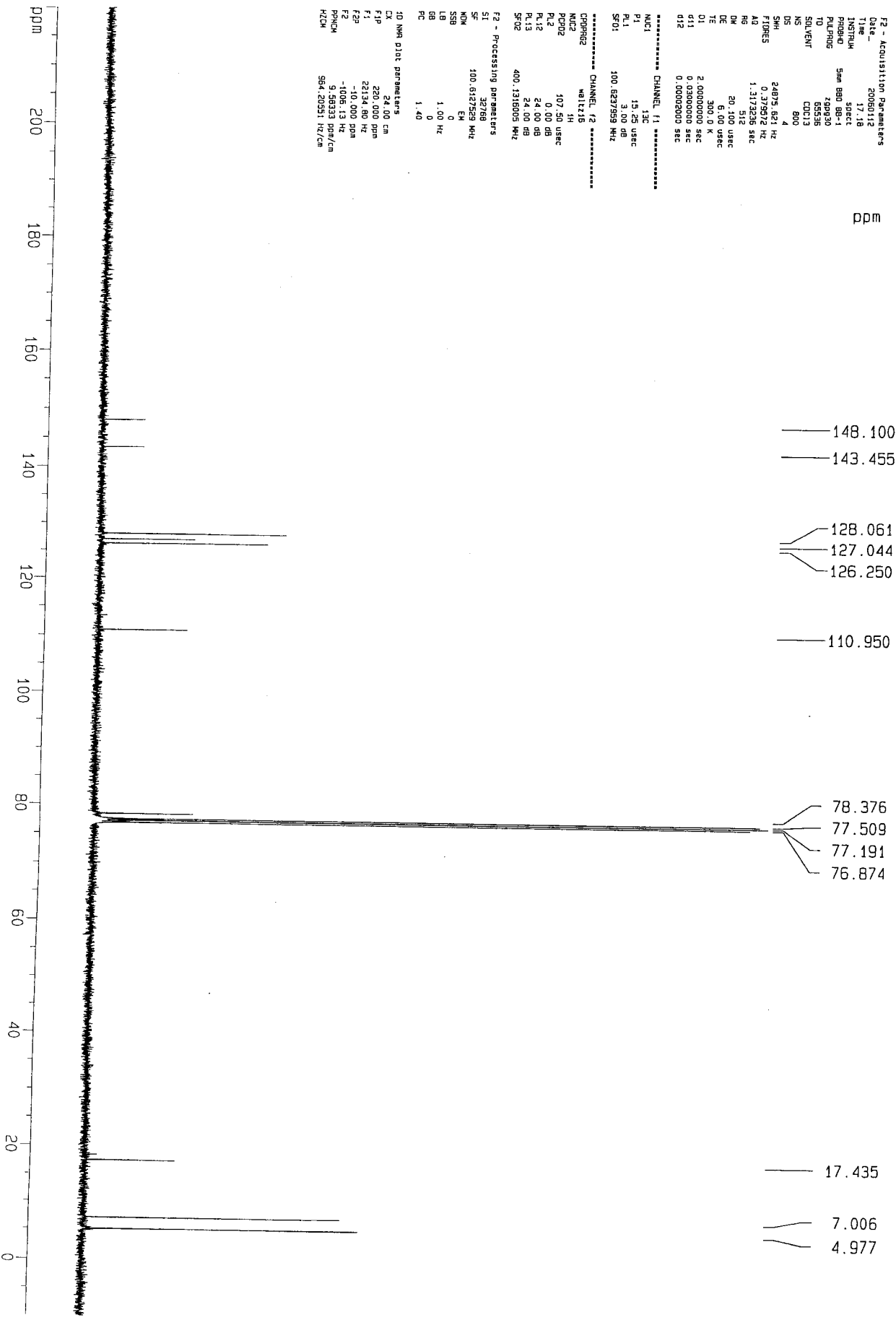
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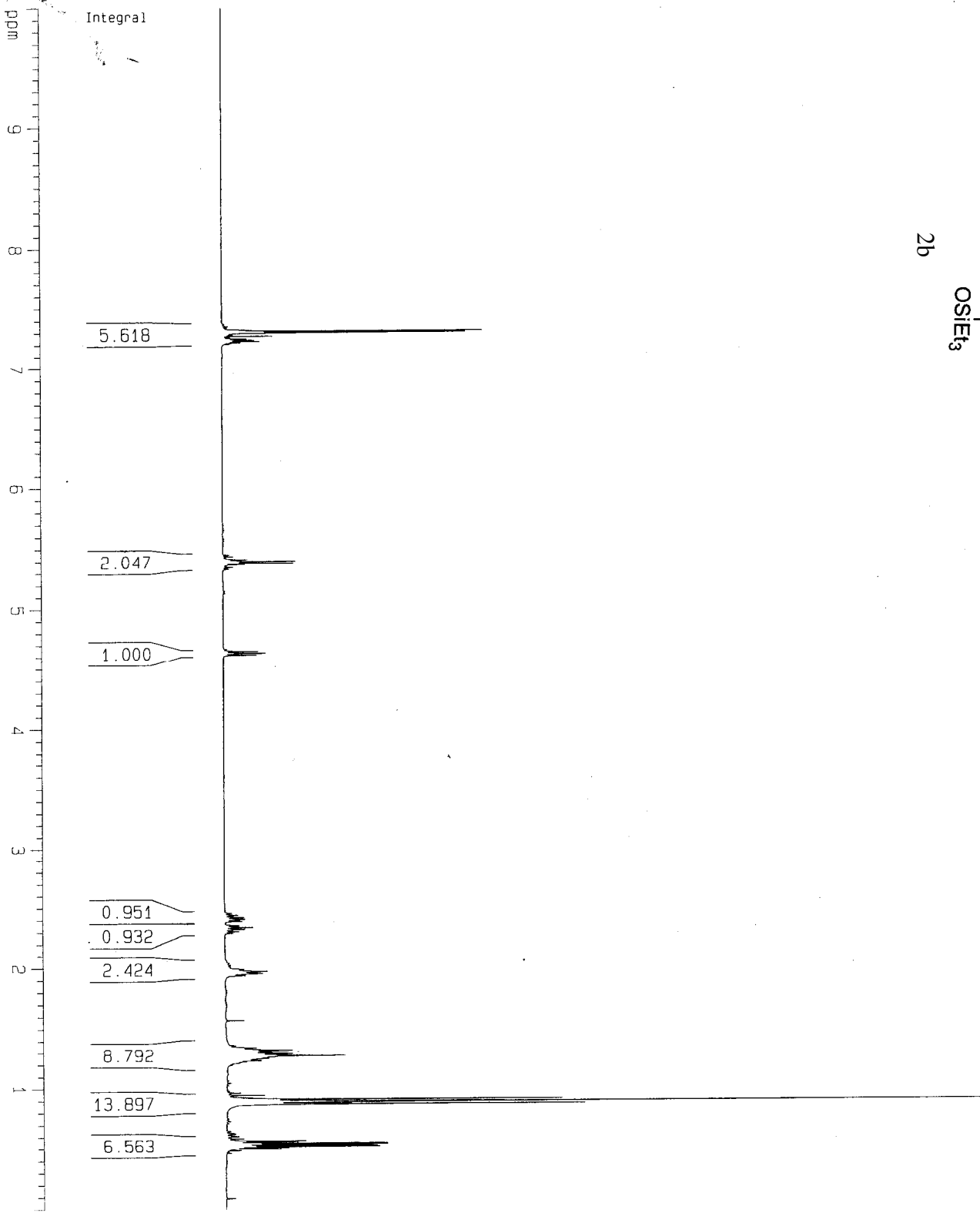
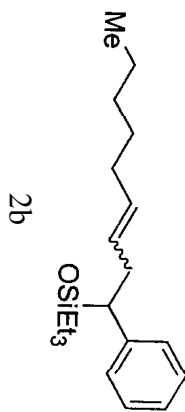
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 LB: 1.00 Hz
 GB: 0
 PC: 1.40

1D NMR dipole parameters
 CX: 2.000 cm
 F1P: 230400 ppm
 F1: 22134.60 Hz
 F2P: -10.000 ppm
 F2: -1006.13 Hz
 PPMICM: 9.56333 ppm/cm
 HZCM: 564.20351 Hz/cm



SN050658 ene product



Current Data Parameters
NAME SNS68ene-H
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20050614
Time 19.59

INSTRUM spect
PROBHD 5 mm QNP 1H
PULPROG zg30

TD 65536
SOLVENT CDCl3
NS 8
DS 4

SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec

RG 128
DM 60.400 usec
DE 5.00 usec
TE 300.0 K

D1 1.00000000 sec

===== CHANNEL f1 =====

NUC1 1H
P1 9.50 usec
PL1 2.00 dB
SF01 400.1324710 MHz

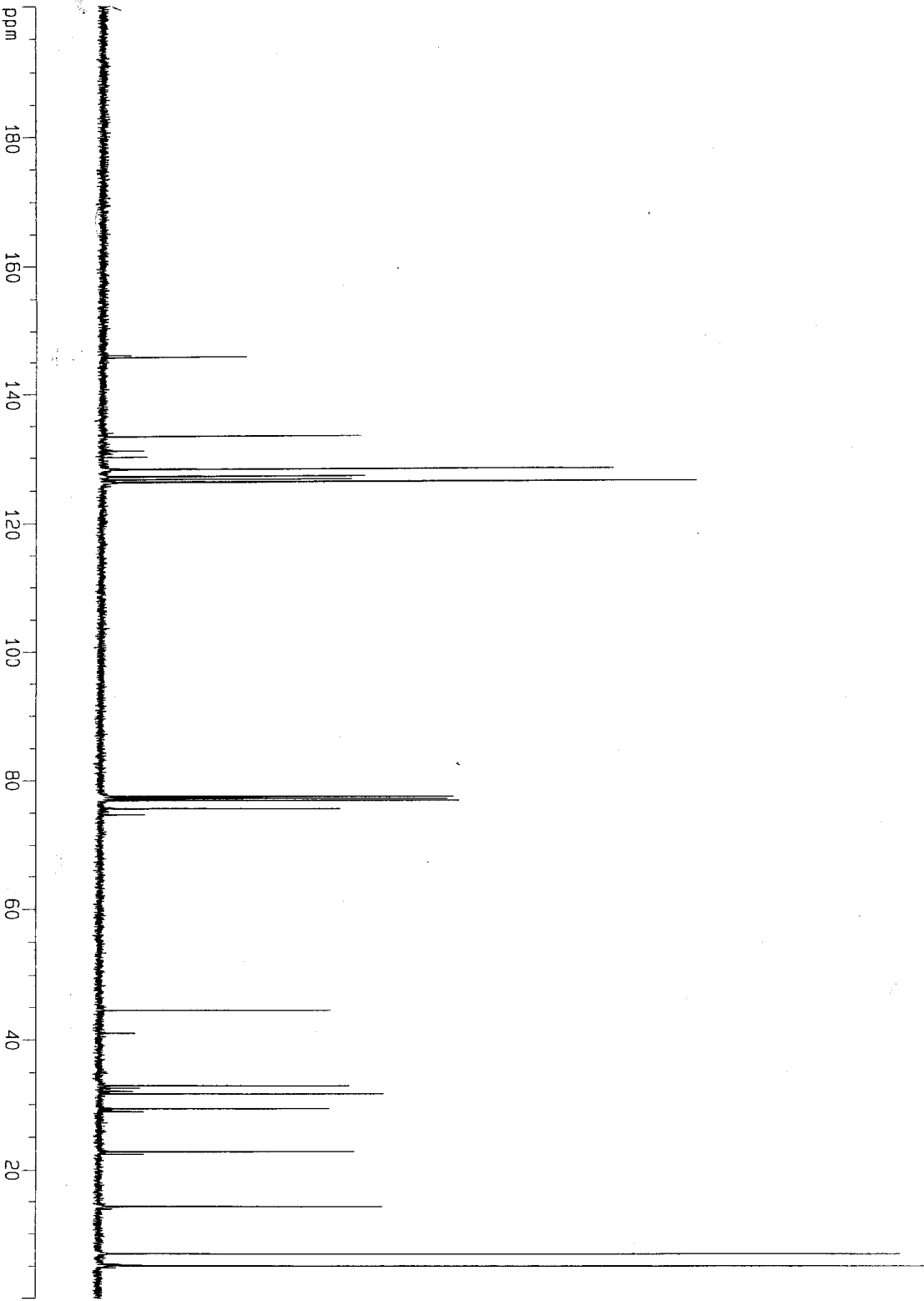
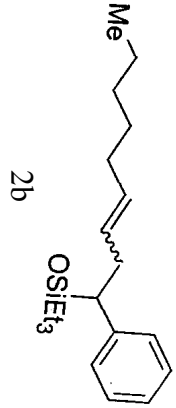
F2 - Processing parameters

SI 32768
SF 400.1300059 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

1D NMR plot parameters

CX 20.00 cm
F1P 10.000 ppm
F1 4001.30 Hz
F2P 0.000 ppm
F2 0.00 Hz
PASCAL 0.50000 ppm/cm
HZCM 200.06500 Hz/cm

octene benzaldehyde TESOTf ene



Current Data Parameters
 NAME octene-ene-C
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20050716
 Time 22.11

INSTRUM spect
 PROCBHD 5mm BBO BB-1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 213
 DS 4

SMH 25125.629 HZ
 FIDRES 0.38387 HZ
 AQ 1.3042164 sec
 RG 1149.4
 DW 19.900 usec
 DE 6.00 usec
 TE 300.0 K

D1 2.00000000 sec
 d11 0.03000000 sec
 d12 0.00002000 sec

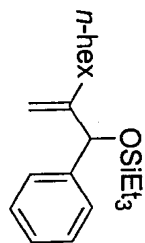
==== CHANNEL f1 =====
 NUC1 13C
 P1 15.25 usec
 PL1 3.00 dB
 SFO1 100.6237959 MHz

==== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 107.50 usec
 PL2 0.00 dB
 PL12 24.00 dB
 PL13 24.00 dB
 SFO2 400.1316005 MHz

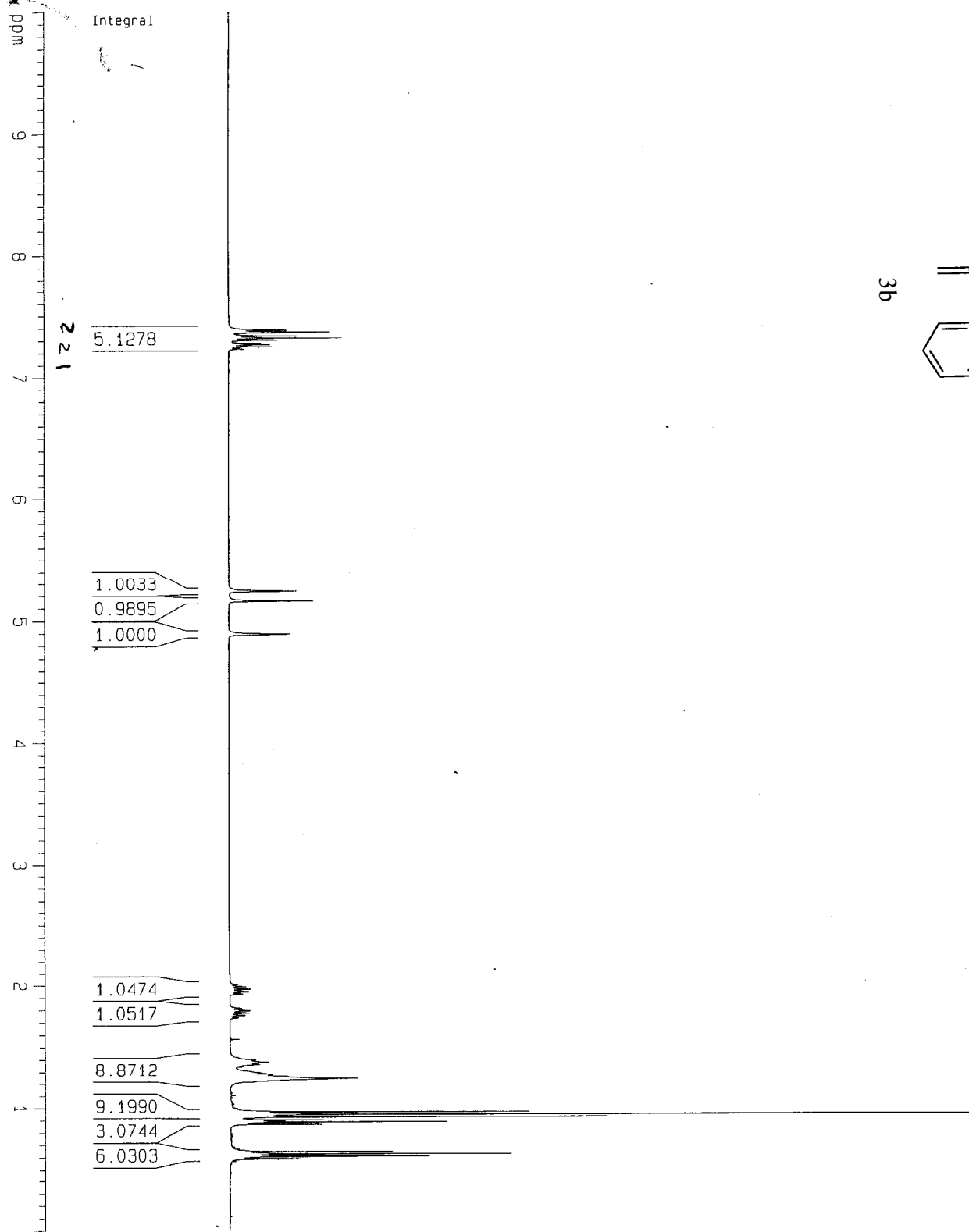
F2 - Processing parameters
 SI 32768
 SF 100.6127476 MHz
 WDM EM
 SSB 0
 LB 1.00 HZ
 GB 0
 PC 1.40

1D NMR plot parameters
 CX 20.00 cm
 F1p 200.000 ppm
 F1 20122.55 HZ
 F2p 0.000 ppm
 F2 0.00 HZ
 PPMCM 10.00000 ppm/cm
 HZCM 1006.12744 HZ/cm

SN050658 allylic alcohol



3b



Current Data Parameters
 NAME SN658neck-c
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20050614
 Time 19.05

INSTRUM spect
 PROBHID 5 mm QNP 1H
 PULPROG zg30

TD 65536
 SOLVENT CDCl3

NS 8
 DS 4

SMH 8278.146 Hz
 FIDRES 0.126314 Hz

AQ 3.9584243 sec
 RG 64

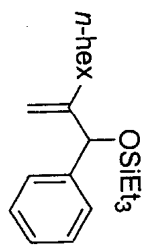
DW 60.400 usec
 DE 6.00 usec

TE 300.0 K
 D1 1.00000000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 9.50 usec
 PL1 2.00 dB
 SF01 400.1324710 MHz

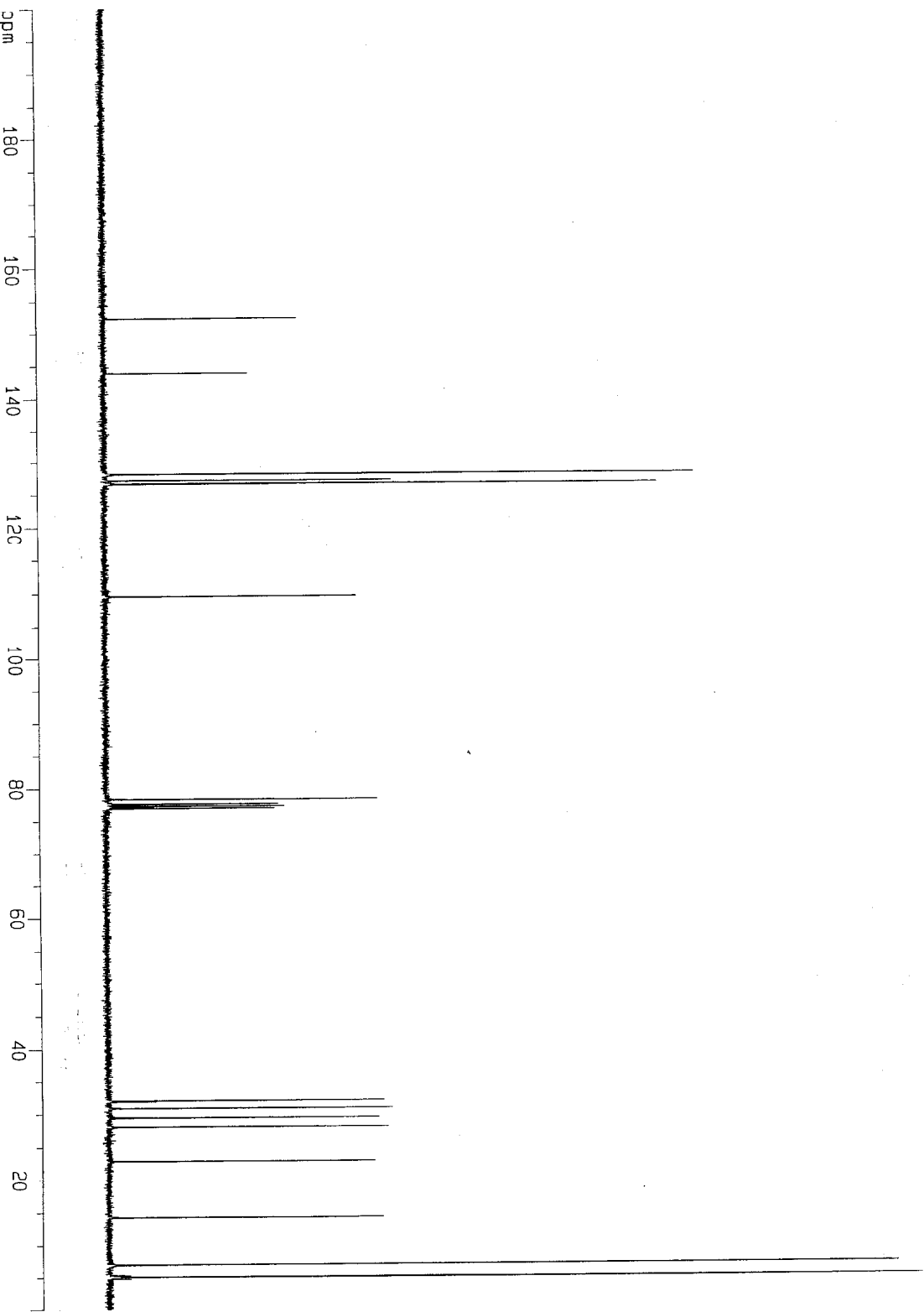
F2 - Processing parameters
 SI 32768
 SF 400.1300000 MHz
 WDM EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 10.000 ppm
 F1 4001.30 Hz
 F2P 0.000 ppm
 F2 0.00 Hz
 PPMCM 0.50000 ppm/cm
 HZCM 200.06500 Hz/cm



3b

octene benzaldehyde TESOTf



Current Data Parameters
 NAME octene-a1-C
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20050716
 Time 21.50

INSTRUM Spect
 PROBHD 5mm BBO B8-1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 95
 DS 4

SMH 25125.629 Hz
 FIDRES 0.38387 Hz
 AQ 1.3042164 sec
 RG 8192

DW 19.900 usec
 DE 6.00 usec
 TE 300.0 K

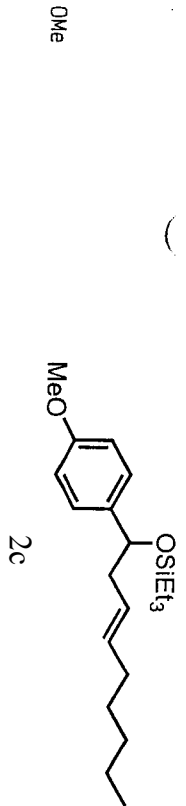
D1 2.00000000 sec
 d11 0.03000000 sec
 d12 0.00020000 sec

===== CHANNEL f1 =====
 NUC1 13C
 P1 15.25 usec
 PL1 3.00 dB
 SFG1 100.6237959 MHz

===== CHANNEL f2 =====
 CPDPRG2 wa1tz16
 NUC2 1H
 PCPD2 107.50 usec
 PL2 0.00 dB
 PL12 24.00 dB
 PL13 24.00 dB
 SF02 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127484 MHz
 WDM EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters
 CX 20.00 cm
 F1p 200.000 ppm
 F1 20122.55 Hz
 F2p 0.000 ppm
 F2 0.00 Hz
 PPMCM :0.00000 ppm/cm
 HZCM 1006.12744 Hz/cm



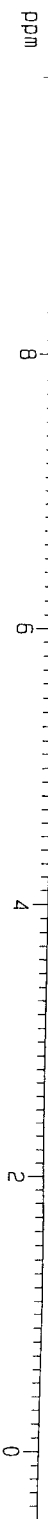
Current Data Parameters
 NAME hcy-8-44p-Ome
 EXPNO 1
 PROCNO 1

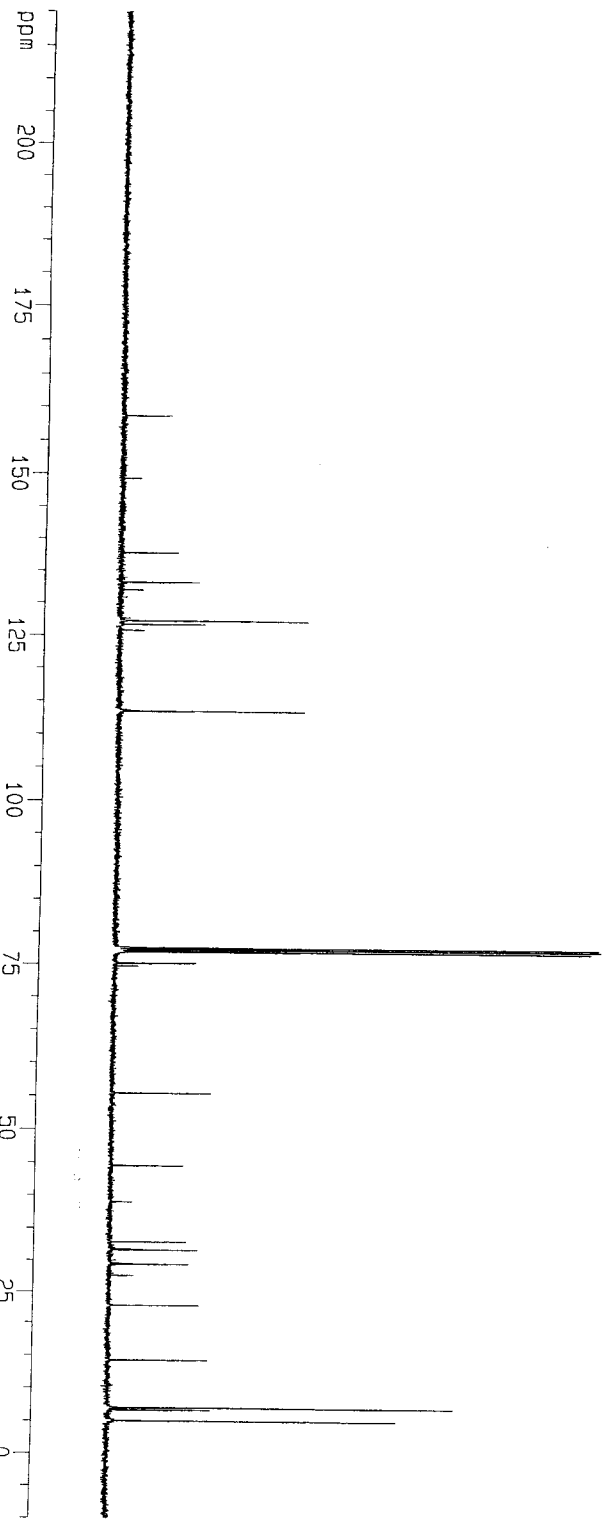
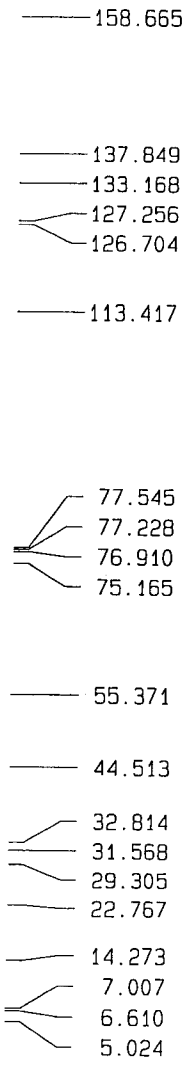
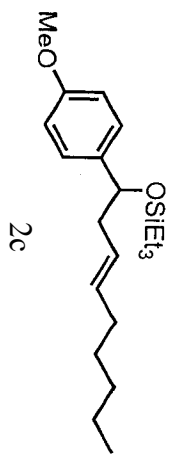
F2 - Acquisition Parameters
 Date_ 20051028
 Time 17.30
 INSTRUM spect
 PROBHD 5 mm QNP 1H/1
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 4
 DS 1
 SMH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 90.5
 DW 60.400 usec
 DE 6.00 usec
 TE 294.8 K
 D1 1.00000000 sec
 MCREST 0.00000000 sec
 MCMRK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 9.88 usec
 PL1 3.00 dB
 SF01 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300054 MHz
 MDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 CY 9.19 cm
 F1P 10.500 ppm
 F1 4201.37 Hz
 F2P -0.500 ppm
 F2 -200.07 Hz
 PPMCM 0.55000 ppm/cm
 HZCM 220.07150 Hz/cm





Current Data Parameters
 NAME hcy-8-44-OMe
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20051028
 Time 16.05
 INSTRUM spect
 PROBHD 5mm BBO BB-1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 1024
 DS 4

SMH 24875.621 HZ
 FIDRES 0.379572 HZ
 AQ 1.3173236 sec
 RG 1024

DE 20.100 usec
 TE 6.00 usec
 D1 300.0 K
 D11 2.00000000 sec
 D12 0.03000000 sec
 D12 0.00002000 sec

===== CHANNEL f1 =====
 NUC1 13C
 P1 15.25 usec
 PL1 3.00 dB
 SF01 100.6237959 MHz

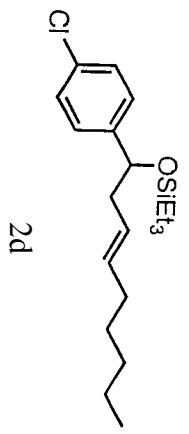
===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 107.50 usec
 PL2 0.00 dB
 PL12 24.00 dB
 PL13 24.00 dB
 SF02 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127499 MHz
 MDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

10 NMR plot parameters
 CX 20.00 cm
 FIP 220.000 ppm
 F1 22134.80 HZ
 F2 -1006.13 HZ
 PPMCM 11.50000 ppm/cm
 HZCM 1157.04563 HZ/cm

15-05-8-2-14

C1



2d

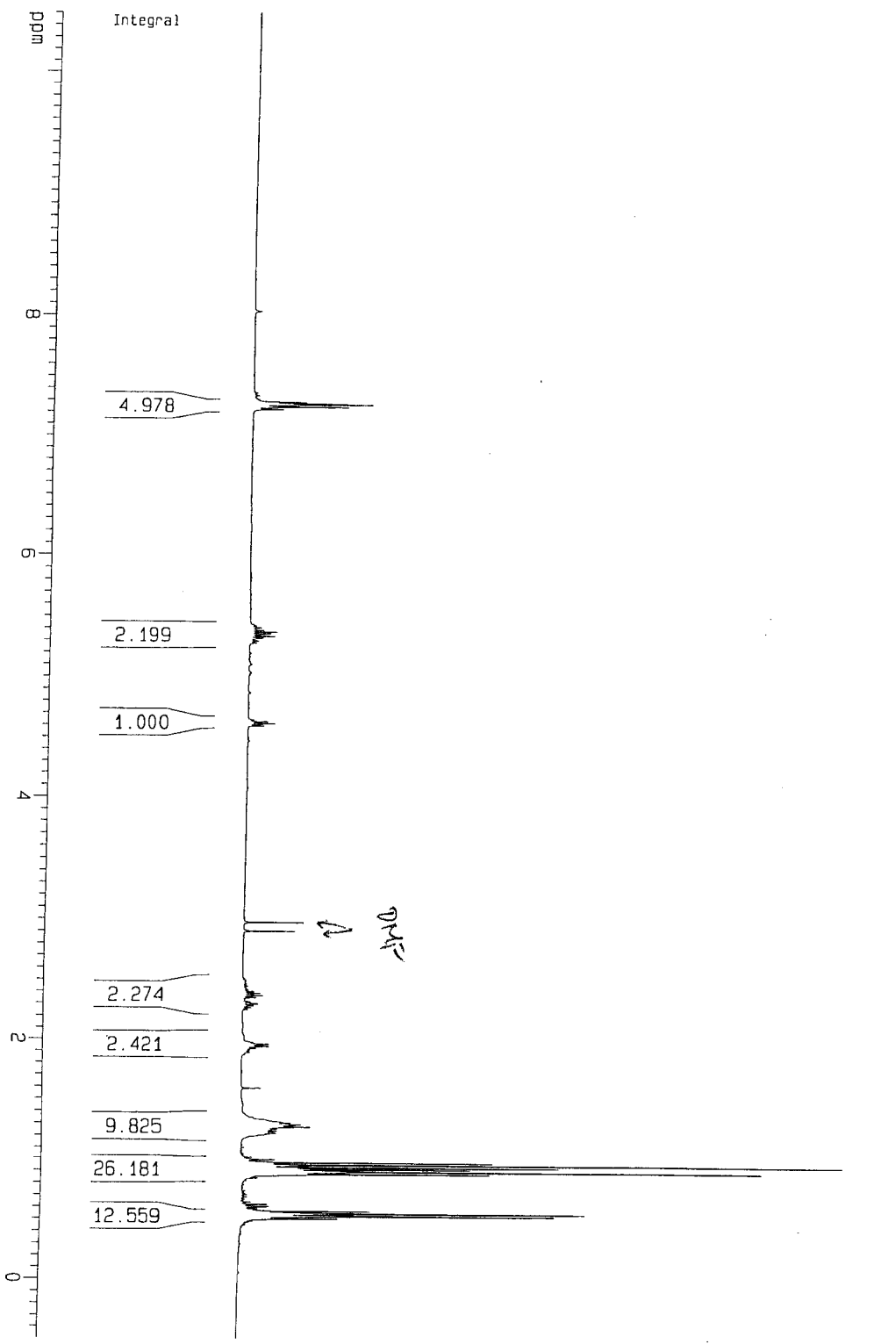
Current Data Parameters
 NAME hcy-8-44p-C1
 EXPNO 1
 PROCNO 1

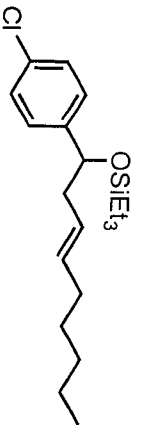
F2 - Acquisition Parameters
 Date_ 20051028
 Time 17.34
 INSTRUM spect
 PROBHD 5 mm QNP 1H/1
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 8
 DS 1
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 90.5
 DW 60.400 usec
 DE 6.00 usec
 TE 294.8 K
 D1 1.00000000 sec
 MCREST 0.00000000 sec
 MCNRK 0.01500000 sec

***** CHANNEL f1 *****
 NUC1 1H
 P1 9.88 usec
 PL1 3.00 dB
 SF01 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300054 MHz
 MDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 CY 9.19 cm
 F1P 10.500 ppm
 F1 4201.37 Hz
 F2P -0.500 ppm
 F2 -200.07 Hz
 PPMCM 0.55000 ppm/cm
 HZCM 220.07150 Hz/cm





C1

2d

144.082
133.726
132.541
128.240
127.507
125.962

77.530
77.214
76.896
74.808

44.370
32.774
31.536
29.248
22.744
14.259
7.010
6.968
6.602
5.104
4.980

Current Data Parameters
NAME hcy-8-44-C1
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters

Date_ 20031028
Time 12.15
INSTRUM spect
PROBHD 5 mm QNP 1H/1
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 800
DS 4
SWH 2390.814 Hz
FIDRES 0.365918 Hz
AQ 1.3664756 sec
RG 1824.6
DM 20.850 usec
DE 6.00 usec
TE 294.8 K
O1 2.00000000 sec
d11 0.03000000 sec
DELTA 1.89999998 sec
MCREST 0.00000000 sec
MCMRK 0.01500000 sec

CHANNEL f1 *****

NUC1 13C
P1 8.50 usec
PL1 3.00 dB
SF01 100.6282898 MHz

CHANNEL f2 *****

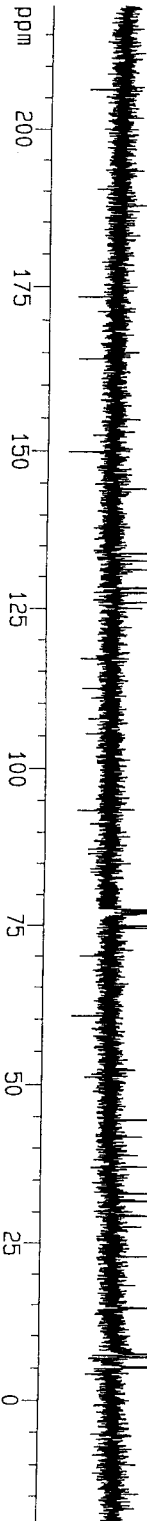
CPDPRG2 waltz16
NUC2 1H
PCPD02 88.01 usec
PL2 3.00 dB
PL12 22.00 dB
PL13 22.00 dB
SF02 400.1316005 MHz

F2 - Processing parameters

SI 32768
SF 100.6127505 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

1D NMR plot parameters

CX 20.00 cm
CY 12.50 cm
F1P 219.353 ppm
F1 22069.67 Hz
F2P -18.995 ppm
F2 -1911.15 Hz
PPMCM 11.91738 ppm/cm
HZCM 1199.04077 Hz/cm



NCH-8-87P. 35c

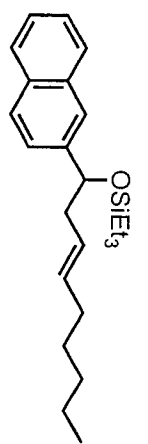
Current Data Parameters
NAME hcy8-87c-p
EXNO 11
PROCNO 1

F2 - Acquisition Parameters
Date 20060109
Time 17:29
INSTRUM spect
PROBHD 5mm BBO BB-1
PULPROG zg30
TD 65536
SOLVENT CDCl3
DS 2
OS 3
SHW 9278.146 Hz
FIDRES 0.126314 Hz
AQ 3.564443 sec
RG 25.4
DM 60.400 usec
DE 6.00 usec
TE 300.0 K
D1 1.00000000 sec

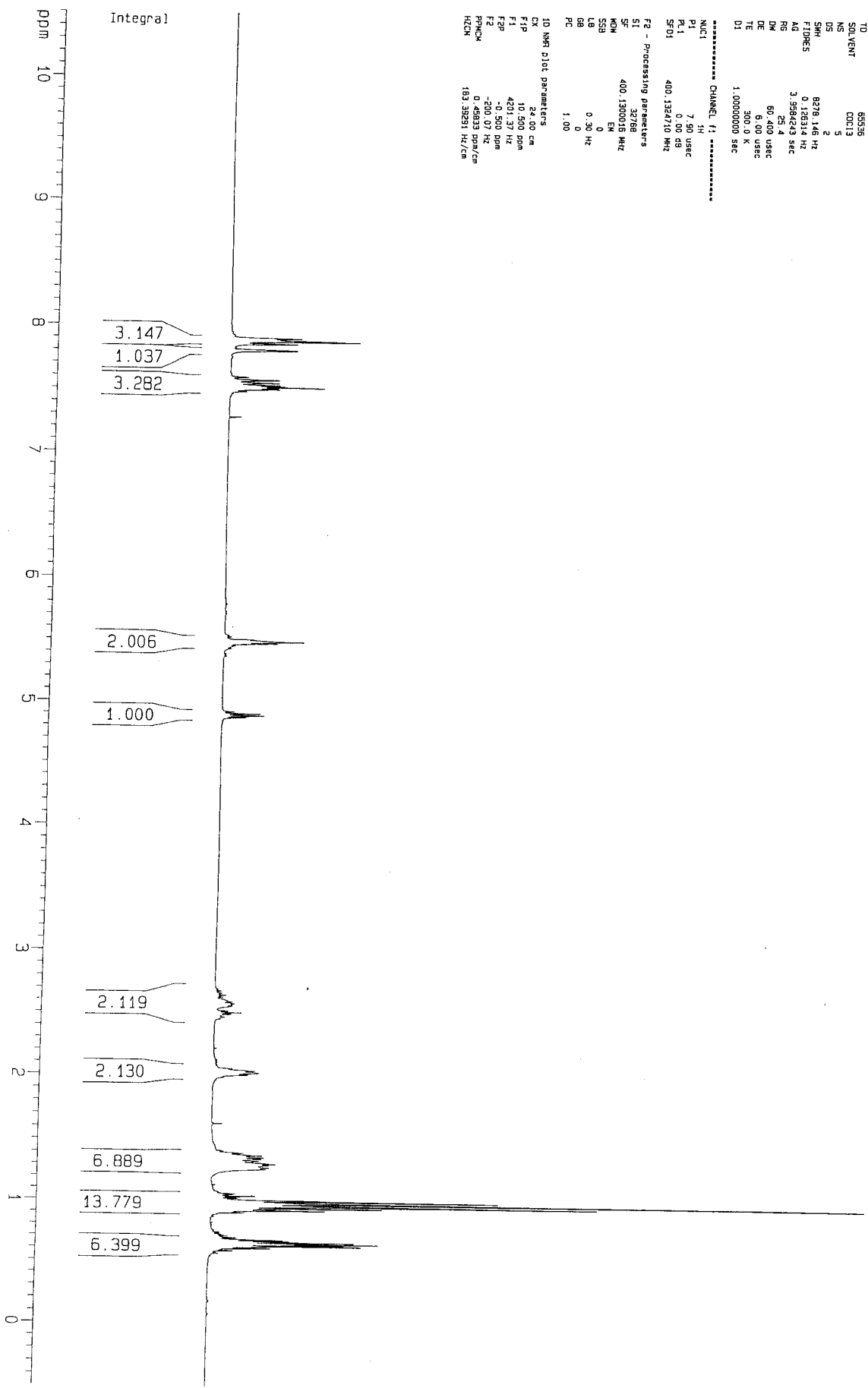
***** CHANNEL f1 *****
NUC1 1H
P1 7.90 usec
R1 0.00 dB
SFO1 400.1362710 MHz

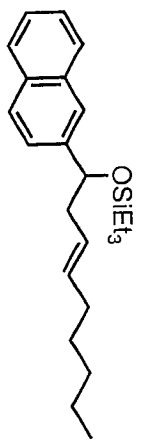
F2 - Processing parameters
SI 32768
SF 400.1300016 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

D0 NMR plot parameters
F1 24.00 cm
F2 10.500 ppm
F3 40.57 Hz
F4 0.50 ppm
F5 -200.07 Hz
PRNCK 0.46933 ppm/cm
HZCK 183.38281 Hz/cm



2e





Current Data Parameters
 NAME ncp4-87n1-p
 EXPNO 22
 PROCNO 1

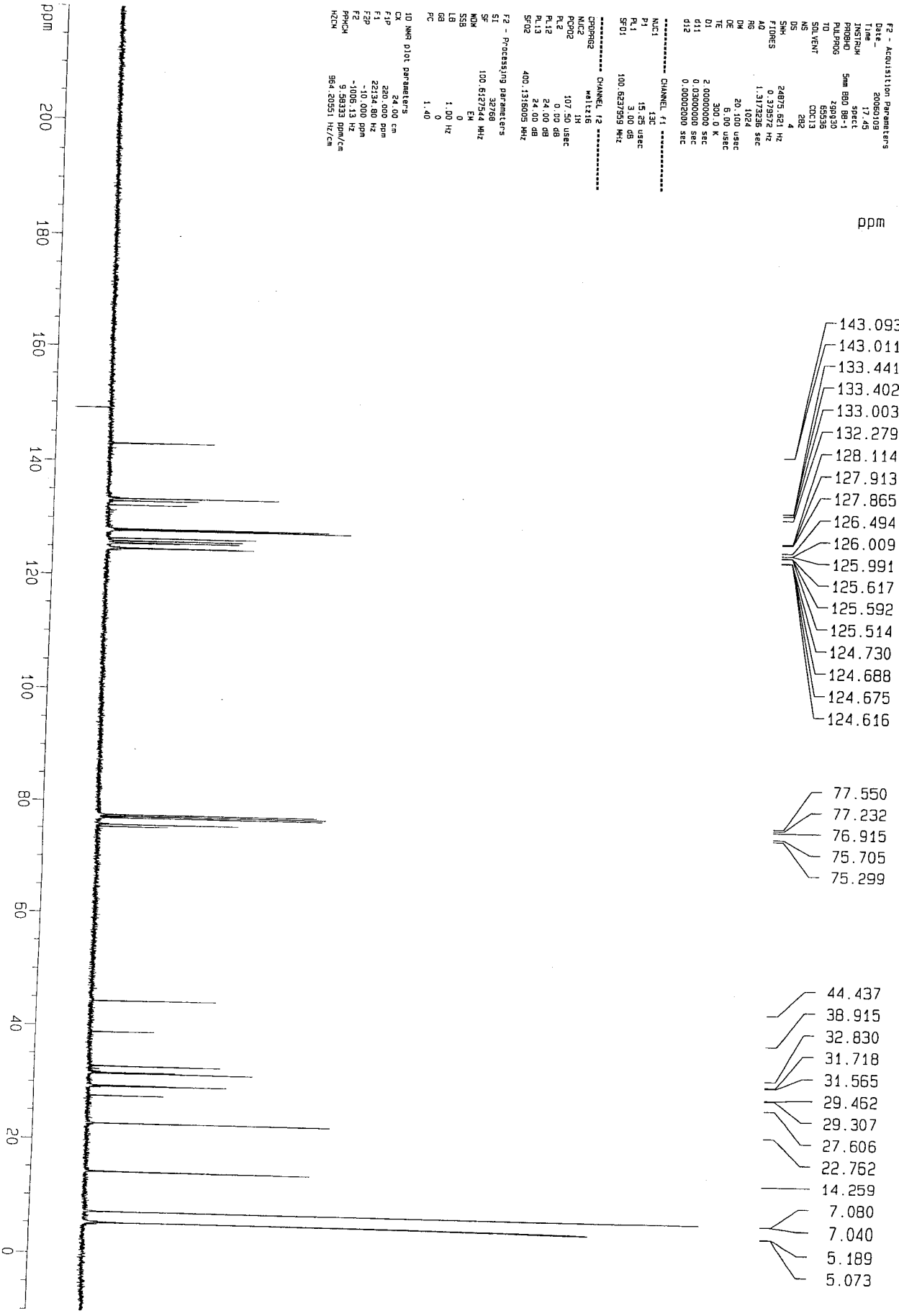
F2 - Acquisition Parameters
 Date_ 20060109
 Time 17:45
 F1 225.130 MHz
 PULPROG sm 890 88-1
 TD 299130
 SFO1 100.627959 MHz
 SOLVENT CDCl3
 NS 2992
 DS 4
 SWH 24875.821 Hz
 FIDRES 0.379572 Hz
 AQ 1.3173286 sec
 RG 1024
 DM 20.100 usec
 DE 6.00 usec
 TE 300.2 K
 D1 2.00000000 sec
 d11 0.03000000 sec
 d12 0.00002000 sec

***** CHANNEL f1 *****
 NUC1 13C
 P1 15.25 usec
 PL1 3.00 dB
 SFO1 100.627959 MHz

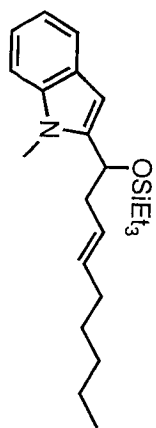
***** CHANNEL f2 *****
 CPDPRG2 waltz16
 NU1 1H
 NU2 13C
 PCPD2 107.50 usec
 PL2 0.00 dB
 PL12 24.00 dB
 PL13 24.00 dB
 SFO2 400.136005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127544 MHz
 MDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR Plot Parameters
 CX 24.00 cm
 F1P 225.130000 MHz
 F1 225.130000 MHz
 F2P -106.13 Hz
 F2 -106.13 Hz
 PPMCK 9.98333 ppm/cm
 HZCM 984.20251 Hz/cm



Me-indole-2-CHO, f1



2f

Current Data Parameters
 NAME hcy-8-52-ind-2
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20051110
 Time 12.10

INSTRUM spect
 PROBHD 5mm BBO BB-1
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3

NS 8
 DS 2
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 18

DW 60.400 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec

===== CHANNEL f1 =====

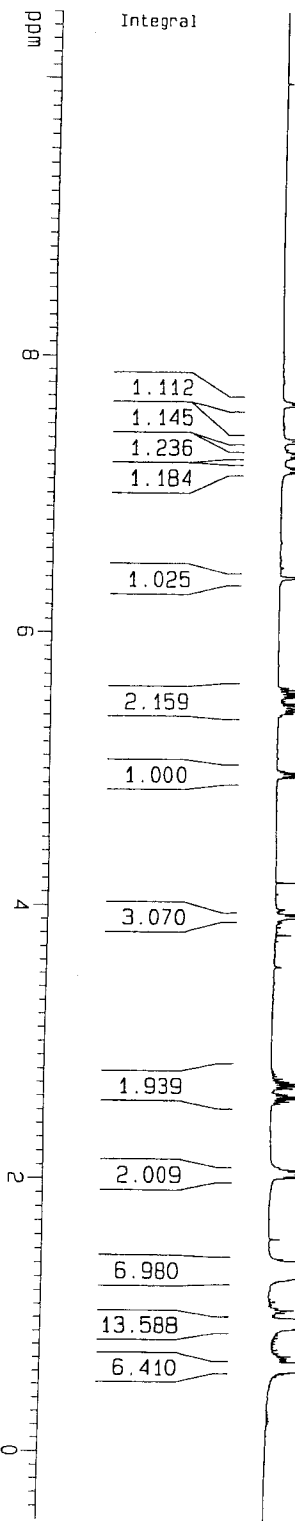
NUC1 1H
 P1 7.90 usec
 PL1 0.00 dB
 SF01 400.1324710 MHz

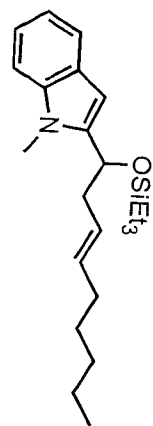
F2 - Processing parameters

SI 32768
 SF 400.1300016 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters

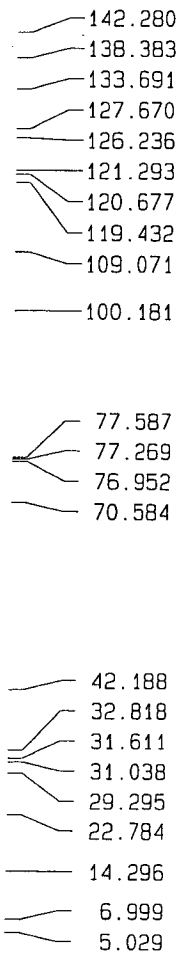
CX 20.00 cm
 F1P 10.500 ppm
 F1 4201.37 Hz
 F2P -0.500 ppm
 F2 -200.07 Hz
 PPMCM 0.55000 ppm/cm
 HZCM 220.07150 Hz/cm





2f

ppm



Current Data Parameters
 NAME hcy-8-52-ind-2
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20051110
 Time 12.17

INSTRUM spect
 PROBRD 5mm BBO BB-1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 434
 DS 4

SMH 24875.621 Hz
 FIDRES 0.379572 Hz
 AQ 1.3173236 sec
 RG 1024

DW 20.100 usec
 DE 6.00 usec
 TE 300.0 K

D1 2.00000000 sec
 d11 0.03000000 sec
 d12 0.00002000 sec

===== CHANNEL f1 =====
 NUC1 13C
 P1 15.25 usec
 PL1 3.00 dB
 SF01 100.6237959 MHz

===== CHANNEL f2 =====
 CPOPRG2 waltz16
 NUC2 1H
 PCPD2 107.50 usec
 PL2 0.00 dB
 PL12 24.00 dB
 PL13 24.00 dB
 SF02 400.1316005 MHz

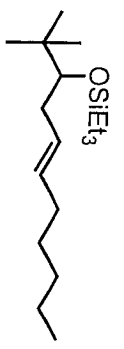
F2 - Processing parameters
 SI 32768
 SF 100.6127529 MHz
 MDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters
 CX 20.00 cm
 F1P 220.000 DPM
 F1 22134.80 Hz
 F2P -10.000 DPM
 F2 -1006.13 Hz
 PPMCH 11.50000 ppm/cm
 HZCM 1157.04663 Hz/cm

124-8-95 - H-pdt.

H-pdt

2g



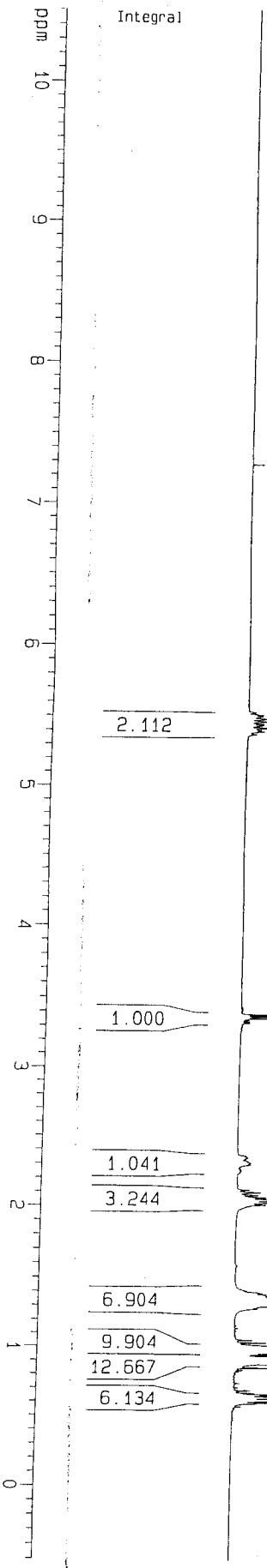
Current Data Parameters
 Name: 124-8-95
 EXPNO: 1
 PROCNO: 1

F2 - Acquisition Parameters
 Date_: 20060120
 Time: 12.41
 INSTRUM: spect
 PROBNM: 5mm BB0-BB-1
 PULPROG: zg30
 TD: 65536
 SOLVENT: CDCl3
 NS: 4
 DS: 2
 SWH: 8278.146 Hz
 FIDRES: 0.28214 Hz
 AQ: 3.58242 sec
 RG: 320
 DM: 60.400 usec
 DE: 6.00 usec
 TE: 300.0 K
 D1: 1.00000000 sec

***** CHANNEL f1 *****
 NUC1: 1H
 P1: 7.50 usec
 PL1: 0.00 dB
 SF01: 400.1324710 MHz

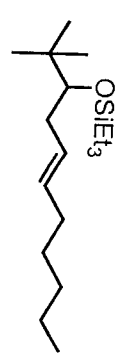
F2 - Processing parameters
 SI: 32768
 SF: 400.130000 MHz
 MDW: 0
 SSB: 0
 LG: 0.30 Hz
 GB: 0
 PC: 1.00

10 NMR plot parameters
 CX: 24.00 cm
 F1P: 10.500 ppm
 F1: 4201.36 Hz
 F2P: -0.500 ppm
 F2: -200.07 Hz
 PRNUC1: 0.45893 ppm/cm
 HZCM: 183.3281 Hz/cm



H-pdt

2g



Current Data Parameters
 NAME: hcv8-shout
 EXPNO: 2
 PROCNO: 1

F2 - Acquisition Parameters
 Date_: 20060120
 Time: 12.55
 INSTRUM: spect
 PROBHD: 5mm BBO BB-1
 PULPROG: zgpg30
 SOLVENT: CDCl₃
 NS: 259
 DS: 4
 SWH: 24875.624 Hz
 FIDRES: 0.37852 Hz
 AQ: 1.317295 sec
 RG: 1149.4
 DM: 20.100 usec
 DE: 6.00 usec
 TE: 300.0 K
 D1: 2.00000000 sec
 d12: 0.03000000 sec
 012: 0.00002000 sec

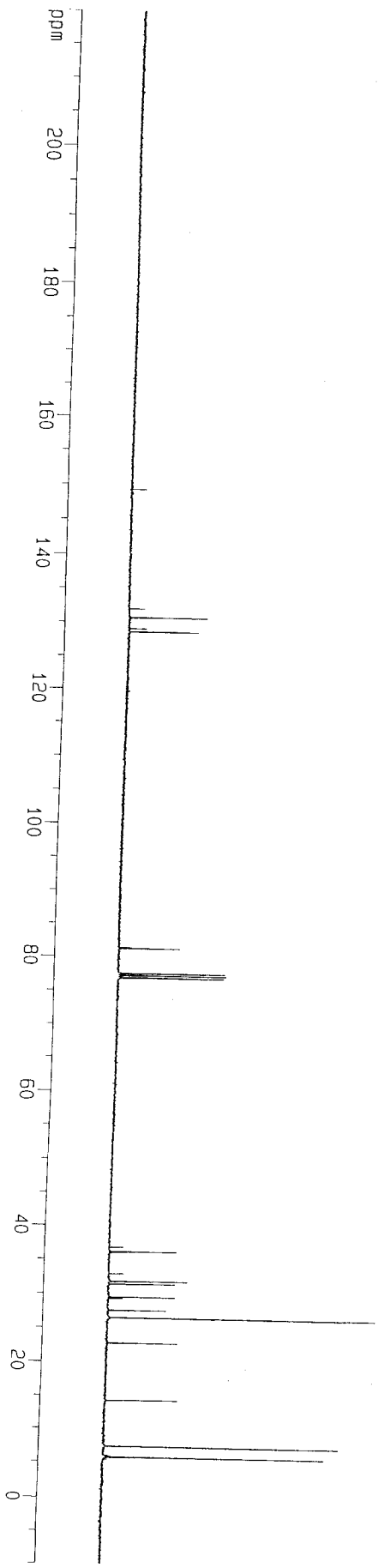
***** CHANNEL f1 *****
 NUC1: ¹³C
 P1: 13.63 usec
 PL1: 0.00 dB
 SFO1: 100.627959 MHz

***** CHANNEL f2 *****
 CPDPRG2: waltz16
 NUC2: ¹H
 PCPD2: 197.50 usec
 PL2: 0.00 dB
 PL12: 24.00 dB
 PL13: 24.00 dB
 SF02: 400.1316005 MHz

F2 - Processing parameters
 SI: 32768
 SF: 100.6127529 MHz
 KW: 64
 SSB: EM
 LB: 1.00 Hz
 GB: 0
 PC: 1.40

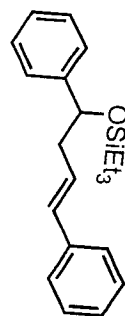
1D NMR plot parameters
 CX: 24.00 cm
 F1P: 220.000 ppm
 F2P: 23134.80 Hz
 F2: -100.000 ppm
 FREQH: 3.5833 ppm/cm
 NZCX: 96.42031 Hz/cm

- 130.612
- 128.506
- 81.205
- 77.479
- 77.161
- 76.844
- 36.155
- 31.793
- 31.355
- 29.463
- 27.601
- 26.480
- 22.775
- 14.241
- 7.293
- 5.693
- 5.671



H-pdt

2h



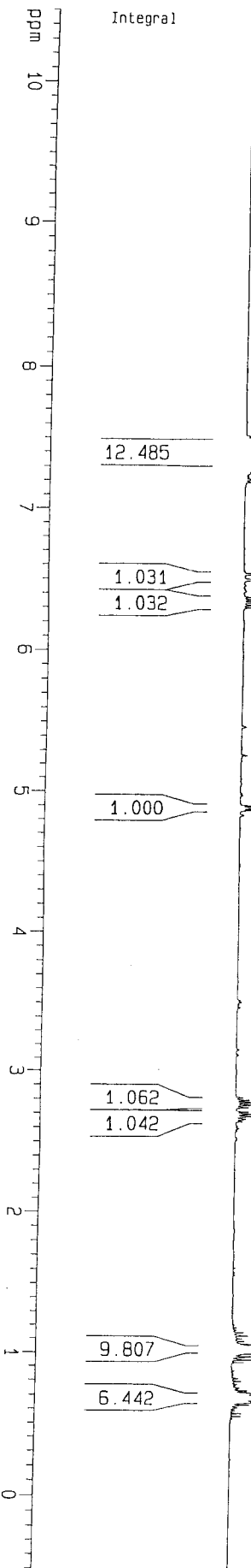
Current Data Parameters
NAME a11yicmtr01/13
EXNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20080206
Time 18:07
INSTRUM spect
PROBHD 5 mm QNP 1H/1
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 5
DS 1
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.5964243 sec
RG 35.9
DM 60.400 usec
DE 6.00 usec
TE 294.2 K
AQC 1.0000000 sec
MDEL 0.0000000 sec
MOR 0.0150000 sec
MORNT 0.0150000 sec

***** CHANNEL f1 *****
NUC1 1H
P1 9.88 usec
PL1 3.00 dB
SFO1 400.1324710 MHz

F2 - Processing parameters
SI 32768
SF 400.1300054 MHz
WDW EM
SSB 0
GB 0.30 Hz
PC 1.00

1D NMR Plot Parameters
CX 24.00 cm
CY 9.71 cm
F1P 10.500 ppm
F1 4201.36 Hz
F2P -0.500 ppm
F2 -200.07 Hz
PRNOM 0.45833 ppm/cm
HZCM 183.35824 Hz/cm



Current Data Parameters
 NAME 01111control13
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060205
 Time 18.10
 INSTRUM spect
 PROBRG 5 aa dnp hv/1
 PULPROG zgpg30
 TD 65536
 SFO1 100.6262626 MHz
 SOLVENT CDCl3
 NS 63
 DS 8
 SWH 2380.814 Hz
 FIDRES 0.365918 Hz
 AQ 1.3664726 sec
 RG 1625.5
 DW 20.650 usec
 DE 6.00 usec
 TE 294.8 K
 D1 2.0000000 sec
 d11 0.0300000 sec
 DELTA 1.8895998 sec
 WDELT 1.0000000 sec
 HCHNK 0.0150000 sec

***** CHANNEL f1 *****
 NUC1 13C
 P1 8.50 usec
 PL1 3.00 dB
 SFO1 100.6262626 MHz

***** CHANNEL f2 *****
 CPDPRG2 waltz16
 NUCL2 1H
 P2 89.01 usec
 PL2 2.00 dB
 PL12 22.00 dB
 PL13 22.00 dB
 SFO2 400.1315005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127505 MHz
 MDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

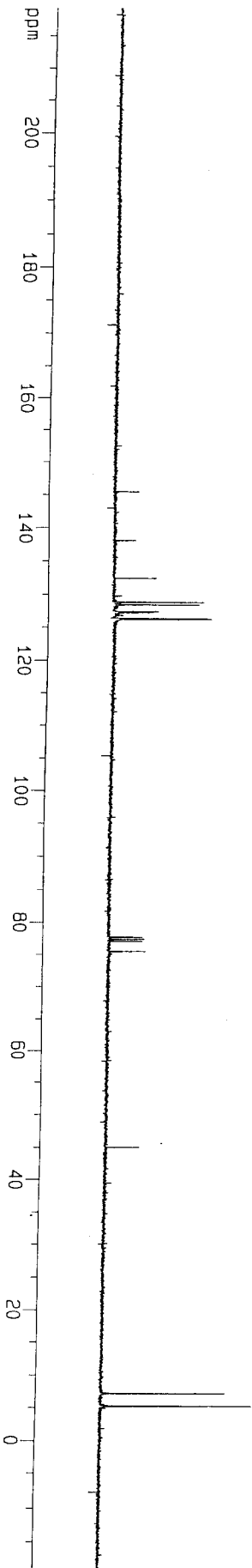
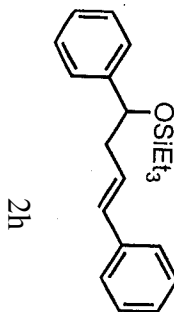
1D NMR plot parameters
 CX 24.00 cm
 CY 2.29 cm
 FIP 219.353 ppm
 F1 22068.67 Hz
 F2P -18.995 ppm
 F2 -1911.15 Hz
 PPMCK 9.93115 ppm/cm
 HZCK 999.20062 Hz/cm

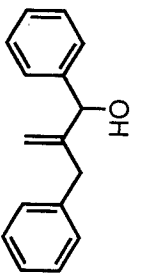
- 145.469
- 138.043
- 132.395
- 128.764
- 128.384
- 127.398
- 127.348
- 127.242
- 126.322
- 126.189

- 77.687
- 77.369
- 77.052
- 75.459

- 45.019

- 7.124
- 5.175
- 5.125





TES deprotection, Apdt

3h (TES group deprotected)

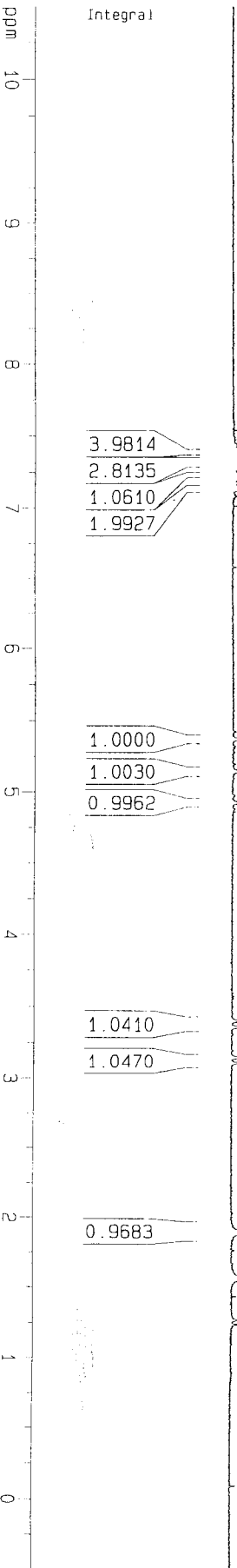
Current Data Parameters
 NAME hc9d-320nuc-4p
 EXPNO 1
 PROCNO 1

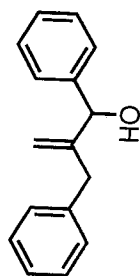
F2 - Acquisition Parameters
 Date_ 20060209
 Time 13:04
 INSTRUM spect
 PROBHD 5mm BBO BB-1
 PULPROG zgpg30
 NO 2830
 SO 65395
 SOLVENT DMS-D13
 NS 2
 DS 2
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.994243 sec
 RG 574.7
 CW 50.400 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec

***** CHANNEL f1 *****
 NUC1 1H
 P1 7.50 usec
 PL 0.10 dB
 SF01 400.1324710 MHz

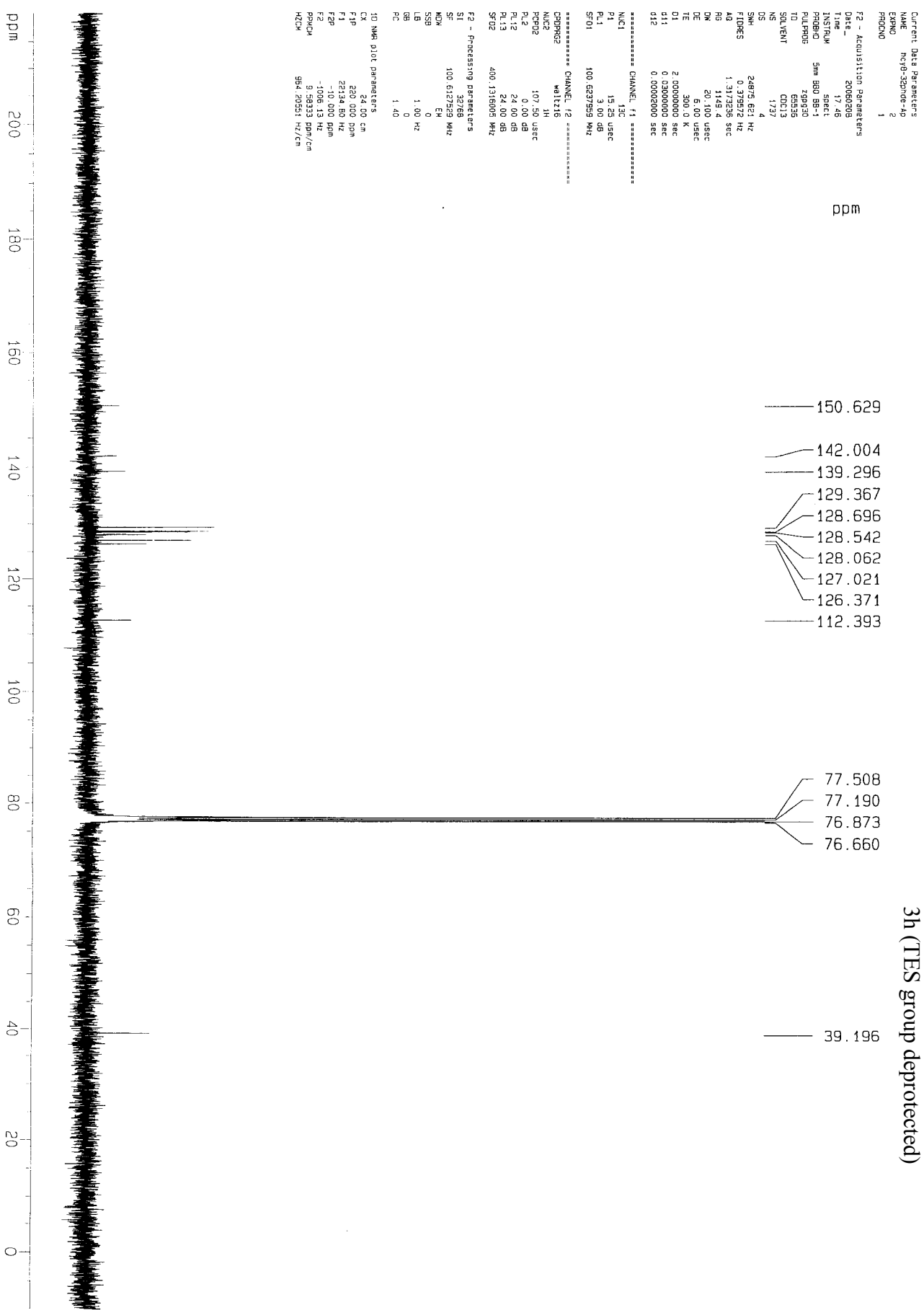
F2 - Processing parameters
 SI 32768
 SF 400.1300016 MHz
 KGM EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CA 24.00 cm
 F1 130.000 dB
 F2 420.000 Hz
 F3P 0.000 Hz
 F2 -200.07 Hz
 PHOVM 0.46833 dB/cm
 HZCM 183.39291 Hz/cm



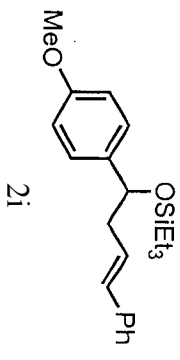


3h (TMS group deprotected)



1
H-pdt
104-8-98

H-pdt



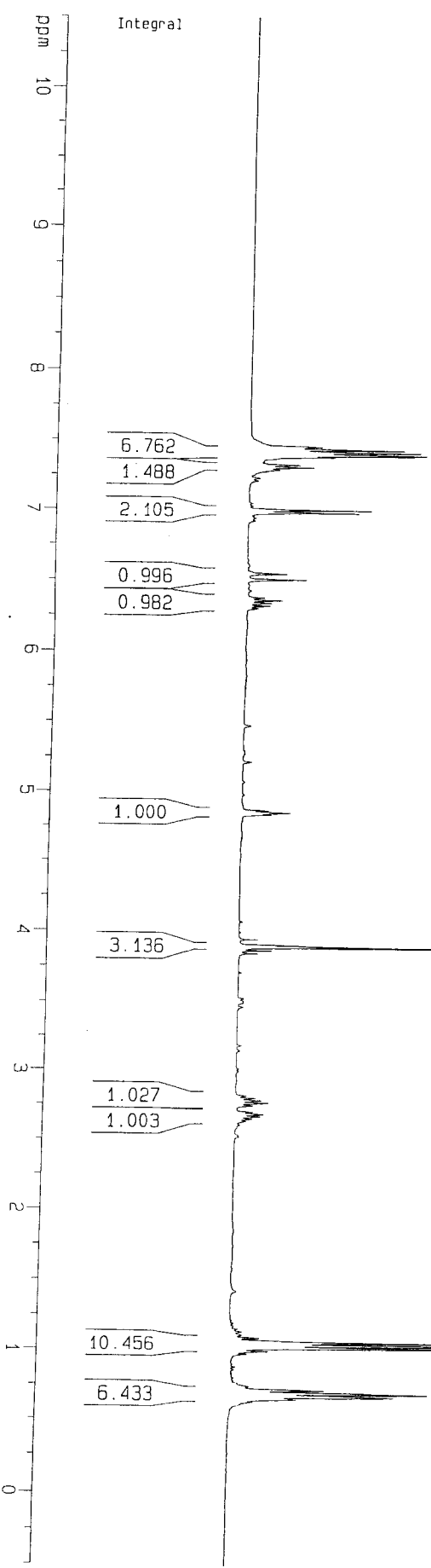
Current Data Parameters
NAME: H-pdt
EXPNO: 1
PROCNO: 1

F2 - Acquisition Parameters
Date_: 20060122
Time: 14.39
INSTRUM: spect
PROBHD: 5mm BBO BB-1
PULPROG: zg30
TD: 65536
SOLVENT: CDCl3
NS: 5
DS: 2
SWH: 8278.146 Hz
FIDRES: 0.162314 Hz
AQ: 3.359242 sec
RG: 60.400
DW: 6.000 usec
DE: 300.0 K
TE: 300.0 K
D1: 1.00000000 sec

***** CHANNEL f1 *****
NUC1: 1H
P1: 7.50 usec
PL1: 0.00 dB
SFO1: 400.1324710 MHz

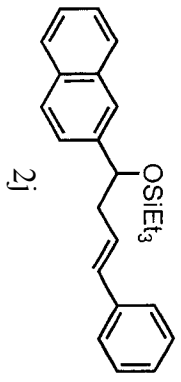
F2 - Processing parameters
SI: 32768
SF: 400.1300918 MHz
WDW: EM
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00

10 NMR plot parameters
CX: 24.00 cm
F1P: 10.500 ppm
F1: 4201.36 Hz
F2P: -0.500 ppm
F2: -200.07 Hz
F2A: 0.05833 ppm/cm
F2M: 183.35291 Hz/cm



NY-8-99 H-pdt

H-pdt



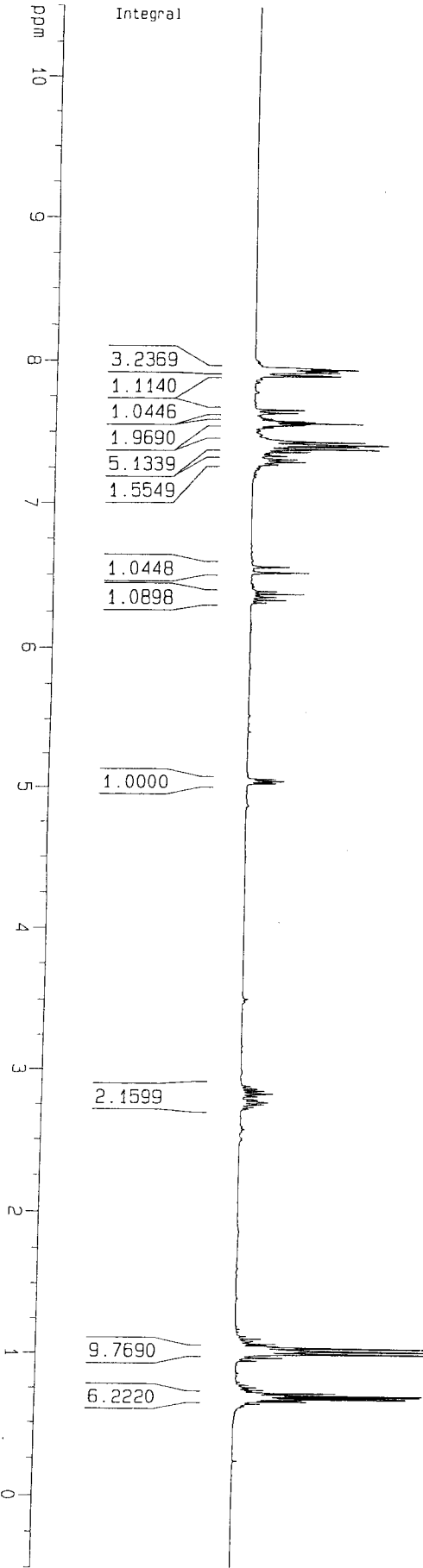
Current Data Parameters
 NAME NY-8-99-Hpdt
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060126
 Time 15:56
 INSTRUM spect
 PROBR4 Sma BBO BB-1
 PULPROG zgpg30
 TO 64536
 SOLVENT CDCl3
 NS 3
 DS 2
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 2072
 DW 60.400 usec
 DE 5.500 usec
 TE 300.0 K
 D1 1.00000000 sec

***** CHANNEL f1 *****
 NUC1 1H
 P1 7.90 usec
 PL1 0.00 dB
 SFO1 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300018 MHz
 MDW 64
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

10 NMR data parameters
 CX 24.00 cm
 F1P 10.500 ppm
 F1 4201.36 Hz
 F2P -0.500 ppm
 F2 -200.07 Hz
 PRNCH 0.45833 ppm/cm
 NUCN 189.59291 Hz/cm



Current Data Parameters
 NAME 103957-N004
 EXPNO 2
 PROCNO 1

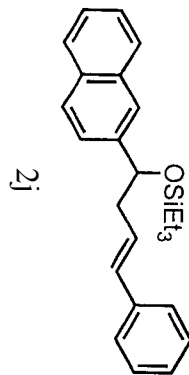
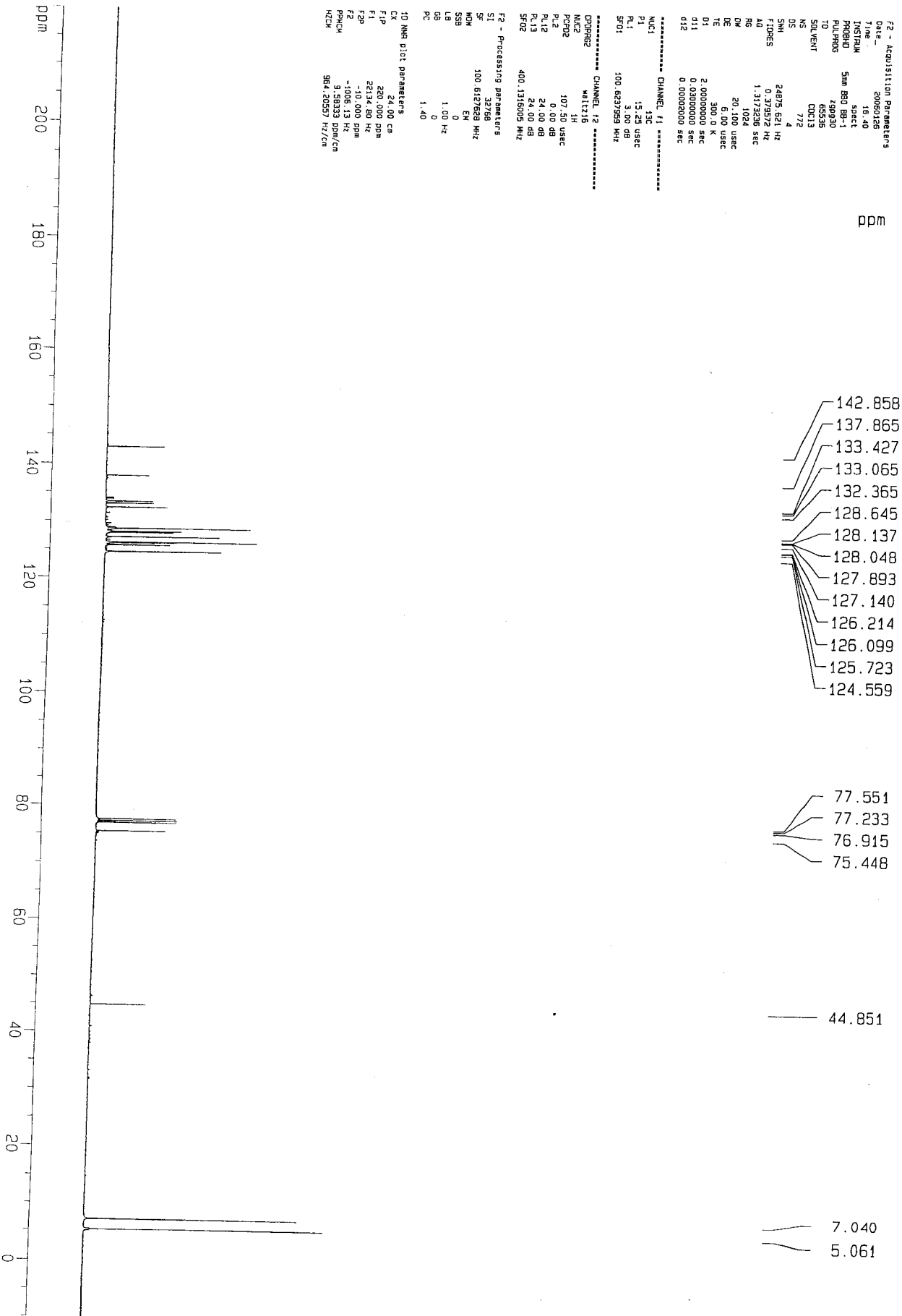
F2 - Acquisition Parameters
 Date_ 20060326
 Time 16:40
 INSTRUM spect
 PROBRD Spin BB9 BB-1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 772
 DS 4
 SWH 24875.621 Hz
 FIDRES 0.379572 Hz
 AQ 1.317259 sec
 RG 64
 DW 20.104 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.00000000 sec
 d11 0.03000000 sec
 d12 0.00002000 sec

***** CHANNEL f1 *****
 NUCL1 13C
 P1 15.25 usec
 F1 3.00 dB
 SFO1 100.6237959 MHz

***** CHANNEL f2 *****
 GPCPRG2 waltz16
 NUCL2 1H
 P2PRG2 107.50 usec
 PL2 0.00 dB
 PL12 24.00 dB
 PL13 24.00 dB
 SFO2 400.1316005 MHz

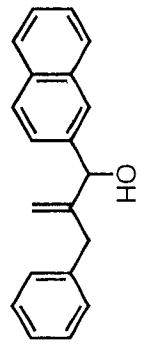
F2 - Processing parameters
 SI 32768
 SF 100.6127628 MHz
 WHW 0
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

ID NMR plot parameters
 CX 24.00 cm
 FIP 220.000 ppm
 F1 22134.80 Hz
 F2P -10.000 ppm
 F2 -1006.13 Hz
 PPHC0 3.58333 DM/cm
 HZCM 964.20557 Hz/cm



MSY-8-104 dr - Apdt

TES deprotection, Apdt



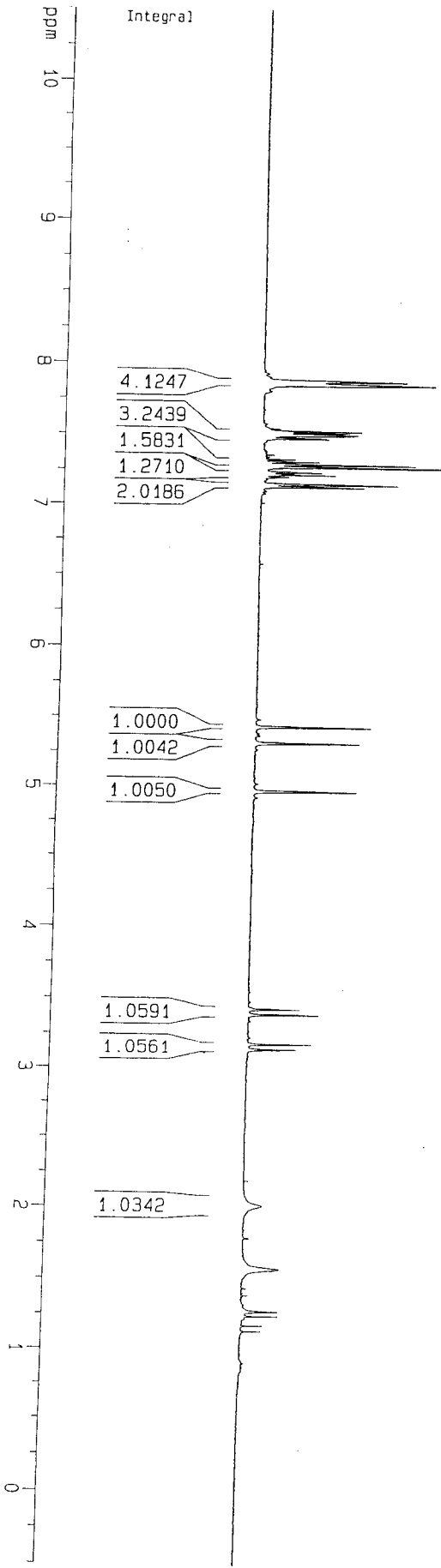
Current Data Parameters
 NAME MSY-104dr-Apdt
 EXPNO 1
 PROCNO 1

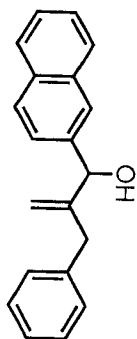
F2 - Acquisition Parameters
 Date_ 20060202
 Time 17.33
 INSTRUM spect
 PROBRD 5mm BBO BB-1
 PULPROG zg30
 ID 65536
 SOLVENT CDCl3
 NS 7
 DS 2
 SM 6278.148 Hz
 FIDRES 0.42314 Hz
 AQ 3.58643 sec
 RG 456.4
 DW 60.400 usec
 DE 6.00 usec
 TE 300.2 K
 D1 1.00000000 sec

***** CHANNEL f1 *****
 NUCL1 1H
 P1 7.50 usec
 PL 0.00 dB
 SF01 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300016 MHz
 WDM EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

10 NMR plot parameters
 DX 24.00 cm
 F1 10.500 ppm
 F2 420.38 Hz
 F3 -30.000 ppm
 PPH0M 0.46933 Hz/cm
 HZCM 183.39291 Hz/cm





3j (TES group deprotected)

- 150.510
- 149.210
- 139.370
- 139.268
- 133.424
- 133.277
- 129.397
- 128.558
- 128.211
- 127.880
- 126.395
- 126.359
- 126.215
- 125.969
- 124.846
- 112.791
- 77.511
- 77.397
- 77.193
- 76.876
- 39.212

```

Current Data Parameters
NAME      hc9-10car-10a
EXPNO    1
PROCNO   1

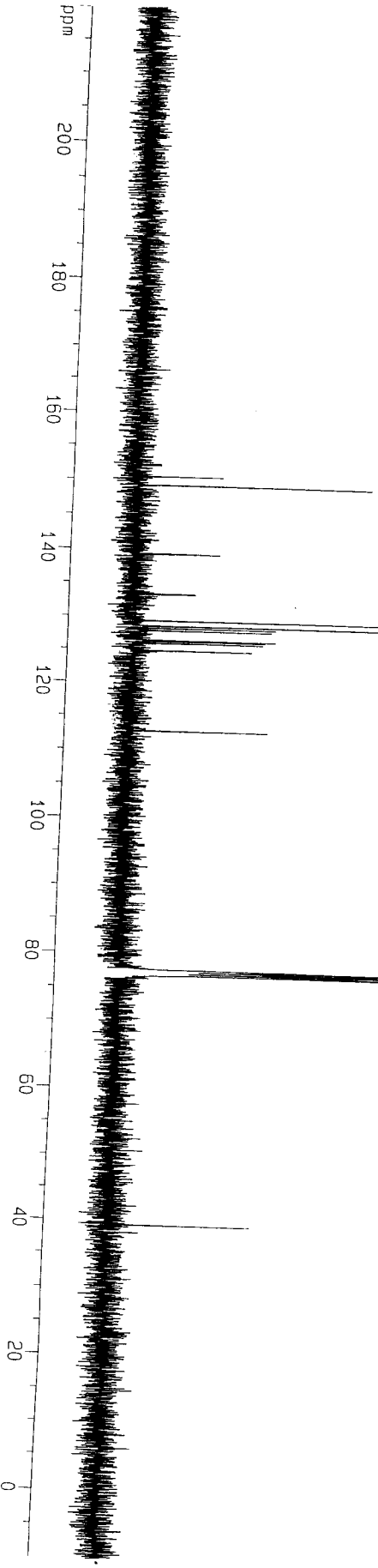
F2 - Acquisition Parameters
Date_    20060203
Time     17.57
INSTRUM  spect
PROBHD   5mm BBO BB-1
PULPROG  zgpg30
TD        65536
SOLVENT  CDCl3
NS        2483
DS        4
SWH       24875.621 Hz
FIDRES   0.32762 Hz
AQ        1.317223 sec
RG        574.7
DM        20.100 usec
DE        6.00 usec
TE        300.0 K
D1        2.00000000 sec
D11       0.02000000 sec
D12       0.00020000 sec

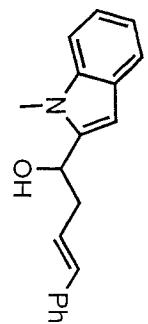
***** CHANNEL f1 *****
NUC1      13C
P1        13.23 usec
PL1       3.00 dB
SFO1      100.627959 MHz

***** CHANNEL f2 *****
CPDPRG2  waltz16
NUC2      1H
P2        107.50 usec
PL2       0.00 dB
PCPD2    24.00 dB
PL12     24.00 dB
PL13     24.00 dB
SFO2     400.1315005 MHz

F2 - Processing parameters
SI        32768
SF        100.6127525 MHz
MGM       EM
SSB       0
LB        1.00 Hz
GB        0
PC        1.40

1D NMR plot parameters
EX        24.00 cm
FIP       220.000 ppm
F1        22134.80 Hz
F2P       -10.000 ppm
F2        -1026.13 Hz
SFO1CM    9.58333 ppm/cm
HZCM      984.20251 Hz/cm
  
```





Me-Indo-2-CHO AllylPh coupling, H pdt deprotection

2k (TES group deprotected)

Current Data Parameters
 NAME: HCV8-706-Hpdt
 EXPNO: 1
 PROCNO: 1

F2 - Acquisition Parameters

Date_: 20051210
 Time: 12.45
 INSTRUM: spect
 PULPROG: 5ms BBD BR-1
 TD: 65536
 SOLVENT: CDCl3
 NS: 5
 DS: 2
 SMI: 8278.146 Hz
 FIDRES: 0.126314 Hz
 AQ: 3.9584243 sec
 RG: 297.4
 DM: 50.400 usec
 DE: 6.00 usec
 TE: 300.0 K
 D1: 1.00000000 sec

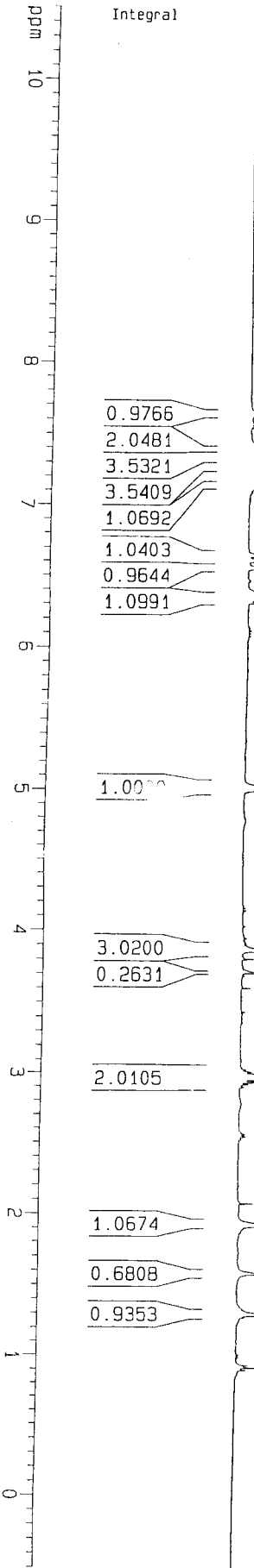
***** CHANNEL f1 *****

NUC1: 1H
 P1: 7.50 usec
 PL1: 0.00 dB
 SFO1: 400.1324710 MHz

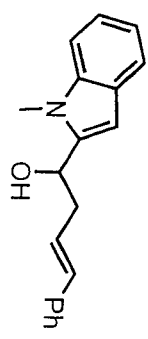
F2 - Processing parameters

SI: 32768
 SF: 400.1300015 MHz
 MDW: EM
 SSB: 0
 LB: 0.30 Hz
 GB: 0
 PC: 1.00

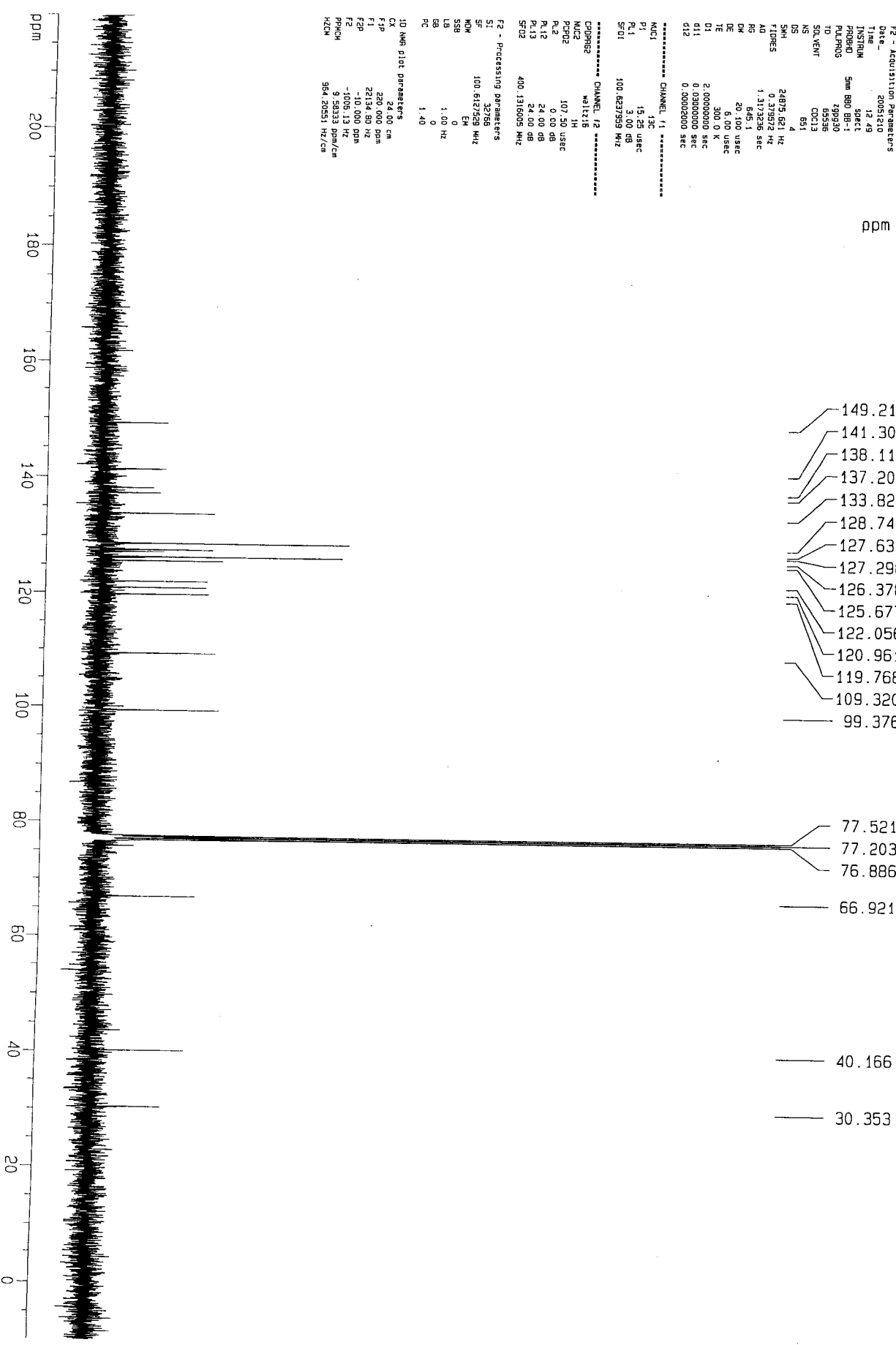
1D NMR plot parameters
 CX: 24.00 cm
 F1P: 10.500 ppm
 F1: 4201.36 Hz
 F2P: -0.500 ppm
 F2: -200.07 Hz
 PPM/CM: 0.45833 ppm/cm
 HZ/CM: 183.39291 Hz/cm



2k (TESS group deprotected)



- 149.210
- 141.305
- 138.119
- 137.200
- 133.821
- 128.741
- 127.632
- 127.298
- 126.378
- 125.677
- 122.056
- 120.961
- 119.768
- 109.320
- 99.376
- 77.521
- 77.203
- 76.886
- 66.921
- 40.166
- 30.353



Current Data Parameters
 NAME NS18-706-INDT
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20051210
 Time 12.49
 INSTRUM spect
 PROBRD 5mm BBO BB-1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 651
 DS 4
 SWH 24875.621 Hz
 FTWRES 0.379572 Hz
 AQ 1.322626 sec
 RG 64
 DW 20.100 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.00000000 sec
 d11 0.03000000 sec
 d12 0.00002000 sec

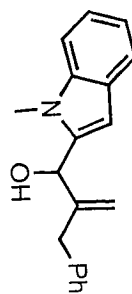
***** CHANNEL f1 *****
 NUCl1 13C
 P1 15.25 usec
 SFO1 100.6237959 MHz

***** CHANNEL f2 *****
 CDPORG2 NA1TC14
 NUCl2 1H
 P2 107.50 usec
 PCPD2 0.00 dB
 PL12 24.00 dB
 PL13 24.00 dB
 SFO2 400.1316005 MHz

F2 - Processing parameters
 SI 32766
 SF 100.6127529 MHz
 SW 4000.000 Hz
 SSB 0
 EM 0
 LB 1.00 Hz
 GB 0
 PC 1.40

10 NMR plot parameters
 CX 24.00 cm
 FIP 220.000 ppm
 F1 22134.80 Hz
 F2P -10.000 ppm
 F2 -1005.13 Hz
 PPRCH 9.58333 ppm/cm
 MICH 954.20051 Hz/cm

CY2PPH f9 deprotection, f6



3K (TES group deprotected)

```

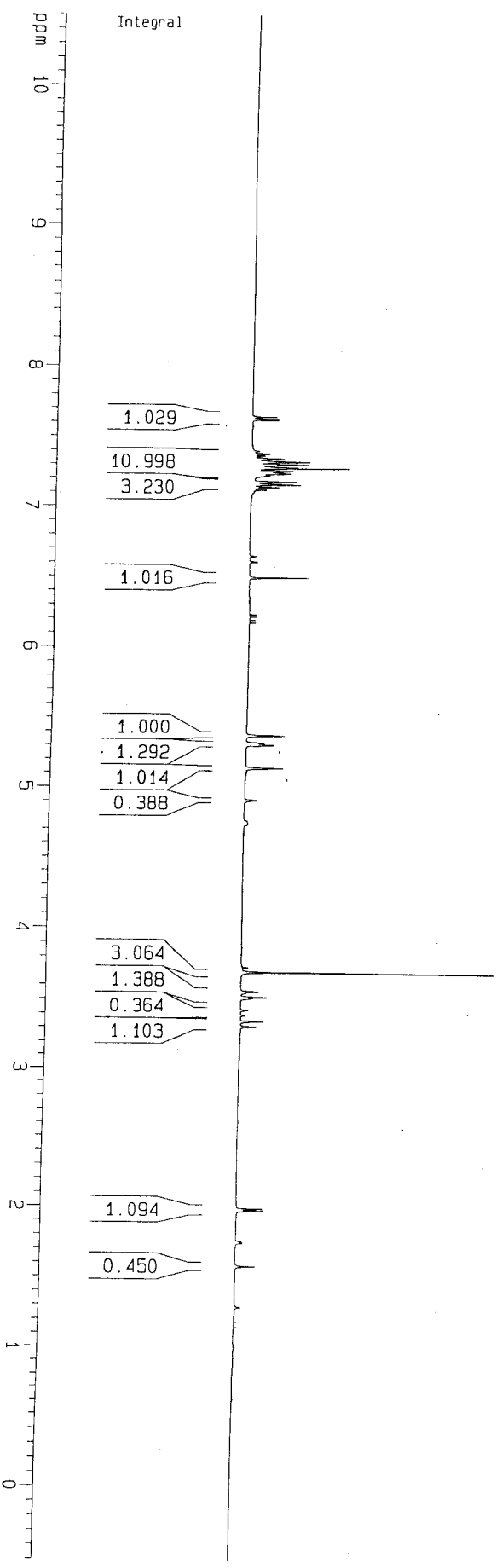
Current Data Parameters
NAME      ncy8-700c19-6
EXPNO    1
PROCNO   1

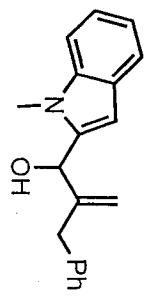
F2 - Acquisition Parameters
Date_    20051212
Time     14.52
INSTRUM  spect
PROBHD   5mm BBO BB-1
PULPROG  zg30
TD        65536
SOLVENT  CDCl3
NS        12
DS        2
SWH       8278.146 Hz
FIDRES    0.182914 Hz
AQ         3.9584443 sec
RG         181
DE         60.400 usec
TE         300.2 K
D1         1.0000000 sec

***** CHANNEL f1 *****
NUC1      1H
P1        7.90 usec
PL1       0.00 dB
SFO1      400.1384710 MHz

F2 - Processing parameters
SI        32768
SF        400.1380018 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB        0
PC        1.00

1D NMR Plot Parameters
CX        24.00 cm
F1P       10.500 ppm
F1        4201.36 Hz
F2P       -0.500 ppm
F2        200.07 Hz
SFOUHZ    4.26833 ppm/cm
HZCM      183.3921 Hz/cm
    
```





3k (TES group deprotected)

```

Current Data Parameters
NAME hc98-700c19-6
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date_ 20031212
Time 15:01
INSTRUM sm
PROBHD 5mm BBO 5B
PULPROG zgpg30
TD 65536
SOLVENT CCl3
NS 593
DS 4
SFO1 24875.621 Hz
FIDRES 0.379572 Hz
AQ 1.3172326 sec
RG 1629.5
DM 20.100 usec
DE 6.00 usec
TE 300.0 K
D1 2.00000000 sec
d12 0.00000000 sec

***** CHANNEL f1 *****
NUC1 13C
P1 15.25 usec
PL1 3.00 dB
SFO1 100.627959 MHz

***** CHANNEL f2 *****
CPDPRG2 waltz16
NUC2 1H
P2 107.50 usec
PL2 0.00 dB
PL12 24.00 dB
PL13 24.00 dB
SFO2 400.131800 MHz

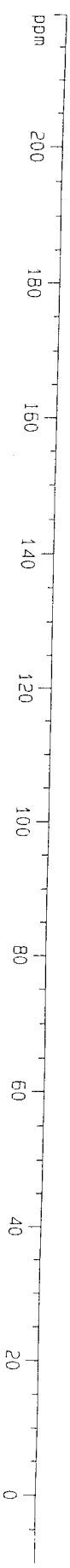
F2 - Processing parameters
SI 32768
SF 100.612759 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

10 NMR 010t parameters
CA 24.00 cm
F1 220.000 ppm
F2 22134.80 Hz
F3 -10.000 ppm
F4 -1006.13 Hz
PNUC1 964.05931 Hz/cm
PNUC2
  
```

- 148.669
- 139.626
- 139.078
- 138.405
- 129.426
- 129.284
- 128.766
- 128.598
- 127.277
- 126.748
- 126.529
- 126.424
- 122.042
- 120.963
- 119.725
- 113.198
- 109.320
- 101.450

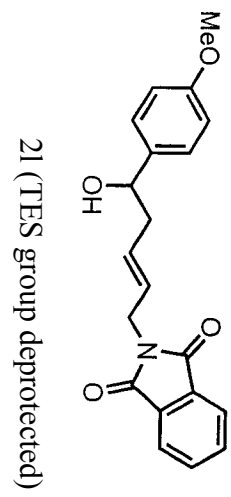
- 77.522
- 77.204
- 76.887
- 69.611

- 40.225
- 30.292



May 8-8-2001

H pdt TES deprotection



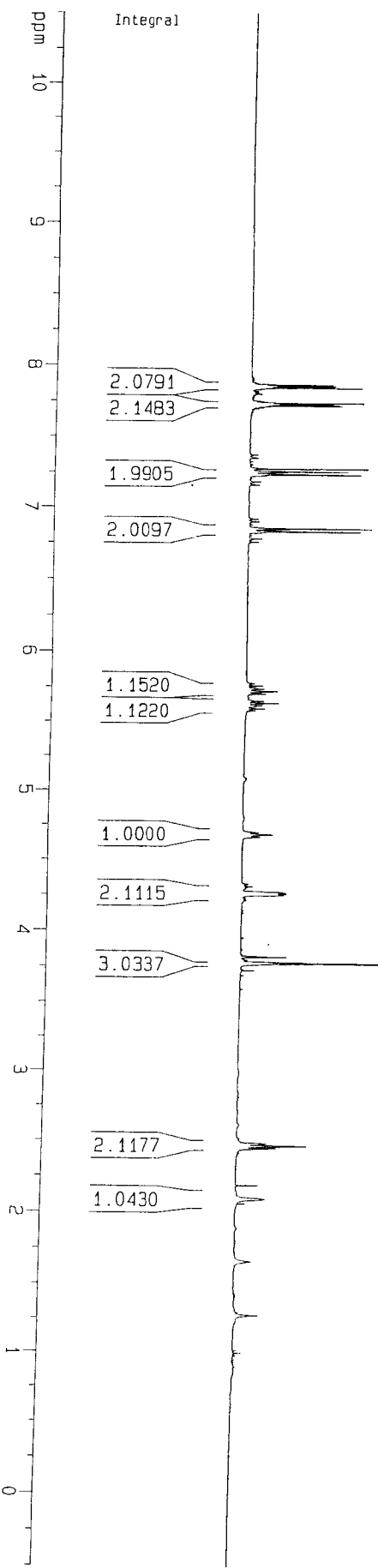
Current Data Parameters
 NAME hcy8-57de-mpdt
 EXPNO 1
 PROCNO 1

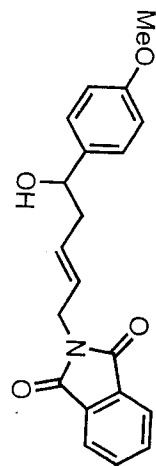
F2 - Acquisition Parameters
 Date_ 2008123
 Time 12:38
 INSTRUM spect
 PROBRD 5mm BBO BB-1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 5
 DS 2
 SWH 8278.148 Hz
 FIDRES 0.128314 Hz
 AQ 3.9584243 sec
 RG 181
 DW 60.400 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec

***** CHANNEL f1 *****
 NUCL1 13
 P1 7.50 usec
 PL1 0.00 dB
 SF01 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300016 MHz
 MD 64
 EM 1
 SSF 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 324.00 cm
 F1P 10.500 ppm
 F1 4201.36 Hz
 F2P -0.500 ppm
 F2 -200.07 Hz
 PRPCK 0.45833 ppm/cm
 HZCM 1813.39291 Hz/cm





21 (TES group deprotected)

- 77.523
- 77.410
- 77.205
- 76.888
- 73.124
- 55.410
- 42.328
- 39.724

- 168.173
- 159.126
- 136.046
- 134.137
- 132.282
- 130.787
- 127.192
- 127.139
- 123.459
- 113.885

ppm

Current Data Parameters
 NAME hcy8-57de-Hot
 EXPNO 2
 PROCNO 1

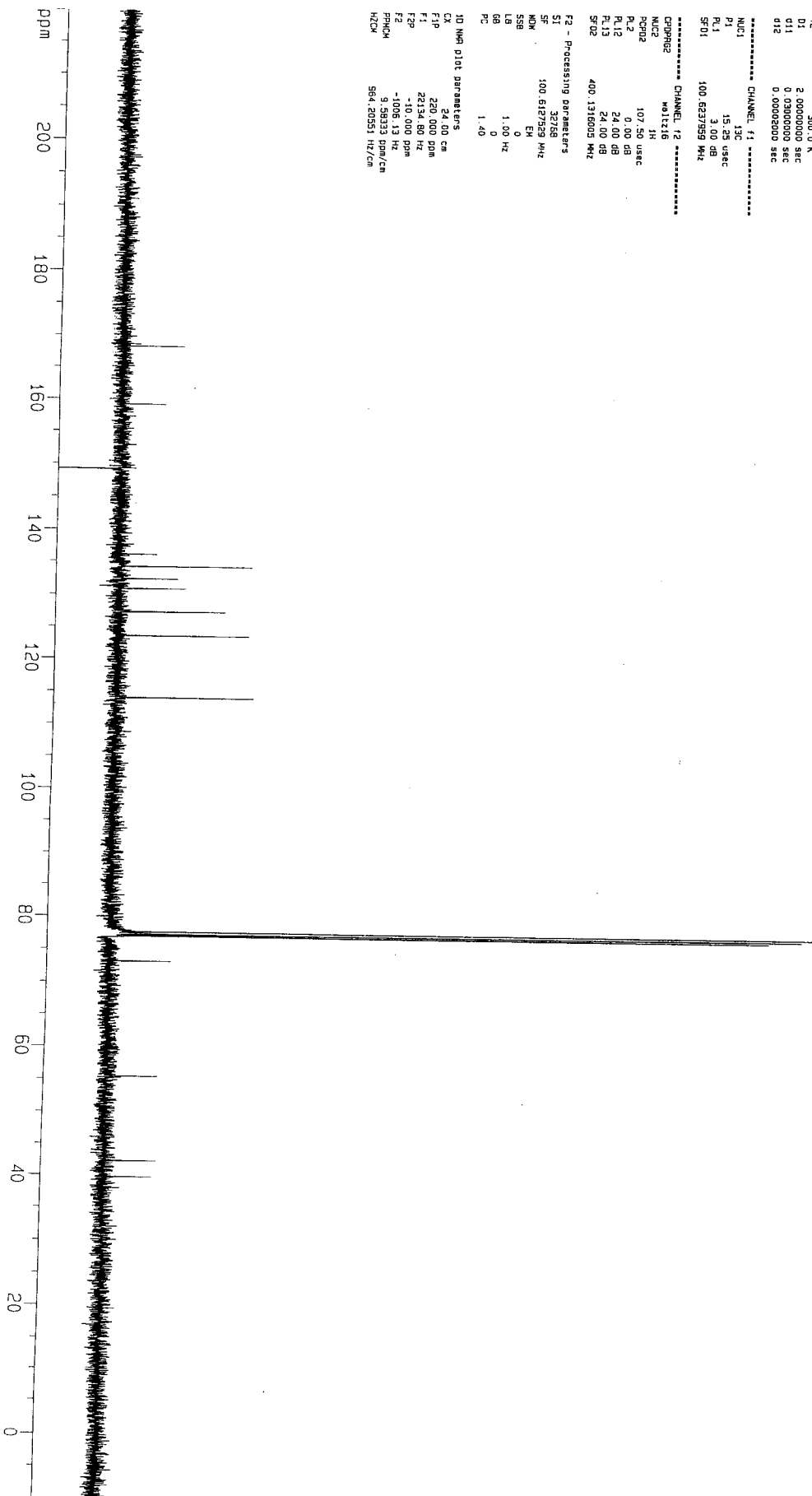
F2 - Acquisition Parameters
 Date_ 20060925
 Time 15.44
 INSTRUM spect
 PROBRW 5mm BBO BB-1
 PULPROG zgpg30
 TO 65536
 SOLVENT CDCl3
 NS 179
 DS 4
 SH 4
 FIDRES 24875.621 Hz
 0.379572 Hz
 1.3173236 sfc
 AQ 0.0
 FWH 20.1024
 DM 20.1024 usfc
 DE 4.00 usfc
 TE 300.0 K
 D1 2.00000000 sec
 d11 0.03000000 sec
 d12 0.00002000 sec

***** CHANNEL f1 *****
 NUC1 13C
 P1 15.25 usfc
 PL1 3.00 dB
 SF01 100.6237559 MHz

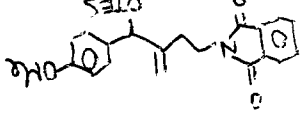
***** CHANNEL f2 *****
 CHOPRG2 MSLC14
 NUC2 1H
 PCPR2 107.50 usfc
 PL2 0.00 dB
 PL12 24.00 dB
 PL13 24.00 dB
 SF02 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127559 MHz
 RM 0
 SM 0
 LS 0
 GB 1.00 Hz
 GB 0
 PC 1.40

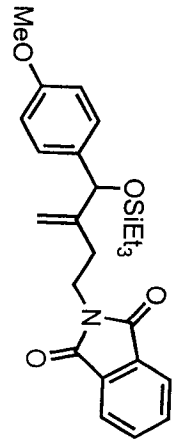
JD NMR plot parameters
 CX 24.00 cm
 FIP 220.000 ppm
 F1 22134.89 Hz
 F2 -10.000 ppm
 F2 -1006.13 Hz
 FWHM 9.158333 ppm/cm
 HZCM 584.20551 Hz/cm



NY-8-113-A.pdf



A-pdt



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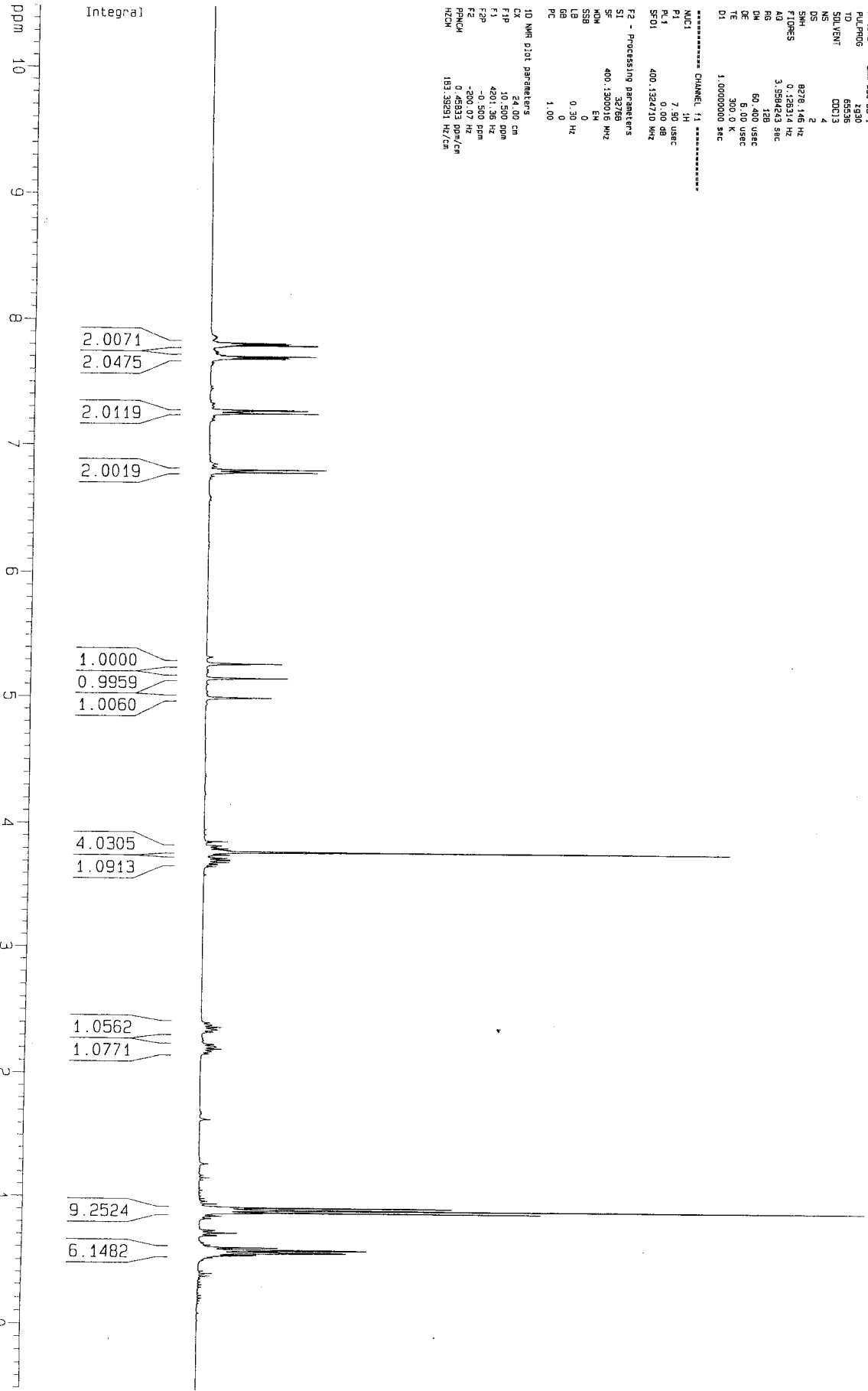
Current Data Parameters
 NAME nc8-113-4pdt
 EXNO 11
 PROCNO 1

F2 - Acquisition Parameters
 Date 20060210
 Time 18:23
 INSTRUM spect
 PROBRD Smm BBO BR-1
 PULPROG zg30
 TD 65536
 SOLVENT DMS-D₆
 DS 4
 DE 2
 SFO 8278.146 Hz
 FIDRES 0.42834 Hz
 AQ 3.558443 sec
 RG 128
 DM 60.400 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec

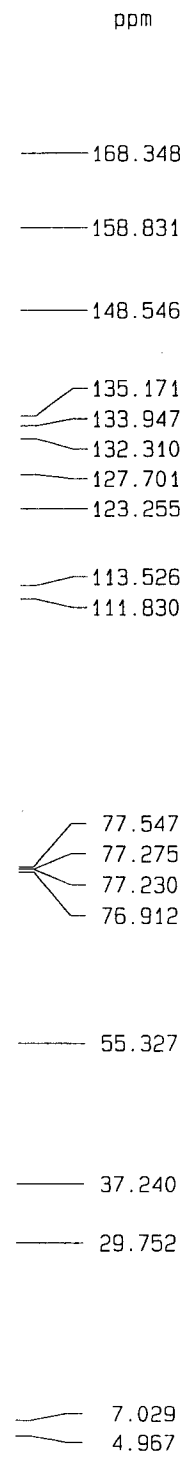
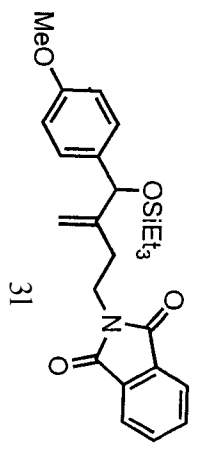
***** CHANNEL f1 *****
 NUC1 ¹H
 P1 7.50 usec
 PL1 0.00 dB
 SFO1 400.1362710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1360016 MHz
 MDW 0
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 24.00 cm
 F1P 10.500 DDM
 F1 4201.36 Hz
 F2 -0.500 ppm
 F2 200.07 Hz
 PRNCH 0.42833 ppm/cm
 HZCN 183.38291 Hz/cm



A-pdt



```

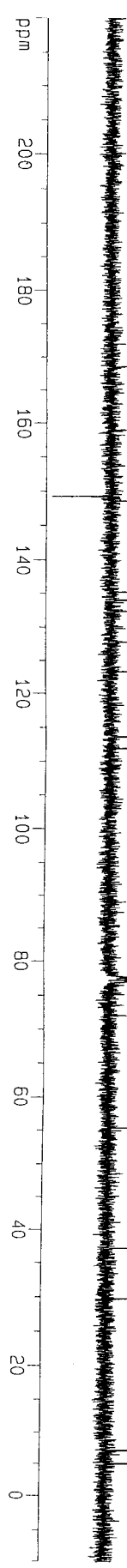
Current Data Parameters
NAME      NCV8-113-Apdt
EXPNO    22
PROCNO   1
F2 - Acquisition Parameters
Date_    20060210
Time     19:28
INSTRUM  spect
PROBHD   5mm BBO
PULPROG  zgpg30
TD        65536
SOLVENT  CDCl3
NS        127
DS        4
SHE      24875.621 Hz
FIDRES   0.379572 Hz
AQ        1.3172285 sec
RG        1024
DM        20.100 usec
DE        6.00 usec
TE        300.2 K
D1        2.000000 sec
d11       0.10000000 sec
d12       0.10000000 sec

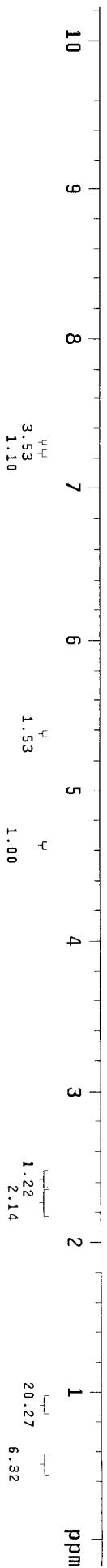
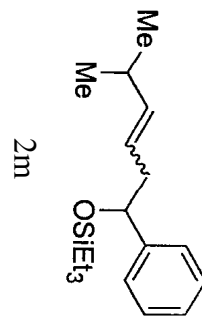
***** CHANNEL f1 *****
NUC1      13C
P1        15.25 usec
PL1       3.00 dB
SFO1     100.6273599 MHz

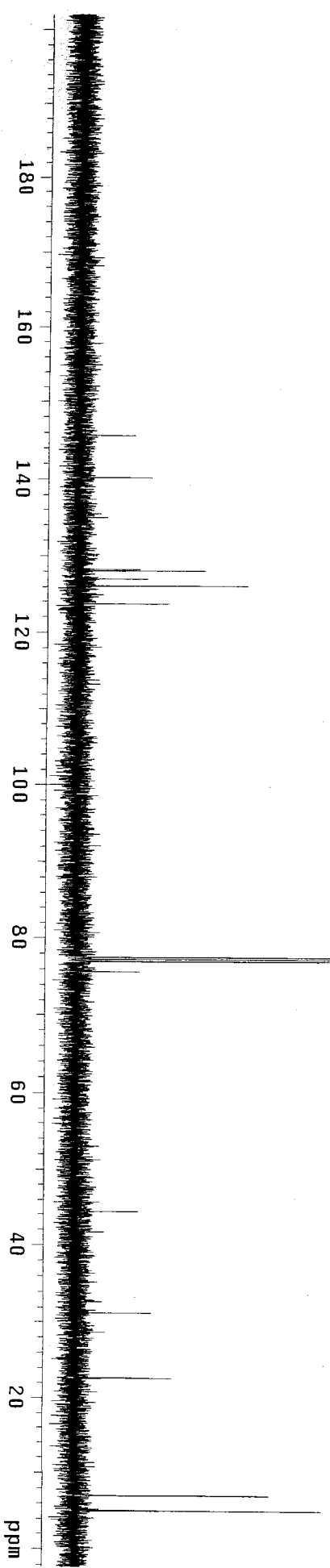
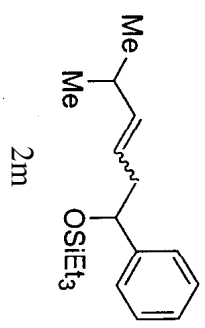
***** CHANNEL f2 *****
CPDPRG2  waltz16
NUC2      1H
PCPD2    107.50 usec
PL2       0.00 dB
PL12     24.00 dB
PL13     24.00 dB
SFO2     400.1310003 MHz

F2 - Processing parameters
SI        32768
SF        100.6127505 MHz
MDW       EM
SSB       0
LB        1.00 Hz
GB        0
PC        1.40

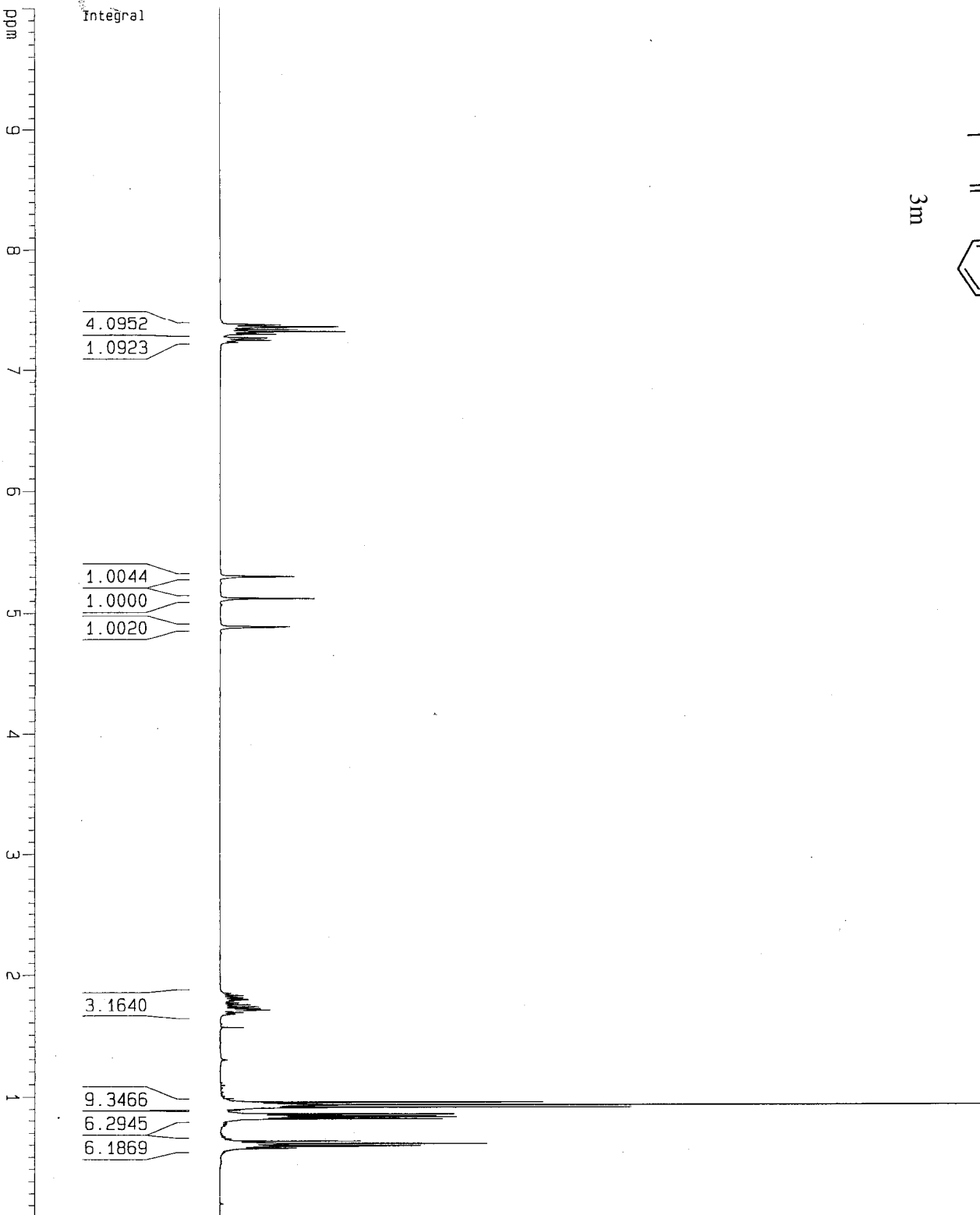
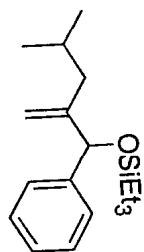
1D NMR plot parameters
CX        24.00 cm
F1P       220.100 GHz
F2P       221.240 MHz
F3P       221.400 MHz
F2 - 1006.13 Hz
PPMCH    9.98333 Hz/cm
KZCN     564.20544 Hz/cm
  
```







SN050726 allylic alcohol



Current Data Parameters
 NAME SN726-ally-H
 EXPNO 1
 PROCNO 1

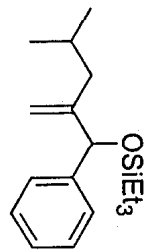
F2 - Acquisition Parameters
 Date_ 20050722
 Time 21:51
 INSTRUM spect
 PROBHD 5mm BBO BB-1
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 32
 DW 60.400 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 7.90 usec
 PL1 0.00 dB
 SF01 400.1324710 MHz

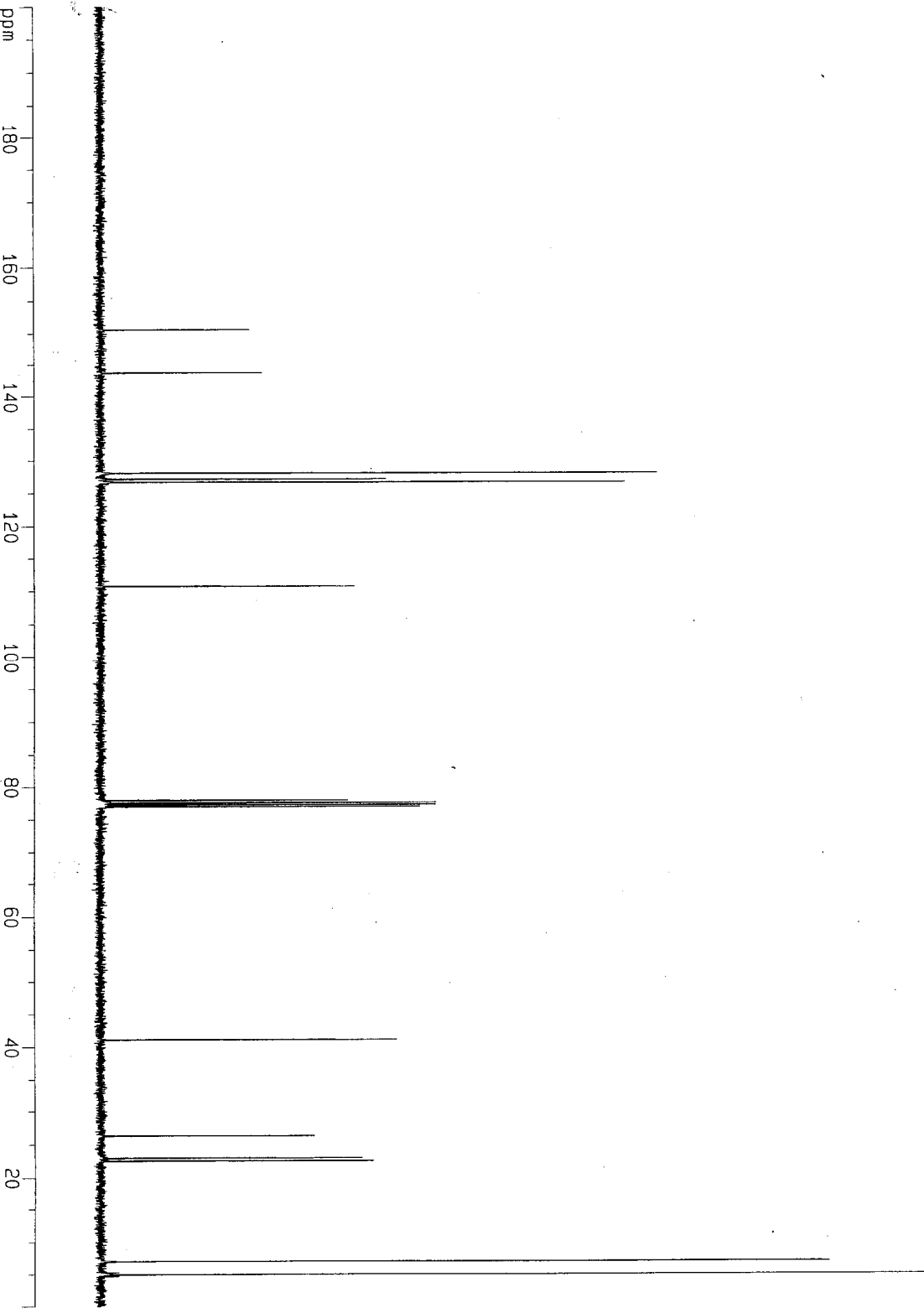
F2 - Processing parameters
 SI 32768
 SF 400.1300056 MHz
 MDM EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 10.000 ppm
 F1 4001.30 Hz
 F2P 0.000 ppm
 F2 0.00 Hz
 PPMCM 0.50000 ppm/cm
 HZCM 200.06500 Hz/cm

SN050726 allylic alcohol



3m



Current Data Parameters
 NAME SN726-ajj-C
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20050722
 Time 21:53

INSTRUM spect
 PROBD 5mm BBO BB-1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 159
 DS 4

SMH 25125.629 Hz
 FIDRES 0.383387 Hz
 AQ 1.3042164 sec
 RG 8192

DW 19.900 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.00000000 sec
 d11 0.03000000 sec
 d12 0.00002000 sec

CHANNEL f1 9
 CHANNEL f2 9

NUC1 13C
 P1 15.25 usec
 PL1 3.00 dB
 SFO1 100.6237959 MHz

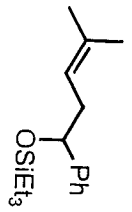
CPDPRG2 waltz16
 CHANNEL f2
 NUC2 1H
 PCPD2 107.50 usec
 PL2 0.00 dB
 PL12 24.00 dB
 PL13 24.00 dB
 SFO2 400.1315005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127499 MHz
 MDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters
 CX 20.00 cm
 F1P 200.000 ppm
 F1 20122.55 Hz
 F2P 0.000 ppm
 F2 0.00 Hz
 PPKCM 10.00000 ppm/cm
 HZCW 1006.12744 Hz/cm

NY-8-90

2n



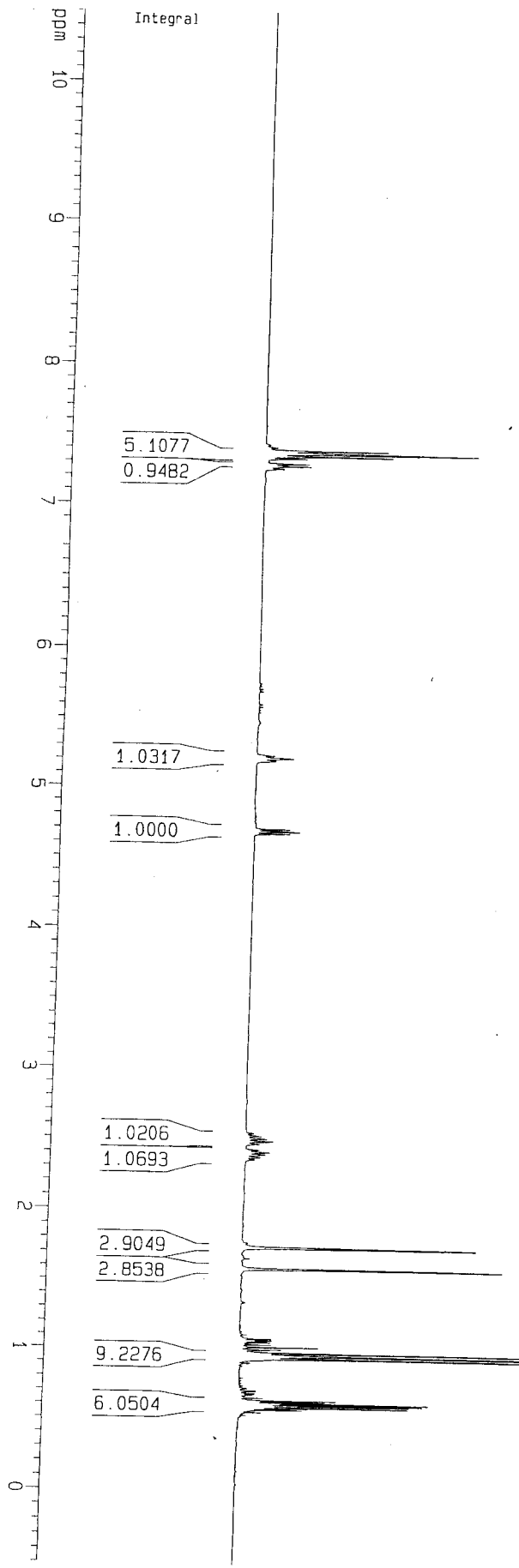
Current Data Parameters
 NAME ncy8-90mg
 EXPNO 1
 PROCNO 1

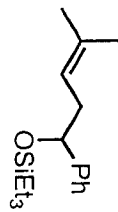
F2 - Acquisition Parameters
 Date_ 20080113
 Time 12 01
 INSTRM spect
 PROBRG Sma 890 BR-1
 PULPROG zgpg
 TO 5.430
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.5984243 sec
 RG 14.3
 DW 60.400 usec
 DE 5.00 usec
 TE 300.0 K
 D1 1.00000000 sec

***** CHANNEL f1 *****
 NUC1 1H
 P1 7.00 usec
 PL1 0.20 dB
 SFO1 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300018 MHz
 NQ 1
 EM 0
 SS 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR 01st Parameters
 CX 24.000 cm
 F1P 14.500 ppm
 F1 4201.36 Hz
 F2p -200.07 Hz
 F2 0.45833 ppm/cm
 FPCW 183.39291 Hz/cm





2n

Current Data Parameters
 NAME: hcyB50pm
 EXPNO: 2
 PROCNO: 1

ppm

- 145.817
- 133.554
- 128.093
- 127.021
- 126.111
- 121.036
- 77.561
- 77.244
- 76.926
- 75.366
- 39.980
- 25.972
- 18.008
- 6.965
- 5.028

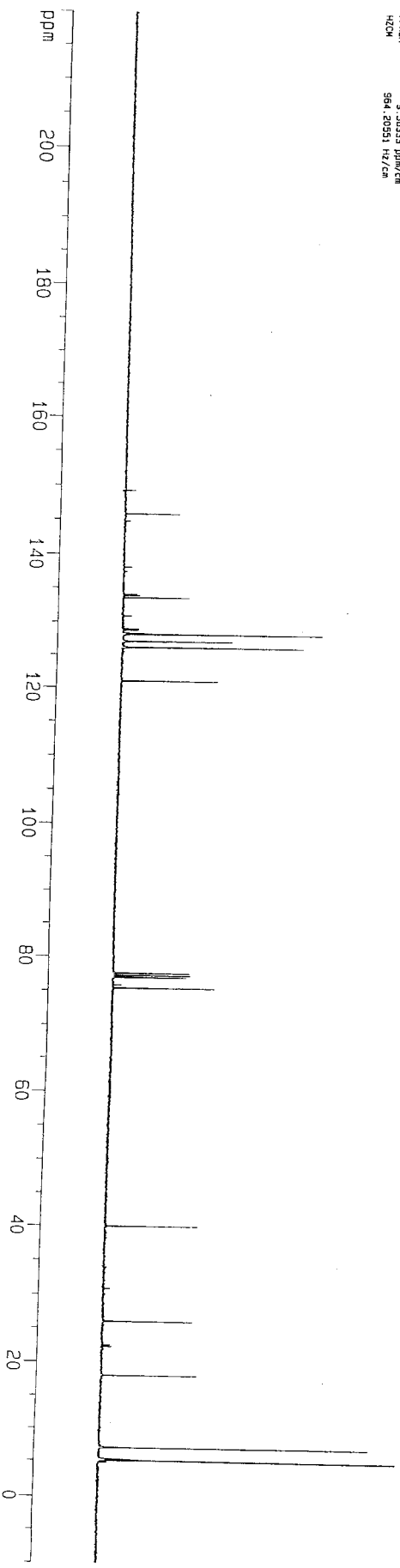
F2 - Acquisition Parameters
 Date_: 20060113
 Time: 12:04
 INSTRUM: spect
 PROBRD: 5mm BBP-BB-1
 PULPROG: zgpg30
 TD: 65536
 SOLVENT: CDCl3
 NS: 151
 DS: 4
 SWH: 24875.621 Hz
 FIDRES: 0.376572 Hz
 AQ: 1.3173236 sec
 RG: 1024
 DW: 20.100 usec
 DE: 6.00 usec
 TE: 309.0 K
 D1: 2.00000000 sec
 d11: 0.03000000 sec
 d12: 0.00002000 sec

***** CHANNEL f1 *****
 NUCL: 13C
 P1: 15.26 usec
 PL1: 3.00 dB
 SF01: 100.6237599 MHz

***** CHANNEL f2 *****
 CPDPRG2: waltz16
 NUCL2: 1H
 P2: 107.50 usec
 PL2: 0.00 dB
 PL12: 24.00 dB
 PL13: 24.00 dB
 SF02: 400.1316005 MHz

F2 - Processing parameters
 SI: 32768
 SF: 100.6127529 MHz
 MW: 64
 SSB: 0
 LB: 1.00 Hz
 GB: 0
 PC: 1.40

ID NMR plot parameters
 CA: 24.00 cm
 FAP: 220.000 ppm
 F1: 22134.80 Hz
 F2: -1005 Hz
 PPMCM: 5.5813 Hz/cm
 HZCM: 564.2051 Hz/cm



120

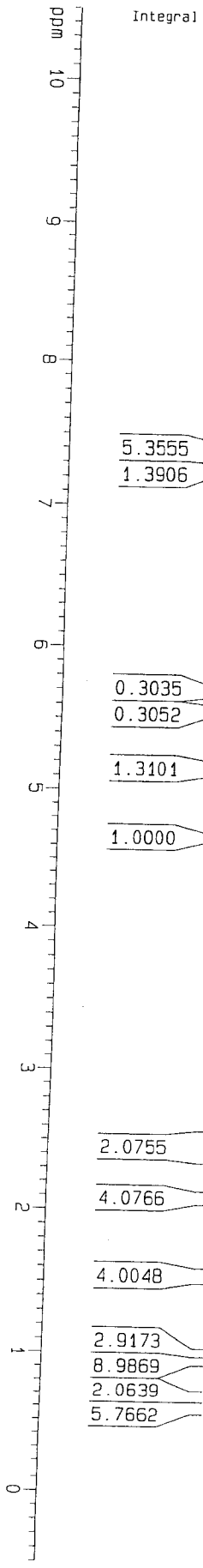
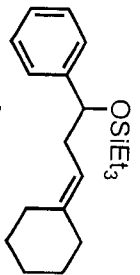
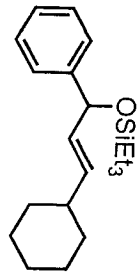
Current Data Parameters
 NAME: HCV9-B2BET
 EXPNO: 1
 PROCNO: 1

F2 - Acquisition Parameters
 Date_: 20080104
 Time: 17.00
 INSTRUM: spect
 P1P2: 5mm BBO BB-1
 PULPROG: zgpg30
 TD: 65536
 SFO1: 400.1324710 MHz
 SOLVENT: CDCl3
 NS: 2
 DS: 2
 SWH: 8278.146 Hz
 FIDRES: 0.126314 Hz
 AQ: 3.9584243 sec
 RG: 15
 DM: 60.400 usec
 DE: 5.00 usec
 TE: 300.0 K
 D1: 1.00000000 sec

***** CHANNEL f1 *****
 NUCL1: 13
 P1: 7.50 usec
 PL1: 0.00 dB
 SFO1: 400.1324710 MHz

F2 - Processing parameters
 SI: 32768
 SF: 400.130016 MHz
 MDW: EN
 SSB: 0
 LB: 0.30 Hz
 GB: 0
 PC: 1.00

1D NMR plot parameters
 CX: 4.00 cm
 F1P: 1.0000 DDM
 F1: 420.32 MHz
 F2P: -0.500 DDM
 F2: -200.07 Hz
 PPMCN: 0.45833 ppm/cm
 HZCN: 183.39291 Hz/cm



Current Data Parameters
 NAME ncis-85e2
 EXPNO 2
 PROCNO 1

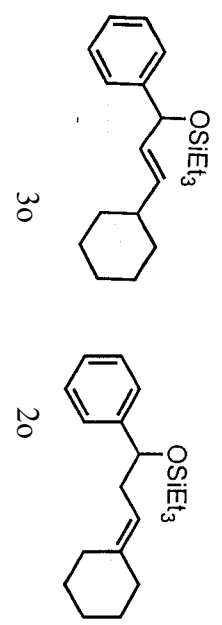
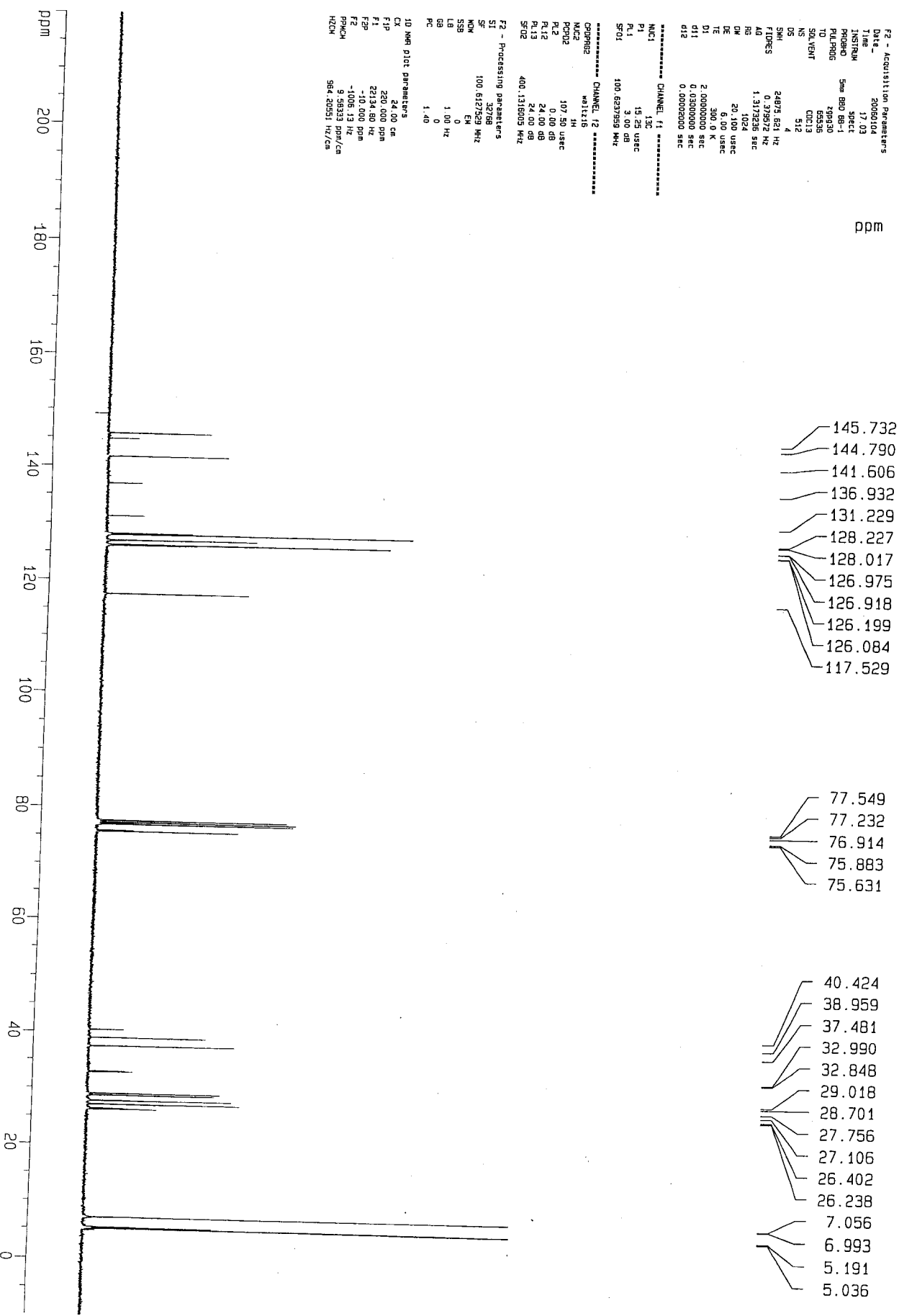
F2 - Acquisition Parameters
 Date_ 20080104
 Time 17.03
 INSTRUM spect
 PROBR0 5mm BBO BB-1
 PULPROG zgpg30
 CH1 299.930
 CH2 655.36
 SOLVENT CDCl3
 NS 512
 DS 4
 SWH 24875.621 Hz
 FIDRES 0.37857 Hz
 AQ 1.317235 sec
 RB 1024
 DE 20.100 usec
 TE 300.0 K
 D1 2.00000000 sec
 d12 0.03000000 sec
 012 0.00000000 sec

***** CHANNEL f1 *****
 NUCl1 13C
 P1 19.23 usec
 PL1 0.00 dB
 SFO1 100.627359 MHz

***** CHANNEL f2 *****
 CPDPRG2 waltz16
 MUC2 HI
 PCPD2 107.50 usec
 RL2 0.00 dB
 RL12 24.00 dB
 RL13 24.00 dB
 SF02 400.1318005 MHz

F2 - Processing parameters
 SI 88
 SF 100.617562 MHz
 KW 64
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

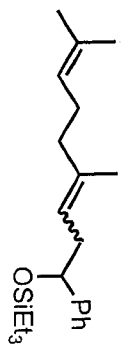
1D NMR plot parameters
 F1P 24.00 cm
 F1 250.000 ppm
 F2P 23134.50 Hz
 F2 100.000 ppm
 PPMCH -1005.1 Hz
 HZCH 9.408213 ppm/cm
 964.20551 Hz/cm



Wey-8-84 H-pdt

H-pdt

2p



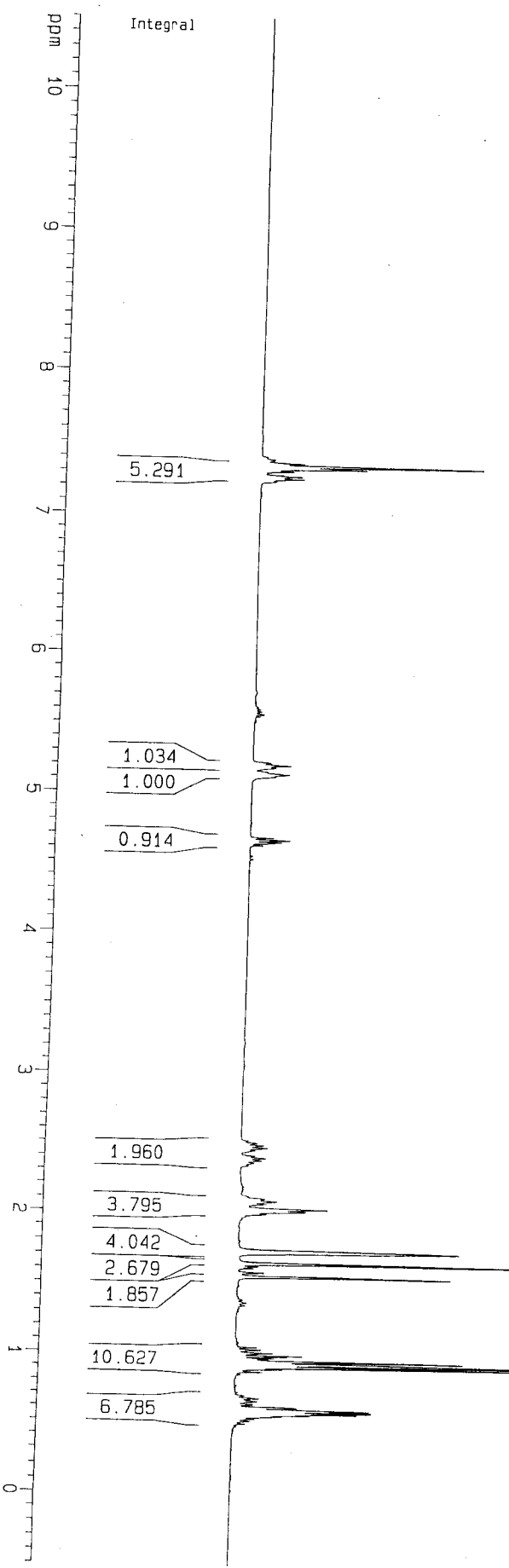
Current Data Parameters
 NAME hcy8-84-h-pdt
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060108
 Time 12:01
 INSTRUM spect
 PROBR4 Shim BBD BB-1
 PULPROG zgpg30
 TD 2930
 SOLVENT CDCl3
 NS 4
 DS 2
 SWH 8278.14 Hz
 FIDRES 0.12634 Hz
 AQ 3.3564243 sec
 RG 32
 DW 60.400 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec

***** CHANNEL f1 *****
 NUC1 1H
 P1 7.50 usec
 PL1 0.00 dB
 SFO1 400.1264710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300016 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

10 NMR Plot Parameters
 CX 24.00 cm
 F1P 14.640 Dm
 F1 4201.36 Hz
 F2P -0.400 Dm
 F2 -200.07 Hz
 PPMACC 0.45833 ppm/cm
 HZCM 183.39291 Hz/cm



Current Data Parameters
 NAME he9-844-pot
 EXNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060106
 Time 12:05
 INSTRUM spect
 PULPROG zgpg30
 TO 2.5536
 SOLVENT CDCl3
 NS 59
 DS 4
 SMH 24675.621 Hz
 FIDRES 0.379572 Hz
 AQ 1.3132356 sec
 RG 1024
 DM 20.100 usec
 DE 6.00 usec
 TE 300.0 K
 D1 0.0300000 sec
 d12 0.0002000 sec

***** CHANNEL f1 *****
 NUCl 13C
 P1 15.25 usec
 PL1 3.00 dB
 SF01 100.627959 MHz

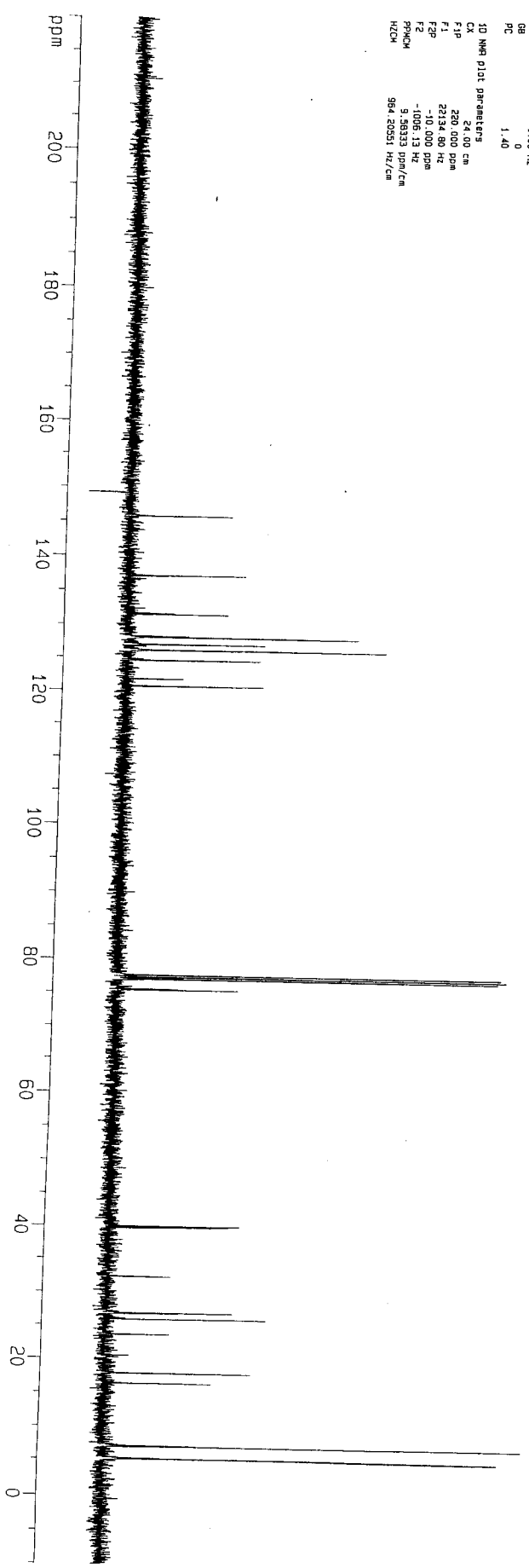
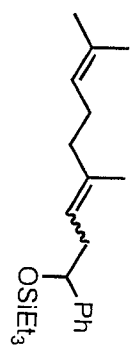
***** CHANNEL f2 *****
 GRPRG2 wa1216
 NUCl 1H
 PCPD2 107.50 usec
 P1 0.00 dB
 PL12 24.00 dB
 PL13 24.00 dB
 SF02 400.130003 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127529 MHz
 MDW 0
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR P104 parameters
 C 24.00 cm
 F10 220.000 ppm
 F1 22134.80 Hz
 F2 -100.000 ppm
 PPM/CM 9.86333 Hz/cm
 HZ/CM 984.20351 Hz/cm

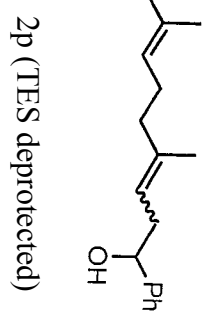
ppm

- 145.817
- 145.763
- 137.192
- 131.479
- 128.228
- 128.099
- 128.042
- 127.035
- 126.998
- 126.132
- 126.083
- 126.032
- 124.599
- 124.536
- 121.745
- 120.676
- 77.531
- 77.213
- 76.896
- 75.518
- 75.330
- 40.034
- 39.837
- 39.686
- 32.284
- 26.831
- 26.716
- 25.916
- 25.893
- 23.638
- 17.841
- 17.819
- 16.343
- 7.037
- 6.978
- 6.950
- 5.160
- 5.017
- 4.993



8-8-12

TES deprotection, H-pdt



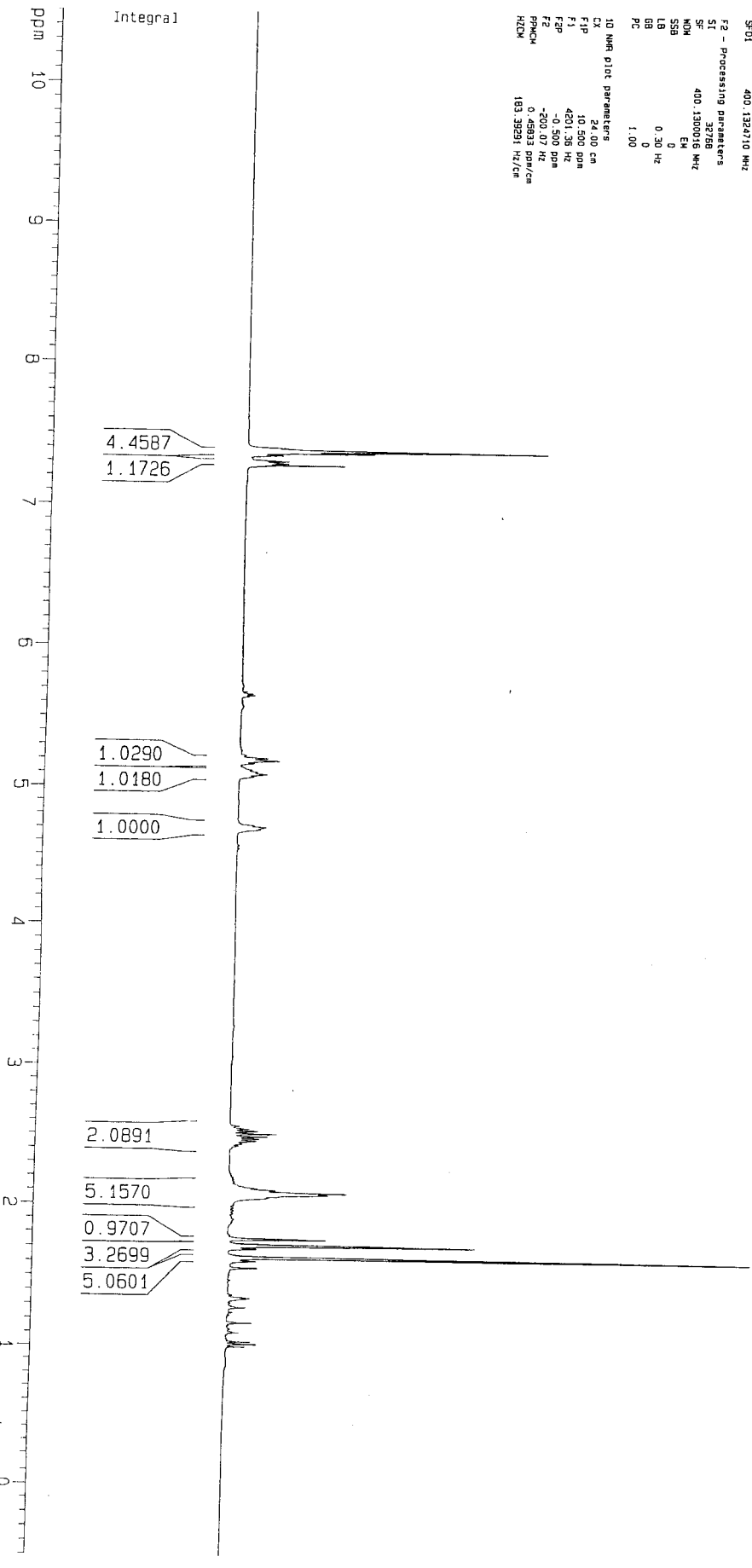
Current Data Parameters
 NAME ncy8-84de-p
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 2006117
 Time 13.39
 INSTRUM spect
 PROBRG 5mm BB1 Bb-1
 PULPROG zg30
 TO 65536
 SOLVENT CDCl3
 NS 3
 DS 2
 SWH 8278.146 Hz
 ZONES 2
 AQ 0.128314 Hz
 RG 3.594249 sec
 DW 59.73
 DE 6.00 usec
 TE 300.0 K
 D1 1.0000000 sec

***** CHANNEL f1 *****
 NUC1 H
 P1 7.90 usec
 PL1 0.00 dB
 SFO1 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.130041 MHz
 NHM 0
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

10 NMR plot parameters
 EX 24.00 cm
 F1P 10.500 ppm
 F1 4201.35 Hz
 F2 -0.500 ppm
 F2P 4200.07 Hz
 SFOCM 0.45853 ppm/cm
 HZCM 183.38291 Hz/cm



Citronellene, Me2AlCl

Current Data Parameters
 NAME 1078-121418
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20080217
 Time 12:26
 INSTRUM spect
 PROBRD 5 mm QNP 1H/1
 PULPROG zg30
 TO 65536
 SOLVENT CDCl3
 NS 8
 DS 1
 SWH 8278.146 Hz
 FIDRES 0.165314 Hz
 AQ 0.156214 sec
 RG 313929.5
 DW 60.400 usec
 DE 6.00 usec
 TE 299.4 K
 D1 1.0000000 sec
 MCREST 0.0000000 sec
 MCNKR 0.0150000 sec

***** CHANNEL f1 *****
 NUC1 1H
 P1 9.89 usec
 PL1 3.00 dB
 SFO1 400.1324710 MHz
 F2 - Processing parameters
 SF 400.1300654 MHz
 SI 32768
 KW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00
 10 MHz plot parameters
 CX 24.00 cm
 CY 7.58 cm
 FIP 10.500 ppm
 F1 4201.38 Hz
 F2 -0.500 ppm
 PRGCM 0.28107 Hz
 PRCM 0.28107 Hz/cm
 HZCM 183.39351 Hz/cm

