SUPPLEMENTAL INFORMATION

Supplemental Figure Legends

Figure S1. Phylogenetic analysis of the putative G α -binding motif found in Calnuc and NUCB2. Rat Calnuc and NUCB2 sequences were used to identify homologous protein sequences by BLAST-Protein search. The sequences corresponding to the G α -binding motif in the identified orthologues were aligned using CLUSTAL W. Conserved residues are shaded in black; similar residues in grey. The result of the alignment showed that Calnuc and NUCB2 G α -binding motif is evolutionarily conserved from invertebrates to humans.



Figure S1

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Figure S2. The interaction between Gai3 and Calnuc is stabilized by an electrostatic contact between Gai3 aa 248 and Calnuc aa 314. (A) K248 of Gai3 is predicted to be located in close proximity to E314 of Calnuc, suggesting that they might establish a molecular point of interaction by electrostatic attraction. The coordinates of Calnuc's putative Ga-binding sequence (aa 309-320, red) were extracted for the Protein Data Bank [PDB#: 1SNL] and threaded over the structure of KB-752 (not shown) in complex with Gai1 (blue) [PDB#: 1Y3A] as described in Fig.1. (B) Mutation of Calnuc's E314 to K restores the defective binding of Calnuc WT to Gai3 K248E. His-Calnuc Δ N wild-type (lane 4) but not His-Calnuc Δ N E314K (lane 5) binding to GST-Gai3 K248E·GDP (Gai3 KE) is dramatically impaired compared to wild-type GST-Gai3·GDP (Gai3 WT, lane 3). No binding of His-Calnuc Δ N WT (lane 2) or His-Calnuc Δ N E314K (lane 5) to GST is detected. 10 µg His-Calnuc Δ N WT (lanes 2-4) or His-Calnuc Δ N E314K (lanes 5-6) were incubated with purified GST (lanes 2 and 6), wild-type GST-Gai3 (lane 3) or GST-Gai3 K248B (lanes 4 and 5) pre-loaded with GDP immobilized on glutathione beads and analyzed as in Fig. **2A.** Input: 1 µg His-Calnuc Δ N WT (lane 1) or His-Calnuc Δ N E314K (lane 7).



Figure S2

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Figure S3. Mutation of W211 or F215 to A in Gai3 abolishes Calnuc binding (A) W211 and F215 in the α 3/ Swll cleft of Gai are predicted to mediate the hydrophobic interaction with L313, F316 and L317 in Calnuc's G α -binding motif. The coordinates of Calnuc's G α -binding sequence (aa 309-320, green) were extracted for the Protein Data Bank [PDB#: 1SNL] and threaded over the structure of KB-752 (not shown) in complex with Gai1 (blue) [PDB#: 1Y3A] as described in **Fig.1**. Gai's W211 and F215 are in close proximity to Calnuc's L313, F316 and L317. **(B)** His-Calnuc Δ N binding to GST-G α i3 W211A·GDP (lane 4) or GST-G α i3 F215A·GDP is virtually abolished compared to wild-type GST-G α i3·GDP (G α i3 WT, lane 3). No binding of His-Calnuc Δ N WT to GST is detected (lane 2). 10 µg His-Calnuc Δ N WT were incubated with purified GST (lanes 2), wild-type GST-G α i3 (lane 3), GST-G α i3 W211A (lanes 4) or GST-G α i3 F215A (lane 5) pre-loaded with GDP immobilized on glutathione beads and analyzed as in **Fig. 2A.** Input (lane 1): 1 µg His-Calnuc Δ N.



Figure S4. Mutation of F316 and F318 to A in Calnuc and NUCB2, respectively, impair activation of Gai3. (A) His-Calnuc Δ N increases the steady-state GTPase activity of His-Gai3 in a dose-dependent manner, and this effect is impaired (~70% reduction) by mutation of His-Calnuc ΔN 's F316 to A (F316A). The steady-state GTPase activity of purified His-G α i3 (50 nM) was determined in the presence of the indicated amounts (0, 6, 16, 30 and 60 µM) of purified wild-type His-Calnuc ΔN (closed circles) or His-Calnuc ΔN F316A mutant (open circles) by guantification of the amount of [y-³²P]GTP (0.5 µM, ~50 c.p.m/ fmol) hydrolyzed in 10 min. Data expressed as % of GTP hydrolyzed by the G protein alone (0 µM) were fitted to a non-linear, one-site hyperbola (solid lines) using Prism 4.0. Results are shown as mean ± S.D of one representative experiment out of three performed in duplicate. The milder effect of this mutant (~70% reduction) compared to the double mutant L313A/ L317A (~100% reduction, Fig 7A) is consistent with their respective binding to Gai3 (Fig 6A) (B) Mutation of F318 to A (F318A) abolishes GST-NUCB2 (173-333) activation of Gai3 steady-state GTPase hydrolysis. The steady-state GTPase activity of purified His-G α i3 (100 nM) was determined exactly as described in A but using 0, 5, 12.5, 25 and 37.5 µM NUCB2. The complete lack of effect of this mutant on the GTPase activity of $G\alpha i3$ is consistent with its lack of binding to the G protein (Fig 6B)



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Figure S5. Model illustrating the common and different features of the G α i interaction with Calnuc or GIV. (A) Alignment of the GBA motif from Calnuc, NUCB2, GIV and the synthetic KB-752 and GSP peptides was performed and analyzed and described in Fig 1A. The bar diagram below illustrates the highly conserved "core" of the motif (red) and the N and Cterminal region which are less conserved (blue). The stars (*) represent a non-conserved amino acid in the "core" (corresponding to Calnuc's E314) or several non-conserved amino acids in the C-terminal region; the w symbol denotes conserved hydrophobic residues within the motif "core". (B) Cartoon illustrating the proposed molecular contacts between Gαi and the GBA motif of Calnuc or GIV. Left (Calnuc): Hydrophobic residues of Gai's SwII (i.e., W211 and F215) interact with hydrophobic residues (conserved) in the "core" of Calnuc's GBA motif (Fig 6A and Fig S3). This interaction is further stabilized by an electrostatic interaction between Gαi3's K248 and Calnuc's E314 (Fig S2) which is not possible for GIV because this residue is not conserved in its GBA motif (position 4 of the consensus sequence shown in A). Right (GIV): Hydrophobic residues of Gai's SwII (i.e., W211 and F215) also interact with hydrophobic residues (conserved) in the "core" of GIV's GBA motif (1). Our published (2) and unpublished data suggest that Gai's W258 probably makes contact with GIV's residues S1689 and N1690, which is not possible for Calnuc (Fig 5B) because these residues in the C-terminal region the GBA motif are not conserved in Calnuc (3 aa downstream of the position 7 of the consensus sequence shown in A).



Figure S5

REFERENCES

- 1. Garcia-Marcos, M., Ghosh, P., and Farquhar, M. G. (2009) *Proc Natl Acad Sci U S A* **106**(9), 3178-3183
- 2. Garcia-Marcos, M., Ghosh, P., Ear, J., and Farquhar, M. G. (2010) *J Biol Chem* **285**(17), 12765-12777