

**Supplemental information for “Mechanical control of cation channels in the myogenic response”**

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## 1. TABLES OF FIXED PARAMETERS

Table S1: Fixed parameters used in vessel wall mechanics model

Fixed parameter	Units	Mesenteric arteriole*	Femoral arteriole*
$C_{pass}$	N/m	1.168	1.416
$C'_{pass}$	unitless	7.240	7.901
$D_{p100}$	$\mu\text{m}$	150.0	172.8
$C_{act}$	N/m	1.108	1.499
$C'_{act}$	unitless	0.843	0.742
$C''_{act}$	unitless	0.406	0.353
$\delta_{wall,ref}$	$\mu\text{m}$	6.16	8.98
$D_{ref}$	$\mu\text{m}$	151	171

\* Fixed parameters used here from optimized fits obtained in (8) to the data in Figure 1 of (6) for the normal control of Wistar-Kyoto rat mesenteric and femoral arterioles except for the reference values of wall thickness and diameter which are direct measurements made in original experimental study.

Table S2: Fixed parameters used VSM crossbridge model

Fixed parameter	Units	Value*
$K_2$	1/s	0.5
$K_3$	1/s	0.4
$K_4$	1/s	0.1
$K_5$	1/s	0.5
$K_7$	1/s	0.1
$(AM+AMp)_{max}$	unitless	0.8
$\gamma$	$\mu\text{M}^3/\text{s}$	17.0

\* All fixed parameter values are from (14)

Table S3: Fixed parameters used in VSM  $\text{Ca}^{2+}$  handling and electrophysiology model

Fixed parameter	Description	Units	Value	Ref
Standard constants and VSM cell properties				
$F$	Faraday constant	C/mole	96485.34	--
$R$	Gas constant	$(\text{C}^*\text{mV})/(\text{K}^*\text{mole})$	8314.472	--
$T$	Temperature	K	293	--
$N_{Av}$	Avogadro's number	1/mole	$6.022 \times 10^{23}$	--

$C_m$	Total membrane capacitance	pF	25	*
$A_m$	Total membrane area	cm <sup>2</sup>	$2.5 \times 10^{-5}$	*
$z_K$	Potassium valence	unitless	1	--
$z_{Na}$	Sodium valence	unitless	1	--
$z_{Ca}$	Calcium valence	unitless	2	--
$z_{Cl}$	Chloride valence	unitless	-1	--
$Ca_e$	External Ca <sup>2+</sup> concentration	mM	2.0	(30) <sup>#</sup>
$Na_e$	External Na <sup>+</sup> concentration	mM	140.0	(30) <sup>#</sup>
$K_e$	External K <sup>+</sup> concentration	mM	5.0	(30) <sup>#</sup>
$Cl_e$	External Cl <sup>-</sup> concentration	mM	129.0	*
$Vol_i$	Cytosolic volume	pL	1.0	(30) <sup>#</sup>
$Vol_{Ca}$	Cytosolic volume available to Ca <sup>2+</sup>	pL	0.7	(30) <sup>#</sup>
$Vol_{SRu}$	SR uptake volume	pL	0.07	(30) <sup>#</sup>
$Vol_{SRr}$	SR release volume	pL	0.007	(19) <sup>#</sup>
<b>L-Type voltage operated Ca<sup>2+</sup> channel (CaL)</b>				
$P_{VOCC}$	CaL whole cell permeability	cm/s	$1.88 \times 10^{-5}$	(16) <sup>§</sup>
<b>Large conductance, Ca<sup>2+</sup>-activated K<sup>+</sup> channel (BK<sub>Ca</sub>)</b>				
$N_{BKCa}$	BK <sub>Ca</sub> channel density	1/cm <sup>2</sup>	$6.6 \times 10^6$	(25) <sup>§</sup>
$\tau_{pf}$	BK <sub>Ca</sub> fast time constant	ms	0.84	(4) <sup>§</sup>
$\tau_{ps}$	BK <sub>Ca</sub> slow time constant	ms	35.9	(4) <sup>§</sup>
$dV_{50,KCaNO}$	BK <sub>Ca</sub> max NO-independent V <sub>50,KCa</sub> shift	mV	46.3	(20) <sup>§</sup>
$dV_{50,KCaGMP}$	BK <sub>Ca</sub> max cGMP-independent V <sub>50,KCa</sub> shift	mV	76	(26) <sup>§</sup>
$P_{BKCa}$	BK <sub>Ca</sub> single channel permeability	cm <sup>3</sup> /s	$3.9 \times 10^{-13}$	(20) <sup>#</sup>
<b>Voltage-dependent K<sup>+</sup> channel (K<sub>v</sub>)</b>				
$g_{Kv}$	K <sub>v</sub> maximal conductance	nS	1.35	(17) <sup>§</sup>
$\tau_{q1}$	K <sub>v</sub> fast inactivation time constant	ms	371	(17) <sup>§</sup>
$\tau_{q2}$	K <sub>v</sub> slow inactivation time constant	ms	2884	(17) <sup>§</sup>
<b>Unspecified K leak channel (K<sub>leak</sub>) including ATP sensitive K<sup>+</sup> channel</b>				
$g_{Kleak}$	K <sub>leak</sub> maximal conductance	nS	0.067	(22) <sup>§</sup>
<b>Non-selective cation channel (NSC)</b>				
$K_{NSC}$	NSC ½ max activation DAG concentration	mM	<b><math>3 \times 10^{-3}</math></b>	(3) <sup>§</sup>
$d_{NSC}^{\min}$	NSC minimum constituent activation	unitless	0.0244	(11) <sup>§</sup>
$P_{NaNSC}$	NSC Na <sup>+</sup> whole cell permeability	cm/s	$5.11 \times 10^{-7}$	(11) <sup>§</sup>
$P_{KNSC}$	NSC K <sup>+</sup> whole cell permeability	cm/s	$5.42 \times 10^{-7}$	(11) <sup>§</sup>

$P_{CaNSC}$	NSC $Ca^{2+}$ whole cell permeability	cm/s	$2.32 \times 10^{-6}$	(11) <sup>§</sup>
Store-operated non-selective cation channel (SOC)				
$g_{CaSOC}$	SOC maximal $Ca^{2+}$ conductance	nS	0.0083	(1) <sup>§</sup>
$g_{NaSOC}$	SOC maximal $Na^{+}$ conductance	nS	0.0575	(2) <sup>§</sup>
$K_{SOC}$	SOC $\frac{1}{2}$ max activation $Ca^{2+}$ concentration	mM	$1 \times 10^{-4}$	(24) <sup>§</sup>
$\tau_{SOC}$	<b>SOC activation time constant</b>	<b>ms</b>	<b>100</b>	*
$Ca^{2+}$ -activated $Cl^{-}$ channel ( $Cl_{Ca}$ )				
$g_{ClCa}$	$Cl_{Ca}$ maximal conductance density	nS/pF	0.23	(18) <sup>§</sup>
$R_{ClcGMPmin}$	$Cl_{Ca}$ cGMP-indep open probability coeff	unitless	0.0132	(18) <sup>§</sup>
$n_{ClCa}$	$Cl_{Ca}$ $Ca^{2+}$ Hill coefficient	unitless	2	(18) <sup>§</sup>
$K_{ClCa}$	$Cl_{Ca}$ $\frac{1}{2}$ max activation $Ca^{2+}$ concentration	mM	$3.65 \times 10^{-4}$	*
$n_{ClcGMP}$	$Cl_{Ca}$ cGMP Hill coefficient	unitless	3.3	(18) <sup>#</sup>
$K_{ClcGMP}$	$Cl_{Ca}$ $\frac{1}{2}$ max activation cGMP concentration	mM	$6.4 \times 10^{-3}$	(18) <sup>#</sup>
Plasma membrane $Ca^{2+}$ -ATPase (PMCA)				
$\bar{I}_{PMCA}$	PMCA maximal current	pA	5.37	*
$K_{mPMCA}$	PMCA Michaelis constant	mM	$1.70 \times 10^{-4}$	(23) <sup>#</sup>
Plasma membrane $Na^{+}$ - $Ca^{2+}$ exchanger (NCX)				
$g_{NCX}$	NCX maximal current	<b>pA</b>	<b>0.000487</b>	*
$d_{NCX}$	NCX scaling factor	unitless	0.0003	(15) <sup>#</sup>
$\gamma_{NCX}$	NCX non-symm V-dep partition parameter	unitless	0.45	(15) <sup>#</sup>
$Na^{+}$ - $K^{+}$ pump (NaK)				
$\bar{I}_{NaK}$	NaK maximal current	pA	2.3083	*
$n_{HKe}$	NaK external $K^{+}$ Hill coefficient	unitless	1.1	(21) <sup>#</sup>
$K_{dKe}$	NaK external $K^{+}$ binding constant	mM	1.6	(21) <sup>#</sup>
$n_{HNai}$	NaK internal $Na^{+}$ Hill coefficient	unitless	1.7	(21) <sup>#</sup>
$K_{dNai}$	NaK internal $Na^{+}$ binding constant	mM	22	(21) <sup>#</sup>
$Q_{10}$	NaK temperature correction	unitless	1.87	*
$Na^{+}$ - $K^{+}$ - $Cl^{-}$ cotransporter (NaKCl)				
$L_{NaKCl}$	NaKCl coefficient	<b><math>\text{mole}^2 \times 10^{-5} / (\text{J}^* \text{s} * \text{m}^2)</math></b>	$1.79 \times 10^{-8}$	*
Sarcoplasmic reticulum $IP_3$ receptor (SRIP <sub>3</sub> )				
$\bar{I}_{IP_3}$	SRIP <sub>3</sub> maximal $Ca^{2+}$ release	<b>1/ms</b>	$2880 \times 10^{-6}$	(10) <sup>#</sup>
$K_{IP_3}$	SRIP <sub>3</sub> dissociation constant	mM	$1.20 \times 10^{-4}$	(5) <sup>#</sup>
$K_{IP_3}^{act}$	SRIP <sub>3</sub> dissociation constant $Ca^{2+}$ activation	mM	$1.70 \times 10^{-4}$	(10) <sup>#</sup>

$k_{IP_3}^{on}$	SRIP <sub>3</sub> Ca <sup>2+</sup> binding rate to inhibition site	1/(ms*mM)	1.4	(5) <sup>#</sup>
$K_{IP_3}^{inh}$	SRIP <sub>3</sub> dissociation constant Ca <sup>2+</sup> inhibition	mM	1x10 <sup>-4</sup>	(10) <sup>#</sup>
Sarcoplasmic reticulum SERCA, uptake, release and internal diffusion				
$\bar{I}_{SERCA}$	SERCA maximal current	pA	6.68	(30) <sup>#</sup>
$K_{mup}$	SERCA Michaelis constant	mM	1x10 <sup>-3</sup>	(19) <sup>#</sup>
$\tau_r$	SR internal diffusion time constant	ms	1000	(30) <sup>#</sup>
$R_{leak}$	SR Ca <sup>2+</sup> leak parameter	unitless	1.07x10 <sup>-5</sup>	(7) <sup>§</sup>
$\tau_{rel}$	SR Ca <sup>2+</sup> release time constant	ms	0.0333	(30) <sup>#</sup>
$\overline{CSQN}$	SR average calsequestrin concentration	mM	15	(12) <sup>#</sup>
$K_{CSQN}$	SR calsequestrin binding constant	mM	0.8	(28) <sup>#</sup>
Sarcoplasmic reticulum four-state ryanodine receptor (RyR)				
$K_{r1}$	RyR Ca <sup>2+</sup> binding to activation site	1/(ms*mM <sup>2</sup> )	2500	(30) <sup>#</sup>
$K_{r2}$	RyR Ca <sup>2+</sup> binding to inactivation site	1/(ms*mM)	1.05	(30) <sup>#</sup>
$K_{mr1}$	RyR Ca <sup>2+</sup> dissociation from activation site	1/ms	0.0076	(30) <sup>#</sup>
$K_{mr2}$	RyR Ca <sup>2+</sup> dissociation from inactivation site	1/ms	0.084	(30) <sup>#</sup>
$\alpha_1$ -adrenoreceptor activation and IP <sub>3</sub> formation				
$k_{rG}$	Receptor recycling rate	1/ms	1.75x10 <sup>-7</sup>	(5) <sup>#</sup>
$\xi_G$	Mobile receptor fraction	unitless	0.85	(5) <sup>#</sup>
$R_{TG}$	Total number of receptors	# receptors	2x10 <sup>4</sup>	(5) <sup>#</sup>
$k_{pG}$	Receptor phosphorylation rate	1/ms	0	*
$K_{IG}$	Unphosphorylated receptor dissoc constant	mM	0.01	(5) <sup>#</sup>
$k_{eG}$	Receptor endocytosis rate	1/ms	6x10 <sup>-6</sup>	(5) <sup>#</sup>
$K_{2G}$	Phosphorylated receptor dissoc constant	mM	0.2	(5) <sup>#</sup>
$k_{aG}$	G-protein activation rate	1/ms	0.17x10 <sup>-3</sup>	(5) <sup>#</sup>
$G_{TG}$	Total # of G-protein molecules	# molecules	1x10 <sup>5</sup>	(5) <sup>#</sup>
$k_{dG}$	G-protein deactivation rate	1/ms	1.5x10 <sup>-3</sup>	(5) <sup>#</sup>
$\alpha_G$	Effective signal gain parameter	1/ms	2.781x10 <sup>-8</sup>	(5) <sup>#</sup>
$K_{cG}$	Ca <sup>2+</sup> -PLC dissociation constant	mM	0.4x10 <sup>-3</sup>	(5) <sup>#</sup>
$k_{degG}$	IP <sub>3</sub> degradation rate	1/ms	1.25x10 <sup>-3</sup>	(5) <sup>#</sup>
$r_{rG}$	PIP <sub>2</sub> replenishment rate	1/ms	0.015x10 <sup>-3</sup>	(5) <sup>#</sup>
$PIP_{2T}$	Total # of PIP <sub>2</sub> molecules	# molecules	5x10 <sup>7</sup>	(5) <sup>#</sup>
sGC activation and cGMP formation				
$V_{cGMPmax}$	Maximal cGMP formation rate	mM/ms	1.26x10 <sup>-7</sup>	(9) <sup>§</sup>

$k_{2sGC}$	sGC fully act E <sub>3</sub> NO-indep rate of formation	1/ms	0.64x10 <sup>-5</sup>	(27) <sup>#</sup>
$k_{3sGC}$	sGC fully act E <sub>3</sub> NO-dep rate of formation	1/(ms*mM)	4.2	(27) <sup>#</sup>
$k_{m1sGC}$	sGC partially act E <sub>2</sub> rate of dissociation	1/ms	15x10 <sup>-3</sup>	(27) <sup>#</sup>
$k_{DsGC}$	sGC fully act E <sub>3</sub> NO-dep rate of dissoc	1/ms	0.4x10 <sup>-3</sup>	(27) <sup>#</sup>
$k_{m2sGC}$	sGC fully act E <sub>3</sub> NO-indep rate of dissoc	1/ms	0.1x10 <sup>-6</sup>	(27) <sup>#</sup>
$k_{1sGC}$	sGC partially act E <sub>2</sub> rate of formation	1/(ms*mM)	2x10 <sup>3</sup>	(27) <sup>#</sup>
$k_{DrsGC}$	cGMP inverse of max $\tau$ of formation rate	1/ms	0.1x10 <sup>-3</sup>	*
$k_{pdecGMP}$	cGMP PDE catalyzed hydrolysis rate	1/ms	0.0695x10 <sup>-3</sup>	(31) <sup>#</sup>
$K_{mpde}$	cGMP PDE catalysis Michaelis constant	mM	1x10 <sup>-3</sup>	*
<hr/>				
Cytosolic Ca <sup>2+</sup> buffering				
$\bar{S}_{CM}$	Calmodulin concentration	mM	0.1	(30) <sup>#</sup>
$K_{dCM}$	Calmodulin-Ca <sup>2+</sup> dissociation constant	mM	2.60x10 <sup>-4</sup>	(30) <sup>#</sup>
$\bar{B}_F$	Generic Ca <sup>2+</sup> buffer concentration	mM	0.1	(10, 29) <sup>§</sup>
$K_{dB}$	Generic Ca <sup>2+</sup> buffer dissociation constant	mM	5.298x10 <sup>-4</sup>	(10, 29) <sup>§</sup>

\* Parameter value assumed or estimated in Kapela et al. (13)

# Parameter value determined directly in paper

§ Parameter value determined from model fits to or analysis of data presented in paper

Parameters, units and values in **bold** indicate corrections and omissions from that originally published in Kapela et al. (13).

## 2. COMPLETE SET OF INTEGRATED MODEL EQUATIONS

### Model 1: Vessel wall mechanics model

The set of equations given below can be used to implicitly solve for a vessel diameter,  $D$ , as a function of the intraluminal pressure,  $P$ , and the activation,  $Act$ , as determined from running Model 2 and Model 3 to steady-state solutions. If these equations are to solve for myogenic or maximally active curves root finding is more easily performed in the stress-strain space rather than the pressure-diameter space due to hysteresis at high intraluminal pressure that often leads to multiple diameters at a given pressure but less often gives multiple strains for a given stress.

Algebraic equations:

$$\delta_{wall} = -\frac{D}{2} + \sqrt{\left(\frac{D}{2}\right)^2 + \frac{CSA_{ref}}{\pi}} \quad [S1]$$

$$\sigma_{total} = \frac{PD}{2\delta_{wall}} \quad [S2]$$

$$\sigma_{pass} = \frac{C_{pass}}{\delta_{wall}} \exp\left[C'_{pass}\left(\frac{D}{D_{p100}} - 1\right)\right] \quad [S3]$$

$$\sigma_{act}^{max} = \frac{C_{act}}{\delta_{wall}} \exp\left[-\left(\frac{D/D_{p100} - C'_{act}}{C''_{act}}\right)^2\right] \quad [S4]$$

$$\sigma_{total} = \sigma_{pass} + Act \sigma_{act}^{max} \quad [S5]$$

Calculation of remaining fixed parameter:

$$CSA_{ref} = \pi\delta_{wall,ref}\left(D_{ref} + \delta_{wall,ref}\right) \quad [S6]$$

### Model 2: VSM crossbridge force generation model

A dynamic VSM crossbridge model is used to determine the steady state VSM activation,  $Act$ , as a function of cytosolic  $Ca^{2+}$ ,  $Ca_i$ . The system of differential equations is run to steady state for a range of  $Ca_i$  and then the Hill expression given below is used to fit the simulation results. This is done to prevent a steady-state simulation to be run repeatedly in the integrated model and at this point we are not interested in the dynamic response of the system.

Algebraic and differential equations:

$$Act = \frac{AMp + AM}{(AMp + AM)_{max}} \quad [S7]$$

$$K_1 = K_6 = \gamma Ca_i^3 \quad [S8]$$

$$\frac{dM}{dt} = -K_1M + K_2Mp + K_7AM \quad [S9]$$

$$\frac{dMp}{dt} = K_4AMp + K_1M - (K_2 + K_3)Mp \quad [S10]$$

$$\frac{dAMp}{dt} = K_3Mp + K_6AM - (K_4 + K_5)AMp \quad [S11]$$

$$\frac{dAM}{dt} = K_5AMp - (K_6 + K_7)AM \quad [S12]$$

Setting of initial conditions:

$$M = Mp = AMp = AM = 0.25 \quad [S13]$$

Hill expression for fit of steady-state VSM activation:

$$Act = \frac{Ca_i^{n_{XB}}}{K_{CaXB}^{n_{XB}} + Ca_i^{n_{XB}}} \quad [S14]$$

### **Model 3: VSM Ca<sup>2+</sup> handling and electrophysiology model**

Kapela et al. have made several clarifications and modifications to the original published model that are incorporated into this current model. These were determined from their model code and communications with the authors. For sGC activation, the time constant for the change in rate of cGMP formation,  $\tau_{sGC}$ , is determined differently as indicated below. Additionally the activation gating for the store-operated cation channel has been changed from a simple algebraic relationship to the first-order kinetics given below. Corrections have been made to fixed parameter values for  $K_{NSC}$  and  $g_{NCX}$  and units on  $g_{NCX}$ ,  $L_{NaKCl}$  and  $\bar{I}_{IP_3}$  as indicated in Table S3. A minimal set of initial conditions is specified and then the remaining initial conditions are calculated as given below. For the  $\alpha_1$ -adrenoreceptor and IP<sub>3</sub> formation equations it was not clear whether the ratio of activities of the ligand-bound to ligand-unbound receptor species,  $\delta_G$ , is a variable or fixed parameter. In this model  $\delta_G$  (here given as  $\delta_{G0}$ ) is estimated based on the initial conditions and held constant throughout the simulation as specified below. Other variables such as the rate of hydrolysis of PIP<sub>2</sub>,  $r_{hG}$ , are initially estimated and then recalculated during the simulation. Standard units of concentration, time, membrane potential, vessel stress and current are mM, ms, mV, kPa and pA in this portion of the integrated model. Note that for the myogenic response both the NE stimulation and NO relaxation mechanisms are inactive and vessel diameter is driven only by step changes in pressure. The model retains these inactive pathways to remain consistent with the previous model by Kapela et al. In the simulations used in this study a single stress- or strain-controlled channel was inserted into the model at a time. The sets of equations below show all formulations of stress- and strain-controlled channels used.

Algebraic and differential equations:

*Reversal potentials:*

$$E_K = \frac{RT}{z_K F} \ln \left( \frac{K_e}{K_i} \right) \quad [S15]$$

$$E_{Na} = \frac{RT}{z_{Na} F} \ln \left( \frac{Na_e}{Na_i} \right) \quad [S16]$$

$$E_{Ca} = \frac{RT}{z_{Ca} F} \ln \left( \frac{Ca_e}{Ca_i} \right) \quad [S17]$$



$$E_{Cl} = \frac{RT}{z_{Cl}F} \ln \left( \frac{Cl_e}{Cl_i} \right) \quad [S18]$$

*Mechanically-controlled supplementary Ca<sup>2+</sup> channel:*

Formulated as a stress-controlled channel (channel variant 4):

$$\bar{h}_{Ca\sigma CC} = \frac{\sigma_{total}^{n_{Ca\sigma CC}}}{\sigma_{50, Ca\sigma CC}^{n_{Ca\sigma CC}} + \sigma_{total}^{n_{Ca\sigma CC}}} \quad [S19]$$

$$I_{Ca\sigma CC} = g_{Ca\sigma CC}^{\max} h_{Ca\sigma CC} (V_m - E_{Ca}) \quad [S20]$$

$$\frac{dh_{Ca\sigma CC}}{dt} = \frac{\bar{h}_{Ca\sigma CC} - h_{Ca\sigma CC}}{\tau_{Ca\sigma CC}} \quad [S21]$$

Formulated as a strain-controlled channel:

$$\bar{h}_{Ca\varepsilon CC} = \frac{\varepsilon_{total}^{n_{Ca\varepsilon CC}}}{\varepsilon_{50, Ca\varepsilon CC}^{n_{Ca\varepsilon CC}} + \varepsilon_{total}^{n_{Ca\varepsilon CC}}} \quad [S22]$$

$$I_{Ca\varepsilon CC} = g_{Ca\varepsilon CC}^{\max} h_{Ca\varepsilon CC} (V_m - E_{Ca}) \quad [S23]$$

$$\frac{dh_{Ca\varepsilon CC}}{dt} = \frac{\bar{h}_{Ca\varepsilon CC} - h_{Ca\varepsilon CC}}{\tau_{Ca\varepsilon CC}} \quad [S24]$$

*Mechanically-controlled supplementary Na<sup>+</sup> channel:*

Formulated as a stress-controlled channel (channel variant 5):

$$\bar{h}_{Na\sigma CC} = \frac{\sigma_{total}^{n_{Na\sigma CC}}}{\sigma_{50, Na\sigma CC}^{n_{Na\sigma CC}} + \sigma_{total}^{n_{Na\sigma CC}}} \quad [S25]$$

$$I_{Na\sigma CC} = g_{Na\sigma CC}^{\max} h_{Na\sigma CC} (V_m - E_{Na}) \quad [S26]$$

$$\frac{dh_{Na\sigma CC}}{dt} = \frac{\bar{h}_{Na\sigma CC} - h_{Na\sigma CC}}{\tau_{Na\sigma CC}} \quad [S27]$$

Formulated as a strain-controlled channel:

$$\bar{h}_{Na\varepsilon CC} = \frac{\varepsilon_{total}^{n_{Na\varepsilon CC}}}{\varepsilon_{50, Na\varepsilon CC}^{n_{Na\varepsilon CC}} + \varepsilon_{total}^{n_{Na\varepsilon CC}}} \quad [S28]$$

$$I_{Na\varepsilon CC} = g_{Na\varepsilon CC}^{\max} h_{Na\varepsilon CC} (V_m - E_{Na}) \quad [S29]$$

$$\frac{dh_{Na\varepsilon CC}}{dt} = \frac{\bar{h}_{Na\varepsilon CC} - h_{Na\varepsilon CC}}{\tau_{Na\varepsilon CC}} \quad [S30]$$

*L-Type voltage operated Ca<sup>2+</sup> channel:*

Formulation from Kapela et al.:

$$\bar{d}_L = \frac{1}{1 + \exp(-V_m/8.3)} \quad [S31]$$

$$\bar{f}_L = \frac{1}{1 + \exp[(V_m + 42)/9.1]} \quad [S32]$$

$$\tau_{dL} = 2.5 \exp\left[-\left(\frac{V_m + 40}{30}\right)^2\right] + 1.15 \quad [S33]$$

$$\tau_{fL} = 65 \exp\left[-\left(\frac{V_m + 35}{25}\right)^2\right] + 45 \quad [S34]$$

$$I_{VOCC} = 10^6 A_m P_{VOCC} d_L f_L \frac{V_m (z_{Ca} F)^2}{RT} \frac{Ca_e - Ca_i \exp(V_m z_{Ca} F / RT)}{1 - \exp(V_m z_{Ca} F / RT)} \quad [S35]$$

$$\frac{dd_L}{dt} = \frac{\bar{d}_L - d_L}{\tau_{dL}} \quad [S36]$$

$$\frac{df_L}{dt} = \frac{\bar{f}_L - f_L}{\tau_{fL}} \quad [S37]$$

Formulated as a channel with vessel stress-control of gating voltage (channel variant 6), which is same as Kapela et al. formulation except the following three equations now replace [S31] and [S32]:

$$V_{CaL\sigma CC}^{off} = V_{CaL\sigma CC}^{offmax} \frac{\sigma_{total}^{n_{CaL\sigma CC}}}{\sigma_{50, CaL\sigma CC}^{n_{CaL\sigma CC}} + \sigma_{total}^{n_{CaL\sigma CC}}} \quad [S38]$$

$$\bar{d}_L = \frac{1}{1 + \exp\left[-(V_m + V_{CaL\sigma CC}^{off})/8.3\right]} \quad [S39]$$

$$\bar{f}_L = \frac{1}{1 + \exp\left[(V_m + V_{CaL\sigma CC}^{off} + 42)/9.1\right]} \quad [S40]$$

Formulated as a channel with vessel strain-control of gating voltage, which is the same as Kapela et al. formulation except the following three equations now replace [S31] and [S32]:

$$V_{CaL\varepsilon CC}^{off} = V_{CaL\varepsilon CC}^{offmax} \frac{\varepsilon_{total}^{n_{CaL\varepsilon CC}}}{\varepsilon_{50, CaL\varepsilon CC}^{n_{CaL\varepsilon CC}} + \varepsilon_{total}^{n_{CaL\varepsilon CC}}} \quad [S41]$$

$$\bar{d}_L = \frac{1}{1 + \exp\left[-(V_m + V_{CaL\varepsilon CC}^{off})/8.3\right]} \quad [S42]$$

$$\bar{f}_L = \frac{1}{1 + \exp\left[(V_m + V_{CaL\varepsilon CC}^{off} + 42)/9.1\right]} \quad [S43]$$

Formulated as a channel with vessel stress-control of conductance (channel variant 7), which is the same as Kapela et al. formulation except for the following 2 equations now replace [S35]:

$$\rho_{CaL\sigma CC} = 1 + (\rho_{\rho CaL\sigma CC}^{\max} - 1) \frac{\sigma_{total}^{n_{\rho CaL\sigma CC}}}{\sigma_{50, \rho CaL\sigma CC}^{n_{\rho CaL\sigma CC}} + \sigma_{total}^{n_{\rho CaL\sigma CC}}} \quad [S44]$$

$$I_{VOCC} = 10^6 A_m P_{VOCC} \rho_{CaL\sigma CC} d_L f_L \frac{V_m (z_{Ca} F)^2}{RT} \frac{Ca_e - Ca_i \exp(V_m z_{Ca} F / RT)}{1 - \exp(V_m z_{Ca} F / RT)} \quad [S45]$$

Formulated as a channel with vessel strain-control of conductance, which is the same as Kapela et al. formulation except for the following 2 equations now replace [S35]:

$$\rho_{CaL\varepsilon CC} = 1 + (\rho_{\rho CaL\varepsilon CC}^{\max} - 1) \frac{\varepsilon_{total}^{n_{\rho CaL\varepsilon CC}}}{\varepsilon_{50, \rho CaL\varepsilon CC}^{n_{\rho CaL\varepsilon CC}} + \varepsilon_{total}^{n_{\rho CaL\varepsilon CC}}} \quad [S46]$$

$$I_{VOCC} = 10^6 A_m P_{VOCC} \rho_{CaL\varepsilon CC} d_L f_L \frac{V_m (z_{Ca} F)^2}{RT} \frac{Ca_e - Ca_i \exp(V_m z_{Ca} F / RT)}{1 - \exp(V_m z_{Ca} F / RT)} \quad [S47]$$

*Large conductance  $Ca^{2+}$ -activated  $K^+$  channel:*

Formulation from Kapela et al.:

$$P_{KCa} = 0.17 p_f + 0.83 p_s \quad [S48]$$

$$R_{NO} = \frac{NO}{NO + 0.2 \times 10^{-3}} \quad [S49]$$

$$R_{cGMP} = \frac{cGMP^2}{cGMP^2 + (1.5 \times 10^{-3})^2} \quad [S50]$$

$$V_{50, KCa} = -41.7 \log(Ca_i) - 128.2 - (dV_{50, KCaNO} R_{NO}) - (dV_{50, KCa cGMP} R_{cGMP}) \quad [S51]$$

$$\bar{p}_o = \frac{1}{1 + \exp\left[-(V_m - V_{50, KCa})/18.25\right]} \quad [S52]$$

$$I_{BKCa} = 10^6 A_m N_{BKCa} P_{BKCa} P_{KCa} \frac{V_m (z_K F)^2}{RT} \frac{K_e - K_i \exp(V_m z_K F / RT)}{1 - \exp(V_m z_K F / RT)} \quad [S53]$$

$$\frac{dp_f}{dt} = \frac{\bar{p}_o - p_f}{\tau_{pf}} \quad [S54]$$

$$\frac{dp_s}{dt} = \frac{\bar{p}_o - p_s}{\tau_{ps}} \quad [S55]$$

Formulated as a channel with vessel stress-control of gating voltage (channel variant 8), which is same as Kapela et al. formulation except the following two equations now replace [S52]:

$$V_{BKCa\sigma CC}^{\text{off}} = V_{BKCa\sigma CC}^{\text{offmax}} \frac{\sigma_{total}^{n_{BKCa\sigma CC}}}{\sigma_{50, BKCa\sigma CC}^{n_{BKCa\sigma CC}} + \sigma_{total}^{n_{BKCa\sigma CC}}} \quad [S56]$$

$$\bar{p}_o = \frac{1}{1 + \exp\left[-(V_m + V_{BKCa\sigma CC}^{\text{off}} - V_{50, KCa})/18.25\right]} \quad [S57]$$

Formulated as a channel with vessel strain-control of gating voltage, which is same as Kapela et al. formulation except the following two equations now replace [S52]:

$$V_{BKCa\varepsilon CC}^{\text{off}} = V_{BKCa\varepsilon CC}^{\text{offmax}} \frac{\varepsilon_{\text{total}}^{n_{BKCa\varepsilon CC}}}{\varepsilon_{50, BKCa\varepsilon CC}^{n_{BKCa\varepsilon CC}} + \varepsilon_{\text{total}}^{n_{BKCa\varepsilon CC}}} \quad [\text{S58}]$$

$$\bar{p}_o = \frac{1}{1 + \exp\left[-(V_m + V_{BKCa\varepsilon CC}^{\text{off}} - V_{50, KCa})/18.25\right]} \quad [\text{S59}]$$

Formulated as a channel with vessel stress-control of conductance (channel variant 9), which is same as Kapela et al. formulation except the following two equations now replace [S53]:

$$\rho_{BKCa\sigma CC} = 1 - (1 - \rho_{\rho_{BKCa\sigma CC}}^{\text{max}}) \frac{\sigma_{\text{total}}^{n_{\rho_{BKCa\sigma CC}}}}{\sigma_{50, \rho_{BKCa\sigma CC}}^{n_{\rho_{BKCa\sigma CC}}} + \sigma_{\text{total}}^{n_{\rho_{BKCa\sigma CC}}}} \quad [\text{S60}]$$

$$I_{BKCa} = 10^6 A_m N_{BKCa} P_{BKCa} \rho_{BKCa\sigma CC} P_{KCa} \frac{V_m (z_K F)^2}{RT} \frac{K_e - K_i \exp(V_m z_K F / RT)}{1 - \exp(V_m z_K F / RT)} \quad [\text{S61}]$$

Formulated as a channel with vessel strain-control of gating voltage, which is same as Kapela et al. formulation except the following two equations now replace [S53]:

$$\rho_{BKCa\varepsilon CC} = 1 - (\rho_{\rho_{BKCa\varepsilon CC}}^{\text{max}} - 1) \frac{\varepsilon_{\text{total}}^{n_{\rho_{BKCa\varepsilon CC}}}}{\varepsilon_{50, \rho_{BKCa\varepsilon CC}}^{n_{\rho_{BKCa\varepsilon CC}}} + \varepsilon_{\text{total}}^{n_{\rho_{BKCa\varepsilon CC}}}} \quad [\text{S62}]$$

$$I_{BKCa} = 10^6 A_m N_{BKCa} P_{BKCa} \rho_{BKCa\varepsilon CC} P_{KCa} \frac{V_m (z_K F)^2}{RT} \frac{K_e - K_i \exp(V_m z_K F / RT)}{1 - \exp(V_m z_K F / RT)} \quad [\text{S63}]$$

*Voltage dependent K<sup>+</sup> channel:*

$$\bar{p}_K = \frac{1}{1 + \exp\left[-(V_m + 11)/15\right]} \quad [\text{S64}]$$

$$\bar{q} = \frac{1}{1 + \exp\left[(V_m + 40)/14\right]} \quad [\text{S65}]$$

$$\tau_{pK} = 61.49 \exp(-0.0268 V_m) \quad [\text{S66}]$$

$$I_{Kv} = g_{Kv} p_k (0.45 q_1 + 0.55 q_2) (V_m - E_K) \quad [\text{S67}]$$

$$\frac{dp_K}{dt} = \frac{\bar{p}_K - p_K}{\tau_{pK}} \quad [\text{S68}]$$

$$\frac{dq_1}{dt} = \frac{\bar{q} - q_1}{\tau_{q1}} \quad [\text{S69}]$$

$$\frac{dq_2}{dt} = \frac{\bar{q} - q_2}{\tau_{q2}} \quad [\text{S70}]$$

*Unspecified K<sup>+</sup> leak channel including ATP-sensitive K<sup>+</sup> channel:*

$$I_{Kleak} = g_{Kleak} (V_m - E_k) \quad [S71]$$

*Non-selective cation channel:*

Formulation from Kapela et al.:

$$DAG = IP_3 \quad [S72]$$

$$Po_{NSC} = 0.4344 + \frac{0.5656}{1 + \exp[-(V_m - 47.12)/24.24]} \quad [S73]$$

$$I_{NaNSC} = 10^6 A_m P_{NaNSC} \left( \frac{DAG}{DAG + K_{NSC}} + d_{NSC}^{\min} \right) Po_{NSC} \quad [S74]$$

$$* \frac{V_m (z_{Na} F)^2}{RT} \frac{Na_e - Na_i \exp(V_m z_{Na} F / RT)}{1 - \exp(V_m z_{Na} F / RT)}$$

$$I_{KNSC} = 10^6 A_m P_{KNSC} \left( \frac{DAG}{DAG + K_{NSC}} + d_{NSC}^{\min} \right) Po_{NSC} \quad [S75]$$

$$* \frac{V_m (z_K F)^2}{RT} \frac{K_e - K_i \exp(V_m z_K F / RT)}{1 - \exp(V_m z_K F / RT)}$$

$$I_{CaNSC} = 10^6 A_m P_{CaNSC} d_{NSC}^{\min} Po_{NSC} \frac{V_m (z_{Ca} F)^2}{RT} \frac{Ca_e - Ca_i \exp(V_m z_{Ca} F / RT)}{1 - \exp(V_m z_{Ca} F / RT)} \quad [S76]$$

$$I_{NSC} = I_{NaNSC} + I_{KNSC} + I_{CaNSC} \quad [S77]$$

Formulated as a channel with vessel stress-control of  $Ca^{2+}$  conductance (channel variant 1), which is same as Kapela et al. formulation except the following two equations now replace [S76]:

$$\rho_{CaNSC\sigma CC} = 1 + (\rho_{CaNSC\sigma CC}^{\max} - 1) \frac{\sigma_{total}^{n_{CaNSC\sigma CC}}}{\sigma_{50, CaNSC\sigma CC}^{n_{CaNSC\sigma CC}} + \sigma_{total}^{n_{CaNSC\sigma CC}}} \quad [S78]$$

$$I_{CaNSC} = 10^6 A_m P_{CaNSC} d_{NSC}^{\min} \rho_{CaNSC\sigma CC} Po_{NSC} \frac{V_m (z_{Ca} F)^2}{RT} \frac{Ca_e - Ca_i \exp(V_m z_{Ca} F / RT)}{1 - \exp(V_m z_{Ca} F / RT)} \quad [S79]$$

Formulated as a channel with vessel strain-control of  $Ca^{2+}$  conductance, which is same as Kapela et al. formulation except the following two equations now replace [S76]:

$$\rho_{CaNSC\varepsilon CC} = 1 + (\rho_{CaNSC\varepsilon CC}^{\max} - 1) \frac{\varepsilon_{total}^{n_{CaNSC\varepsilon CC}}}{\varepsilon_{50, CaNSC\varepsilon CC}^{n_{CaNSC\varepsilon CC}} + \varepsilon_{total}^{n_{CaNSC\varepsilon CC}}} \quad [S80]$$

$$I_{CaNSC} = 10^6 A_m P_{CaNSC} d_{NSC}^{\min} \rho_{CaNSC\varepsilon CC} Po_{NSC} \frac{V_m (z_{Ca} F)^2}{RT} \frac{Ca_e - Ca_i \exp(V_m z_{Ca} F / RT)}{1 - \exp(V_m z_{Ca} F / RT)} \quad [S81]$$

Formulated as a channel with vessel stress-control of  $Na^+$  conductance (channel variant 2), which is same as Kapela et al. formulation except the following two equations now replace [S74]:

$$\rho_{NaNSC\sigma CC} = 1 + (\rho_{NaNSC\sigma CC}^{\max} - 1) \frac{\sigma_{total}^{n_{NaNSC\sigma CC}}}{\sigma_{50,NaNSC\sigma CC}^{n_{NaNSC\sigma CC}} + \sigma_{total}^{n_{NaNSC\sigma CC}}} \quad [S82]$$

$$I_{NaNSC} = 10^6 A_m P_{NaNSC} \left( \frac{DAG}{DAG + K_{NSC}} + d_{NSC}^{\min} \right) \rho_{NaNSC\sigma CC} P_{O_{NSC}} \\ * \frac{V_m (z_{Na} F)^2}{RT} \frac{Na_e - Na_i \exp(V_m z_{Na} F / RT)}{1 - \exp(V_m z_{Na} F / RT)} \quad [S83]$$

Formulated as a channel with vessel stress-control of  $Na^+$  conductance, which is same as Kapela et al. formulation except the following two equations now replace [S74]:

$$\rho_{NaNSC\varepsilon CC} = 1 + (\rho_{NaNSC\varepsilon CC}^{\max} - 1) \frac{\varepsilon_{total}^{n_{NaNSC\varepsilon CC}}}{\varepsilon_{50,NaNSC\varepsilon CC}^{n_{NaNSC\varepsilon CC}} + \varepsilon_{total}^{n_{NaNSC\varepsilon CC}}} \quad [S84]$$

$$I_{NaNSC} = 10^6 A_m P_{NaNSC} \left( \frac{DAG}{DAG + K_{NSC}} + d_{NSC}^{\min} \right) \rho_{NaNSC\varepsilon CC} P_{O_{NSC}} \\ * \frac{V_m (z_{Na} F)^2}{RT} \frac{Na_e - Na_i \exp(V_m z_{Na} F / RT)}{1 - \exp(V_m z_{Na} F / RT)} \quad [S85]$$

Formulated as a channel with vessel stress-control of  $Ca^{2+}$  and  $Na^+$  gating voltage (channel variant 3), which is same as Kapela et al. formulation except the following two equations now replace [S73]:

$$V_{NSC\sigma CC}^{off} = V_{NSC\sigma CC}^{off \max} \frac{\sigma_{total}^{n_{NSC\sigma CC}}}{\sigma_{50,NSC\sigma CC}^{n_{NSC\sigma CC}} + \sigma_{total}^{n_{NSC\sigma CC}}} \quad [S86]$$

$$P_{O_{NSC}} = 0.4344 + \frac{0.5656}{1 + \exp\left[-(V_m + V_{NSC\sigma CC}^{off} - 47.12)/24.24\right]} \quad [S87]$$

Formulated as a channel with vessel strain-control of  $Ca^{2+}$  and  $Na^+$  gating voltage, which is same as Kapela et al. formulation except the following two equations now replace [S73]:

$$V_{NSC\varepsilon CC}^{off} = V_{NSC\varepsilon CC}^{off \max} \frac{\varepsilon_{total}^{n_{NSC\varepsilon CC}}}{\varepsilon_{50,NSC\varepsilon CC}^{n_{NSC\varepsilon CC}} + \varepsilon_{total}^{n_{NSC\varepsilon CC}}} \quad [S88]$$

$$P_{O_{NSC}} = 0.4344 + \frac{0.5656}{1 + \exp\left[-(V_m + V_{NSC\varepsilon CC}^{off} - 47.12)/24.24\right]} \quad [S89]$$

*Store-operated non-selective cation channel:*

$$\bar{P}_{SOC} = \frac{1}{1 + Ca_{SRu} / K_{SOC}} \quad [S90]$$

$$I_{NaSOC} = g_{NaSOC} P_{SOC} (V_m - E_{Na}) \quad [S91]$$

$$I_{CaSOC} = g_{CaSOC} P_{SOC} (V_m - E_{Ca}) \quad [S92]$$

$$I_{SOC} = I_{NaSOC} + I_{CaSOC} \quad [S93]$$

$$\frac{dP_{SOC}}{dt} = \frac{\bar{P}_{SOC} - P_{SOC}}{\tau_{SOC}} \quad [S94]$$

$Ca^{2+}$ -activated  $Cl$  channel:

$$\alpha_{Cl} = \frac{cGMP^{n_{ClcGMP}}}{cGMP^{n_{ClcGMP}} + K_{ClcGMP}^{n_{ClcGMP}}} \quad [S95]$$

$$K_{ClCa cGMP} = 4 \times 10^{-4} (1 - 0.9 \alpha_{Cl}) \quad [S96]$$

$$P_{Cl} = R_{ClcGMP}^{\min} \frac{Ca_i^{n_{ClCa}}}{Ca_i^{n_{ClCa}} + K_{ClCa}^{n_{ClCa}}} + \alpha_{Cl} \frac{Ca_i^{n_{ClCa}}}{Ca_i^{n_{ClCa}} + K_{ClCa cGMP}^{n_{ClCa}}} \quad [S97]$$

$$I_{ClCa} = C_m g_{ClCa} P_{Cl} (V_m - E_{Cl}) \quad [S98]$$

Plasma membrane  $Ca^{2+}$ -ATPase:

$$I_{PMCA} = \bar{I}_{PMCA} \frac{Ca_i}{Ca_i + K_{mPMCA}} \quad [S99]$$

Plasma membrane  $Na^+$ - $Ca^{2+}$  exchanger:

$$R_{NCXcGMP} = 1 + 0.55 \frac{cGMP}{cGMP + 4.5 \times 10^{-2}} \quad [S100]$$

$$\phi_F = \exp\left(\frac{\gamma_{NCX} V_m F}{RT}\right) \quad [S101]$$

$$\phi_R = \exp\left[\frac{(\gamma_{NCX} - 1) V_m F}{RT}\right] \quad [S102]$$

$$I_{NCX} = g_{NCX} R_{NCXcGMP} \frac{Na_i^3 Ca_e \phi_F - Na_e^3 Ca_i \phi_R}{1 + d_{NCX} (Na_e^3 Ca_i + Na_i^3 Ca_e)} \quad [S103]$$

$Na^+$ - $K^+$  pump:

$$I_{NaK} = C_m \bar{I}_{NaK} Q_{NaK} \frac{K_e^{n_{HKe}}}{K_e^{n_{HKe}} + K_{dKe}^{n_{HKe}}} \frac{Na_i^{n_{HNai}}}{Na_i^{n_{HNai}} + K_{dNai}^{n_{HNai}}} \frac{V_m + 150}{V_m + 200} \quad [S104]$$

$Na^+$ - $K^+$ - $Cl$  cotransporter:

$$R_{NaKClcGMP} = 1 + 3.5 \frac{cGMP}{cGMP + 6.4 \times 10^{-3}} \quad [S105]$$

$$I_{ClNaKCl} = -A_m R_{NaKClcGMP} L_{NaKCl} z_{Cl} FRT \ln \left[ \left( \frac{Na_e}{Na_i} \right) \left( \frac{K_e}{K_i} \right) \left( \frac{Cl_e}{Cl_i} \right)^2 \right] \quad [S106]$$

$$I_{NaNaKCl} = -\frac{I_{ClNaKCl}}{2} \quad [S107]$$

$$I_{KNaKCl} = -\frac{I_{ClNaKCl}}{2} \quad [S108]$$

*Sarcoplasmic reticulum IP<sub>3</sub> receptor:*

$$I_{IP_3} = \bar{I}_{IP_3} Vol_{Ca} z_{Ca} F \left( \frac{IP_3}{IP_3 + K_{IP_3}} \frac{Ca_i}{Ca_i + K_{IP_3}^{act}} h_{IP_3} \right)^3 (Ca_{SRu} - Ca_i) \quad [S109]$$

$$\frac{dh_{IP_3}}{dt} = k_{IP_3}^{on} \left[ K_{IP_3}^{inh} - (Ca_i + K_{IP_3}^{inh}) h_{IP_3} \right] \quad [S110]$$

*Sarcoplasmic reticulum SERCA, internal diffusion and release:*

$$I_{SERCA} = \bar{I}_{SERCA} \frac{Ca_i}{Ca_i + K_m^{up}} \quad [S111]$$

$$I_{tr} = \frac{Vol_{SRu} z_{Ca} F}{\tau_{tr}} (Ca_{SRu} - Ca_{SRr}) \quad [S112]$$

$$I_{rel} = \frac{Vol_{SRr} z_{Ca} F}{\tau_{rel}} (R_{10}^2 + R_{leak}) (Ca_{SRr} - Ca_i) \quad [S113]$$

$$\frac{dCa_{SRu}}{dt} = \frac{I_{SERCA} - I_{tr} - I_{IP_3}}{Vol_{SRu} z_{Ca} F} \quad [S114]$$

$$\frac{dCa_{SRr}}{dt} = \frac{I_{tr} - I_{rel}}{(Vol_{SRr} z_{Ca} F) \left[ 1 + \frac{CSQN K_{CSQN}}{(K_{CSQN} + Ca_{SRr})^2} \right]} \quad [S115]$$

*Sarcoplasmic reticulum ryanodine receptor:*

$$R_{00} = 1 - R_{10} - R_{11} - R_{01} \quad [S116]$$

$$\frac{dR_{10}}{dt} = K_{r1} Ca_i^2 R_{00} - (K_{mr1} + K_{r2} Ca_i) R_{10} + K_{mr2} R_{11} \quad [S117]$$

$$\frac{dR_{11}}{dt} = K_{r2} Ca_i R_{10} - (K_{mr1} + K_{mr2}) R_{11} + K_{r1} Ca_i^2 R_{01} \quad [S118]$$

$$\frac{dR_{01}}{dt} = K_{r2} Ca_i R_{00} + K_{mr1} R_{11} - (K_{mr2} + K_{r1} Ca_i^2) R_{01} \quad [S119]$$

*α<sub>1</sub>-adrenoreceptor activation and IP<sub>3</sub> formation:*

$$\rho_{rG} = \frac{NE R_G^S}{\xi_G R_{TG} (K_{1G} + NE)} \quad [S120]$$

$$r_{hG} = \alpha_G G \frac{Ca_i}{Ca_i + K_{cG}} \quad [S121]$$



$$\frac{dR_G^S}{dt} = k_{rG}\xi_G R_{TG} - \left( k_{rG} + \frac{k_{pG}NE}{K_{1G} + NE} \right) R_G^S - k_{rG} R_{PG}^S \quad [S122]$$

$$\frac{dR_{PG}^S}{dt} = NE \left( \frac{k_{pG}R_G^S}{K_{1G} + NE} - \frac{k_{eG}R_{PG}^S}{K_{2G} + NE} \right) \quad [S123]$$

$$\frac{dG}{dt} = k_{aG}(\delta_{G0} + \rho_{rG})(G_{TG} - G) - k_{dG}G \quad [S124]$$

$$\frac{dIP_3}{dt} = \frac{r_{hG}PIP_2}{\gamma_G} - k_{degG}IP_3 \quad [S125]$$

$$\frac{dPIP_2}{dt} = -(r_{hG} + r_{rG})PIP_2 - r_{rG}\gamma_G IP_3 + r_{rG}PIP_{2T} \quad [S126]$$

*sGC activation and cGMP formation:*

$$\bar{V}_{cGMP} = V_{cGMP}^{\max} \frac{B5_{sGC}NO + NO^2}{A0_{sGC} + A1_{sGC}NO + NO^2} \quad [S127]$$

$$\tau_{sGC} = \begin{cases} \frac{1}{k_{3sGC}NO + k_{DrsGC}} & \text{if } \bar{V}_{cGMP} - V_{cGMP} \geq 0 \\ \frac{1}{k_{m2sGC} + k_{DrsGC}} & \text{otherwise} \end{cases} \quad [S128]$$

$$\frac{dV_{cGMP}}{dt} = \frac{\bar{V}_{cGMP} - V_{cGMP}}{\tau_{sGC}} \quad [S129]$$

$$\frac{dcGMP}{dt} = V_{cGMP} - k_{pdecGMP} \left( \frac{cGMP^2}{cGMP + K_{mpde}} \right) \quad [S130]$$

*Ionic balances:*

Note: For vessel stress-controlled supplementary  $Ca^{2+}$  channel (channel variant 4 with stress),  $I_{Ca\sigma CC}$  is calculated from equation [S20], otherwise it is 0. For vessel strain-controlled supplementary  $Ca^{2+}$  channel (channel variant 4 with strain),  $I_{Ca\varepsilon CC}$  is calculated from equation [S23], otherwise it is 0. For vessel stress-controlled supplementary  $Na^+$  channel (channel variant 5 with stress),  $I_{Na\sigma CC}$  is calculated from equation [S26], otherwise it is 0. For vessel strain-controlled supplementary  $Na^+$  channel (channel variant 5 with strain),  $I_{Na\varepsilon CC}$  is calculated from equation [S29], otherwise it is 0.

$$I_{Catotm} = I_{CaSOC} + I_{VOCC} - 2I_{NCX} + I_{PMCA} + I_{CaNSC} + I_{Ca\sigma CC} + I_{Ca\varepsilon CC} \quad [S131]$$

$$I_{Natotm} = I_{NaNaKCl} + I_{NaSOC} + 3I_{NaK} + 3I_{NCX} + I_{NaNSC} + I_{Na\sigma CC} + I_{Na\varepsilon CC} \quad [S132]$$

$$I_{Ktotm} = I_{KNaKCl} + I_{BKCa} + I_{Kv} + I_{KNSC} + I_{Kleak} - 2I_{NaK} \quad [S133]$$

$$I_{Cltotm} = I_{ClNaKCl} + I_{ClCa} \quad [S134]$$

$$\frac{dCa_i}{dt} = -\frac{I_{Catom} + I_{SERCA} - I_{rel} - I_{IP3}}{Vol_{Ca} z_{Ca} F} \left( \frac{1}{1 + \frac{\bar{S}_{CM} K_{dCM}}{(K_{dCM} + Ca_i)^2} + \frac{\bar{B}_F K_{dB}}{(K_{dB} + Ca_i)^2}} \right) \quad [S135]$$

$$\frac{dNa_i}{dt} = -\frac{I_{Natom}}{Vol_i z_{Na} F} \quad [S136]$$

$$\frac{dK_i}{dt} = -\frac{I_{Ktotm}}{Vol_i z_K F} \quad [S137]$$

$$\frac{dCl_i}{dt} = -\frac{I_{Cltom}}{Vol_i z_{Cl} F} \quad [S138]$$

*Membrane potential:*

Note: Mechanically controlled supplementary channel currents ( $I_{Ca\sigma CC}$ ,  $I_{Ca\epsilon CC}$ ,  $I_{Na\sigma CC}$  and  $I_{Na\epsilon CC}$ ) are calculated as for the ionic balances as noted above.

$$\frac{dV_m}{dt} = -\frac{1}{C_m} (I_{VOCC} + I_{BKCa} + I_{Kv} + I_{Kleak} + I_{NSC} + I_{SOC} + I_{ClCa} + I_{PMCA} + I_{NCX} + I_{NaK} + I_{Ca\sigma CC} + I_{Ca\epsilon CC} + I_{Na\sigma CC} + I_{Na\epsilon CC} - I_{stim}) \quad [S139]$$

Calculation of remaining fixed parameters:

$Na^+ - K^+$  pump:

$$Q_{NaK} = Q_{10}^{\frac{T-309.15}{10}} \quad [S140]$$

$\alpha_1$ -adrenoreceptor activation and  $IP_3$  formation:

$$\gamma_G = 10^{-15} N_{Avo} Vol_i \quad [S141]$$

$$\delta_{G0} = \frac{k_{dG} G_0}{k_{aG} (G_{TG} - G_0)} \quad [S142]$$

$sGC$  activation and  $cGMP$  formation:

$$B5_{sGC} = k_{2sGC} / k_{3sGC} \quad [S143]$$

$$A0_{sGC} = \frac{(k_{m1sGC} + k_{2sGC}) k_{DsGC} + k_{m1sGC} k_{m2sGC}}{k_{1sGC} k_{3sGC}} \quad [S144]$$

$$A1_{sGC} = \frac{(k_{1sGC} + k_{3sGC}) k_{DsGC} + (k_{2sGC} + k_{m2sGC}) k_{1sGC}}{k_{1sGC} k_{3sGC}} \quad [S145]$$

Setting and calculation of initial conditions:

*Vessel wall stress- or strain-activated  $Ca^{2+}$  channel:*

$$h_{Ca\sigma CC} = 0 \quad \text{or} \quad h_{Ca\epsilon CC} = 0 \quad [S146]$$

*Vessel wall stress- or strain-activated  $Na^+$  channel:*

$$h_{Na\sigma CC} = 0 \quad \text{or} \quad h_{Na\epsilon CC} = 0 \quad [S147]$$

*L-Type voltage operated  $Ca^{2+}$  channel:*

$$d_{L0} = \frac{1}{1 + \exp(-V_{m0}/8.3)} \quad [S148]$$

$$f_{L0} = \frac{1}{1 + \exp[(V_{m0} + 42)/9.1]} \quad [S149]$$

*Large conductance Ca<sup>2+</sup>-activated K<sup>+</sup> channel:*

$$R_{NO0} = \frac{NO_0}{NO_0 + 0.2 \times 10^{-3}} \quad [S150]$$

$$R_{cGMP0} = \frac{cGMP_0^2}{cGMP_0^2 + (0.55 \times 10^{-3})^2} \quad [S151]$$

$$V_{50,KCa0} = -41.7 \log(Ca_{i0}) - 128.2 - dV_{50,KCaNO0} R_{NO0} - dV_{50,KCaGMP0} R_{cGMP0} \quad [S152]$$

$$p_{f0} = \frac{1}{1 + \exp[-(V_{m0} - V_{50,KCa0})/18.25]} \quad [S153]$$

$$p_{s0} = \frac{1}{1 + \exp[-(V_{m0} - V_{50,KCa0})/18.25]} \quad [S154]$$

*Voltage dependent K<sup>+</sup> channel:*

$$p_{K0} = \frac{1}{1 + \exp[-(V_{m0} + 11)/15]} \quad [S155]$$

$$q_{10} = \frac{1}{1 + \exp[(V_{m0} + 40)/14]} \quad [S156]$$

$$q_{20} = \frac{1}{1 + \exp[(V_{m0} + 40)/14]} \quad [S157]$$

*Store-operated non-selective cation channel:*

$$P_{SOC0} = 0 \quad [S158]$$

*Sarcoplasmic reticulum IP<sub>3</sub> receptor:*

$$h_{IP30} = \frac{K_{IP3}^{inh}}{Ca_{i0} + K_{IP3}^{inh}} \quad [S159]$$

*Sarcoplasmic reticulum SERCA, internal diffusion and release:*

$$Ca_{SRi0} = 0.66 \quad [S160]$$

$$Ca_{SRr0} = 0.57 \quad [S161]$$

*Sarcoplasmic reticulum ryanodine receptor:*

$$R_{100} = 0.0033 \quad [S162]$$

$$R_{110} = 4 \times 10^{-6} \quad [S163]$$

$$R_{010} = 0.9955 \quad [S164]$$

*α<sub>1</sub>-adrenoreceptor activation and IP<sub>3</sub> formation:*

$$r_{hG0} = \frac{k_{degG} \gamma_G IP_{30}}{PIP_{20}} \quad [S165]$$

$$R_{G0}^S = R_{TG} \xi_G \quad [S166]$$

$$R_{PG0}^S = 0 \quad [S167]$$

$$G_0 = \frac{r_{hG0} (K_{cG} + Ca_{i0})}{\alpha_G Ca_{i0}} \quad [S168]$$

$$IP_{30} = 0 \quad [S169]$$

$$PIP_{20} = PIP_{2T} - (1 + k_{degG}/r_{rG}) \gamma_G IP_{30} \quad [S170]$$

*sGC activation and cGMP formation:*

$$V_{cGMP0} = 0 \quad [S171]$$

$$cGMP_0 = 0 \quad [S172]$$

*Ionic balances:*

$$Ca_{i0} = 68 \times 10^{-6} \quad [S173]$$

$$Na_{i0} = 8.4 \quad [S174]$$

$$K_{i0} = 140 \quad [S175]$$

$$Cl_{i0} = 59.4 \quad [S176]$$

*Membrane potential:*

$$V_{m0} = -59.4 \quad [S177]$$

### 3. TABLES OF OPTIMIZED PARAMETERS

Table S4: Optimized parameters to fit of steady-state VSM crossbridge activation

Optimized parameter	Description	Units	Value
$n_{XB}$	Sensitivity of VSM activation to $Ca^{2+}$	unitless	3.1984
$K_{CaXB}$	$\frac{1}{2}$ activation VSM $Ca^{2+}$ concentration	$\mu M$	0.2066

Table S5: Optimized parameters of myogenic response fits with nine different hypothetical vessel stress-controlled channel variants

Optimized parameter	Description	Units	Value	
			Mesenteric	Femoral
NSC with stress-controlled $Ca^{2+}$ conductance (CaNSC $\sigma$ CC)				
$\rho_{CaNSC\sigma CC}^{\max}$	CaNSC $\sigma$ CC max ratio of $\sigma$ -control to normal conduct	unitless	5.628	14.610
$n_{CaNSC\sigma CC}$	CaNSC $\sigma$ CC sensitivity to stress	unitless	3.475	1.790
$\sigma_{50,CaNSC\sigma CC}$	CaNSC $\sigma$ CC $\frac{1}{2}$ activation stress	kPa	82.690	76.746
NSC with stress-controlled $Na^{+}$ conductance (NaNSC $\sigma$ CC)				
$\rho_{NaNSC\sigma CC}^{\max}$	NaNSC $\sigma$ CC max ratio of $\sigma$ -control to normal conduct	unitless	6.997	7.931
$n_{NaNSC\sigma CC}$	NaNSC $\sigma$ CC sensitivity to stress	unitless	4.161	2.742
$\sigma_{50,NaNSC\sigma CC}$	NaNSC $\sigma$ CC $\frac{1}{2}$ activation stress	kPa	82.245	34.107
NSC with stress-controlled gating voltage (NSC $\sigma$ CC)				
$V_{NSC\sigma CC}^{off \max}$	NSC $\sigma$ CC max offset of gating voltage	mV	403.429	665.142
$n_{NSC\sigma CC}$	NSC $\sigma$ CC sensitivity to stress	unitless	2.299	1.136
$\sigma_{50,NSC\sigma CC}$	NSC $\sigma$ CC $\frac{1}{2}$ activation stress	kPa	91.222	82.789
Supplementary $Ca^{2+}$ channel with stress-controlled conductance (Ca $\sigma$ CC)				
$g_{Ca\sigma CC}^{\max}$	Ca $\sigma$ CC maximal conductance	pA/mV	2.365	3.926
$n_{Ca\sigma CC}$	Ca $\sigma$ CC sensitivity to stress	unitless	1.647	1.755
$\sigma_{50,Ca\sigma CC}$	Ca $\sigma$ CC $\frac{1}{2}$ activation stress	kPa	1881.08	764.942
Supplementary $Na^{+}$ channel stress-controlled conductance (Na $\sigma$ CC)				
$g_{Na\sigma CC}^{\max}$	Na $\sigma$ CC maximal conductance	pA/mV	4.272	5.139
$n_{Na\sigma CC}$	Na $\sigma$ CC sensitivity to stress	unitless	2.707	1.948
$\sigma_{50,Na\sigma CC}$	Na $\sigma$ CC $\frac{1}{2}$ activation stress	kPa	300.937	211.178

L-type  $Ca^{2+}$  channel with stress-controlled gating voltage (CaL $\sigma$ CC)

$V_{CaL\sigma CC}^{off\ max}$	CaL $\sigma$ CC maximal membrane potential offset	mV	32.490	71.636
$n_{CaL\sigma CC}$	CaL $\sigma$ CC sensitivity to stress	unitless	2.875	1.346
$\sigma_{50,Ca\sigma CC}$	CaL $\sigma$ CC $\frac{1}{2}$ activation stress	kPa	74.463	68.855
L-type Ca <sup>2+</sup> channel with stress-controlled conductance ( $\rho$ CaL $\sigma$ CC)				
$\rho_{\rho CaL\sigma CC}^{max}$	$\rho$ CaL $\sigma$ CC max ratio of $\sigma$ -control to normal conduct	unitless	7.134	20.000
$n_{\rho CaL\sigma CC}$	$\rho$ CaL $\sigma$ CC sensitivity to stress	unitless	3.194	1.535
$\sigma_{50,\rho CaL\sigma CC}$	$\rho$ CaL $\sigma$ CC $\frac{1}{2}$ activation stress	kPa	76.946	84.928
Large conductance Ca <sup>2+</sup> -activated K <sup>+</sup> channel with stress-controlled gating voltage (BKCa $\sigma$ CC)				
$V_{BKCa\sigma CC}^{off\ max}$	BKCa $\sigma$ CC maximal membrane potential offset	mV	20.00	19.866
$n_{BKCa\sigma CC}$	BKCa $\sigma$ CC sensitivity to stress	unitless	2.000	2.504
$\sigma_{50,BKCa\sigma CC}$	BKCa $\sigma$ CC $\frac{1}{2}$ activation stress	kPa	54.598	1010.50
Large conductance Ca <sup>2+</sup> -activated K <sup>+</sup> channel with stress-controlled conductance ( $\rho$ BKCa $\sigma$ CC)				
$\rho_{\rho BKCa\sigma CC}^{max}$	$\rho$ BKCa $\sigma$ CC max ratio of $\sigma$ -control to normal conduct	unitless	0.000	0.000
$n_{\rho BKCa\sigma CC}$	$\rho$ BKCa $\sigma$ CC sensitivity to stress	unitless	2.027	1.711
$\sigma_{50,\rho BKCa\sigma CC}$	$\rho$ BKCa $\sigma$ CC $\frac{1}{2}$ activation stress	kPa	49.620	1.648

#### 4. COMPARISON OF THIN AND THICK WALL FORMULATION

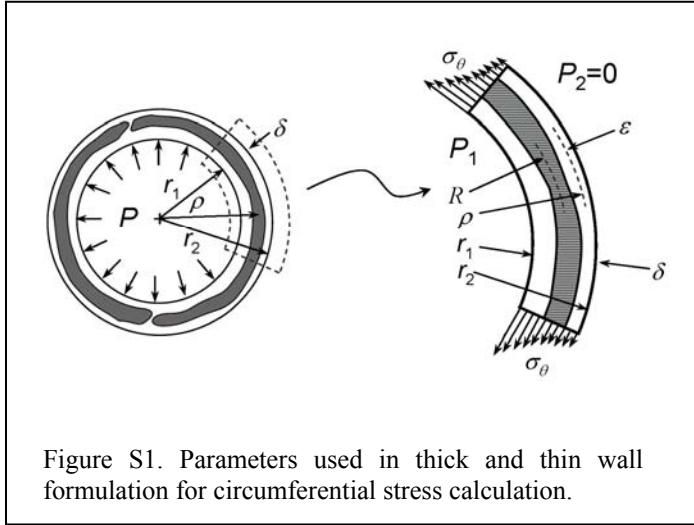


Figure S1. Parameters used in thick and thin wall formulation for circumferential stress calculation.

The thin wall vessel formulation for calculating average vessel wall stress (also known as the Law of Laplace) assumes the vessel wall is thin enough compared to the internal radius to have roughly constant circumferential stress across the vessel wall. In many cases this is true however when the arterioles analyzed in this study contract to very small internal diameters the circumferential stress on the inner and outer surfaces of the vessel wall may deviate from that approximated with the thin wall formulation. To estimate the error between the thin and thick walled

circumferential stress formulation for each steady state diameter reached in these experimental studies we first cast the two formulations as follows:

Thick wall circumferential stress formulation:

$$\sigma_{\theta,TKW}(\rho) = \frac{P_1}{1 - r_1^2/r_2^2} \left( \frac{r_1^2}{r_2^2} + \frac{r_1^2}{\rho^2} \right) \quad [S178]$$

where  $\sigma_{\theta,TKW}$  is the thick wall formulation circumferential stress,  $P_1$  is the intraluminal pressure,  $r_1$  is the inner radius,  $r_2$  is the outer radius and  $\rho$  is the vessel wall position where we are calculating the stress. If we let:

$$r_1 = R - \delta/2; \quad r_2 = R + \delta/2; \quad \rho = R + \varepsilon \quad [S179]$$

where  $R$  is the midline radius,  $\delta$  is the vessel wall thickness and  $\varepsilon$  varies from  $-\delta/2$  to  $\delta/2$  then we have for the thick walled stress at the normalized distance  $\varepsilon/R$ :

$$\frac{\sigma_{\theta,TKW}(\varepsilon/R)}{P_1} = \frac{\left[ \frac{1 - \delta/R + \frac{1}{4}(\delta^2/R^2)}{1 + \delta/R + \frac{1}{4}(\delta^2/R^2)} \right] + \left[ \frac{1 - \delta/R + \frac{1}{4}(\delta^2/R^2)}{1 + 2(\varepsilon/R) + (\varepsilon^2/R^2)} \right]}{1 - \left[ \frac{1 - \delta/R + \frac{1}{4}(\delta^2/R^2)}{1 + \delta/R + \frac{1}{4}(\delta^2/R^2)} \right]} \quad [S180]$$

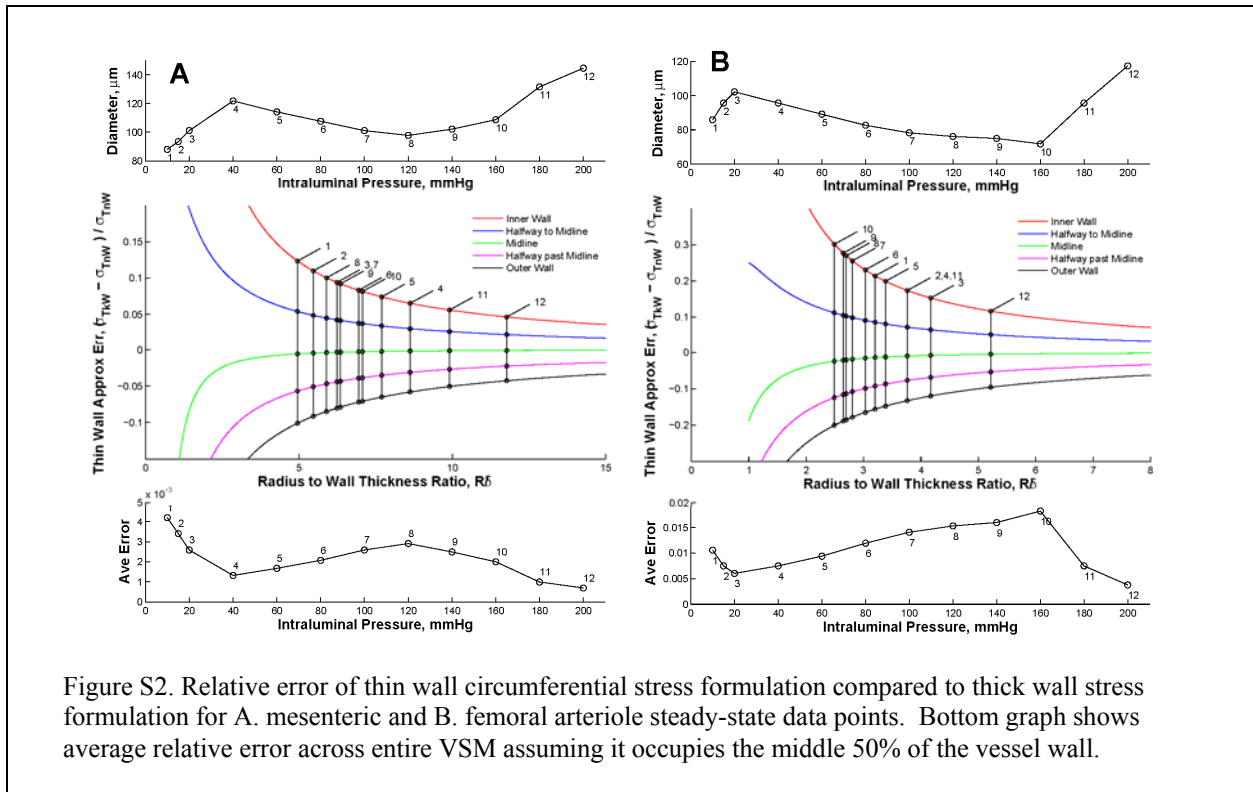
Thin wall circumferential stress formulation:

$$\sigma_{\theta,TnW} = \frac{P_1 r_1}{\delta} = \frac{P_1 (R - \delta/2)}{\delta} \quad [S181]$$

where  $\sigma_{\theta,TnW}$  is the thin wall formulation circumferential stress. Rearranging we have:

$$\frac{\sigma_{\theta, ThW}}{P_1} = \frac{1}{\delta/R} - \frac{1}{2} \quad [S182]$$

The standard rule of thumb for the thin walled approximation to be valid is a radius to wall thickness ratio,  $R/\delta$  of 20. In the mesenteric arteriole dataset the  $R/\delta$  ranges from 5 to 12 and with the femoral arteriole dataset the range is 2.5 to 5.25 so the thin walled approximation appears to be not valid in this case. However in our problem we are trying to estimate the stress across the VSM cell that spans the midline of the vessel wall where the error between thin wall and thick wall stress formulations is much smaller. Shown below in Figure S2A, B we see the error between the thick walled and thin walled vessel formulation for each data point in the mesenteric and femoral datasets. Error is given for inner wall, at 25%, at 50%, at 75% and outer vessel wall stress for each steady state diameter. If we assume the VSM cell occupies the middle 50% of the vessel wall we see that the average stress over the VSM cell varies by less than 0.5% and 2% between the two formulations over the mesenteric and femoral datasets respectively. More accurate stress values can be obtained with the thick walled vessel formulation if more detailed knowledge about how and where the stress is sensed in VSM cells. Additionally even if in this study we are underestimating or overestimating the circumferential stress we are not altering the trend in circumferential stress with changes in intraluminal pressure. Therefore free parameters in the model may change but the conclusions remain unaltered concerning which stress-controlled channel variants are able to describe the steady state myogenic response.





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