

Supporting Information

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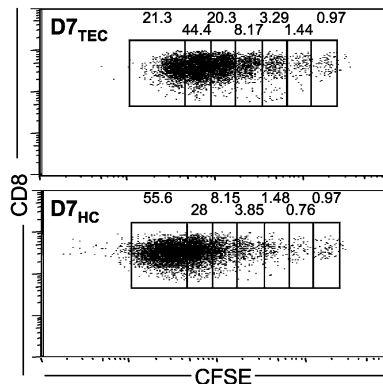


Fig. S1. D7 CD8⁺ T cells selected on HCs demonstrate higher proliferative capacity on in vitro stimulation than those selected on TECs. Splenocytes from D7_{TEC} and D7_{HC} were labeled with 10 μ M CFSE and incubated for 65 h with LemA pulsed stimulator cells. Proliferation of D7_{TEC} or D7_{HC} was measured by CFSE dilution. Events shown are gated on live D7_{TEC} or D7_{HC}. CFSE, carboxyfluorescein succinimidyl ester.

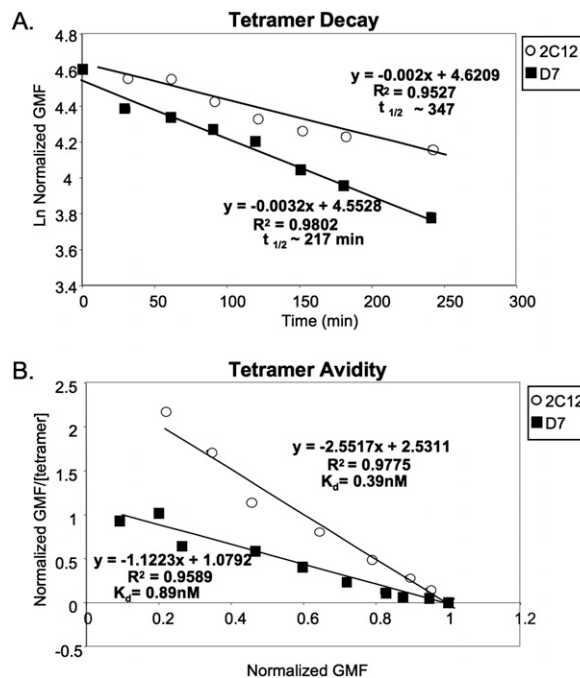


Fig. S2. Comparison of TCR affinity between D7 and iNKT cells. (A) D7 Tg mouse splenocytes and N38-2C12 iNKT cell hybridomas were incubated with 10 μ M allophycocyanin-conjugated M3/LemA and CD1d/ α -GalCer tetramers, respectively, for 45 min at 4 $^{\circ}$ C and then incubated at 37 $^{\circ}$ C with anti-TCR- β and excess anti-M3 or anti-CD1d mAb, respectively. At 20-min intervals, an aliquot of cells was washed, fixed with 4% paraformaldehyde, and analyzed via flow cytometry, gating on T cells. The natural log of percentage geometrical mean fluorescence of tetramer at each time point (compared with 0 min) was plotted against time. The half-life of each tetramer was derived from the slope as $t_{1/2} = \ln 2/\text{slope}$. α -GalCer, α galactosylceramide; GMF, geometrical mean fluorescence, Ln, natural log. (B) D7 and iNKT cells were stained using 0.1–50 nM of the respective tetramers. Cells were stained for 45 min at 4 $^{\circ}$ C, washed, and analyzed with a FACSCanto II, gating on T cells. Equilibrium binding data were plotted as a Scatchard plot of GMF/concentration of tetramer used in staining against GMF. Tetramer avidity, K_D, is derived from the slope as K_D = 1/slope.

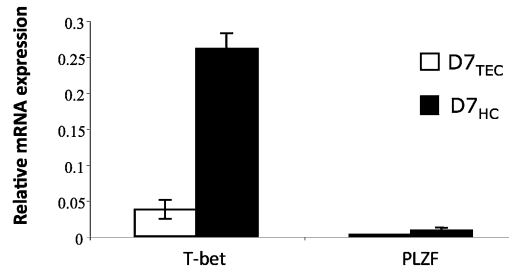


Fig. S3. D7 T cells selected on HCs express higher levels of *T-bet* than D7 T cells selected on TECs. Real-time RT-PCR of *T-bet* and *PLZF* mRNA of D7 T cells sorted from D7_{TEC} and D7_{HC} chimeric mice. The bar graph represents the mean \pm SEM of triplicate determinants of *T-bet* and *PLZF* expression normalized to *GAPDH* expression levels. *T-bet* primers are: forward, 5'-CCAGCACCAGACAGAGATGA-3'; reverse, 5'-GCTTCCCAATGAAACTTC-3'. *PLZF* primers are: forward, 5'-CCAGTTCTCAAAGGAGATG-3'; reverse, 5'-TTCCCACACAGCAGACAGAAG-3'.

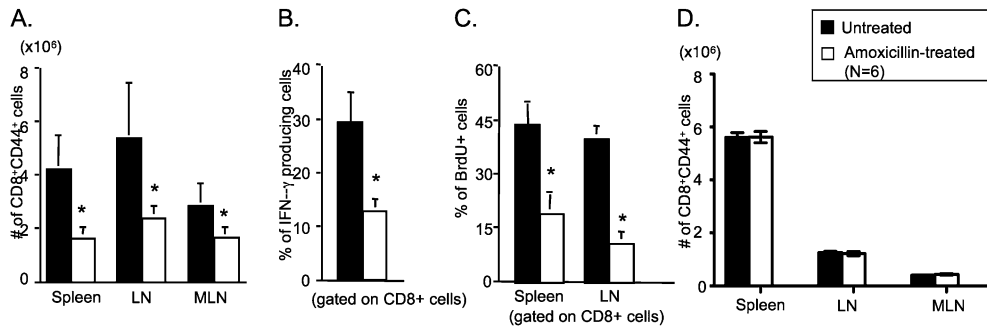


Fig. S4. Suppressed "innate-like" phenotype and effector functions in D7 mice fed with amoxicillin. Pregnant D7 Tg mice were treated with 3 mg/mL filter-sterilized amoxicillin solution at least 5 d before delivery. Treatment was continued until pups were 7 wk of age, when mice were killed and lymphoid tissues were harvested. (A) Number of activated (CD44^{hi}) CD8⁺ T cells from amoxicillin-treated and untreated 7-wk-old mice were compared via flow cytometry. * $P < 0.05$. LN, lymph node; MLN, mesenteric lymph node. (B) Pooled nucleated cells from the spleen and axillary and inguinal lymph nodes of treated and untreated 7-wk-old D7 mice were cultured with LemA for 8 h. Intracellular IFN- γ was analyzed by flow cytometry. * $P < 0.05$. (C) Relative proliferative capacity of CD8⁺ T cells isolated from treated and untreated mice following treatment with LemA was analyzed via BrdU incorporation assay. * $P < 0.05$. (D) As a control, the number of CD44^{hi}CD8⁺ T cells from treated and untreated histocompatibility-Y antigen (H-Y) Tg mice (expressing a TCR specific for an MHC Ia molecule) were compared by flow cytometry.